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## (54) METHODS AND COMPOSITIONS FOR WEED CONTROL

VERFAHREN UND ZUSAMMENSETZUNGEN ZUR UNKRAUTBEKÄMPFUNG

PROCÉDÉS ET COMPOSITIONS DE LUTTE CONTRE LES MAUVAISES HERBES

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**Description**

[0001] Sequences of the sequence listing that are not cited in the claims are submitted for comparative purposes.

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[0002] The methods and compositions generally relate to the field of weed management. More specifically, relate to 7,8-dihydropteroate synthase inhibitors (DHPS) genes in plants and compositions containing polynucleotide molecules for modulating and/or regulating their expression. Further provided are methods and compositions useful for weed control.

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**BACKGROUND**

[0003] Weeds are plants that compete with cultivated plants in an agronomic environment and cost farmers billions of dollars annually in crop losses and the expense of efforts to keep weeds under control. Weeds also serve as hosts for crop diseases and insect pests. The losses caused by weeds in agricultural production environments include decreases in crop yield, reduced crop quality, increased irrigation costs, increased harvesting costs, reduced land value, injury to livestock, and crop damage from insects and diseases harbored by the weeds. The principal means by which weeds cause these effects are: 1) competing with crop plants for water, nutrients, sunlight and other essentials for growth and development, 2) production of toxic or irritant chemicals that cause human or animal health problem, 3) production of immense quantities of seed or vegetative reproductive parts or both that contaminate agricultural products and perpetuate the species in agricultural lands, and 4) production on agricultural and nonagricultural lands of vast amounts of vegetation that must be disposed of. Herbicide tolerant weeds are a problem with nearly all herbicides in use, there is a need to effectively manage these weeds. There are over 365 weed biotypes currently identified as being herbicide resistant to one or more herbicides by the Herbicide Resistance Action Committee (HRAC), the North American Herbicide Resistance Action Committee (NAHRAC), and the Weed Science Society of America (WSSA).

[0004] The 7,8-dihydropteroate synthase (DHPS) is an enzyme involved in folic acid synthesis which is needed for purine nucleotide biosynthesis. This enzyme is the target of herbicides that include the carbamate chemical family.

**BRIEF DESCRIPTION OF THE FIGURES**

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[0005] The following drawings form part of the present specification and are included to further demonstrate certain methods, compositions or results. They may be better understood by reference to one or more of these drawings in combination with the detailed description of specific embodiments presented herein. The invention can be more fully understood from the following description of the figures:

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FIGURE 1. Treatment with oligonucleotide pools followed by Prowl (pendimethalin)

FIGURE 2. Treatment of Palmer Amaranth with 3 oligonucleotide pools followed by Prowl herbicide at 13.45 kg/ha (12 lb/ac) rate

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**SUMMARY**

[0006] In one aspect, the invention provides a method of plant control comprising: treating a plant with a composition comprising a double-stranded RNA (dsRNA) polynucleotide and a transfer agent, wherein said dsRNA polynucleotide is identical or complementary to an RNA sequence of a 7,8-dihydropteroate synthase (DHPS) gene sequence, wherein said DHPS gene sequence is selected from the group consisting of SEQ ID NOs:1, 2, 5, 9, 11, 18, 19, and a polynucleotide fragment thereof with at least 18 contiguous nucleotides from said DHPS gene sequence, wherein said transfer agent is an organosilicone composition or an organosilicone compound contained therein and conditions the surface of said plant for permeation by said dsRNA polynucleotide, and whereby said plant's growth, development, or reproductive ability is suppressed or delayed or said plant is more sensitive to a DHPS inhibitor herbicide or a mitosis inhibitor herbicide as a result of said dsRNA polynucleotide containing composition relative to an untreated plant.

[0007] In a related aspect, the present invention provides a composition comprising a dsRNA polynucleotide and a transfer agent, wherein said dsRNA polynucleotide is identical or complementary to an RNA sequence of a DHPS gene sequence, wherein said DHPS gene sequence is selected from the group consisting of SEQ ID NOs:1, 2, 5, 9, 11, 18, 19, and a polynucleotide fragment thereof with at least 18 contiguous nucleotides from said DHPS gene sequence, wherein said transfer agent is an organosilicone composition or an organosilicone compound contained therein and conditions the surface of a plant for permeation by said dsRNA polynucleotide, and whereby said plant treated with said composition has its growth, development, or reproductive ability regulated, suppressed or delayed or said plant is more sensitive to a DHPS inhibitor herbicide or a mitosis inhibitor herbicide as a result of said dsRNA polynucleotide containing

composition relative to an untreated plant.

[0008] In a further embodiment the present invention also provides a method of reducing expression of a DHPS gene in a plant comprising: external application to said plant of a composition comprising a dsRNA polynucleotide and a transfer agent, wherein said dsRNA polynucleotide is identical or complementary to an RNA sequence of a DHPS gene sequence, wherein said DHPS gene sequence is selected from the group consisting of SEQ ID NOs: 1, 2, 5, 9, 11, 18, 19, and a polynucleotide fragment thereof with at least 18 contiguous nucleotides from said DHPS gene sequence, wherein said transfer agent is an organosilicone composition or an organosilicone compound contained therein and conditions the surface of said plant for permeation by said dsRNA polynucleotide, and whereby said expression of said DHPS gene is reduced relative to an untreated plant.

[0009] Alternatively, the present invention provides methods of identifying dsRNA polynucleotides useful in modulating DHPS gene expression when externally treating a plant comprising: a) providing a plurality of dsRNA polynucleotides that comprise a region identical or complementary to a polynucleotide fragment of at least 18 contiguous nucleotides in length that is identical or complementary to a DHPS gene sequence selected from the group consisting of SEQ ID NOs: 1, 2, 5, 9, 11, 18, and 19; b) externally treating said plant with a composition comprising one or more of said dsRNA polynucleotides and a transfer agent, wherein said transfer agent is an organosilicone composition or an organosilicone compound contained therein and conditions the surface of said plant for permeation by said one or more of said dsRNA polynucleotides; c) analyzing said plant, or extract for modulation of DHPS gene expression, and d) said plant treated with said composition has its growth, development or reproductive ability regulated, suppressed, or delayed or said plant is more sensitive to a DHPS inhibitor herbicide or a mitosis inhibitor herbicide as a result of said composition, relative to an untreated plant.

[0010] In yet another embodiment, the present invention provides an agricultural chemical composition comprising an admixture of a dsRNA polynucleotide, a transfer agent, a DHPS inhibitor herbicide or a mitosis inhibitor herbicide, and a co-herbicide, wherein said dsRNA polynucleotide is identical or complementary to a portion of an RNA sequence of a DHPS gene sequence, wherein said DHPS gene sequence is selected from the group consisting of SEQ ID NOs: 1, 2, 5, 9, 11, 18, 19, and a polynucleotide fragment thereof with at least 18 contiguous nucleotides from said DHPS gene sequence, wherein said transfer agent is an organosilicone composition or an organosilicone compound contained therein and conditions a surface of a plant for permeation by said dsRNA polynucleotide, and whereby said plant treated with said composition has its growth, development, or reproductive ability regulated, suppressed, or delayed or said plant is more sensitive to said DHPS inhibitor herbicide or said mitosis inhibitor herbicide as a result of said dsRNA polynucleotide containing composition, relative to an untreated plant.

[0011] In a further aspect the invention provides an agricultural chemical composition comprising an admixture of a dsRNA polynucleotide, a transfer agent, a DHPS inhibitor herbicide, and a pesticide, wherein said dsRNA polynucleotide is identical or complementary to a portion of an RNA sequence of a DHPS gene sequence, wherein said DHPS gene sequence is selected from the group consisting of SEQ ID NOs: 1, 2, 5, 9, 11, 18, 19, and a polynucleotide fragment thereof with at least 18 contiguous nucleotides from said DHPS gene sequence, wherein said transfer agent is an organosilicone composition or an organosilicone compound contained therein and conditions a surface of a plant for permeation by said dsRNA polynucleotide, whereby said plant treated with said composition has its growth, development, or reproductive ability regulated, suppressed, or delayed or said plant is more sensitive to said DHPS inhibitor herbicide or a mitosis inhibitor herbicide as a result of said dsRNA polynucleotide containing composition relative to an untreated plant.

## DETAILED DESCRIPTION

[0012] Provided are methods and compositions containing a polynucleotide that provide for regulation, repression or delay of 7,8-dihydropteroate synthase (DHPS) gene expression and enhanced control of weedy plant species and importantly DHPS inhibitor resistant weed biotypes. Aspects of the method can be applied to manage various weedy plants in agronomic and other cultivated environments.

[0013] The following definitions and methods are provided to better define the present invention and to guide those of ordinary skill in the art. Unless otherwise noted, terms are to be understood according to conventional usage by those of ordinary skill in the relevant art. Where a term is provided in the singular, the inventors also contemplate aspects described by the plural of that term.

[0014] By "non-transcribable" polynucleotides is meant that the polynucleotides do not comprise a complete polymerase II transcription unit.

[0015] As used herein "solution" refers to homogeneous mixtures and non-homogeneous mixtures such as suspensions, colloids, micelles, and emulsions.

[0016] Weedy plants are plants that compete with cultivated plants, those of particular importance include, but are not limited to important invasive and noxious weeds and herbicide resistant biotypes in crop production, such as, *Amaranthus* species -*A. albus*, *A. blitoides*, *A. hybridus*, *A. palmeri*, *A. powelli*, *A. retroflexus*, *A. spinosus*, *A. tuberculatus*, and *A.*

*viridis; Ambrosia* species - *A. trifida, A. artemisifolia; Lolium* species -*L. multiflorum, L. rigidum, L perenne; Digitaria* species -*D. insularis; Euphorbia* species -*E. heterophylla; Kochia* species -*K. scoparia; Sorghum* species -*S. halepense; Conyza* species -*C. bonariensis, C. canadensis, C. sumatrensis; Chloris* species -*C. truncate; Echinochola* species - *E. colona, E. crus-galli; Eleusine* species -*E. indica; Poa* species -*P. annua; Plantago* species -*P. lanceolata; Avena* species -*A. fatua; Chenopodium* species - *C. album; Setaria* species - *S. viridis, Abutilon theophrasti, Ipomoea* species, *Sesbania*, species, *Cassia* species, *Sida* species, *Brachiaria*, species and *Solanum* species.

**[0017]** Additional weedy plant species found in cultivated areas include *Alopecurus myosuroides, Avena sterilis, Avena sterilis ludoviciana, Brachiaria plantaginea, Bromus diandrus, Bromus rigidus, Cynosurus echinatus, Digitaria ciliaris, Digitaria ischaemum, Digitaria sanguinalis, Echinochloa oryzicola, Echinochloa phyllopogon, Eriochloa punctata, Hordeum glaucum, Hordeum leporinum, Ischaemum rugosum, Leptochloa chinensis, Lolium persicum, , Phalaris minor, Phalaris paradoxa, Rottboellia exalta, Setaria faberi, Setaria viridis var, robusta-alba schreiber, Setaria viridis var, robusta -purpurea, Snowdenia polystachaea, Sorghum sudanese, Alisma plantago-aquatica, Amaranthus lavidus, Amaranthus quitensis, Ammania auriculata, Ammania coccinea, Anthemis cotula, Apera spica-venti, Bacopa rotundifolia, Bidens pilosa, Bidens subalternans, Brassica tournefortii, Bromus tectorum, Camelina microcarpa, Chrysanthemum coronarium, Cuscuta campestris, Cyperus difformis, Damasonium minus, Descurainia sophia, Diplotaxis tenuifolia, Echium plantagineum, Elatine triandra var, pedicellata, Euphorbia heterophylla, Fallopia convolvulus, Fimbristylis miliacea, Galeopsis tetrahit, Galium spurium, Helianthus annuus, Iva xanthifolia, Ixophorus unisetus, Ipomoea indica, Ipomoea purpurea, Ipomoea sepia, Ipomoea aquatic, Ipomoea triloba, Lactuca serriola, Limnocharis flava, Limnophila erecta, Limnophila sessiliflora, Lindernia dubia, Lindernia dubia var, major, Lindernia micrantha, Lindernia procumbens, Mesembryanthemum crystallinum, Monochoria korsakowii, Monochoria vaginalis, Neslia paniculata, Papaver rhoes, Parthenium hysterophorus, Pentzia suffruticosa, Phalaris minor, Raphanus raphanistrum, Raphanus sativus, Rapistrum rugosum, Rotala indica var, uliginosa, Sagittaria guyanensis, Sagittaria montevidensis, Sagittaria pygmaea, Salsola iberica, Scirpusjuncoides var, ohwianus, Scirpus mucronatus, Setaria lutescens, Sida spinosa, Sinapis arvensis, Sisymbrium orientale, Sisymbrium thellungi, Solanum ptycanthum, Sonchus asper, Sonchus oleraceus, Sorghum bicolor, Stellaria media, Thlaspi arvense, Xanthium strumarium, Arctotheca calendula, Conyza sumatrensis, Crassocephalum crepidioides, Cynopephala carthageneris, Epilobium adenocaulon, Erigeron philadelphicus, Landoltia punctata, Lepidium virginicum, Monochoria korsakowii, Solanum americanum, Solanum nigrum, Vulpia bromoides, Youngia japonica, Hydrilla verticillata, Carduus nutans, Carduus pycnocephalus, Centaurea solstitialis, Cirsium arvense, Commelina diffusa, Convolvulus arvensis, Daucus carota, Digitaria ischaemum, Echinochloa crus-pavonis, Fimbristylis miliacea, Galeopsis tetrahit, Galium spurium, Limnophila erecta, Matricaria perforata, Papaver rhoes, Ranunculus acris, Soliva sessilis, Sphenoclea zeylanica, Stellaria media, Nassella trichotoma, Stipa neesiana, Agrostis stolonifera, Polygonum aviculare, Alopecurus japonicus, Beckmannia syzigachne, Bromus tectorum, Chloris inflate, Echinochloa erecta, Portulaca oleracea, and Senecio vulgaris. It is believed that all plants contain an DHPS gene in their genome, which can be isolated and polynucleotides made according to the methods that are useful for regulating, suppressing or delaying the expression of the target DHPS gene in the plants and the growth or development of the treated plants.*

**[0018]** Some cultivated plants may also be weedy plants when they occur in unwanted environments. For example, corn plants growing in a soybean field. Transgenic crops with one or more herbicide tolerances will need specialized methods of management to control weeds and volunteer crop plants.

**[0019]** A "trigger" or "trigger polynucleotide" is a polynucleotide molecule that is homologous or complementary to a target gene polynucleotide. The trigger polynucleotide molecules modulate expression of the target gene when topically applied to a plant surface with a transfer agent, whereby a plant treated with said composition has its growth or development or reproductive ability regulated, suppressed or delayed or said plant is more sensitive to a DHPS inhibitor herbicide or mitosis inhibitor herbicide as a result of said polynucleotide containing composition relative to a plant not treated with a composition containing the trigger molecule.

**[0020]** It is contemplated that the compositions will contain one or more polynucleotides and one or more herbicides that include but not limited to DHPS gene trigger polynucleotides and a DHPS inhibitor herbicide and anyone or more additional herbicide target gene trigger polynucleotides and the related herbicides and anyone or more additional essential gene trigger polynucleotides. Essential genes are genes in a plant that provide key enzymes or other proteins, for example, a biosynthetic enzyme, metabolizing enzyme, receptor, signal transduction protein, structural gene product, transcription factor, or transport protein; or regulating RNAs, such as, microRNAs, that are essential to the growth or survival of the organism or cell or involved in the normal growth and development of the plant (Meinke, et al., Trends Plant Sci. 2008 Sep;13(9):483-91). The suppression of an essential gene enhances the effect of a herbicide that affects the function of a gene product different than the suppressed essential gene. The compositions can include various trigger polynucleotides that modulate the expression of an essential gene other than DHPS.

**[0021]** Herbicides, for which transgenes for plant tolerance have been demonstrated, include but are not limited to: auxin-like herbicides, glyphosate, glufosinate, sulfonylureas, imidazolinones, bromoxynil, delapon, dicamba, cyclohexanedione, protoporphyrionogen oxidase inhibitors, 4-hydroxyphenyl-pyruvate-dioxygenase inhibitors herbicides. For example, transgenes and their polynucleotide molecules that encode proteins involved in herbicide tolerance are known

in the art, and include, but are not limited to an 5-enolpyruvylshikimate-3-phosphate synthase (EPSPS), for example, as more fully described in U.S. Pat. Nos. 7,807,791 (SEQ ID NO:5); 6,248,876 B1; 5,627,061; 5,804,425; 5,633,435; 5,145,783; 4,971,908; 5,392,910; 5,188,642; 4,940,835; 5,866,775; 6,225,114 B1; 6,130,366; 5,390,667; 4,535,060; 4,769,061; 5,633,448; 5,510,471; U.S. Pat. No. RE36,449; U.S. Pat. Nos. RE37,287 E; and 5,491,288; tolerance to 5 sulfonylurea and/or imidazolinone, for example, as described more fully in U.S. Pat. Nos. 5,605,011; 5,013,659; 5,141,870; 5,767,361; 5,739,180; 5,304,732; 4,761,373; 5,339,107; 5,928,937; and 5,378,824; and international publication WO 96/33270; tolerance to hydroxyphenylpyruvatedioxygenases inhibitiong herbicides in plants are described in U.S. Pat. Nos. 6,245,968 B1; 6,268,549, and 6,069,115; US Pat.Pub. 20110191897 and US7,392,379 SEQ ID NO:3; US7,935,869; US7,304,209, SEQ ID NO:1, 3,5 and 15; aryloxyalkanoate dioxygenase polynucleotides, which confer 10 tolerance to 2,4-D and other phenoxy auxin herbicides as well as to aryloxyphenoxypropionate herbicides as described, for example, in WO2005/107437; US7,838,733 SEQ ID NO:5;) and dicamba-tolerance polynucleotides as described, for example, in Herman et al. (2005) J. Biol. Chem. 280: 24759-24767. Other examples of herbicide-tolerance traits 15 include those conferred by polynucleotides encoding an exogenous phosphinothricin acetyltransferase, as described in U.S. Pat. Nos. 5,969,213; 5,489,520; 5,550,398; 5,874,265; 5,919,675; 5,561,236; 5,648,477; 5,639,024; 6,177,616; and 5,879,903. Plants containing an exogenous phosphinothricin acetyltransferase can exhibit improved tolerance to 20 glufosinate herbicides, which inhibit the enzyme glutamine synthase. Additionally, herbicide-tolerance polynucleotides include those conferred by polynucleotides conferring altered protoporphyrinogen oxidase (protox) activity, as described in U.S. Pat. Nos. 6,288,306 B1; 6,282,837 B1; and 5,767,373; and WO 01/12825. Plants containing such polynucleotides can exhibit improved tolerance to any of a variety of herbicides which target the protox enzyme (also referred to as 25 protox inhibitors). Polynucleotides encoding a glyphosate oxidoreductase and a glyphosate-N-acetyl transferase (GOX) described in U.S. Patent 5,393,175 and GAT described in U.S. Patent publication 20030083480, dicamba monooxygenase U.S. Patent publication 20030135879 ; a polynucleotide molecule encoding bromoxynil nitrilase (*Bxn* described in U.S. Patent No. 4,810,648 for Bromoxynil tolerance 30 25 ; a polynucleotide molecule encoding phytoene desaturase (*crtl*) described in Misawa et al, (1993) Plant J. 4:833-840 and Misawa et al, (1994) Plant J. 6:481-489 for norflurazon tolerance; a polynucleotide molecule encoding acetohydroxyacid synthase (AHAS, aka ALS) described in Sathasiivan et al. (1990) Nucl. Acids Res. 18:398-2193 for tolerance to sulfonylurea herbicides; and the *bar* gene described in DeBlock, et al. (1987) EMBO J. 6:2513-2519 for glufosinate and bialaphos tolerance. The transgenic coding regions and regulatory elements of the herbicide tolerance genes are 35 targets in which polynucleotide triggers and herbicides can be included in the compositions.

**[0022]** DHPS inhibitor herbicides include but are not limited to carbamates and asulam. Mitosis inhibitor herbicides include but are not limited to dinitroaniline herbicides for example benfluralin, butralin, dinitramine, ethalfluralin, oryzalin, pendimethalin, and trifluralin. Additional mitosis inhibitor herbicides also include but are not limited to Phosphoroamidates, Pyridines, Benzamides, and Benzeneddicarboxylic acids.

**[0023]** Numerous herbicides with similar or different modes of action (herein referred to as co-herbicides) are available that can be added to the compositions, for example, members of the herbicide families that include but are not limited to amide herbicides, aromatic acid herbicides, arsenical herbicides, benzothiazole herbicides, benzoylcyclohexanedione herbicides, benzofuranyl alkylsulfonate herbicides, carbamate herbicides, cyclohexene oxime herbicides, cyclopropyl-isoxazole herbicides, dicarboximide herbicides, dinitroaniline herbicides, dinitrophenol herbicides, diphenyl ether herbicides, dithiocarbamate herbicides, halogenated aliphatic herbicides, imidazolinone herbicides, inorganic herbicides, nitrile herbicides, organophosphorus herbicides, oxadiazolone herbicides, oxazole herbicides, phenoxy herbicides, phenylenediamine herbicides, pyrazole herbicides, pyridazine herbicides, pyridazinone herbicides, pyridine herbicides, pyrimidinediamine herbicides, pyrimidinyloxybenzylamine herbicides, quaternary ammonium herbicides, thiocarbamate herbicides, thiocarbonate herbicides, thiourea herbicides, triazine herbicides, triazinone herbicides, triazole herbicides, 40 triazolone herbicides, triazolopyrimidine herbicides, uracil herbicides, and urea herbicides. In particular, the rates of use of the added herbicides can be reduced in compositions comprising the polynucleotides. Use rate reductions of the additional added herbicides can be 10-25 percent, 26-50 percent, 51-75 percent or more can be achieved that enhance the activity of the polynucleotides and herbicide composition and is contemplated. Representative co-herbicides of the families include but are not limited to acetochlor, acifluorfen, acifluorfen-sodium, aclonifen, acrolein, alachlor, alloxydim, allyl alcohol, ametryn, amicarbazone, amidosulfuron, aminopyralid, amitrole, ammonium sulfamate, anilofos, asulam, atraton, atrazine, azimsulfuron, BCPC, beflobutamid, benazolin, benfluralin, benfuresate, bensulfuron, bensulfuron-methyl, bensulide, bentazone, benzfendizone, benzobicyclon, benzofenap, bifenoxy, bilanafos, bispyribac, bispyribac-sodium, borax, bromacil, bromobutide, bromoxynil, butachlor, butafenacil, butamifos, butralin, butroxydim, butylate, cacodylic acid, calcium chlorate, cafenstrole, carbetamide, carfentrazone, carfentrazone-ethyl, CDEA, CEPC, chlorflurenol, chlorflurenol-methyl, chlорidazon, chlorimuron, chlorimuron-ethyl, chloroacetic acid, chlorotoluron, chlorpropham, chlorsulfuron, chlorthal, chlorthal-dimethyl, cinidon-ethyl, cinmethylin, cinosulfuron, cisanilide, clethodim, clodinafop, 45 clodinafop-propargyl, clomazone, clomeprop, clopyralid, cloransulam, cloransulam-methyl, CMA, 4-CPB, CPMF, 4-CPP, CPPC, cresol, cumyluron, cyanamide, cyanazine, cycloate, cyclosulfamuron, cycloxydim, cyhalofop, cyhalofop-butyl,

2,4-D, 3,4-DA, daimuron, dalapon, dazomet, 2,4-DB, 3,4-DB, 2,4-DEB, desmedipham, dicamba, dichlobenil, ortho-dichlorobenzene, para-dichlorobenzene, dichlorprop, dichlorprop-P, diclofop, diclofop-methyl, diclosulam, difenzoquat, difenzoquat metilsulfate, diflufenican, diflufenzopyr, dimefuron, dimepiperate, dimethachlor, dimethametryn, dimethenamid, dimethenamid-P, dimethylaminoc acid, dinitramine, dinoterm, diphenamid, diquat, diquat dibromide, dithiopyr, diuron, DNOC, 3,4-DP, DSMA, EBEP, endothal, EPTC, esprocarb, ethalfluralin, ethametsulfuron, ethametsulfuron-methyl, ethofumesate, ethoxyfen, ethoxysulfuron, etobenzanid, fenoxaprop-P, fenoxaprop-P-ethyl, fentrazamide, ferrous sulfate, flamprop-M, flazasulfuron, florasulam, fluazifop, fluazifop-butyl, fluazifop-P, fluazifop-P-butyl, flucarbazone, flucarbazone-sodium, flucetosulfuron, fluchloralin, flufenacet, flufenpyr, flufenpyr-ethyl, flumetsulam, flumiclorac, flumiclorac-pentyl, flumioxazin, fluometuron, fluoroglycofen, fluoroglycofen-ethyl, flupropanate, flupyrifurfuron, flupyrifurfuron-methyl-sodium, flurenol, fluridone, fluorochloridone, fluoroxypryn, flurtamone, fluthiacet, fluthiacet-methyl, fomesafen, foramsulfuron, fosamine, glufosinate, glufosinate-ammonium, glyphosate, halosulfuron, halosulfuron-methyl, haloxyfop, haloxyfop-P, HC-252, hexazinone, imazamethabenz, imazamethabenz-methyl, imazamox, imazapic, imazapyr, imazaquin, imazethapyr, imazosulfuron, indanofan, iodomethane, iodosulfuron, iodosulfuron-methyl-sodium, ioxynil, isoproturon, isouron, isoxaben, isoxachlortole, isoxaflutole, karbutilate, lactofen, lenacil, linuron, MAA, MAMA, MCPA, MCPA-thioethyl, MCPB, mecoprop, mecoprop-P, mefenacet, mefluidide, mesosulfuron, mesosulfuron-methyl, mesotrione, metam, metamifop, metamitron, metazachlor, methabenzthiazuron, methylarsonic acid, methyldymron, methyl isothiocyanate, metobenzuron, metolachlor, S-metolachlor, metosulam, metoxuron, metribuzin, metsulfuron, metsulfuron-methyl, MK-66, molinate, monolinuron, MSMA, naproanilide, napropamide, naptalam, neburon, nicosulfuron, nonanoic acid, norflurazon, oleic acid (fatty acids), orbencarb, orthosulfamuron, oryzalin, oxadiargyl, oxadiazon, oxasulfuron, oxaziclofone, oxyfluorfen, paraquat, paraquat dichloride, pebulate, pendimethalin, penoxsulam, pentachlorophenol, pentanochlor, pentozacone, pethoxamid, petroleum oils, phenmedipham, phenmedipham-ethyl, picloram, picolinafen, pinoxaden, piperophos, potassium arsenite, potassium azide, pretilachlor, primisulfuron, primisulfuron-methyl, prodiamine, profluazol, profoxydim, prometon, prometryn, propachlor, propanil, propaquizafop, propazine, prophan, propisochlor, propoxycarbazone, propoxycarbazone-sodium, propyzamide, prosulfocarb, prosulfuron, pyraclonil, pyraflufen, pyraflufen-ethyl, pyrazolynate, pyrazosulfuron, pyrazosulfuron-ethyl, pyrazoxyfen, pyribenzoxim, pyributicarb, pyridafol, pyridate, pyriftalid, pyriminobac, pyriminobac-methyl, pyrimisulfan, pyrithiobac, pyrithiobac-sodium, quinclorac, quinmerac, quinooclamine, quizalofop, quizalofop-P, rimsulfuron, sethoxydim, siduron, simazine, simetryn, SMA, sodium arsenite, sodium azide, sodium chloride, sulcotrione, sulfentrazone, sulfometuron, sulfometuron-methyl, sulfosate, sulfosulfuron, sulfuric acid, tar oils, 2,3,6-TBA, TCA, TCA-sodium, tebuthiuron, tepraloxydim, terbacil, terbumeton, terbutylazine, terbutryn, thenylchlor, thiazopyr, thifensulfuron, thifensulfuron-methyl, thiobencarb, tiocarbazil, topramezone, tralkoxydim, tri-allate, triasulfuron, triaziflam, tribenuron, tribenuron-methyl, tricamba, triclopyr, trietazine, trifloxysulfuron, trifloxysulfuron-sodium, trifluralin, triflusulfuron, triflusulfuron-methyl, trihydroxytriazine, tritosulfuron, [3-[2-chloro-4-fluoro-5-(methyl-6-trifluoromethyl-2,4-dioxo-2,3,4-tetrahydropyrimidin-3-yl)phenoxy]-2-pyridyloxy]acetic acid ethyl ester (CAS RN 353292-3-6), 4-[(4,5-dihydro-3-methoxy-4-methyl-5-oxo)-H-2,4-triazol-ylcarbonyl-sulfamoyl]-5-methylthiophene-3-carboxylic acid (BAY636), BAY747 (CAS RN 33504-84-2), topramezone (CAS RN 2063-68-8), 4-hydroxy-3-[[2-[(2-methoxyethoxy)methyl]-6-(trifluoro-methyl)-3-pyridinyl]carbonyl]-bicyclo[3.2.]oct-3-en-2-one (CAS RN 35200-68-5), and 4-hydroxy-3-[[2-(3-methoxypropyl)-6-(difluoromethyl)-3-pyridinyl]carbon-yl]-bicyclo[3.2.]oct-3-en-2-one. Additionally, including herbicidal compounds of unspecified modes of action as described in CN101279950A, CN101279951A, DE10000600A1, DE10116399A1, DE102004054666A1, DE102005014638A1, DE102005014906A1, DE102007012168A1, DE102010042866A1, DE10204951A1, DE10234875A1, DE10234876A1, DE10256353A1, DE10256354A1, DE10256367A1, EP1157991A2, EP1238586A1, EP2147919A1, EP2160098A2, JP03968012B2, JP2001253874A, JP2002080454A, JP2002138075A, JP2002145707A, JP2002220389A, JP2003064059A, JP2003096059A, JP2004051628A, JP2004107228A, JP2005008583A, JP2005239675A, JP2005314407A, JP2006232824A, JP2006282552A, JP2007153847A, JP2007161701A, JP2007182404A, JP2008074840A, JP2008074841A, JP2008133207A, JP2008133218A, JP2008169121A, JP2009067739A, JP2009114128A, JP2009126792A, JP2009137851A, US20060111241A1, US20090036311A1, US20090054240A1, US20090215628A1, US20100099561A1, US20100152443A1, US20110105329A1, US20110201501A1, WO2001055066A2, WO2001056975A1, WO2001056979A1, WO2001090071A2, WO2001090080A1, WO2002002540A1, WO2002028182A1, WO2002040473A1, WO2002044173A2, WO2003000679A2, WO2003006422A1, WO2003013247A1, WO2003016308A1, WO2003020704A1, WO2003022051A1, WO2003022831A1, WO2003022843A1, WO2003029243A2, WO2003037085A1, WO2003037878A1, WO2003045878A2, WO2003050087A2, WO2003051823A1, WO2003051824A1, WO2003051846A2, WO2003076409A1, WO2003087067A1, WO2003090539A1, WO2003091217A1, WO2003093269A2, WO2003104206A2, WO2004002947A1, WO2004002981A2, WO2004011429A1, WO2004029060A1, WO2004035545A2, WO2004035563A1, WO2004035564A1, WO2004037787A1, WO2004067518A1, WO2004067527A1, WO2004077950A1, WO2005000824A1, WO2005007627A1, WO2005040152A1, WO2005047233A1, WO2005047281A1, WO2005061443A2, WO2005061464A1, WO2005068434A1, WO2005070889A1, WO2005089551A1, WO2005095335A1, WO2006006569A1, WO2006024820A1,

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 WO2010119906A1, WO2010130970A1, WO2011003776A2, WO2011035874A1, WO2011065451A1

**[0024]** An agronomic field in need of plant control is treated by application of the composition directly to the surface of the growing plants, such as by a spray. For example, the method is applied to control weeds in a field of crop plants by spraying the field with the composition.. The composition can be provided as a tank mix, a sequential treatment of components (generally the polynucleotide containing composition followed by the herbicide), or a simultaneous treatment or mixing of one or more of the components of the composition from separate containers. Treatment of the field can occur as often as needed to provide weed control and the components of the composition can be adjusted to target specific weed species or weed families through utilization of specific polynucleotides or polynucleotide compositions capable of selectively targeting the specific species or plant family to be controlled. The composition can be applied at effective use rates according to the time of application to the field, for example, preplant, at planting, post planting, postharvest. DHPS inhibitor herbicides can be applied to a field at rates of 500 to 3000 g ai/ha (active ingredient per hectare) or more. The polynucleotides of the composition can be applied at rates of 1 to 30 grams per acre depending on the number of trigger molecules needed for the scope of weeds in the field.

**[0025]** Crop plants in which weed control is needed include but are not limited to, i) corn, soybean, cotton, canola, sugar beet, alfalfa, sugarcane, rice, and wheat; ii) vegetable plants including, but not limited to, tomato, sweet pepper, hot pepper, melon, watermelon, cucumber, eggplant, cauliflower, broccoli, lettuce, spinach, onion, peas, carrots, sweet corn, Chinese cabbage, leek, fennel, pumpkin, squash or gourd, radish, Brussels sprouts, tomatillo, garden beans, dry beans, or okra; iii) culinary plants including, but not limited to, basil, parsley, coffee, or tea; or , iv) fruit plants including but not limited to apple, pear, cherry, peach, plum, apricot, banana, plantain, table grape, wine grape, citrus, avocado, mango, or berry; v) a tree grown for ornamental or commercial use, including, but not limited to, a fruit or nut tree; or, vi) an ornamental plant (e. g., an ornamental flowering plant or shrub or turf grass). The methods and compositions provided herein can also be applied to plants produced by a cutting, cloning, or grafting process (i. e., a plant not grown from a seed) include fruit trees and plants that include, but are not limited to, citrus, apples, avocados, tomatoes, eggplant, cucumber, melons, watermelons, and grapes as well as various ornamental plants.

### Pesticidal Mixtures

**[0026]** The polynucleotide compositions may also be used as mixtures with various agricultural chemicals and/or insecticides, miticides and fungicides, pesticidal and biopesticidal agents. Examples include but are not limited to azinphos-methyl, acephate, isoxathion, isofenphos, ethion, etrimfos, oxydemeton-methyl, oxydeprofos, quinalphos, chlorpyrifos, chlorpyrifos-methyl, chlorgenvinphos, cyanophos, dioxabenzofos, dichlorvos, disulfoton, dimethylvinphos, dimethoate, sulprofos, diazinon, thiometon, tetrachlorvinphos, temephos, tebupirimfos, terbufos, naled, vamidothion, pyraclofos, pyridafenthion, pirimiphos-methyl, fenitrothion, fenthion, phentoate, flupyrazophos, prothiofos, propaphos, profenofos, phoxime, phosalone, phosmet, formothion, phorate, malathion, mecarbam, mesulfenfos, methamidophos, methidathion, parathion, methyl parathion, monocrotophos, trichlorphon, EPN, isazophos, isamidofofos, cadusafos, di-amidaphos, dichlofenthion, thionazin, fenamiphos, fosthiazate, fosthietan, phosphocarb, DSP, ethoprophos, alanycarb, aldicarb, isoprocarb, ethiofencarb, carbaryl, carbosulfan, xylylcarb, thiodicarb, pirimicarb, fenobucarb, furathiocarb, propoxur, bendiocarb, benfuracarb, methomyl, metolcarb, XMC, carbofuran, aldoxycarb, oxamyl, acrinathrin, allethrin, esfenvalerate, empenthrin, cycloprothrin, cyhalothrin, gamma-cyhalothrin, lambda-cyhalothrin, cyfluthrin, beta-cyfluthrin, cypermethrin, alpha-cypermethrin, zeta-cypermethrin, silafluofen, tetramethrin, tefluthrin, deltamethrin, tralomethrin, bifenthrin, phenothrin, fenvalerate, fenpropothrin, furamethrin, prallethrin, flucythrinate, fluvalinate, flubrocyclhrinate, permethrin, resmethrin, ethofenprox, cartap, thiocyclam, bensultap, acetamiprid, imidacloprid, clothianidin, dinotefuran, thiacycloprid, thiamethoxam, nitenpyram, chlorfluazuron, diflubenzuron, teflubenzuron, triflumuron, novaluron, noviflumuron, bistrifluoron, fluazuron, flucycloxuron, flufenoxuron, hexaflumuron, lufenuron, chromafenozide, tebufenozide, halofenozone, methoxyfenozide, diofenolan, cyromazine, pyriproxyfen, buprofezin, methoprene, hydroprene, kinoprene, triazamate, endosulfan, chlorfenson, chlorbenzilate, dicofol, bromopropylate, acetoprole, fipronil, ethiprole, pyrethrin, rotenone, nicotine sulphate, BT (*Bacillus Thuringiensis*) agent, spinosad, abamectin, acequinocyl, amidoflumet, amitraz,

etoxazole, chinomethionat, clofentezine, fenbutatin oxide, dienochlor, cyhexatin, spirodiclofen, spiromesifen, tetradifon, tebufenpyrad, binapacryl, bifenazate, pyridaben, pyrimidifen, fenazaquin, fenothiocarb, fenpyroximate, fluacrypyrim, fluazinam, flufenzin, hexythiazox, propargite, benzamate, polynactin complex, milbemectin, lufenuron, mecarbam, methiocarb, mevinphos, halfenprox, azadirachtin, diafenthiuron, indoxacarb, emamectin benzoate, potassium oleate, sodium oleate, chlorfenapyr, tolfenpyrad, pymetrozine, fenoxy carb, hydramethylnon, hydroxy propyl starch, pyridalyl, flufenim, flubendiamide, flonicamid, metaflumizole, lepimectin, TPIC, albendazole, oxbendazole, oxfendazole, trichlamide, fensulfothion, fenbendazole, levamisole hydrochloride, morantel tartrate, dazomet, metam-sodium, triadimefon, hexaconazole, propiconazole, ipconazole, prochloraz, triflumizole, tebuconazole, epoxiconazole, difenoconazole, flusazole, triadimenol, cyproconazole, metconazole, fluquinconazole, bitertanol, tetaconazole, triticonazole, flutriafol, penconazole, diniconazole, fenbuconazole, bromuconazole, imibenconazole, simeconazole, myclobutanil, hymexazole, imazalil, furametypy, thifluzamide, etridiazole, oxpoconazole, oxpoconazole fumarate, perfurazoate, prothioconazole, pyrifenoxy, fenarimol, nuarimol, bupirimate, mepanipyrim, cyprodinil, pyrimethanil, metalaxyl, mefenoxam, oxadixyl, benalaxyl, thiophanate, thiophanate-methyl, benomyl, carbendazim, fuberidazole, thiabendazole, manzeb, propineb, zineb, metiram, maneb, ziram, thiuram, chlorothalonil, ethaboxam, oxycarboxin, carboxin, flutolanil, silthofam, mepronil, dimethomorph, fenpropidin, fenpropimorph, spiroxamine, tridemorph, dodemorph, flumorph, azoxystrobin, kresoxim-methyl, metominostrobin, orysastrobin, fluoxastrobin, trifloxystrobin, dimoxystrobin, pyraclostrobin, picoxystrobin, iprodione, procymidone, vinclozolin, chlozolinate, flusulfamide, dazomet, methyl isothiocyanate, chloropicrin, methasulfo-carb, hydroxyisoxazole, potassium hydroxyisoxazole, echlomezol, D-D, carbam, basic copper chloride, basic copper sulfate, copper nonylphenolsulfonate, oxine copper, DBEDC, anhydrous copper sulfate, copper sulfate pentahydrate, cupric hydroxide, inorganic sulfur, wettable sulfur, lime sulfur, zinc sulfate, fentin, sodium hydrogen carbonate, potassium hydrogen carbonate, sodium hypochlorite, silver, edifenphos, tolclofos-methyl, fosetyl, iprobenfos, dinocap, pyrazophos, carpropamid, fthalide, tricyclazole, pyroquilon, diclocymet, fenoxyanil, kasugamycin, validamycin, polyoxins, blasticiden S, oxytetracycline, mildiomycin, streptomycin, rape seed oil, machine oil, benthiavalicarbisopropyl, iprovalicarb, propamocarb, diethofencarb, fluoroimide, fludioxanil, fenpiclonil, quinoxyfen, oxolinic acid, chlorothalonil, captan, folpet, probenazole, acibenzolar-S-methyl, tiadinil, cyflufenamid, fenhexamid, diflumetorim, metrafenone, picobenzamide, proquinazid, famoxadone, cyazofamid, fenamidone, zoxamide, boscalid, cymoxanil, dithianon, fluazinam, dichlofluanide, triforine, isoprothiolane, ferimzone, diclomezine, tecloftalam, pencycuron, chinomethionat, iminoctadine acetate, iminoctadine albesilate, ambam, polycarbamate, thiadiazine, chloroneb, nickel dimethyldithiocarbamate, guazatine, dodecylguanidine-acetate, quintozen, tolylfluanid, anilazine, nitrothalisopropyl, fenitropan, dimethirimol, benthiazole, harpin protein, flumetover, mandipropamide and penthiopyrad.

## Polynucleotides

**[0027]** As used herein, the term "DNA", "DNA molecule", "DNA polynucleotide molecule" refers to a single-stranded DNA (ssDNA) or double-stranded DNA (dsDNA) molecule of genomic or synthetic origin, such as, a polymer of deoxyribonucleotide bases or a DNA polynucleotide molecule. As used herein, the term "DNA sequence", "DNA nucleotide sequence" or "DNA polynucleotide sequence" refers to the nucleotide sequence of a DNA molecule. As used herein, the term "RNA", "RNA molecule", "RNA polynucleotide molecule" refers to a single-stranded RNA (ssRNA) or double-stranded RNA (dsRNA) molecule of genomic or synthetic origin, such as, a polymer of ribonucleotide bases that comprise single or double stranded regions. Unless otherwise stated, nucleotide sequences in the text of this specification are given, when read from left to right, in the 5' to 3' direction. The nomenclature used herein is that required by Title 37 of the United States Code of Federal Regulations § 1.822 and set forth in the tables in WIPO Standard ST.25 (1998), Appendix 2, Tables 1 and 3.

**[0028]** As used herein, "polynucleotide" refers to a DNA or RNA molecule containing multiple nucleotides and generally refers both to "oligonucleotides" (a polynucleotide molecule of typically 50 or fewer nucleotides in length) and polynucleotides of 51 or more nucleotides. Embodiments include compositions including oligonucleotides having a length of 18-25 nucleotides (18-mers, 19-mers, 20-mers, 21-mers, 22-mers, 23-mers, 24-mers, or 25-mers), for example, oligonucleotides of Table 3 (SEQ ID NO:907-1175) or fragments thereof or medium-length polynucleotides having a length of 26 or more nucleotides (polynucleotides of 26, 27, 28, 29, 30, 39, 32, 33, 34, 35, 36, 37, 38, 39, 40, 41, 42, 43, 44, 45, 39, 47, 48, 49, 50, 51, 52, 53, 54, 55, 56, 57, 58, 59, 60, about 65, about 70, about 75, about 80, about 85, about 90, about 95, about 100, about 110, about 120, about 130, about 140, about 150, about 160, about 170, about 180, about 190, about 200, about 210, about 220, about 230, about 240, about 250, about 260, about 270, about 280, about 290, or about 300 nucleotides), for example, polynucleotides of Table 2 (SEQ ID NO: 55-906) or fragments thereof or long polynucleotides having a length greater than about 300 nucleotides (for example, polynucleotides of between about 300 to about 400 nucleotides, between about 400 to about 500 nucleotides, between about 500 to about 600 nucleotides, between about 600 to about 700 nucleotides, between about 700 to about 800 nucleotides, between about 800 to about 900 nucleotides, between about 900 to about 1000 nucleotides, between about 300 to about 500 nucleotides, between about 300 to about 600 nucleotides, between about 300 to about 700 nucleotides, between about 300 to about 800

nucleotides, between about 300 to about 900 nucleotides, or about 1000 nucleotides in length, or even greater than about 1000 nucleotides in length, for example up to the entire length of a target gene including coding or non-coding or both coding and non-coding portions of the target gene), for example, polynucleotides of Table 1 (SEQ ID NO:1-54), wherein the selected polynucleotides or fragments thereof homologous or complementary to SEQ ID NO:1-54 suppresses, represses or otherwise delay the expression of the target DHPS gene. A target gene comprises any polynucleotide molecule in a plant cell or fragment thereof for which the modulation of the expression of the target gene is provided by the methods and compositions. Where a polynucleotide is double-stranded, its length can be similarly described in terms of base pairs. Oligonucleotides and polynucleotides can be made that are essentially identical or essentially complementary to adjacent genetic elements of a gene, for example, spanning the junction region of an intron and exon, the junction region of a promoter and a transcribed region, the junction region of a 5' leader and a coding sequence, the junction of a 3' untranslated region and a coding sequence.

**[0029]** Polynucleotide compositions used in the various embodiments include compositions including oligonucleotides or polynucleotides or a mixture of both, including RNA or DNA or RNA/DNA hybrids or chemically modified oligonucleotides or polynucleotides or a mixture thereof. In some embodiments, the polynucleotide may be a combination of ribonucleotides and deoxyribonucleotides, for example, synthetic polynucleotides consisting mainly of ribonucleotides but with one or more terminal deoxyribonucleotides or synthetic polynucleotides consisting mainly of deoxyribonucleotides but with one or more terminal dideoxyribonucleotides. In some embodiments, the polynucleotide includes non-canonical nucleotides such as inosine, thiouridine, or pseudouridine. In some embodiments, the polynucleotide includes chemically modified nucleotides. Examples of chemically modified oligonucleotides or polynucleotides are well known in the art; see, for example, US Patent Publication 20110171287, US Patent Publication 20110171176, and US Patent Publication 20110152353, US Patent Publication, 20110152339, US Patent Publication 20110160082 . For example, including but not limited to the naturally occurring phosphodiester backbone of an oligonucleotide or polynucleotide can be partially or completely modified with phosphorothioate, phosphorodithioate, or methylphosphonate internucleotide linkage modifications, modified nucleoside bases or modified sugars can be used in oligonucleotide or polynucleotide synthesis, and oligonucleotides or polynucleotides can be labeled with a fluorescent moiety (for example, fluorescein or rhodamine) or other label (for example, biotin).

**[0030]** The polynucleotides can be single- or double-stranded RNA or single- or double-stranded DNA or double-stranded DNA/RNA hybrids or modified analogues thereof, and can be of oligonucleotide lengths or longer. In more specific embodiments, the polynucleotides that provide single-stranded RNA in the plant cell are selected from the group consisting of (a) a single-stranded RNA molecule (ssRNA), (b) a single-stranded RNA molecule that self-hybridizes to form a double-stranded RNA molecule, (c) a double-stranded RNA molecule (dsRNA), (d) a single-stranded DNA molecule (ssDNA), (e) a single-stranded DNA molecule that self-hybridizes to form a double-stranded DNA molecule, and (f) a single-stranded DNA molecule including a modified Pol III gene that is transcribed to an RNA molecule, (g) a double-stranded DNA molecule (dsDNA), (h) a double-stranded DNA molecule including a modified Pol III gene that is transcribed to an RNA molecule, (i) a double-stranded, hybridized RNA/DNA molecule, or combinations thereof. In some embodiments these polynucleotides include chemically modified nucleotides or non-canonical nucleotides. In some embodiments, the oligonucleotides may be blunt-ended or may comprise a 3' overhang of from 1-5 nucleotides of at least one or both of the strands. Other configurations of the oligonucleotide are known in the field and are contemplated herein. In embodiments of the method the polynucleotides include double-stranded DNA formed by intramolecular hybridization, double-stranded DNA formed by intermolecular hybridization, double-stranded RNA formed by intramolecular hybridization, or double-stranded RNA formed by intermolecular hybridization. In one embodiment the polynucleotides include single-stranded DNA or single-stranded RNA that self-hybridizes to form a hairpin structure having an at least partially double-stranded structure including at least one segment that will hybridize to RNA transcribed from the gene targeted for suppression. Not intending to be bound by any mechanism, it is believed that such polynucleotides are or will produce single-stranded RNA with at least one segment that will hybridize to RNA transcribed from the gene targeted for suppression. In certain other embodiments the polynucleotides further includes a promoter, generally a promoter functional in a plant, for example, a pol II promoter, a pol III promoter, a pol IV promoter, or a pol V promoter.

**[0031]** The term "gene" refers to components that comprise chromosomal DNA, plasmid DNA, cDNA, intron and exon DNA, artificial DNA polynucleotide, or other DNA that encodes a peptide, polypeptide, protein, or RNA transcript molecule, and the genetic elements flanking the coding sequence that are involved in the regulation of expression, such as, promoter regions, 5' leader regions, 3' untranslated region that may exist as native genes or transgenes in a plant genome. The gene or a fragment thereof is isolated and subjected to polynucleotide sequencing methods that determines the order of the nucleotides that comprise the gene. Any of the components of the gene are potential targets for a trigger oligonucleotide and polynucleotides.

**[0032]** The trigger polynucleotide molecules are designed to modulate expression by inducing regulation or suppression of an endogenous DHPS gene in a plant and are designed to have a nucleotide sequence essentially identical or essentially complementary to the nucleotide sequence of an endogenous DHPS gene of a plant or to the sequence of RNA transcribed from an endogenous DHPS gene of a plant, including a transgene in a plant that provides for a herbicide

resistant DHPS enzyme, which can be coding sequence or non-coding sequence. Effective molecules that modulate expression are referred to as "a trigger molecule, or trigger polynucleotides". By "essentially identical" or "essentially complementary" is meant that the trigger polynucleotides (or at least one strand of a double-stranded polynucleotide or portion thereof, or a portion of a single strand polynucleotide) are designed to hybridize to the endogenous gene noncoding sequence or to RNA transcribed (known as messenger RNA or an RNA transcript) from the endogenous gene to effect regulation or suppression of expression of the endogenous gene. Trigger molecules are identified by "tiling" the gene targets with partially overlapping probes or non-overlapping probes of antisense or sense polynucleotides that are essentially identical or essentially complementary to the nucleotide sequence of an endogenous gene. Multiple target sequences can be aligned and sequence regions with homology in common are identified as potential trigger molecules for the multiple targets. Multiple trigger molecules of various lengths, for example 18-25 nucleotides, 26-50 nucleotides, 51-100 nucleotides, 101-200 nucleotides, 201-300 nucleotides or more can be pooled into a few treatments in order to investigate polynucleotide molecules that cover a portion of a gene sequence (for example, a portion of a coding versus a portion of a noncoding region, or a 5' versus a 3' portion of a gene) or an entire gene sequence including coding and noncoding regions of a target gene. Polynucleotide molecules of the pooled trigger molecules can be divided into smaller pools or single molecules in order to identify trigger molecules that provide the desired effect.

**[0033]** The target gene RNA and DNA polynucleotide molecules (Table 1, SEQ ID NO:1-54) are sequenced by any number of available methods and equipment. Some of the sequencing technologies are available commercially, such as the sequencing-by-hybridization platform from Affymetrix Inc. (Sunnyvale, Calif.) and the sequencing-by-synthesis platforms from 454 Life Sciences (Bradford, Conn.), Illumina/Solexa (Hayward, Calif.) and Helicos Biosciences (Cambridge, Mass.), and the sequencing-by-ligation platform from Applied Biosystems (Foster City, Calif.), as described below. In addition to the single molecule sequencing performed using sequencing-by-synthesis of Helicos Biosciences, other single molecule sequencing technologies are encompassed by the method and include the SMRT™ technology of Pacific Biosciences, the Ion Torrent™ technology, and nanopore sequencing being developed for example, by Oxford Nanopore Technologies. A DHPS target gene comprising DNA or RNA can be isolated using primers or probes essentially complementary or essentially homologous to SEQ ID NO:1-54 or a fragment thereof. A polymerase chain reaction (PCR) gene fragment can be produced using primers essentially complementary or essentially homologous to SEQ ID NO:1-54 or a fragment thereof. that is useful to isolate a DHPS gene from a plant genome.

**[0034]** Embodiments of functional single-stranded polynucleotides functional have sequence complementarity that need not be 100 percent, but is at least sufficient to permit hybridization to RNA transcribed from the target gene or DNA of the target gene to form a duplex to permit a gene silencing mechanism. Thus, in embodiments, a polynucleotide fragment is designed to be essentially identical to, or essentially complementary to, a sequence of 18 or more contiguous nucleotides in either the target DHPS gene sequence or messenger RNA transcribed from the target gene. By "essentially identical" is meant having 100 percent sequence identity or at least about 83, 84, 85, 86, 87, 88, 89, 90, 91, 92, 93, 94, 95, 96, 97, 98, or 99 percent sequence identity when compared to the sequence of 18 or more contiguous nucleotides in either the target gene or RNA transcribed from the target gene; by "essentially complementary" is meant having 100 percent sequence complementarity or at least about 83, 84, 85, 86, 87, 88, 89, 90, 91, 92, 93, 94, 95, 96, 97, 98, or 99 percent sequence complementarity when compared to the sequence of 18 or more contiguous nucleotides in either the target gene or RNA transcribed from the target gene. In some embodiments, polynucleotide molecules are designed to have 100 percent sequence identity with or complementarity to one allele or one family member of a given target gene (coding or non-coding sequence of a gene); in other embodiments the polynucleotide molecules are designed to have 100 percent sequence identity with or complementarity to multiple alleles or family members of a given target gene. The trigger polynucleotide sequences in the sequence listing SEQ ID NO: 1-1222 or table 1, 2 or 3 maybe complementary or homologous to a portion of the DHPS target gene sequence.

**[0035]** In certain embodiments, the polynucleotides used in the compositions that are essentially identical or essentially complementary to the target gene or transcript will comprise the predominant nucleic acid in the composition. Thus in certain embodiments, the polynucleotides that are essentially identical or essentially complementary to the target gene or transcript will comprise at least about 50%, 75%, 95%, 98% or 100% of the nucleic acids provided in the composition by either mass or molar concentration. However, in certain embodiments, the polynucleotides that are essentially identical or essentially complementary to the target gene or transcript can comprise at least about 1% to about 50%, about 10% to about 50%, about 20% to about 50%, or about 30% to about 50% of the nucleic acids provided in the composition by either mass or molar concentration. Also provided are compositions where the polynucleotides that are essentially identical or essentially complementary to the target gene or transcript can comprise at least about 1% to 100%, about 10% to 100%, about 20% to about 100%, about 30% to about 50%, or about 50% to a 100% of the nucleic acids provided in the composition by either mass or molar concentration.

**[0036]** "Identity" refers to the degree of similarity between two polynucleic acid or protein sequences. An alignment of the two sequences is performed by a suitable computer program. A widely used and accepted computer program for performing sequence alignments is CLUSTALW v1.6 (Thompson, et al. Nucl. Acids Res., 22: 3973-3980, 1994). The number of matching bases or amino acids is divided by the total number of bases or amino acids, and multiplied by 100

to obtain a percent identity. For example, if two 580 base pair sequences had 145 matched bases, they would be 25 percent identical. If the two compared sequences are of different lengths, the number of matches is divided by the shorter of the two lengths. For example, if there are 100 matched amino acids between a 200 and a 400 amino acid protein, they are 50 percent identical with respect to the shorter sequence. If the shorter sequence is less than 150 bases or 50 amino acids in length, the number of matches are divided by 150 (for nucleic acid bases) or 50 (for amino acids), and multiplied by 100 to obtain a percent identity.

**[0037]** Trigger molecules for specific gene family members can be identified from coding and/or non-coding sequences of gene families of a plant or multiple plants, by aligning and selecting 200-300 polynucleotide fragments from the least homologous regions amongst the aligned sequences and evaluated using topically applied polynucleotides (as sense or anti-sense ssDNA or ssRNA, dsRNA, or dsDNA) to determine their relative effectiveness in inducing the herbicidal phenotype. The effective segments are further subdivided into 50-60 polynucleotide fragments, prioritized by least homology, and reevaluated using topically applied polynucleotides. The effective 50-60 polynucleotide fragments are subdivided into 19-30 polynucleotide fragments, prioritized by least homology, and again evaluated for induction of the yield/quality phenotype. Once relative effectiveness is determined, the fragments are utilized singly, or again evaluated in combination with one or more other fragments to determine the trigger composition or mixture of trigger polynucleotides for providing the yield/quality phenotype.

**[0038]** Trigger molecules for broad activity can be identified from coding and/or non-coding sequences of gene families of a plant or multiple plants, by aligning and selecting 200-300 polynucleotide fragments from the most homologous regions amongst the aligned sequences and evaluated using topically applied polynucleotides (as sense or anti-sense ssDNA or ssRNA, dsRNA, or dsDNA) to determine their relative effectiveness in inducing the yield/quality phenotype. The effective segments are subdivided into 50-60 polynucleotide fragments, prioritized by most homology, and reevaluated using topically applied polynucleotides. The effective 50-60 polynucleotide fragments are subdivided into 19-30 polynucleotide fragments, prioritized by most homology, and again evaluated for induction of the yield/quality phenotype. Once relative effectiveness is determined, the fragments may be utilized singly, or in combination with one or more other fragments to determine the trigger composition or mixture of trigger polynucleotides for providing the yield/quality phenotype.

**[0039]** Methods of making polynucleotides are well known in the art. Chemical synthesis, in vivo synthesis and in vitro synthesis methods and compositions are known in the art and include various viral elements, microbial cells, modified polymerases, and modified nucleotides. Commercial preparation of oligonucleotides often provides two deoxyribonucleotides on the 3' end of the sense strand. Long polynucleotide molecules can be synthesized from commercially available kits, for example, kits from Applied Biosystems/Ambion (Austin, TX) have DNA ligated on the 5' end in a microbial expression cassette that includes a bacterial T7 polymerase promoter that makes RNA strands that can be assembled into a dsRNA and kits provided by various manufacturers that include T7 RiboMax Express (Promega, Madison, WI), AmpliScribe T7-Flash (Epicentre, Madison, WI), and TranscriptAid T7 High Yield (Fermentas, Glen Burnie, MD). dsRNA molecules can be produced from microbial expression cassettes in bacterial cells (Ongvarrasopone et al. ScienceAsia 33:35-39; Yin, Appl. Microbiol. Biotechnol 84:323-333, 2009; Liu et al., BMC Biotechnology 10:85, 2010) that have regulated or deficient RNase III enzyme activity or the use of various viral vectors to produce sufficient quantities of dsRNA. DHPS gene fragments are inserted into the microbial expression cassettes in a position in which the fragments are express to produce ssRNA or dsRNA useful in the methods described herein to regulate expression on a target DHPS gene. Long polynucleotide molecules can also be assembled from multiple RNA or DNA fragments. In some embodiments design parameters such as Reynolds score (Reynolds et al. Nature Biotechnology 22, 326 - 330 (2004), Tuschl rules (Pei and Tuschl, Nature Methods 3(9): 670-676, 2006), i-score (Nucleic Acids Res 35: e123, 2007), i-Score Designer tool and associated algorithms (Nucleic Acids Res 32: 936-948, 2004. Biochem Biophys Res Commun 316: 1050-1058, 2004, Nucleic Acids Res 32: 893-901, 2004, Cell Cycle 3: 790-5, 2004, Nat Biotechnol 23: 995-1001, 2005, Nucleic Acids Res 35: e27, 2007, BMC Bioinformatics 7: 520, 2006, Nucleic Acids Res 35: e123, 2007, Nat Biotechnol 22: 326-330, 2004) are known in the art and may be used in selecting polynucleotide sequences effective in gene silencing. In some embodiments random design or empirical selection of polynucleotide sequences is used in selecting polynucleotide sequences effective in gene silencing. In some embodiments the sequence of a polynucleotide is screened against the genomic DNA of the intended plant to minimize unintentional silencing of other genes.

**[0040]** The polynucleotide compositions are useful in compositions, such as solutions of polynucleotide molecules, at low concentrations, alone or in combination with other components either in the same solution or in separately applied solutions that provide a permeability-enhancing agent. While there is no upper limit on the concentrations and dosages of polynucleotide molecules that can useful in the methods, lower effective concentrations and dosages will generally be sought for efficiency. The concentrations can be adjusted in consideration of the volume of spray or treatment applied to plant leaves or other plant part surfaces, such as flower petals, stems, tubers, fruit, anthers, pollen, or seed. In one embodiment, a useful treatment for herbaceous plants using 25-mer oligonucleotide molecules is about 1 nanomole (nmol) of oligonucleotide molecules per plant, for example, from about 0.05 to 1 nmol per plant. Other embodiments for herbaceous plants include useful ranges of about 0.05 to about 100 nmol, or about 0.1 to about 20 nmol, or about 1

nmol to about 10 nmol of polynucleotides per plant. Very large plants, trees, or vines may require correspondingly larger amounts of polynucleotides. When using long dsRNA molecules that can be processed into multiple oligonucleotides, lower concentrations can be used. To illustrate embodiments, the factor 1X, when applied to oligonucleotide molecules is arbitrarily used to denote a treatment of 0.8 nmol of polynucleotide molecule per plant; 10X, 8 nmol of polynucleotide molecule per plant; and 100X, 80 nmol of polynucleotide molecule per plant.

**[0041]** The trigger polynucleotide and oligonucleotide molecule compositions are useful in compositions, such as liquids that comprise these polynucleotide molecules, alone or in combination with other components, for example one or more herbicide molecules, either in the same liquid or in separately applied liquids that also provide a transfer agent. As used herein, a transfer agent is an agent that, when combined with a polynucleotide in a composition that is topically applied to a target plant surface, enables the polynucleotide to enter a plant cell. In certain embodiments, a transfer agent is an agent that conditions the surface of plant tissue, e. g., leaves, stems, roots, flowers, or fruits, to permeation by the polynucleotide molecules into plant cells. The transfer of polynucleotides into plant cells can be facilitated by the prior or contemporaneous application of a polynucleotide-transferring agent to the plant tissue. In some embodiments the transferring agent is applied subsequent to the application of the polynucleotide composition. The polynucleotide transfer agent enables a pathway for polynucleotides through cuticle wax barriers, stomata and/or cell wall or membrane barriers into plant cells. Suitable transfer agents to facilitate transfer of the polynucleotide into a plant cell include agents that increase permeability of the exterior of the plant or that increase permeability of plant cells to oligonucleotides or polynucleotides. Such agents to facilitate transfer of the composition into a plant cell include a chemical agent, or a physical agent, or combinations thereof. Chemical agents for conditioning or transfer include (a) surfactants, (b) an organic solvent or an aqueous solution or aqueous mixtures of organic solvents, (c) oxidizing agents, (d) acids, (e) bases, (f) oils, (g) enzymes, or combinations thereof. Embodiments of the method can optionally include an incubation step, a neutralization step (e.g., to neutralize an acid, base, or oxidizing agent, or to inactivate an enzyme), a rinsing step, or combinations thereof. Embodiments of agents or treatments for conditioning of a plant to permeation by polynucleotides include emulsions, reverse emulsions, liposomes, and other micellar-like compositions. Embodiments of agents or treatments for conditioning of a plant to permeation by polynucleotides include counter-ions or other molecules that are known to associate with nucleic acid molecules, e. g., inorganic ammonium ions, alkyl ammonium ions, lithium ions, polyamines such as spermine, spermidine, or putrescine, and other cations. Organic solvents useful in conditioning a plant to permeation by polynucleotides include DMSO, DMF, pyridine, N-pyrrolidine, hexamethylphosphoramide, acetonitrile, dioxane, polypropylene glycol, other solvents miscible with water or that will dissolve phosphonucleotides in non-aqueous systems (such as is used in synthetic reactions). Naturally derived or synthetic oils with or without surfactants or emulsifiers can be used, e. g., plant-sourced oils, crop oils (such as those listed in the 9<sup>th</sup> Compendium of Herbicide Adjuvants, publicly available on the worldwide web (internet) at [herbicide.adjuvants.com](http://herbicide.adjuvants.com) can be used, e. g., paraffinic oils, polyol fatty acid esters, or oils with short-chain molecules modified with amides or polyamines such as polyethylene-neimine or N-pyrrolidine. Transfer agents include, but are not limited to, organosilicone preparations.

**[0042]** Ligands can be tethered to a polynucleotide, for example a dsRNA, ssRNA, dsDNA or ssDNA. Ligands in general can include modifiers, e.g., for enhancing uptake; diagnostic compounds or reporter groups e.g., for monitoring distribution; cross-linking agents; nuclease-resistance conferring moieties; and natural or unusual nucleobases. General examples include lipophiles, lipids (e.g., cholesterol, a bile acid, or a fatty acid (e.g., lithocholic-oleyl, lauroyl, docosanyl, stearoyl, palmitoyl, myristoyl oleoyl, linoleoyl), steroids (e.g., uvaol, hecigenin, diosgenin), terpenes (e.g., triterpenes, e.g., sarsasapogenin, Friedelin, epifriedelanol derivatized lithocholic acid), vitamins (e.g., folic acid, vitamin A, biotin, pyridoxal), carbohydrates, proteins, protein binding agents, integrin targeting molecules, polycationics, peptides, polyamines, and peptide mimics. The ligand may also be a recombinant or synthetic molecule, such as a synthetic polymer, e.g., polyethylene glycol (PEG), PEG-40K, PEG-20K and PEG-5K. Other examples of ligands include lipophilic molecules, e.g., cholesterol, cholic acid, adamantine acetic acid, 1-pyrene butyric acid, dihydrotestosterone, glycerol (e.g., esters and ethers thereof, e.g., C.sub.10, C.sub.11, C.sub.12, C.sub.13, C.sub.14, C.sub.15, C.sub.16, C.sub.17, C.sub.18, C.sub.19, or C.sub.20 alkyl; e.g., lauroyl, docosanyl, stearoyl, oleoyl, linoleoyl 1,3-bis-O(hexadecyl)glycerol, 1,3-bis-O(octaadecyl)glycerol), geranyloxyhexyl group, hexadecylglycerol, borneol, menthol, 1,3-propanediol, heptadecyl group, palmitic acid, myristic acid, O3-(oleoyl)lithocholic acid, O3-(oleoyl)cholenic acid, dodecanoyle, lithocholyl, 5-beta-cholanyl, N,N-distearyl-lithocholamide, 1,2-di-O-stearoylglyceride, dimethoxytrityl, or phenoxazine) and PEG (e.g., PEG-5K, PEG-20K, PEG-40K). Preferred lipophilic moieties include lipid, sterols, oleyl, retinyl, or cholesteryl residues.

**[0043]** Conjugating a ligand to a dsRNA can enhance its cellular absorption, lipophilic compounds that have been conjugated to oligonucleotides include 1-pyrene butyric acid, 1,3-bis-O-(hexadecyl)glycerol, and menthol. One example of a ligand for receptor-mediated endocytosis is folic acid. Folic acid enters the cell by folate-receptor-radiated endocytosis. dsRNA compounds bearing folic acid would be efficiently transported into the cell via the folate-receptor-mediated endocytosis. Other ligands that have been conjugated to oligonucleotides include polyethylene glycols, carbohydrate clusters, cross-linking agents, porphyrin conjugates, delivery peptides and lipids such as cholesterol. In certain instances, conjugation of a cationic ligand to oligonucleotides results in improved resistance to nucleases. Representative examples

of cationic ligands are propylammonium and dimethylpropylammonium. Interestingly, antisense oligonucleotides were reported to retain their high binding affinity to mRNA when the cationic ligand was dispersed, throughout the oligonucleotide. See M. Manoharan Antisense & Nucleic Acid Drug Development 2002, 12, 103 and references therein.

**[0044]** A biologic delivery can be accomplished by a variety of methods including, without limitation, (1) loading liposomes with a dsRNA acid molecule provided herein and (2) complexing a dsRNA molecule with lipids or liposomes to form nucleic acid-lipid or nucleic acid-liposome complexes. The liposome can be composed of cationic and neutral lipids commonly used to transfect cells in vitro. Cationic lipids can complex (e.g., charge-associate) with negatively charged, nucleic acids to form liposomes. Examples of cationic liposomes include, without limitation, lipofectin, lipofectamine, lipofectace, and DOTAP. Procedures for forming liposomes are well known in the art. Liposome compositions can be formed, for example, from phosphatidylcholine, dimyristoyl phosphatidylcholine, dipalmitoyl phosphatidylcholine, dimyristoyl phosphatidyl glycerol, dioleoyl phosphatidylethanolamine or liposomes comprising dihydrophosphingomyelin (DHSM). Numerous lipophilic agents are commercially available, including Lipofectin.RTM. (Invitrogen/Life Technologies, Carlsbad, Calif.) and Effectene™ (Qiagen, Valencia, Calif.). In addition, systemic delivery methods can be optimized using commercially available cationic lipids such as DDAB or DOTAP, each of which can be mixed with a neutral lipid such as DOPE or cholesterol. In some cases, liposomes such as those described by Templeton et al. Nature Biotechnology, 15:647-652 (1997) can be used. In other embodiments, polycations such as polyethyleneimine can be used to achieve delivery in vivo and ex vivo (Boletta et al., J. Am Soc. Nephrol. 7:1728, 1996). Additional information regarding the use of liposomes to deliver nucleic acids can be found in U.S. Pat. No. 6,271,359, PCT Publication WO 96/40964 and Morrissey, D. et al., 2005. Nature Biotechnol. 23(8):1002-7.

**[0045]** In certain embodiments, an organosilicone preparation that is commercially available as Silwet® L-77 surfactant having CAS Number 27306-78-1 and EPA Number: CAL.REG.NO. 5905-50073-AA, and currently available from Momentive Performance Materials, Albany, New York can be used to prepare a polynucleotide composition. In certain embodiments where a Silwet L-77 organosilicone preparation is used as a pre-spray treatment of plant leaves or other plant surfaces, freshly made concentrations in the range of about 0.015 to about 2 percent by weight (wt percent) (e.g., about 0.01, 0.015, 0.02, 0.025, 0.03, 0.035, 0.04, 0.045, 0.05, 0.055, 0.06, 0.065, 0.07, 0.075, 0.08, 0.085, 0.09, 0.095, 0.1, 0.2, 0.3, 0.4, 0.5, 0.6, 0.7, 0.8, 0.9, 1.0, 1.1, 1.2, 1.3, 1.4, 1.5, 1.6, 1.7, 1.8, 1.9, 2.0, 2.1, 2.2, 2.3, 2.5 wt percent) are efficacious in preparing a leaf or other plant surface for transfer of polynucleotide molecules into plant cells from a topical application on the surface. In certain embodiments of the methods and compositions provided herein, a composition that comprises a polynucleotide molecule and an organosilicone preparation comprising Silwet L-77 in the range of about 0.015 to about 2 percent by weight (wt percent) (e.g., about 0.01, 0.015, 0.02, 0.025, 0.03, 0.035, 0.04, 0.045, 0.05, 0.055, 0.06, 0.065, 0.07, 0.075, 0.08, 0.085, 0.09, 0.095, 0.1, 0.2, 0.3, 0.4, 0.5, 0.6, 0.7, 0.8, 0.9, 1.0, 1.1, 1.2, 1.3, 1.4, 1.5, 1.6, 1.7, 1.8, 1.9, 2.0, 2.1, 2.2, 2.3, 2.5 wt percent) is used or provided.

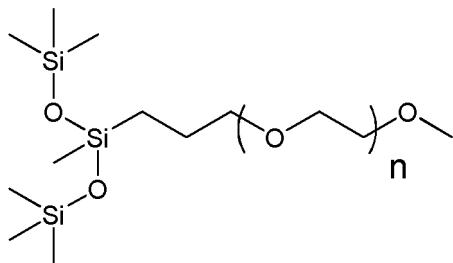
**[0046]** In certain embodiments, any of the commercially available organosilicone preparations provided such as the following Breakthru S 321, Breakthru S 200 Cat# 67674-67-3, Breakthru OE 441 Cat#68937-55-3, Breakthru S 278 Cat #27306-78-1, Breakthru S 243, Breakthru S 233 Cat#134180-76-0, available from manufacturer Evonik Goldschmidt (Germany), Silwet® HS 429, Silwet® HS 312, Silwet® HS 508, Silwet® HS 604 (Momentive Performance Materials, Albany, New York) can be used as transfer agents in a polynucleotide composition. In certain embodiments where an organosilicone preparation is used as a pre-spray treatment of plant leaves or other surfaces, freshly made concentrations in the range of about 0.015 to about 2 percent by weight (wt percent) (e.g., about 0.01, 0.015, 0.02, 0.025, 0.03, 0.035, 0.04, 0.045, 0.05, 0.055, 0.06, 0.065, 0.07, 0.075, 0.08, 0.085, 0.09, 0.095, 0.1, 0.2, 0.3, 0.4, 0.5, 0.6, 0.7, 0.8, 0.9, 1.0, 1.1, 1.2, 1.3, 1.4, 1.5, 1.6, 1.7, 1.8, 1.9, 2.0, 2.1, 2.2, 2.3, 2.5 wt percent) are efficacious in preparing a leaf or other plant surface for transfer of polynucleotide molecules into plant cells from a topical application on the surface. In certain embodiments of the methods and compositions provided herein, a composition that comprises a polynucleotide molecule and an organosilicone preparation in the range of about 0.015 to about 2 percent by weight (wt percent) (e.g., about 0.01, 0.015, 0.02, 0.025, 0.03, 0.035, 0.04, 0.045, 0.05, 0.055, 0.06, 0.065, 0.07, 0.075, 0.08, 0.085, 0.09, 0.095, 0.1, 0.2, 0.3, 0.4, 0.5, 0.6, 0.7, 0.8, 0.9, 1.0, 1.1, 1.2, 1.3, 1.4, 1.5, 1.6, 1.7, 1.8, 1.9, 2.0, 2.1, 2.2, 2.3, 2.5 wt percent) is used or provided.

**[0047]** Organosilicone preparations used in the methods and compositions provided herein can comprise one or more effective organosilicone compounds. As used herein, the phrase "effective organosilicone compound" is used to describe any organosilicone compound that is found in an organosilicone preparation that enables a polynucleotide to enter a plant cell. In certain embodiments, an effective organosilicone compound can enable a polynucleotide to enter a plant cell in a manner permitting a polynucleotide mediated suppression of a target gene expression in the plant cell. In general, effective organosilicone compounds include, but are not limited to, compounds that can comprise: i) a trisiloxane head group that is covalently linked to, ii) an alkyl linker including, but not limited to, an n-propyl linker, that is covalently linked to, iii) a poly glycol chain, that is covalently linked to, iv) a terminal group. Trisiloxane head groups of such effective organosilicone compounds include, but are not limited to, heptamethyltrisiloxane. Alkyl linkers can include, but are not limited to, an n-propyl linker. Poly glycol chains include, but are not limited to, polyethylene glycol or polypropylene glycol. Poly glycol chains can comprise a mixture that provides an average chain length "n" of about "7.5". In certain embodiments,

the average chain length "n" can vary from about 5 to about 14. Terminal groups can include, but are not limited to, alkyl groups such as a methyl group. Effective organosilicone compounds are believed to include, but are not limited to, trisiloxane ethoxylate surfactants or polyalkylene oxide modified heptamethyl trisiloxane.

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10



15 (Compound I: polyalkyleneoxide heptamethyltrisiloxane, average n=7.5).

[0048] In certain embodiments, an organosilicone preparation that comprises an organosilicone compound comprising a trisiloxane head group is used in the methods and compositions provided herein. In certain embodiments, an organosilicone preparation that comprises an organosilicone compound comprising a heptamethyltrisiloxane head group is used in the methods and compositions provided herein. In certain embodiments, an organosilicone composition that comprises Compound I is used in the methods and compositions provided herein. In certain embodiments, an organosilicone composition that comprises Compound I is used in the methods and compositions provided herein. In certain embodiments of the methods and compositions provided herein, a composition that comprises a polynucleotide molecule and one or more effective organosilicone compound in the range of about 0.015 to about 2 percent by weight (wt percent) (e. g., about 0.01, 0.015, 0.02, 0.025, 0.03, 0.035, 0.04, 0.045, 0.05, 0.055, 0.06, 0.065, 0.07, 0.075, 0.08, 0.085, 0.09, 0.095, 0.1, 0.2, 0.3, 0.4, 0.5, 0.6, 0.7, 0.8, 0.9, 1.0, 1.1, 1.2, 1.3, 1.4, 1.5, 1.6, 1.7, 1.8, 1.9, 2.0, 2.1, 2.2, 2.3, 2.5 wt percent) is used or provided.

[0049] Compositions include but are not limited components that are one or more polynucleotides essentially identical to, or essentially complementary to a DHPS gene sequence (promoter, intron, exon, 5' untranslated region, 3' untranslated region), a transfer agent that provides for the polynucleotide to enter a plant cell, a herbicide that complements the action of the polynucleotide, one or more additional herbicides that further enhance the herbicide activity of the composition or provide an additional mode of action different from the complementing herbicide, various salts and stabilizing agents that enhance the utility of the composition as an admixture of the components of the composition.

[0050] The methods include one or more applications of a polynucleotide composition and one or more applications of a permeability-enhancing agent for conditioning of a plant to permeation by polynucleotides. When the agent for conditioning to permeation is an organosilicone composition or compound contained therein, embodiments of the polynucleotide molecules are double-stranded RNA oligonucleotides, single-stranded RNA oligonucleotides, double-stranded RNA polynucleotides, single-stranded RNA polynucleotides, double-stranded DNA oligonucleotides, single-stranded DNA oligonucleotides, double-stranded DNA polynucleotides, single-stranded DNA polynucleotides, chemically modified RNA or DNA oligonucleotides or polynucleotides or mixtures thereof.

[0051] Compositions and methods are useful for modulating the expression of an endogenous DHPS gene or transgenic DHPS gene (for example US Patent No. 6,121,513) in a plant cell. In various embodiments, a DHPS gene includes coding (protein-coding or translatable) sequence, non-coding (non-translatable) sequence, or both coding and non-coding sequence. Compositions can include polynucleotides and oligonucleotides designed to target multiple genes, or multiple segments of one or more genes. The target gene can include multiple consecutive segments of a target gene, multiple non-consecutive segments of a target gene, multiple alleles of a target gene, or multiple target genes from one or more species.

[0052] An aspect provides a method for modulating expression of a DHPS gene in a plant including (a) conditioning of a plant to permeation by polynucleotides and (b) treatment of the plant with the polynucleotide molecules, wherein the polynucleotide molecules include at least one segment of 18 or more contiguous nucleotides cloned from or otherwise identified from the target DHPS gene in either anti-sense or sense orientation, whereby the polynucleotide molecules permeate the interior of the plant and induce modulation of the target gene. The conditioning and polynucleotide application can be performed separately or in a single step. When the conditioning and polynucleotide application are performed in separate steps, the conditioning can precede or can follow the polynucleotide application within minutes, hours, or days. In some embodiments more than one conditioning step or more than one polynucleotide molecule application can be performed on the same plant. In embodiments of the method, the segment can be cloned or identified from (a) coding (protein-encoding), (b) non-coding (promoter and other gene related molecules), or (c) both coding and non-coding parts of the target gene. Non-coding parts include DNA, such as promoter regions or the RNA transcribed by the DNA that provide RNA regulatory molecules, including but not limited to: introns, 5' or 3' untranslated regions,

and microRNAs (miRNA), *trans*-acting siRNAs, natural anti-sense siRNAs, and other small RNAs with regulatory function or RNAs having structural or enzymatic function including but not limited to: ribozymes, ribosomal RNAs, t-RNAs, aptamers, and riboswitches.

[0053] The following examples are included to demonstrate examples of certain preferred embodiments.

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## EXAMPLES

**Example 1.** Polynucleotides related to the DHPS gene sequences.

[0054] The target DHPS polynucleotide molecule naturally occurs in the genome of plants that include but are not limited to *Amaranthus palmeri*, *Amaranthus rudis*, *Amaranthus hybrids*, *Amaranthus lividus*, *Amaranthus viridis*, *Ambrosia trifida*, *Conyza canadensis*, *Digitaria sanguinalis*, *Euphorbia heterophylla*, *Kochia scoparia*, *Lolium multiflorum*, and include molecules related to the expression of a polypeptide identified as a DHPS, that include genomic DNA (gDNA) and coding cDNAs comprising coding and noncoding regions of a DHPS gene and fragments thereof as shown in Table 1.

[0055] Polynucleotide molecules were extracted from these plant species by methods standard in the field, for example, total RNA was extracted using Trizol Reagent (Invitrogen Corp, Carlsbad, CA Cat. No. 15596-018), following the manufacturer's protocol or modifications thereof by those skilled in the art of polynucleotide extraction that may enhance recover or purity of the extracted RNA. Briefly, start with 1 gram of ground plant tissue for extraction. Prealiquot 10 milliliters (mL) Trizol reagent to 15 mL conical tubes, add ground powder to tubes and shake to homogenize. Incubate the homogenized samples for 5 minutes (min) at room temperature (RT) and then add 3 mL of chloroform. Shakes tubes vigorously by hand for 15-30 seconds (sec) and incubate at RT for 3 min. Centrifuge the tubes at 7,000 revolutions per minute (rpm) for 10 min at 4 degrees C. Transfer the aqueous phase to a new 1.5 mL tube and add 1 volume of cold isopropanol. Incubate the samples for 20-30 min at RT and centrifuge at 10,000 rpm for 10 min at 4 degrees C. Wash pellet with Sigma-grade 80 percent ethanol. Remove the supernatant and briefly air-dry the pellet. Dissolve the RNA pellet in approximately 200 microliters of DEPC treated water. Heat briefly at 65C to dissolve pellet and vortex or pipet to resuspend RNA pellet and then adjust RNA concentration to 1-2 microgram/microliter.

[0056] DNA was extracted using EZNA SP Plant DNA Mini kit (Omega Bioteck, Norcross GA, Cat#D5511) and Lysing Matrix E tubes (Q-Biogen, Cat#6914), following the manufacturer's protocol or modifications thereof by those skilled in the art of polynucleotide extraction that may enhance recover or purity of the extracted DNA. Briefly, aliquot ground tissue to a Lysing Matrix E tube on dry ice, add 800 $\mu$ l Buffer SP1 to each sample, homogenize in a bead beater for 35-45sec, incubate on ice for 45-60 sec, centrifuge at  $\geq$ 14000 rpm for 1min at RT, add 10 microliter RNase A to the lysate, incubate at 65°C for 10min, centrifuge for 1min at RT, add 280 $\mu$ l Buffer SP2 and vortex to mix, incubate the samples on ice for 5min, centrifuge at  $\geq$ 10,000g for 10 min at RT, transfer the supernatant to a homogenizer column in a 2ml collection tube, centrifuge at 10,000g for 2 min at RT, transfer the cleared lysate into a 1.5ml microfuge tube, add 1.5 volumes Buffer SP3 to the cleared lysate, vortex immediately to obtain a homogeneous mixture, transfer up to 650 $\mu$ l supernatant to the Hi-Bind column, centrifuge at 10,000g for 1min, repeat, apply 100 $\mu$ l 65°C Elution Buffer to the column, centrifuge at 10,000g for 5min at RT.

[0057] Next-generation DNA sequencers, such as the 454-FLX (Roche, Branford, CT), the SOLiD (Applied Biosystems,), and the Genome Analyzer (HiSeq2000, Illumina, San Diego, CA) were used to provide polynucleotide sequence from the DNA and RNA extracted from the plant tissues. Raw sequence data was assembled into contigs. The contig sequence was used to identify trigger molecules that can be applied to a plant to enable regulation of the gene expression.

[0058] The target DNA sequence isolated from genomic (gDNA) and coding DNA (cDNA) from the various weedy plant species for the DHPS gene and the assembled contigs as set forth in SEQ ID NOs 1-54 and Table 1.

**Example 2.** Polynucleotides related to the trigger molecules

[0059] The gene sequences and fragments of Table 1 were divided into 200 polynucleotide (200-mer) lengths with 25 polynucleotide overlapping regions and are shown in Table 2, SEQ ID NO:55-906. These polynucleotides are tested to select the most efficacious trigger regions across the length of any target sequence. The trigger polynucleotides are constructed as sense or anti-sense ssDNA or ssRNA, dsRNA, or dsDNA, or dsDNA/RNA hybrids and combined with an organosilicone based transfer agent to provide a polynucleotide preparation. The polynucleotides are combined into sets of two to three polynucleotides per set, using 4-8 nmol of each polynucleotide. Each polynucleotide set is prepared with the transfer agent and applied to a plant or a field of plants in combination with a DHPS inhibitor containing herbicide, or followed by a DHPS inhibitor treatment one to three days after the polynucleotide application, to determine the effect on the plant's susceptibility to an DHPS inhibitor. The effect is measured as stunting the growth and/or killing of the plant and is measured 8-14 days after treatment with the polynucleotide set and DHPS inhibitor. The most efficacious sets are identified and the individual polynucleotides are tested in the same methods as the sets are and the most efficacious single 200-mer identified. The 200-mer sequence is divided into smaller sequences of 50-70-mer regions with 10-15

polynucleotide overlapping regions and the polynucleotides tested individually. The most efficacious 50-70-mer is further divided into smaller sequences of 25-mer regions with a 12 to 13 polynucleotide overlapping region and tested for efficacy in combination with DHPS inhibitor treatment. By this method it is possible to identify an oligonucleotide or several oligonucleotides that are the most efficacious trigger molecule to effect plant sensitivity to a DHPS inhibitor or modulation of an DHPS gene expression. The modulation of DHPS gene expression is determined by the detection of DHPS siRNA molecules specific to a DHPS gene or by an observation of a reduction in the amount of DHPS RNA transcript produced relative to an untreated plant or by merely observing the anticipated phenotype of the application of the trigger with the DHPS inhibitor containing herbicide. Detection of siRNA can be accomplished, for example, using kits such as mirVana (Ambion, Austin TX) and mirPremier (Sigma-Aldrich, St Louis, MO).

[0060] The target DNA sequence isolated from genomic (gDNA) and coding DNA (cDNA) from the various weedy plant species for the DHPS gene and the assembled contigs as set forth in SEQ ID NOs 1-54 were divided into polynucleotide fragments as shown in Table 2 and as set forth in SEQ ID NOs 55-906.

[0061] The gene sequences and fragments of Table 1 were compared and 21-mers of contiguous polynucleotides were identified that had homology across the various DHPS gene sequences. The purpose is to identify trigger molecules that are useful as herbicidal molecules or in combination with a DHPS inhibitor herbicide across a broad range of weed species. The sequences shown in Table 3 represent the 21-mers that were present in the DHPS gene of at least eight of the weed species of Table 1. It is contemplated that additional 21-mers can be selected from the sequences of Table 1 that are specific for a single weed species or a few weeds species within a genus or trigger molecules that are at least 18 contiguous nucleotides, at least 19 contiguous nucleotides, at least 20 contiguous nucleotides or at least 21 contiguous nucleotides in length and at least 85 percent identical to an DHPS gene sequence selected from the group consisting of SEQ ID NOs:1-54 or fragment thereof.

[0062] By this method it is possible to identify an oligonucleotide or several oligonucleotides that are the most efficacious trigger molecule to effect plant sensitivity to DHPS inhibitor or modulation of DHPS gene expression. The modulation of DHPS gene expression is determined by the detection of DHPS siRNA molecules specific to DHPS gene or by an observation of a reduction in the amount of DHPS RNA transcript produced relative to an untreated plant. Detection of siRNA can be accomplished, for example, using kits such as mirVana (Ambion, Austin TX) and mirPremier (Sigma-Aldrich, St Louis, MO).

[0063] The target DNA sequence isolated from genomic (gDNA) and coding DNA (cDNA) from the various weedy plant species for the DHPS gene and the assembled contigs as set forth in SEQ ID NOs 1-54 were divided into fragments as shown in Table 3 and as set forth in SEQ ID NOs 907-1175.

**Example 3.** Methods related to treating plants or plant parts with a topical mixture of the trigger molecules.

[0064] Glyphosate-sensitive Palmer amaranth (*A. palmeri* R-22) plants were grown in the greenhouse (30 / 20 C day/night T; 14 hour photoperiod) in 10.16 cm (4 inch) square pots containing Sun Crro® Redi-Earth and 3.5 kg/cubic meter Osmocote® 14-14-14 fertilizer. Palmer amaranth plants at 5 to 10 cm in height were pre-treated with a mixture of short (25mer) dsDNA trigger oligonucleotides targeting DHPS coding or noncoding regions using 4 nmol of each oligonucleotide and pooling 5-6 oligonucleotides in each treatment, formulated in 20 millimolar sodium phosphate buffer (pH 6.8) containing 2% ammonium sulfate and 1% Silwet L-77. Plants were treated manually by pipetting 10 µL of polynucleotide solution on four fully expanded mature leaves, for a total of 40 microliters of solution per plant. There were eight treatment pools, DHPS1-6, DHPS7-12, DHPS13-18, DHPS19-24, DHPS25-30, DHPS31-36, DHPS37-42, and DHPS43-47 (Table 4). Twenty-four hours later, the plants were treated with pendimethalin (Prowl®, BASF, this herbicide functions as a mitosis inhibitor similar to asulam a known DHPS inhibitor, other dinitroaniline herbicides that function to inhibit mitosis include but are not limited to Sonalan® (ethalfluralin), Squadron1®, Steel1®, Treflan®/Tri-lin®/Tri-4) at a rate of 13.45 kg/ha (12 lb/ac). Four replications of each treatment was conducted. Plant height was determined just before polynucleotide treatment and at intervals upto fourteen days after herbicide treatments to determine effect of the oligonucleotide and herbicide treatments. The results were expressed as percent reduction in height relative to the untreated control (no formulation and no trigger molecules), another treatment was the formulation control which is herbicide plus buffer plus ammonium plus Silwet. Figure 1 illustrates the results of this test. Three of the pooled oligonucleotides demonstrated an enhancement of the herbicide activity, these are DHPS1-6, DHPS7-12, and DHPS13-18. Further testing of single oligonucleotides from DHPS1-6 and DHPS7-12 demonstrated that DHPS 1 (SEQ ID NO:1176) and DHPS11(SEQ ID NO:1186) had the highest activity among the oligonucleotides in those respective pools with these 2 oligonucleotides providing greater than 15 percent increase in herbicide injury.

**Table 4.** DHPS dsDNA oligonucleotides.

trigger name	SEQ ID NO:	Sense sequence	Antisense sequence
DHPS1	1176	TTTATTCTAAAGTTGCttcGGAGG	CCTCCGAAGCAACTTACAATAAAA

(continued)

trigger name	SEQ ID NO:	Sense sequence	Antisense sequence
DHPS2	1177	CCGTCAGAAGGGGGCACACATTGT	ACAATGTGTGCCCTCTTGACGG
DHPS3	1178	GAATGATGTCTCtaGTGGgAAACTC	GAGTTTCCCCTAGAGACATCATT
DHPS4	1179	GATTCCGAGATGTTAATGTTGTTG	CAACAAACATTAAACATCTCGGAATC
DHPS5	1180	CGGACCTTAAAGTTCTTATAGC	GCTATATAAGGAACCTTAAGGTCG
DHPS6	1181	AATGCACATGCGAGGAGATCCGACT	AGTCGGATCTCCTCGCATGTGCATT
DHPS7	1182	TCAATGCAAAACTCTGAGAACATTGA	TCAAGTTCTCAGAGTTTGATTGA
DHPS8	1183	CCTACAATGATGTTGTAAGCAAGT	ACTTGCTTACAAACATCATTGAGG
DHPS9	1184	GGCTTCGGAGTTGAGTTAGGGTc	GACCCTAGAACTCAACTCCGAAGCC
DHPS10	1185	ATAGATGCAGAATTATCGGGAATTC	GAATTCCCGATAATTCTGCATCTAT
DHPS11	1186	CTGCTTGGAGGATAGTTATTGATCC	GGATCAATAACTATCCTCCAAGCAG
DHPS12	1187	CGGCATCGGATTTCTAAGAACATCG	CGTATTCTAGAAAATCCGATGCCG
DHPS13	1188	AATCAAATTTGGAAATTCTTAGTG	CACTAAGAATTCCAAATTGATTG
DHPS14	1189	GTTTACAAAAGATACTGGGAAGAGAT	ATCTCTTCCGTATTTGAAAC
DHPS15	1190	AGCTAAGAAGAGTTGGCGGTGGCT	AGCCACGCCAAACTCTTCTTAGCT
DHPS16	1191	CATTGCCCTTGCTAATTGGACCTT	AAGGTCCAATTAGCAAGGGCAATG
DHPS17	1192	CAAGAAAGAGGTTCTGGCGAGAT	ATCTGCCAGAACCTCTTCTTG
DHPS18	1193	TTGTGAGGCCCTGTAGCAGCTGAC	GTCAGCTGCTACAGGGCGCTCACAA
DHPS19	1194	AGGGATCCTGCTACCATTGCTTCTA	TAGAAGCAATGGTAGCAGGATCCCT
DHPS20	1195	TAACTGCTGGAGTTTAGGTGGTGC	GCACCACCTAAACTCCAGCAGTTA
DHPS21	1196	AAACATTGTAAGAGTACATAATGTt	AACATTATGTAAGTACTTTACAATGTT
DHPS22	1197	AGGGATAACCTTGATGCTGTCAAGT	ACTTGACAGCATCAAGGTTATCCCT
DHPS23	1198	TATGTGATGCCATACTCGGAAAAAC	GTTCCTCCGAGTATGGCATCACATA
DHPS24	1199	TGATTAACTGCTTGTGTACCCACC	GGTGGTACAACAAAGCAGTTAACATCA
DHPS25	1200	TTGTGAATGATGTCTAGTGGAAAGGT	ACCTTCACTAGACATCATTACAA
DHPS26	1201	TCGATTGGGATTATGATGAAGTCCG	CGGACTTCATCATAATCCCAATCGA
DHPS27	1202	TTAGTTGCTCGAAAGtaGAGCTTT	AAAGCTCTACTTCGAGCAAACCTAA
DHPS28	1203	AGCCCCTGTAAGGctATAGTCTTACA	TGTAAGAACTATAGCCTTACGGGCT
DHPS29	1204	GATGTAGAATCATTGGCTATTGCC	GGCAAATAGCCAATGATTCTACATC
DHPS30	1205	TATTCTGCTTAGAAGATCATAGCA	TGCTATGATCTCTAAGCAGAAATA
DHPS31	1206	TTGATCCAGACTTGGATTCCATGT	ACATGGAATCCAAAGTCTGGATCAA
DHPS32	1207	AGAATTATGTTGCGACTCATGGTCG	CGACCATGAGTCGCAACATAATTCT
DHPS33	1208	AATTGGCCCTTGAGGAATTGTTGA	TCAACAACTCCTCAAAGGGCCAATT
DHPS34	1209	AGTTCCCTGAGTCTATGGTCATC	GATTGAACCATAAGACTCAAGGAACT
DHPS35	1210	GGCTCGGTTGATGAAGGTTGGACT	AGTCCAACCTTCATCGAACCGAGCC
DHPS36	1211	GTGTCGAGACAAGATGGGCAAAGA	TCTTGACCCATCTGTCTCGACAC
DHPS37	1212	GACCAATGGACAAGAAGTCGACCTT	AAGGTCGACTTCTGTCCATTGGTC
DHPS38	1213	GGGCAAGCTTGATCgaGTTACTTGG	CCAAGTAACTCGATCAAGCTGCC
DHPS39	1214	ACAAcGCAGATAACAGCAGGCTCTGG	CCAGAGCCTGCTGTTATCTGCTTGT

(continued)

trigger name	SEQ ID NO:	Sense sequence	Antisense sequence
DHPS40	1215	CGACcATTAGACATACATATCATTt	AAATGATATGTATGTCTAATGGTCG
DHPS41	1216	GTTTTTtGAAGTCAGATCTGCAATC	GATTGCAGATCTGACTTCAAAAAAC
DHPS42	1217	AATCAGTTGCAATGGACAAaCCATA	TATGGTTGTCCATTGCAACTGATT
DHPS43	1218	TACGGTTaTTagTTGTTCCCTGTCAC	GTGACAGGAACAACATAATAACCGTA
DHPS44	1219	TgTTTGAGTTGAATTAGATCATGCA	TGCATGATCTAATTCAACTCAAACA
DHPS45	1220	AACATTGTAAGAACATTGATCATGT	ACATGATCAAGATTCTTACAATGTT
DHPS46	1221	GAAGCTATGTGATACTGTACTTCTC	GAGAAGTACAGTATCACATAGCTTC
DHPS47	1222	AAAATGAAATCTTGATATGATGTT	AACATCATATCAAAGATTCATTTT

**Example 4.** A method to control weeds in a field.

[0065] A method to control weeds in a field comprises the use of trigger polynucleotides that can modulate the expression of a DHPS gene in one or more target weed plant species. In Table 3, an analysis of DHPS gene sequences from multiple plant species provided a collection of 21-mer polynucleotides that were common to at least 4 of the species and can be used in compositions to affect the growth or develop or sensitivity to DHPS inhibitor herbicide to control multiple weed species in a field. Other oligonucleotide segments can be selected from the disclosed gene sequences that are more specific to a particular weed species, for example, an oligonucleotide that has a sequence that is homologous or complementary to a DHPS gene of three weeds species, or of two weed species, or of only one weed species. A composition containing 1 or 2 or 3 or 4 or more of the polynucleotides of Table 3 would enable broad activity of the composition against the multiple weed species that occur in a field environment.

[0066] The method includes creating a composition that comprises components that include at least one polynucleotide of Table 3 (SEQ ID NO:907-1175) or any other effective gene expression modulating polynucleotide essentially identical or essentially complementary to SEQ ID NO:1-54 or fragment thereof, a transfer agent that mobilizes the polynucleotide into a plant cell and a DHPS inhibiting herbicide and optionally a polynucleotide that modulates the expression of an essential gene and optionally a co-herbicide that has a different mode of action relative to a DHPS inhibitor. The polynucleotide of the composition includes a dsRNA, ssDNA or dsDNA or a combination thereof. A composition containing a polynucleotide can have a use rate of about 1 to 30 grams or more per acre depending on the size of the polynucleotide and the number of polynucleotides in the composition. The composition may include one or more additional co-herbicides as needed to provide effective multi-species weed control. Crop safety can be enhanced by reducing the amount of effective herbicide needed to control weeds in the field. A field of crop plants or a turf grass environment in need of weedy plant control is treated by spray application of the composition. The composition can be provided as a tank mix, a sequential treatment of components (generally the polynucleotide followed by the herbicide), a simultaneous treatment or mixing of one or more of the components of the composition from separate containers. Treatment of the field can occur as often as needed to provide weed control and the components of the composition can be adjusted to target specific weed species or weed families.

**Example 5.** Herbicidal Compositions comprising pesticidal agents

[0067] A method of controlling weeds and plant pest and pathogens in a field of DHPS inhibitor tolerant crop plants is provided, wherein the method comprises applying a composition comprising a DHPS trigger oligonucleotide, a DHPS inhibitor composition and an admixture of a pest control agent. For example, the admixture comprises insecticides, fungicides, nematocides, bactericides, acaricides, growth regulators, chemosterilants, semiochemicals, repellents, attractants, pheromones, feeding stimulants or other biologically active compounds or biological agents, such as, microorganisms.

[0068] For example, the admixture comprises a fungicide compound for use on a DHPS inhibitor tolerant crop plant to prevent or control plant disease caused by a plant fungal pathogen. The fungicide compound of the admixture may be a systemic or contact fungicide or mixtures of each. More particularly the fungicide compound includes, but is not limited to members of the chemical groups strobilurins, triazoles, chloronitriles, carboxamides and mixtures thereof. The composition may additionally have an admixture comprising an insecticidal compound or agent.

[0069] The DHPS trigger oligonucleotides and DHPS inhibitor or mitosis inhibitor herbicide (for example, asulam, or other dinitroaniline herbicides) tank mixes with fungicides, insecticides or both are tested for use in soybean and corn

for control of foliar diseases and pests. Testing is conducted to develop a method for use of mixtures of the trigger oligonucleotides and asulam formulation and various commercially available fungicides for weed control and pest control. The field plots are planted with soybeans or corn. All plots receive a post plant application of the DHPS trigger + asulam about 3 weeks after planting. The mixtures of trigger + asulam or trigger + asulam + fungicide + insecticides are used to treat the plots at the R1 stage of soybean development (first flowering) or tassel stage of corn. Data is taken for percent weed control at 7 and 21 days after R1 treatment, soybean safety (% necrosis, chlorosis, growth rate): 5 days after treatment, disease rating, pest ratings and yield (bushels/Acre). These mixtures and treatments are designed to provide simultaneous weed and pest control, such as fungal pest control, for example, leaf rust disease; and insect pest control, for example, aphids, armyworms, loopers, beetles, stinkbugs, and leaf hoppers.

**[0070]** Agricultural chemicals are provided in containers suitable for safe storage, transportation and distribution, stability of the chemical compositions, mixing with solvents and instructions for use. A container of a mixture of a trigger oligonucleotide + herbicide + fungicide compound, or a mixture of a trigger oligonucleotide + herbicide compound and an insecticide compound, or a trigger oligonucleotide + a herbicide compound and a fungicide compound and an insecticide compound (for example, lambda-cyhalothrin, Warrier®). The container may further provide instructions on the effective use of the mixture. Containers can be of any material that is suitable for the storage of the chemical mixture. Containers can be of any material that is suitable for the shipment of the chemical mixture. The material can be of cardboard, plastic, metal, or a composite of these materials. The container can have a volume of 0.5 liter, 1 liter, 2 liter, 3-5 liter, 5-10 liter, 10-20 liter, 20-50 liter or more depending upon the need. A tank mix of a trigger oligonucleotide + herbicide compound and a fungicide compound is provided, methods of application to the crop to achieve an effective dose of each compound are known to those skilled in the art and can be refined and further developed depending on the crop, weather conditions, and application equipment used.

**[0071]** Insecticides, fungicides, nematocides, bactericides, acaricides, growth regulators, chemosterilants, semiochemicals, repellents, attractants, pheromones, feeding stimulants or other biologically active compounds can be added to the trigger oligonucleotide to form a multi-component pesticide giving an even broader spectrum of agricultural protection. Examples of such agricultural protectants with which compounds of this invention can be formulated are: insecticides such as abamectin, acephate, azinphos-methyl, bifenthrin, buprofezin, carbofuran, chlорfenapyr, chlорpyrifos, chlорpyrifos-methyl, cyfluthrin, beta-cyfluthrin, cyhalothrin, lambda-cyhalothrin, deltamethrin, diafenthuron, diazinon, diflubenzuron, dimethoate, esfenvalerate, fenoxycarb, fenpropathrin, fenvalerate, fipronil, flucythrinate, tau-fluvalinate, fonophos, imidacloprid, isofenphos, malathion, metaldehyde, methamidophos, methidathion, methomyl, methoprene, methoxychlor, methyl 7-chloro-2,5-dihydro-2-[[N-(methoxycarbonyl)-N-[4-(trifluoromethoxy)phenyl ]amino]carbonyl]indeno[1,2-e][1,3,4]oxadiazine-4a(3H)-carboxylate (DPX-JW062), monocrotophos, oxamyl, parathion, parathion-methyl, permethrin, phorate, phosalone, phosmet, phosphamidon, pirimicarb, profenofos, rotenone, sulprofos, tebufenozone, tefluthrin, terbufos, tetrachlorvinphos, thiocarb, tralomethrin, trichlorfon and triflumuron; most preferably a DHPS inhibitor compound is formulated with a fungicide compound or combinations of fungicides, such as azoxystrobin, benomyl, blasticidin-S, Bordeaux mixture (tribasic copper sulfate), bromuconazole, captan, carbendazim, chloroneb, chlorothalonil, copper oxychloride, copper salts, cymoxanil, cyproconazole, cyprodinil (CGA 219417), diclomezine, dicloran, difenoconazole, dimethomorph, diniconazole, diniconazole-M, dodine, edifenphos, epoxiconazole (BAS 480F), famoxadone, fenarimol, fenbuconazole, fenpropiconazole, fenpropidin, fenpropimorph, fluazinam, fluquinconazole, flusilazole, flutolanil, flutriafol, folpet, fosetyl-aluminum, furaxyl, hexaconazole, ipconazole, iprobenfos, iprodione, isoprothiolane, kasugamycin, kresoxim-methyl, mancozeb, maneb, mepronil, metalaxyl, metconazole, S-methyl 7-benzothiazolecarbothioate (CGA 245704), myclobutanil, neo-azosin (ferric methanearsonate), oxadixyl, penconazole, pencycuron, probenazole, prochloraz, propiconazole, pyrifenox, pyroquilon, quinoxyfen, spiroxamine (KWG4168), sulfur, tebuconazole, tetaconazole, thiabendazole, thiophanate-methyl, thiram, triadimefon, triadimenol, tricyclazole, trifloxystrobin, triticonazole, validamycin and vinclozolin; combinations of fungicides are common for example, cyproconazole and azoxystrobin, difenoconazole, and metalaxyl-M, fludioxonil and metalaxyl-M, mancozeb and metalaxyl-M, copper hydroxide and metalaxyl-M, cyprodinil and fludioxonil, cyproconazole and propiconazole; commercially available fungicide formulations for control of Asian soybean rust disease include, but are not limited to Quadris® (Syngenta Corp), Bravo® (Syngenta Corp), Echo 720® (Sipcam Agro Inc), Headline® 2.09EC (BASF Corp), Tilt® 3.6EC (Syngenta Corp), PropiMax™ 3.6EC (Dow AgroSciences), Bumper® 41.8EC (MakhteshimAgan), Folicur® 3.6F (Bayer CropScience), Laredo® 25EC (Dow AgroSciences), Laredo™ 25EW (Dow AgroSciences), Stratego® 2.08F (Bayer Corp), Domark™ 125SL (Sipcam Agro USA), and Pristine®38%WDG (BASF Corp) these can be combined with DHPS inhibitor compositions as described in the present invention to provide enhanced protection from fungal disease; nematocides such as aldoxycarb and fenamiphos; bactericides such as streptomycin; acaricides such as amitraz, chinomethionat, chlorobenzilate, cyhexatin, dicofol, dienochlor, etoxazole, fenazaquin, fenbutatin oxide, fenpropathrin, fenpyroximate, hexythiazox, propargite, pyridaben and tebufenpyrad; and biological agents such as *Bacillus thuringiensis*, *Bacillus thuringiensis* delta endotoxin, baculovirus, and entomopathogenic bacteria, virus and fungi.

**Table 1**

SEQ ID NO	SPECIES	TYPE	LENGTH
1	Amaranthus hybridus	cDNAContig	672
2	Amaranthus lividus	cDNAContig	458
3	Amaranthus palmeri	gDNAContig	5590
4	Amaranthus palmeri	gDNAContig	5515
5	Amaranthus palmeri	cDNAContig	1431
6	Amaranthus palmeri	cDNAContig	1431
7	Amaranthus palmeri	cDNAContig	1163
8	Amaranthus rudis	cDNAContig	1431
9	Amaranthus rudis	gDNAContig	1059
10	Amaranthus rudis	gDNAContig	792
11	Amaranthus rudis	gDNAContig	764
12	Amaranthus rudis	gDNAContig	755
13	Amaranthus rudis	cDNAContig	602
14	Amaranthus rudis	gDNAContig	584
15	Amaranthus rudis	gDNAContig	570
16	Amaranthus rudis	cDNAContig	411
17	Amaranthus rudis	gDNAContig	260
18	Amaranthus viridis	cDNAContig	1504
19	Amaranthus viridis	cDNAContig	655
20	Ambrosia trifida	cDNAContig	1413
21	Ambrosia trifida	gDNAContig	1328
22	Ambrosia trifida	gDNAContig	486
23	Ambrosia trifida	cDNAContig	486
24	Ambrosia trifida	cDNAContig	410
25	Ambrosia trifida	gDNAContig	381
26	Ambrosia trifida	gDNAContig	304
27	Ambrosia trifida	gDNAContig	255
28	Ambrosia trifida	gDNAContig	201
29	Ambrosia trifida	cDNAContig	101
30	Conyza canadensis	gDNAContig	5446
31	Conyza canadensis	gDNAContig	5446
32	Conyza canadensis	cDNAContig	1401
33	Conyza canadensis	cDNAContig	1262
34	Euphorbia heterophylla	gDNAContig	4643
35	Euphorbia heterophylla	gDNAContig	4556
36	Euphorbia heterophylla	cDNAContig	1425
37	Euphorbia heterophylla	cDNAContig	1425

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(continued)

SEQ ID NO	SPECIES	TYPE	LENGTH
38	Euphorbia heterophylla	cDNAContig	681
39	Euphorbia heterophylla	gDNAContig	444
40	Euphorbia heterophylla	gDNAContig	379
41	Digitaria sanguinalis	gDNAContig	4031
42	Digitaria sanguinalis	cDNAContig	1428
43	Kochia scoparia	gDNAContig	4007
44	Kochia scoparia	gDNAContig	3505
45	Kochia scoparia	cDNAContig	1446
46	Kochia scoparia	cDNAContig	1446
47	Lolium multiflorum	gDNAContig	3920
48	Lolium multiflorum	cDNAContig	1467
49	Lolium multiflorum	gDNAContig	834
50	Lolium multiflorum	cDNAContig	834
51	Lolium multiflorum	gDNAContig	212
52	Lolium multiflorum	cDNAContig	210
53	Lolium multiflorum	gDNAContig	201
54	Lolium multiflorum	cDNAContig	150

**Table 2**

SEQ ID NO	Species	Name   Reference	Type	Start	End
55	Amaranthus hybridus	DHP_A1_1	cDNAContig	1	200
56	Amaranthus hybridus	DHP_A1_1	cDNAContig	1	200
57	Amaranthus hybridus	DHP_A1_2	cDNAContig	176	375
58	Amaranthus hybridus	DHP_A1_2	cDNAContig	176	375
59	Amaranthus hybridus	DHP_A1_3	cDNAContig	351	550
60	Amaranthus hybridus	DHP_A1_3	cDNAContig	351	550
61	Amaranthus lividus	DHP_A2_1	cDNAContig	1	200
62	Amaranthus lividus	DHP_A2_1	cDNAContig	1	200
63	Amaranthus lividus	DHP_A2_2	cDNAContig	176	375
64	Amaranthus lividus	DHP_A2_2	cDNAContig	176	375
65	Amaranthus palmeri	DHP_A4_1	gDNAContig	1	200
66	Amaranthus palmeri	DHP_A4_1	gDNAContig	1	200
67	Amaranthus palmeri	DHP_A4_2	gDNAContig	176	375
68	Amaranthus palmeri	DHP_A4_2	gDNAContig	176	375
69	Amaranthus palmeri	DHP_A4_3	gDNAContig	351	550
70	Amaranthus palmeri	DHP_A4_3	gDNAContig	351	550
71	Amaranthus palmeri	DHP_A4_4	gDNAContig	526	725

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(continued)

	SEQ ID NO	Species	Name   Reference	Type	Start	End
5	72	Amaranthus palmeri	DHP_A4_4	gDNAContig	526	725
10	73	Amaranthus palmeri	DHP_A4_5	gDNAContig	701	900
15	74	Amaranthus palmeri	DHP_A4_5	gDNAContig	701	900
20	75	Amaranthus palmeri	DHP_A4_6	gDNAContig	876	1075
25	76	Amaranthus palmeri	DHP_A4_6	gDNAContig	876	1075
30	77	Amaranthus palmeri	DHP_A4_7	gDNAContig	1051	1250
35	78	Amaranthus palmeri	DHP_A4_7	gDNAContig	1051	1250
40	79	Amaranthus palmeri	DHP_A4_8	gDNAContig	1226	1425
45	80	Amaranthus palmeri	DHP_A4_8	gDNAContig	1226	1425
50	81	Amaranthus palmeri	DHP_A4_9	gDNAContig	1401	1600
55	82	Amaranthus palmeri	DHP_A4_9	gDNAContig	1401	1600
	83	Amaranthus palmeri	DHP_A4_10	gDNAContig	1576	1775
	84	Amaranthus palmeri	DHP_A4_10	gDNAContig	1576	1775
	85	Amaranthus palmeri	DHP_A4_11	gDNAContig	1751	1950
	86	Amaranthus palmeri	DHP_A4_11	gDNAContig	1751	1950
	87	Amaranthus palmeri	DHP_A4_12	gDNAContig	1926	2125
	88	Amaranthus palmeri	DHP_A4_12	gDNAContig	1926	2125
	89	Amaranthus palmeri	DHP_A4_13	gDNAContig	2101	2300
	90	Amaranthus palmeri	DHP_A4_13	gDNAContig	2101	2300
	91	Amaranthus palmeri	DHP_A4_14	gDNAContig	2276	2475
	92	Amaranthus palmeri	DHP_A4_14	gDNAContig	2276	2475
	93	Amaranthus palmeri	DHP_A4_15	gDNAContig	2451	2650
	94	Amaranthus palmeri	DHP_A4_15	gDNAContig	2451	2650
	95	Amaranthus palmeri	DHP_A4_16	gDNAContig	2626	2825
	96	Amaranthus palmeri	DHP_A4_16	gDNAContig	2626	2825
	97	Amaranthus palmeri	DHP_A4_17	gDNAContig	2801	3000
	98	Amaranthus palmeri	DHP_A4_17	gDNAContig	2801	3000
	99	Amaranthus palmeri	DHP_A4_18	gDNAContig	2976	3175
	100	Amaranthus palmeri	DHP_A4_18	gDNAContig	2976	3175
	101	Amaranthus palmeri	DHP_A4_19	gDNAContig	3151	3350
	102	Amaranthus palmeri	DHP_A4_19	gDNAContig	3151	3350
	103	Amaranthus palmeri	DHP_A4_20	gDNAContig	3326	3525
	104	Amaranthus palmeri	DHP_A4_20	gDNAContig	3326	3525
	105	Amaranthus palmeri	DHP_A4_21	gDNAContig	3501	3700
	106	Amaranthus palmeri	DHP_A4_21	gDNAContig	3501	3700
	107	Amaranthus palmeri	DHP_A4_22	gDNAContig	3676	3875
	108	Amaranthus palmeri	DHP_A4_22	gDNAContig	3676	3875
	109	Amaranthus palmeri	DHP_A4_23	gDNAContig	3851	4050

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(continued)

	SEQ ID NO	Species	Name   Reference	Type	Start	End
5	110	Amaranthus palmeri	DHP_A4_23	gDNAContig	3851	4050
10	111	Amaranthus palmeri	DHP_A4_24	gDNAContig	4026	4225
15	112	Amaranthus palmeri	DHP_A4_24	gDNAContig	4026	4225
20	113	Amaranthus palmeri	DHP_A4_25	gDNAContig	4201	4400
25	114	Amaranthus palmeri	DHP_A4_25	gDNAContig	4201	4400
30	115	Amaranthus palmeri	DHP_A4_26	gDNAContig	4376	4575
35	116	Amaranthus palmeri	DHP_A4_26	gDNAContig	4376	4575
40	117	Amaranthus palmeri	DHP_A4_27	gDNAContig	4551	4750
45	118	Amaranthus palmeri	DHP_A4_27	gDNAContig	4551	4750
50	119	Amaranthus palmeri	DHP_A4_28	gDNAContig	4726	4925
55	120	Amaranthus palmeri	DHP_A4_28	gDNAContig	4726	4925
	121	Amaranthus palmeri	DHP_A4_29	gDNAContig	4901	5100
	122	Amaranthus palmeri	DHP_A4_29	gDNAContig	4901	5100
	123	Amaranthus palmeri	DHP_A4_30	gDNAContig	5076	5275
	124	Amaranthus palmeri	DHP_A4_30	gDNAContig	5076	5275
	125	Amaranthus palmeri	DHP_A4_31	gDNAContig	5251	5450
	126	Amaranthus palmeri	DHP_A4_31	gDNAContig	5251	5450
	127	Amaranthus palmeri	DHP_A7_1	cDNAContig	1	200
	128	Amaranthus palmeri	DHP_A7_1	cDNAContig	1	200
	129	Amaranthus palmeri	DHP_A7_2	cDNAContig	176	375
	130	Amaranthus palmeri	DHP_A7_2	cDNAContig	176	375
	131	Amaranthus palmeri	DHP_A7_3	cDNAContig	351	550
	132	Amaranthus palmeri	DHP_A7_3	cDNAContig	351	550
	133	Amaranthus palmeri	DHP_A7_4	cDNAContig	526	725
	134	Amaranthus palmeri	DHP_A7_4	cDNAContig	526	725
	135	Amaranthus palmeri	DHP_A7_5	cDNAContig	701	900
	136	Amaranthus palmeri	DHP_A7_5	cDNAContig	701	900
	137	Amaranthus palmeri	DHP_A7_6	cDNAContig	876	1075
	138	Amaranthus palmeri	DHP_A7_6	cDNAContig	876	1075
	139	Amaranthus palmeri	DHP_A5_1	cDNAContig	1	200
	140	Amaranthus palmeri	DHP_A5_1	cDNAContig	1	200
	141	Amaranthus palmeri	DHP_A5_2	cDNAContig	176	375
	142	Amaranthus palmeri	DHP_A5_2	cDNAContig	176	375
	143	Amaranthus palmeri	DHP_A5_3	cDNAContig	351	550
	144	Amaranthus palmeri	DHP_A5_3	cDNAContig	351	550
	145	Amaranthus palmeri	DHP_A5_4	cDNAContig	526	725
	146	Amaranthus palmeri	DHP_A5_4	cDNAContig	526	725
	147	Amaranthus palmeri	DHP_A5_5	cDNAContig	701	900

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(continued)

	SEQ ID NO	Species	Name   Reference	Type	Start	End
5	148	Amaranthus palmeri	DHP_A5_5	cDNAContig	701	900
10	149	Amaranthus palmeri	DHP_A5_6	cDNAContig	876	1075
15	150	Amaranthus palmeri	DHP_A5_6	cDNAContig	876	1075
20	151	Amaranthus palmeri	DHP_A5_7	cDNAContig	1051	1250
25	152	Amaranthus palmeri	DHP_A5_7	cDNAContig	1051	1250
30	153	Amaranthus palmeri	DHP_A5_8	cDNAContig	1226	1425
35	154	Amaranthus palmeri	DHP_A5_8	cDNAContig	1226	1425
40	155	Amaranthus palmeri	DHP_A6_1	cDNAContig	1	200
45	156	Amaranthus palmeri	DHP_A6_1	cDNAContig	1	200
50	157	Amaranthus palmeri	DHP_A6_2	cDNAContig	176	375
55	158	Amaranthus palmeri	DHP_A6_2	cDNAContig	176	375
	159	Amaranthus palmeri	DHP_A6_3	cDNAContig	351	550
	160	Amaranthus palmeri	DHP_A6_3	cDNAContig	351	550
	161	Amaranthus palmeri	DHP_A6_4	cDNAContig	526	725
	162	Amaranthus palmeri	DHP_A6_4	cDNAContig	526	725
	163	Amaranthus palmeri	DHP_A6_5	cDNAContig	701	900
	164	Amaranthus palmeri	DHP_A6_5	cDNAContig	701	900
	165	Amaranthus palmeri	DHP_A6_6	cDNAContig	876	1075
	166	Amaranthus palmeri	DHP_A6_6	cDNAContig	876	1075
	167	Amaranthus palmeri	DHP_A6_7	cDNAContig	1051	1250
	168	Amaranthus palmeri	DHP_A6_7	cDNAContig	1051	1250
	169	Amaranthus palmeri	DHP_A6_8	cDNAContig	1226	1425
	170	Amaranthus palmeri	DHP_A6_8	cDNAContig	1226	1425
	171	Amaranthus palmeri	DHP_A3_1	gDNAContig	1	200
	172	Amaranthus palmeri	DHP_A3_1	gDNAContig	1	200
	173	Amaranthus palmeri	DHP_A3_2	gDNAContig	176	375
	174	Amaranthus palmeri	DHP_A3_2	gDNAContig	176	375
	175	Amaranthus palmeri	DHP_A3_3	gDNAContig	351	550
	176	Amaranthus palmeri	DHP_A3_3	gDNAContig	351	550
	177	Amaranthus palmeri	DHP_A3_4	gDNAContig	526	725
	178	Amaranthus palmeri	DHP_A3_4	gDNAContig	526	725
	179	Amaranthus palmeri	DHP_A3_5	gDNAContig	701	900
	180	Amaranthus palmeri	DHP_A3_5	gDNAContig	701	900
	181	Amaranthus palmeri	DHP_A3_6	gDNAContig	876	1075
	182	Amaranthus palmeri	DHP_A3_6	gDNAContig	876	1075
	183	Amaranthus palmeri	DHP_A3_7	gDNAContig	1051	1250
	184	Amaranthus palmeri	DHP_A3_7	gDNAContig	1051	1250
	185	Amaranthus palmeri	DHP_A3_8	gDNAContig	1226	1425

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(continued)

	SEQ ID NO	Species	Name   Reference	Type	Start	End
5	186	Amaranthus palmeri	DHP_A3_8	gDNAContig	1226	1425
10	187	Amaranthus palmeri	DHP_A3_9	gDNAContig	1401	1600
15	188	Amaranthus palmeri	DHP_A3_9	gDNAContig	1401	1600
20	189	Amaranthus palmeri	DHP_A3_10	gDNAContig	1576	1775
25	190	Amaranthus palmeri	DHP_A3_10	gDNAContig	1576	1775
30	191	Amaranthus palmeri	DHP_A3_11	gDNAContig	1751	1950
35	192	Amaranthus palmeri	DHP_A3_11	gDNAContig	1751	1950
40	193	Amaranthus palmeri	DHP_A3_12	gDNAContig	1926	2125
45	194	Amaranthus palmeri	DHP_A3_12	gDNAContig	1926	2125
50	195	Amaranthus palmeri	DHP_A3_13	gDNAContig	2101	2300
55	196	Amaranthus palmeri	DHP_A3_13	gDNAContig	2101	2300
	197	Amaranthus palmeri	DHP_A3_14	gDNAContig	2276	2475
	198	Amaranthus palmeri	DHP_A3_14	gDNAContig	2276	2475
	199	Amaranthus palmeri	DHP_A3_15	gDNAContig	2451	2650
	200	Amaranthus palmeri	DHP_A3_15	gDNAContig	2451	2650
	201	Amaranthus palmeri	DHP_A3_16	gDNAContig	2626	2825
	202	Amaranthus palmeri	DHP_A3_16	gDNAContig	2626	2825
	203	Amaranthus palmeri	DHP_A3_17	gDNAContig	2801	3000
	204	Amaranthus palmeri	DHP_A3_17	gDNAContig	2801	3000
	205	Amaranthus palmeri	DHP_A3_18	gDNAContig	2976	3175
	206	Amaranthus palmeri	DHP_A3_18	gDNAContig	2976	3175
	207	Amaranthus palmeri	DHP_A3_19	gDNAContig	3151	3350
	208	Amaranthus palmeri	DHP_A3_19	gDNAContig	3151	3350
	209	Amaranthus palmeri	DHP_A3_20	gDNAContig	3326	3525
	210	Amaranthus palmeri	DHP_A3_20	gDNAContig	3326	3525
	211	Amaranthus palmeri	DHP_A3_21	gDNAContig	3501	3700
	212	Amaranthus palmeri	DHP_A3_21	gDNAContig	3501	3700
	213	Amaranthus palmeri	DHP_A3_22	gDNAContig	3676	3875
	214	Amaranthus palmeri	DHP_A3_22	gDNAContig	3676	3875
	215	Amaranthus palmeri	DHP_A3_23	gDNAContig	3851	4050
	216	Amaranthus palmeri	DHP_A3_23	gDNAContig	3851	4050
	217	Amaranthus palmeri	DHP_A3_24	gDNAContig	4026	4225
	218	Amaranthus palmeri	DHP_A3_24	gDNAContig	4026	4225
	219	Amaranthus palmeri	DHP_A3_25	gDNAContig	4201	4400
	220	Amaranthus palmeri	DHP_A3_25	gDNAContig	4201	4400
	221	Amaranthus palmeri	DHP_A3_26	gDNAContig	4376	4575
	222	Amaranthus palmeri	DHP_A3_26	gDNAContig	4376	4575
	223	Amaranthus palmeri	DHP_A3_27	gDNAContig	4551	4750

(continued)

	SEQ ID NO	Species	Name   Reference	Type	Start	End
5	224	Amaranthus palmeri	DHP_A3_27	gDNAContig	4551	4750
10	225	Amaranthus palmeri	DHP_A3_28	gDNAContig	4726	4925
15	226	Amaranthus palmeri	DHP_A3_28	gDNAContig	4726	4925
20	227	Amaranthus palmeri	DHP_A3_29	gDNAContig	4901	5100
25	228	Amaranthus palmeri	DHP_A3_29	gDNAContig	4901	5100
30	229	Amaranthus palmeri	DHP_A3_30	gDNAContig	5076	5275
35	230	Amaranthus palmeri	DHP_A3_30	gDNAContig	5076	5275
40	231	Amaranthus palmeri	DHP_A3_31	gDNAContig	5251	5450
45	232	Amaranthus palmeri	DHP_A3_31	gDNAContig	5251	5450
50	233	Amaranthus rudis	DHP_A10_1	gDNAContig	1	200
55	234	Amaranthus rudis	DHP_A10_1	gDNAContig	1	200
	235	Amaranthus rudis	DHP_A10_2	gDNAContig	176	375
	236	Amaranthus rudis	DHP_A10_2	gDNAContig	176	375
	237	Amaranthus rudis	DHP_A10_3	gDNAContig	351	550
	238	Amaranthus rudis	DHP_A10_3	gDNAContig	351	550
	239	Amaranthus rudis	DHP_A10_4	gDNAContig	526	725
	240	Amaranthus rudis	DHP_A10_4	gDNAContig	526	725
	241	Amaranthus rudis	DHP_A13_1	cDNAContig	1	200
	242	Amaranthus rudis	DHP_A13_1	cDNAContig	1	200
	243	Amaranthus rudis	DHP_A13_2	cDNAContig	176	375
	244	Amaranthus rudis	DHP_A13_2	cDNAContig	176	375
	245	Amaranthus rudis	DHP_A13_3	cDNAContig	351	550
	246	Amaranthus rudis	DHP_A13_3	cDNAContig	351	550
	247	Amaranthus rudis	DHP_A14_1	gDNAContig	1	200
	248	Amaranthus rudis	DHP_A14_1	gDNAContig	1	200
	249	Amaranthus rudis	DHP_A14_2	gDNAContig	176	375
	250	Amaranthus rudis	DHP_A14_2	gDNAContig	176	375
	251	Amaranthus rudis	DHP_A14_3	gDNAContig	351	550
	252	Amaranthus rudis	DHP_A14_3	gDNAContig	351	550
	253	Amaranthus rudis	DHP_A17_1	gDNAContig	1	200
	254	Amaranthus rudis	DHP_A17_1	gDNAContig	1	200
	255	Amaranthus rudis	DHP_A16_1	cDNAContig	1	200
	256	Amaranthus rudis	DHP_A16_1	cDNAContig	1	200
	257	Amaranthus rudis	DHP_A16_2	cDNAContig	176	375
	258	Amaranthus rudis	DHP_A16_2	cDNAContig	176	375
	259	Amaranthus rudis	DHP_A8_1	cDNAContig	1	200
	260	Amaranthus rudis	DHP_A8_1	cDNAContig	1	200
	261	Amaranthus rudis	DHP_A8_2	cDNAContig	176	375

(continued)

	SEQ ID NO	Species	Name   Reference	Type	Start	End
5	262	Amaranthus rufis	DHP_A8_2	cDNAContig	176	375
10	263	Amaranthus rufis	DHP_A8_3	cDNAContig	351	550
15	264	Amaranthus rufis	DHP_A8_3	cDNAContig	351	550
20	265	Amaranthus rufis	DHP_A8_4	cDNAContig	526	725
25	266	Amaranthus rufis	DHP_A8_4	cDNAContig	526	725
30	267	Amaranthus rufis	DHP_A8_5	cDNAContig	701	900
35	268	Amaranthus rufis	DHP_A8_5	cDNAContig	701	900
40	269	Amaranthus rufis	DHP_A8_6	cDNAContig	876	1075
45	270	Amaranthus rufis	DHP_A8_6	cDNAContig	876	1075
50	271	Amaranthus rufis	DHP_A8_7	cDNAContig	1051	1250
55	272	Amaranthus rufis	DHP_A8_7	cDNAContig	1051	1250
	273	Amaranthus rufis	DHP_A8_8	cDNAContig	1226	1425
	274	Amaranthus rufis	DHP_A8_8	cDNAContig	1226	1425
	275	Amaranthus rufis	DHP_A12_1	gDNAContig	1	200
	276	Amaranthus rufis	DHP_A12_1	gDNAContig	1	200
	277	Amaranthus rufis	DHP_A12_2	gDNAContig	176	375
	278	Amaranthus rufis	DHP_A12_2	gDNAContig	176	375
	279	Amaranthus rufis	DHP_A12_3	gDNAContig	351	550
	280	Amaranthus rufis	DHP_A12_3	gDNAContig	351	550
	281	Amaranthus rufis	DHP_A12_4	gDNAContig	526	725
	282	Amaranthus rufis	DHP_A12_4	gDNAContig	526	725
	283	Amaranthus rufis	DHP_A15_1	gDNAContig	1	200
	284	Amaranthus rufis	DHP_A15_1	gDNAContig	1	200
	285	Amaranthus rufis	DHP_A15_2	gDNAContig	176	375
	286	Amaranthus rufis	DHP_A15_2	gDNAContig	176	375
	287	Amaranthus rufis	DHP_A15_3	gDNAContig	351	550
	288	Amaranthus rufis	DHP_A15_3	gDNAContig	351	550
	289	Amaranthus rufis	DHP_A11_1	gDNAContig	1	200
	290	Amaranthus rufis	DHP_A11_1	gDNAContig	1	200
	291	Amaranthus rufis	DHP_A11_2	gDNAContig	176	375
	292	Amaranthus rufis	DHP_A11_2	gDNAContig	176	375
	293	Amaranthus rufis	DHP_A11_3	gDNAContig	351	550
	294	Amaranthus rufis	DHP_A11_3	gDNAContig	351	550
	295	Amaranthus rufis	DHP_A11_4	gDNAContig	526	725
	296	Amaranthus rufis	DHP_A11_4	gDNAContig	526	725
	297	Amaranthus rufis	DHP_A9_1	gDNAContig	1	200
	298	Amaranthus rufis	DHP_A9_1	gDNAContig	1	200
	299	Amaranthus rufis	DHP_A9_2	gDNAContig	176	375

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	SEQ ID NO	Species	Name   Reference	Type	Start	End
5	300	Amaranthus rudis	DHP_A9_2	gDNAContig	176	375
10	301	Amaranthus rudis	DHP_A9_3	gDNAContig	351	550
15	302	Amaranthus rudis	DHP_A9_3	gDNAContig	351	550
20	303	Amaranthus rudis	DHP_A9_4	gDNAContig	526	725
25	304	Amaranthus rudis	DHP_A9_4	gDNAContig	526	725
30	305	Amaranthus rudis	DHP_A9_5	gDNAContig	701	900
35	306	Amaranthus rudis	DHP_A9_5	gDNAContig	701	900
40	307	Amaranthus viridis	DHP_A19_1	cDNAContig	1	200
45	308	Amaranthus viridis	DHP_A19_1	cDNAContig	1	200
50	309	Amaranthus viridis	DHP_A19_2	cDNAContig	176	375
55	310	Amaranthus viridis	DHP_A19_2	cDNAContig	176	375
	311	Amaranthus viridis	DHP_A19_3	cDNAContig	351	550
	312	Amaranthus viridis	DHP_A19_3	cDNAContig	351	550
	313	Amaranthus viridis	DHP_A18_1	cDNAContig	1	200
	314	Amaranthus viridis	DHP_A18_1	cDNAContig	1	200
	315	Amaranthus viridis	DHP_A18_2	cDNAContig	176	375
	316	Amaranthus viridis	DHP_A18_2	cDNAContig	176	375
	317	Amaranthus viridis	DHP_A18_3	cDNAContig	351	550
	318	Amaranthus viridis	DHP_A18_3	cDNAContig	351	550
	319	Amaranthus viridis	DHP_A18_4	cDNAContig	526	725
	320	Amaranthus viridis	DHP_A18_4	cDNAContig	526	725
	321	Amaranthus viridis	DHP_A18_5	cDNAContig	701	900
	322	Amaranthus viridis	DHP_A18_5	cDNAContig	701	900
	323	Amaranthus viridis	DHP_A18_6	cDNAContig	876	1075
	324	Amaranthus viridis	DHP_A18_6	cDNAContig	876	1075
	325	Amaranthus viridis	DHP_A18_7	cDNAContig	1051	1250
	326	Amaranthus viridis	DHP_A18_7	cDNAContig	1051	1250
	327	Amaranthus viridis	DHP_A18_8	cDNAContig	1226	1425
	328	Amaranthus viridis	DHP_A18_8	cDNAContig	1226	1425
	329	Ambrosia trifida	DHP_A21_1	gDNAContig	1	200
	330	Ambrosia trifida	DHP_A21_1	gDNAContig	1	200
	331	Ambrosia trifida	DHP_A21_2	gDNAContig	176	375
	332	Ambrosia trifida	DHP_A21_2	gDNAContig	176	375
	333	Ambrosia trifida	DHP_A21_3	gDNAContig	351	550
	334	Ambrosia trifida	DHP_A21_3	gDNAContig	351	550
	335	Ambrosia trifida	DHP_A21_4	gDNAContig	526	725
	336	Ambrosia trifida	DHP_A21_4	gDNAContig	526	725
	337	Ambrosia trifida	DHP_A21_5	gDNAContig	701	900

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(continued)

	SEQ ID NO	Species	Name   Reference	Type	Start	End
5	338	Ambrosia trifida	DHP_A21_5	gDNAContig	701	900
	339	Ambrosia trifida	DHP_A21_6	gDNAContig	876	1075
10	340	Ambrosia trifida	DHP_A21_6	gDNAContig	876	1075
	341	Ambrosia trifida	DHP_A21_7	gDNAContig	1051	1250
15	342	Ambrosia trifida	DHP_A21_7	gDNAContig	1051	1250
	343	Ambrosia trifida	DHP_A28_1	gDNAContig	1	200
	344	Ambrosia trifida	DHP_A28_1	gDNAContig	1	200
20	345	Ambrosia trifida	DHP_A26_1	gDNAContig	1	200
	346	Ambrosia trifida	DHP_A26_1	gDNAContig	1	200
	347	Ambrosia trifida	DHP_A24_1	cDNAContig	1	200
25	348	Ambrosia trifida	DHP_A24_1	cDNAContig	1	200
	349	Ambrosia trifida	DHP_A24_2	cDNAContig	176	375
	350	Ambrosia trifida	DHP_A24_2	cDNAContig	176	375
	351	Ambrosia trifida	DHP_A27_1	gDNAContig	1	200
30	352	Ambrosia trifida	DHP_A27_1	gDNAContig	1	200
	353	Ambrosia trifida	DHP_A23_1	cDNAContig	1	200
	354	Ambrosia trifida	DHP_A23_1	cDNAContig	1	200
35	355	Ambrosia trifida	DHP_A23_2	cDNAContig	176	375
	356	Ambrosia trifida	DHP_A23_2	cDNAContig	176	375
	357	Ambrosia trifida	DHP_A29_1	cDNAContig	1	101
	358	Ambrosia trifida	DHP_A29_1	cDNAContig	1	101
40	359	Ambrosia trifida	DHP_A20_1	cDNAContig	1	200
	360	Ambrosia trifida	DHP_A20_1	cDNAContig	1	200
	361	Ambrosia trifida	DHP_A20_2	cDNAContig	176	375
45	362	Ambrosia trifida	DHP_A20_2	cDNAContig	176	375
	363	Ambrosia trifida	DHP_A20_3	cDNAContig	351	550
	364	Ambrosia trifida	DHP_A20_3	cDNAContig	351	550
	365	Ambrosia trifida	DHP_A20_4	cDNAContig	526	725
50	366	Ambrosia trifida	DHP_A20_4	cDNAContig	526	725
	367	Ambrosia trifida	DHP_A20_5	cDNAContig	701	900
	368	Ambrosia trifida	DHP_A20_5	cDNAContig	701	900
	369	Ambrosia trifida	DHP_A20_6	cDNAContig	876	1075
55	370	Ambrosia trifida	DHP_A20_6	cDNAContig	876	1075
	371	Ambrosia trifida	DHP_A20_5	cDNAContig	1051	1250
	372	Ambrosia trifida	DHP_A20_5	cDNAContig	1051	1250
	373	Ambrosia trifida	DHP_A25_1	gDNAContig	1	200
	374	Ambrosia trifida	DHP_A25_1	gDNAContig	1	200
	375	Ambrosia trifida	DHP_A25_2	gDNAContig	176	375

(continued)

	SEQ ID NO	Species	Name   Reference	Type	Start	End
5	376	Ambrosia trifida	DHP_A25_2	gDNAContig	176	375
	377	Conyza canadensis	DHP_A33_1	cDNAContig	1	200
	378	Conyza canadensis	DHP_A33_1	cDNAContig	1	200
	379	Conyza canadensis	DHP_A33_2	cDNAContig	176	375
	380	Conyza canadensis	DHP_A33_2	cDNAContig	176	375
	381	Conyza canadensis	DHP_A33_3	cDNAContig	351	550
	382	Conyza canadensis	DHP_A33_3	cDNAContig	351	550
10	383	Conyza canadensis	DHP_A33_4	cDNAContig	526	725
	384	Conyza canadensis	DHP_A33_4	cDNAContig	526	725
	385	Conyza canadensis	DHP_A33_5	cDNAContig	701	900
	386	Conyza canadensis	DHP_A33_5	cDNAContig	701	900
	387	Conyza canadensis	DHP_A33_6	cDNAContig	876	1075
	388	Conyza canadensis	DHP_A33_6	cDNAContig	876	1075
	389	Conyza canadensis	DHP_A33_7	cDNAContig	1051	1250
15	390	Conyza canadensis	DHP_A33_7	cDNAContig	1051	1250
	391	Conyza canadensis	DHP_A32_1	cDNAContig	1	200
	392	Conyza canadensis	DHP_A32_1	cDNAContig	1	200
	393	Conyza canadensis	DHP_A32_2	cDNAContig	176	375
	394	Conyza canadensis	DHP_A32_2	cDNAContig	176	375
	395	Conyza canadensis	DHP_A32_3	cDNAContig	351	550
	396	Conyza canadensis	DHP_A32_3	cDNAContig	351	550
20	397	Conyza canadensis	DHP_A32_4	cDNAContig	526	725
	398	Conyza canadensis	DHP_A32_4	cDNAContig	526	725
	399	Conyza canadensis	DHP_A32_5	cDNAContig	701	900
	400	Conyza canadensis	DHP_A32_5	cDNAContig	701	900
	401	Conyza canadensis	DHP_A32_6	cDNAContig	876	1075
	402	Conyza canadensis	DHP_A32_6	cDNAContig	876	1075
	403	Conyza canadensis	DHP_A32_7	cDNAContig	1051	1250
25	404	Conyza canadensis	DHP_A32_7	cDNAContig	1051	1250
	405	Conyza canadensis	DHP_A30_1	gDNAContig	1	200
	406	Conyza canadensis	DHP_A30_1	gDNAContig	1	200
	407	Conyza canadensis	DHP_A30_2	gDNAContig	176	375
	408	Conyza canadensis	DHP_A30_2	gDNAContig	176	375
	409	Conyza canadensis	DHP_A30_3	gDNAContig	351	550
	410	Conyza canadensis	DHP_A30_3	gDNAContig	351	550
30	411	Conyza canadensis	DHP_A30_4	gDNAContig	526	725
	412	Conyza canadensis	DHP_A30_4	gDNAContig	526	725
	413	Conyza canadensis	DHP_A30_5	gDNAContig	701	900
35						
40						
45						
50						
55						

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	SEQ ID NO	Species	Name   Reference	Type	Start	End
5	414	<i>Conyza canadensis</i>	DHP_A30_5	gDNAContig	701	900
10	415	<i>Conyza canadensis</i>	DHP_A30_6	gDNAContig	876	1075
15	416	<i>Conyza canadensis</i>	DHP_A30_6	gDNAContig	876	1075
20	417	<i>Conyza canadensis</i>	DHP_A30_7	gDNAContig	1051	1250
25	418	<i>Conyza canadensis</i>	DHP_A30_7	gDNAContig	1051	1250
30	419	<i>Conyza canadensis</i>	DHP_A30_8	gDNAContig	1226	1425
35	420	<i>Conyza canadensis</i>	DHP_A30_8	gDNAContig	1226	1425
40	421	<i>Conyza canadensis</i>	DHP_A30_9	gDNAContig	1401	1600
45	422	<i>Conyza canadensis</i>	DHP_A30_9	gDNAContig	1401	1600
50	423	<i>Conyza canadensis</i>	DHP_A30_10	gDNAContig	1576	1775
55	424	<i>Conyza canadensis</i>	DHP_A30_10	gDNAContig	1576	1775
	425	<i>Conyza canadensis</i>	DHP_A30_11	gDNAContig	1751	1950
	426	<i>Conyza canadensis</i>	DHP_A30_11	gDNAContig	1751	1950
	427	<i>Conyza canadensis</i>	DHP_A30_12	gDNAContig	1926	2125
	428	<i>Conyza canadensis</i>	DHP_A30_12	gDNAContig	1926	2125
	429	<i>Conyza canadensis</i>	DHP_A30_13	gDNAContig	2101	2300
	430	<i>Conyza canadensis</i>	DHP_A30_13	gDNAContig	2101	2300
	431	<i>Conyza canadensis</i>	DHP_A30_14	gDNAContig	2276	2475
	432	<i>Conyza canadensis</i>	DHP_A30_14	gDNAContig	2276	2475
	433	<i>Conyza canadensis</i>	DHP_A30_15	gDNAContig	2451	2650
	434	<i>Conyza canadensis</i>	DHP_A30_15	gDNAContig	2451	2650
	435	<i>Conyza canadensis</i>	DHP_A30_16	gDNAContig	2626	2825
	436	<i>Conyza canadensis</i>	DHP_A30_16	gDNAContig	2626	2825
	437	<i>Conyza canadensis</i>	DHP_A30_17	gDNAContig	2801	3000
	438	<i>Conyza canadensis</i>	DHP_A30_17	gDNAContig	2801	3000
	439	<i>Conyza canadensis</i>	DHP_A30_18	gDNAContig	2976	3175
	440	<i>Conyza canadensis</i>	DHP_A30_18	gDNAContig	2976	3175
	441	<i>Conyza canadensis</i>	DHP_A30_19	gDNAContig	3151	3350
	442	<i>Conyza canadensis</i>	DHP_A30_19	gDNAContig	3151	3350
	443	<i>Conyza canadensis</i>	DHP_A30_20	gDNAContig	3326	3525
	444	<i>Conyza canadensis</i>	DHP_A30_20	gDNAContig	3326	3525
	445	<i>Conyza canadensis</i>	DHP_A30_21	gDNAContig	3501	3700
	446	<i>Conyza canadensis</i>	DHP_A30_21	gDNAContig	3501	3700
	447	<i>Conyza canadensis</i>	DHP_A30_22	gDNAContig	3676	3875
	448	<i>Conyza canadensis</i>	DHP_A30_22	gDNAContig	3676	3875
	449	<i>Conyza canadensis</i>	DHP_A30_23	gDNAContig	3851	4050
	450	<i>Conyza canadensis</i>	DHP_A30_23	gDNAContig	3851	4050
	451	<i>Conyza canadensis</i>	DHP_A30_24	gDNAContig	4026	4225

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	SEQ ID NO	Species	Name   Reference	Type	Start	End
5	452	<i>Conyza canadensis</i>	DHP_A30_24	gDNAContig	4026	4225
10	453	<i>Conyza canadensis</i>	DHP_A30_25	gDNAContig	4201	4400
15	454	<i>Conyza canadensis</i>	DHP_A30_25	gDNAContig	4201	4400
20	455	<i>Conyza canadensis</i>	DHP_A30_26	gDNAContig	4376	4575
25	456	<i>Conyza canadensis</i>	DHP_A30_26	gDNAContig	4376	4575
30	457	<i>Conyza canadensis</i>	DHP_A30_27	gDNAContig	4551	4750
35	458	<i>Conyza canadensis</i>	DHP_A30_27	gDNAContig	4551	4750
40	459	<i>Conyza canadensis</i>	DHP_A30_28	gDNAContig	4726	4925
45	460	<i>Conyza canadensis</i>	DHP_A30_28	gDNAContig	4726	4925
50	461	<i>Conyza canadensis</i>	DHP_A30_29	gDNAContig	4901	5100
55	462	<i>Conyza canadensis</i>	DHP_A30_29	gDNAContig	4901	5100
	463	<i>Conyza canadensis</i>	DHP_A30_30	gDNAContig	5076	5275
	464	<i>Conyza canadensis</i>	DHP_A30_30	gDNAContig	5076	5275
	465	<i>Conyza canadensis</i>	DHP_A31_1	gDNAContig	1	200
	466	<i>Conyza canadensis</i>	DHP_A31_1	gDNAContig	1	200
	467	<i>Conyza canadensis</i>	DHP_A31_2	gDNAContig	176	375
	468	<i>Conyza canadensis</i>	DHP_A31_2	gDNAContig	176	375
	469	<i>Conyza canadensis</i>	DHP_A31_3	gDNAContig	351	550
	470	<i>Conyza canadensis</i>	DHP_A31_3	gDNAContig	351	550
	471	<i>Conyza canadensis</i>	DHP_A31_4	gDNAContig	526	725
	472	<i>Conyza canadensis</i>	DHP_A31_4	gDNAContig	526	725
	473	<i>Conyza canadensis</i>	DHP_A31_5	gDNAContig	701	900
	474	<i>Conyza canadensis</i>	DHP_A31_5	gDNAContig	701	900
	475	<i>Conyza canadensis</i>	DHP_A31_6	gDNAContig	876	1075
	476	<i>Conyza canadensis</i>	DHP_A31_6	gDNAContig	876	1075
	477	<i>Conyza canadensis</i>	DHP_A31_7	gDNAContig	1051	1250
	478	<i>Conyza canadensis</i>	DHP_A31_7	gDNAContig	1051	1250
	479	<i>Conyza canadensis</i>	DHP_A31_8	gDNAContig	1226	1425
	480	<i>Conyza canadensis</i>	DHP_A31_8	gDNAContig	1226	1425
	481	<i>Conyza canadensis</i>	DHP_A31_9	gDNAContig	1401	1600
	482	<i>Conyza canadensis</i>	DHP_A31_9	gDNAContig	1401	1600
	483	<i>Conyza canadensis</i>	DHP_A31_10	gDNAContig	1576	1775
	484	<i>Conyza canadensis</i>	DHP_A31_10	gDNAContig	1576	1775
	485	<i>Conyza canadensis</i>	DHP_A31_11	gDNAContig	1751	1950
	486	<i>Conyza canadensis</i>	DHP_A31_11	gDNAContig	1751	1950
	487	<i>Conyza canadensis</i>	DHP_A31_12	gDNAContig	1926	2125
	488	<i>Conyza canadensis</i>	DHP_A31_12	gDNAContig	1926	2125
	489	<i>Conyza canadensis</i>	DHP_A31_13	gDNAContig	2101	2300

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	SEQ ID NO	Species	Name   Reference	Type	Start	End
5	490	<i>Conyza canadensis</i>	DHP_A31_13	gDNAContig	2101	2300
10	491	<i>Conyza canadensis</i>	DHP_A31_14	gDNAContig	2276	2475
15	492	<i>Conyza canadensis</i>	DHP_A31_14	gDNAContig	2276	2475
20	493	<i>Conyza canadensis</i>	DHP_A31_15	gDNAContig	2451	2650
25	494	<i>Conyza canadensis</i>	DHP_A31_15	gDNAContig	2451	2650
30	495	<i>Conyza canadensis</i>	DHP_A31_16	gDNAContig	2626	2825
35	496	<i>Conyza canadensis</i>	DHP_A31_16	gDNAContig	2626	2825
40	497	<i>Conyza canadensis</i>	DHP_A31_17	gDNAContig	2801	3000
45	498	<i>Conyza canadensis</i>	DHP_A31_17	gDNAContig	2801	3000
50	499	<i>Conyza canadensis</i>	DHP_A31_18	gDNAContig	2976	3175
55	500	<i>Conyza canadensis</i>	DHP_A31_18	gDNAContig	2976	3175
5	501	<i>Conyza canadensis</i>	DHP_A31_19	gDNAContig	3151	3350
10	502	<i>Conyza canadensis</i>	DHP_A31_19	gDNAContig	3151	3350
15	503	<i>Conyza canadensis</i>	DHP_A31_20	gDNAContig	3326	3525
20	504	<i>Conyza canadensis</i>	DHP_A31_20	gDNAContig	3326	3525
25	505	<i>Conyza canadensis</i>	DHP_A31_21	gDNAContig	3501	3700
30	506	<i>Conyza canadensis</i>	DHP_A31_21	gDNAContig	3501	3700
35	507	<i>Conyza canadensis</i>	DHP_A31_22	gDNAContig	3676	3875
40	508	<i>Conyza canadensis</i>	DHP_A31_22	gDNAContig	3676	3875
45	509	<i>Conyza canadensis</i>	DHP_A31_23	gDNAContig	3851	4050
50	510	<i>Conyza canadensis</i>	DHP_A31_23	gDNAContig	3851	4050
55	511	<i>Conyza canadensis</i>	DHP_A31_24	gDNAContig	4026	4225
5	512	<i>Conyza canadensis</i>	DHP_A31_24	gDNAContig	4026	4225
10	513	<i>Conyza canadensis</i>	DHP_A31_25	gDNAContig	4201	4400
15	514	<i>Conyza canadensis</i>	DHP_A31_25	gDNAContig	4201	4400
20	515	<i>Conyza canadensis</i>	DHP_A31_26	gDNAContig	4376	4575
25	516	<i>Conyza canadensis</i>	DHP_A31_26	gDNAContig	4376	4575
30	517	<i>Conyza canadensis</i>	DHP_A31_27	gDNAContig	4551	4750
35	518	<i>Conyza canadensis</i>	DHP_A31_27	gDNAContig	4551	4750
40	519	<i>Conyza canadensis</i>	DHP_A31_28	gDNAContig	4726	4925
45	520	<i>Conyza canadensis</i>	DHP_A31_28	gDNAContig	4726	4925
50	521	<i>Conyza canadensis</i>	DHP_A31_29	gDNAContig	4901	5100
55	522	<i>Conyza canadensis</i>	DHP_A31_29	gDNAContig	4901	5100
5	523	<i>Conyza canadensis</i>	DHP_A31_30	gDNAContig	5076	5275
10	524	<i>Conyza canadensis</i>	DHP_A31_30	gDNAContig	5076	5275
15	525	<i>Digitaria sanguinalis</i>	DHP_A41_1	gDNAContig	1	200
20	526	<i>Digitaria sanguinalis</i>	DHP_A41_1	gDNAContig	1	200
25	527	<i>Digitaria sanguinalis</i>	DHP_A41_2	gDNAContig	176	375

(continued)

	SEQ ID NO	Species	Name   Reference	Type	Start	End
5	528	Digitaria sanguinalis	DHP_A41_2	gDNAContig	176	375
10	529	Digitaria sanguinalis	DHP_A41_3	gDNAContig	351	550
15	530	Digitaria sanguinalis	DHP_A41_3	gDNAContig	351	550
20	531	Digitaria sanguinalis	DHP_A41_4	gDNAContig	526	725
25	532	Digitaria sanguinalis	DHP_A41_4	gDNAContig	526	725
30	533	Digitaria sanguinalis	DHP_A41_5	gDNAContig	701	900
35	534	Digitaria sanguinalis	DHP_A41_5	gDNAContig	701	900
40	535	Digitaria sanguinalis	DHP_A41_6	gDNAContig	876	1075
45	536	Digitaria sanguinalis	DHP_A41_6	gDNAContig	876	1075
50	537	Digitaria sanguinalis	DHP_A41_7	gDNAContig	1051	1250
55	538	Digitaria sanguinalis	DHP_A41_7	gDNAContig	1051	1250
	539	Digitaria sanguinalis	DHP_A41_8	gDNAContig	1226	1425
	540	Digitaria sanguinalis	DHP_A41_8	gDNAContig	1226	1425
	541	Digitaria sanguinalis	DHP_A41_9	gDNAContig	1401	1600
	542	Digitaria sanguinalis	DHP_A41_9	gDNAContig	1401	1600
	543	Digitaria sanguinalis	DHP_A41_10	gDNAContig	1576	1775
	544	Digitaria sanguinalis	DHP_A41_10	gDNAContig	1576	1775
	545	Digitaria sanguinalis	DHP_A41_11	gDNAContig	1751	1950
	546	Digitaria sanguinalis	DHP_A41_11	gDNAContig	1751	1950
	547	Digitaria sanguinalis	DHP_A41_12	gDNAContig	1926	2125
	548	Digitaria sanguinalis	DHP_A41_12	gDNAContig	1926	2125
	549	Digitaria sanguinalis	DHP_A41_13	gDNAContig	2101	2300
	550	Digitaria sanguinalis	DHP_A41_13	gDNAContig	2101	2300
	551	Digitaria sanguinalis	DHP_A41_14	gDNAContig	2276	2475
	552	Digitaria sanguinalis	DHP_A41_14	gDNAContig	2276	2475
	553	Digitaria sanguinalis	DHP_A41_15	gDNAContig	2451	2650
	554	Digitaria sanguinalis	DHP_A41_15	gDNAContig	2451	2650
	555	Digitaria sanguinalis	DHP_A41_16	gDNAContig	2626	2825
	556	Digitaria sanguinalis	DHP_A41_16	gDNAContig	2626	2825
	557	Digitaria sanguinalis	DHP_A41_17	gDNAContig	2801	3000
	558	Digitaria sanguinalis	DHP_A41_17	gDNAContig	2801	3000
	559	Digitaria sanguinalis	DHP_A41_18	gDNAContig	2976	3175
	560	Digitaria sanguinalis	DHP_A41_18	gDNAContig	2976	3175
	561	Digitaria sanguinalis	DHP_A41_19	gDNAContig	3151	3350
	562	Digitaria sanguinalis	DHP_A41_19	gDNAContig	3151	3350
	563	Digitaria sanguinalis	DHP_A41_20	gDNAContig	3326	3525
	564	Digitaria sanguinalis	DHP_A41_20	gDNAContig	3326	3525
	565	Digitaria sanguinalis	DHP_A41_21	gDNAContig	3501	3700

(continued)

	SEQ ID NO	Species	Name   Reference	Type	Start	End
5	566	Digitaria sanguinalis	DHP_A41_21	gDNAContig	3501	3700
10	567	Digitaria sanguinalis	DHP_A41_22	gDNAContig	3676	3875
15	568	Digitaria sanguinalis	DHP_A41_22	gDNAContig	3676	3875
20	569	Digitaria sanguinalis	DHP_A42_1	cDNAContig	1	200
25	570	Digitaria sanguinalis	DHP_A42_1	cDNAContig	1	200
30	571	Digitaria sanguinalis	DHP_A42_2	cDNAContig	176	375
35	572	Digitaria sanguinalis	DHP_A42_2	cDNAContig	176	375
40	573	Digitaria sanguinalis	DHP_A42_3	cDNAContig	351	550
45	574	Digitaria sanguinalis	DHP_A42_3	cDNAContig	351	550
50	575	Digitaria sanguinalis	DHP_A42_4	cDNAContig	526	725
55	576	Digitaria sanguinalis	DHP_A42_4	cDNAContig	526	725
	577	Digitaria sanguinalis	DHP_A42_5	cDNAContig	701	900
	578	Digitaria sanguinalis	DHP_A42_5	cDNAContig	701	900
	579	Digitaria sanguinalis	DHP_A42_6	cDNAContig	876	1075
	580	Digitaria sanguinalis	DHP_A42_6	cDNAContig	876	1075
	581	Digitaria sanguinalis	DHP_A42_7	cDNAContig	1051	1250
	582	Digitaria sanguinalis	DHP_A42_7	cDNAContig	1051	1250
	583	Digitaria sanguinalis	DHP_A42_8	cDNAContig	1226	1425
	584	Digitaria sanguinalis	DHP_A42_8	cDNAContig	1226	1425
	585	Euphorbia heterophylla	DHP_A35_1	gDNAContig	1	200
	586	Euphorbia heterophylla	DHP_A35_1	gDNAContig	1	200
	587	Euphorbia heterophylla	DHP_A35_2	gDNAContig	176	375
	588	Euphorbia heterophylla	DHP_A35_2	gDNAContig	176	375
	589	Euphorbia heterophylla	DHP_A35_3	gDNAContig	351	550
	590	Euphorbia heterophylla	DHP_A35_3	gDNAContig	351	550
	591	Euphorbia heterophylla	DHP_A35_4	gDNAContig	526	725
	592	Euphorbia heterophylla	DHP_A35_4	gDNAContig	526	725
	593	Euphorbia heterophylla	DHP_A35_5	gDNAContig	701	900
	594	Euphorbia heterophylla	DHP_A35_5	gDNAContig	701	900
	595	Euphorbia heterophylla	DHP_A35_6	gDNAContig	876	1075
	596	Euphorbia heterophylla	DHP_A35_6	gDNAContig	876	1075
	597	Euphorbia heterophylla	DHP_A35_7	gDNAContig	1051	1250
	598	Euphorbia heterophylla	DHP_A35_7	gDNAContig	1051	1250
	599	Euphorbia heterophylla	DHP_A35_8	gDNAContig	1226	1425
	600	Euphorbia heterophylla	DHP_A35_8	gDNAContig	1226	1425
	601	Euphorbia heterophylla	DHP_A35_9	gDNAContig	1401	1600
	602	Euphorbia heterophylla	DHP_A35_9	gDNAContig	1401	1600
	603	Euphorbia heterophylla	DHP_A35_10	gDNAContig	1576	1775

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	SEQ ID NO	Species	Name   Reference	Type	Start	End
5	604	Euphorbia heterophylla	DHP_A35_10	gDNAContig	1576	1775
10	605	Euphorbia heterophylla	DHP_A35_11	gDNAContig	1751	1950
15	606	Euphorbia heterophylla	DHP_A35_11	gDNAContig	1751	1950
20	607	Euphorbia heterophylla	DHP_A35_12	gDNAContig	1926	2125
25	608	Euphorbia heterophylla	DHP_A35_12	gDNAContig	1926	2125
30	609	Euphorbia heterophylla	DHP_A35_13	gDNAContig	2101	2300
35	610	Euphorbia heterophylla	DHP_A35_13	gDNAContig	2101	2300
40	611	Euphorbia heterophylla	DHP_A35_14	gDNAContig	2276	2475
45	612	Euphorbia heterophylla	DHP_A35_14	gDNAContig	2276	2475
50	613	Euphorbia heterophylla	DHP_A35_15	gDNAContig	2451	2650
55	614	Euphorbia heterophylla	DHP_A35_15	gDNAContig	2451	2650
	615	Euphorbia heterophylla	DHP_A35_16	gDNAContig	2626	2825
	616	Euphorbia heterophylla	DHP_A35_16	gDNAContig	2626	2825
	617	Euphorbia heterophylla	DHP_A35_17	gDNAContig	2801	3000
	618	Euphorbia heterophylla	DHP_A35_17	gDNAContig	2801	3000
	619	Euphorbia heterophylla	DHP_A35_18	gDNAContig	2976	3175
	620	Euphorbia heterophylla	DHP_A35_18	gDNAContig	2976	3175
	621	Euphorbia heterophylla	DHP_A35_19	gDNAContig	3151	3350
	622	Euphorbia heterophylla	DHP_A35_19	gDNAContig	3151	3350
	623	Euphorbia heterophylla	DHP_A35_20	gDNAContig	3326	3525
	624	Euphorbia heterophylla	DHP_A35_20	gDNAContig	3326	3525
	625	Euphorbia heterophylla	DHP_A35_21	gDNAContig	3501	3700
	626	Euphorbia heterophylla	DHP_A35_21	gDNAContig	3501	3700
	627	Euphorbia heterophylla	DHP_A35_22	gDNAContig	3676	3875
	628	Euphorbia heterophylla	DHP_A35_22	gDNAContig	3676	3875
	629	Euphorbia heterophylla	DHP_A35_23	gDNAContig	3851	4050
	630	Euphorbia heterophylla	DHP_A35_23	gDNAContig	3851	4050
	631	Euphorbia heterophylla	DHP_A35_24	gDNAContig	4026	4225
	632	Euphorbia heterophylla	DHP_A35_24	gDNAContig	4026	4225
	633	Euphorbia heterophylla	DHP_A35_25	gDNAContig	4201	4400
	634	Euphorbia heterophylla	DHP_A35_25	gDNAContig	4201	4400
	635	Euphorbia heterophylla	DHP_A37_1	cDNAContig	1	200
	636	Euphorbia heterophylla	DHP_A37_1	cDNAContig	1	200
	637	Euphorbia heterophylla	DHP_A37_2	cDNAContig	176	375
	638	Euphorbia heterophylla	DHP_A37_2	cDNAContig	176	375
	639	Euphorbia heterophylla	DHP_A37_3	cDNAContig	351	550
	640	Euphorbia heterophylla	DHP_A37_3	cDNAContig	351	550
	641	Euphorbia heterophylla	DHP_A37_4	cDNAContig	526	725

(continued)

	SEQ ID NO	Species	Name   Reference	Type	Start	End
5	642	Euphorbia heterophylla	DHP_A37_4	cDNAContig	526	725
10	643	Euphorbia heterophylla	DHP_A37_5	cDNAContig	701	900
15	644	Euphorbia heterophylla	DHP_A37_5	cDNAContig	701	900
20	645	Euphorbia heterophylla	DHP_A37_6	cDNAContig	876	1075
25	646	Euphorbia heterophylla	DHP_A37_6	cDNAContig	876	1075
30	647	Euphorbia heterophylla	DHP_A37_7	cDNAContig	1051	1250
35	648	Euphorbia heterophylla	DHP_A37_7	cDNAContig	1051	1250
40	649	Euphorbia heterophylla	DHP_A37_8	cDNAContig	1226	1425
45	650	Euphorbia heterophylla	DHP_A37_8	cDNAContig	1226	1425
50	651	Euphorbia heterophylla	DHP_A36_1	cDNAContig	1	200
55	652	Euphorbia heterophylla	DHP_A36_1	cDNAContig	1	200
	653	Euphorbia heterophylla	DHP_A36_2	cDNAContig	176	375
	654	Euphorbia heterophylla	DHP_A36_2	cDNAContig	176	375
	655	Euphorbia heterophylla	DHP_A36_3	cDNAContig	351	550
	656	Euphorbia heterophylla	DHP_A36_3	cDNAContig	351	550
	657	Euphorbia heterophylla	DHP_A36_4	cDNAContig	526	725
	658	Euphorbia heterophylla	DHP_A36_4	cDNAContig	526	725
	659	Euphorbia heterophylla	DHP_A36_5	cDNAContig	701	900
	660	Euphorbia heterophylla	DHP_A36_5	cDNAContig	701	900
	661	Euphorbia heterophylla	DHP_A36_6	cDNAContig	876	1075
	662	Euphorbia heterophylla	DHP_A36_6	cDNAContig	876	1075
	663	Euphorbia heterophylla	DHP_A36_7	cDNAContig	1051	1250
	664	Euphorbia heterophylla	DHP_A36_7	cDNAContig	1051	1250
	665	Euphorbia heterophylla	DHP_A36_8	cDNAContig	1226	1425
	666	Euphorbia heterophylla	DHP_A36_8	cDNAContig	1226	1425
	667	Euphorbia heterophylla	DHP_A40_1	gDNAContig	1	200
	668	Euphorbia heterophylla	DHP_A40_1	gDNAContig	1	200
	669	Euphorbia heterophylla	DHP_A40_2	gDNAContig	176	375
	670	Euphorbia heterophylla	DHP_A40_2	gDNAContig	176	375
	671	Euphorbia heterophylla	DHP_A39_1	gDNAContig	1	200
	672	Euphorbia heterophylla	DHP_A39_1	gDNAContig	1	200
	673	Euphorbia heterophylla	DHP_A39_2	gDNAContig	176	375
	674	Euphorbia heterophylla	DHP_A39_2	gDNAContig	176	375
	675	Euphorbia heterophylla	DHP_A38_1	cDNAContig	1	200
	676	Euphorbia heterophylla	DHP_A38_1	cDNAContig	1	200
	677	Euphorbia heterophylla	DHP_A38_2	cDNAContig	176	375
	678	Euphorbia heterophylla	DHP_A38_2	cDNAContig	176	375
	679	Euphorbia heterophylla	DHP_A38_3	cDNAContig	351	550

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	SEQ ID NO	Species	Name   Reference	Type	Start	End
5	680	Euphorbia heterophylla	DHP_A38_3	cDNAContig	351	550
10	681	Euphorbia heterophylla	DHP_A34_1	gDNAContig	1	200
15	682	Euphorbia heterophylla	DHP_A34_1	gDNAContig	1	200
20	683	Euphorbia heterophylla	DHP_A34_2	gDNAContig	176	375
25	684	Euphorbia heterophylla	DHP_A34_2	gDNAContig	176	375
30	685	Euphorbia heterophylla	DHP_A34_3	gDNAContig	351	550
35	686	Euphorbia heterophylla	DHP_A34_3	gDNAContig	351	550
40	687	Euphorbia heterophylla	DHP_A34_4	gDNAContig	526	725
45	688	Euphorbia heterophylla	DHP_A34_4	gDNAContig	526	725
50	689	Euphorbia heterophylla	DHP_A34_5	gDNAContig	701	900
55	690	Euphorbia heterophylla	DHP_A34_5	gDNAContig	701	900
	691	Euphorbia heterophylla	DHP_A34_6	gDNAContig	876	1075
	692	Euphorbia heterophylla	DHP_A34_6	gDNAContig	876	1075
	693	Euphorbia heterophylla	DHP_A34_7	gDNAContig	1051	1250
	694	Euphorbia heterophylla	DHP_A34_7	gDNAContig	1051	1250
	695	Euphorbia heterophylla	DHP_A34_8	gDNAContig	1226	1425
	696	Euphorbia heterophylla	DHP_A34_8	gDNAContig	1226	1425
	697	Euphorbia heterophylla	DHP_A34_9	gDNAContig	1401	1600
	698	Euphorbia heterophylla	DHP_A34_9	gDNAContig	1401	1600
	699	Euphorbia heterophylla	DHP_A34_10	gDNAContig	1576	1775
	700	Euphorbia heterophylla	DHP_A34_10	gDNAContig	1576	1775
	701	Euphorbia heterophylla	DHP_A34_11	gDNAContig	1751	1950
	702	Euphorbia heterophylla	DHP_A34_11	gDNAContig	1751	1950
	703	Euphorbia heterophylla	DHP_A34_12	gDNAContig	1926	2125
	704	Euphorbia heterophylla	DHP_A34_12	gDNAContig	1926	2125
	705	Euphorbia heterophylla	DHP_A34_13	gDNAContig	2101	2300
	706	Euphorbia heterophylla	DHP_A34_13	gDNAContig	2101	2300
	707	Euphorbia heterophylla	DHP_A34_14	gDNAContig	2276	2475
	708	Euphorbia heterophylla	DHP_A34_14	gDNAContig	2276	2475
	709	Euphorbia heterophylla	DHP_A34_15	gDNAContig	2451	2650
	710	Euphorbia heterophylla	DHP_A34_15	gDNAContig	2451	2650
	711	Euphorbia heterophylla	DHP_A34_16	gDNAContig	2626	2825
	712	Euphorbia heterophylla	DHP_A34_16	gDNAContig	2626	2825
	713	Euphorbia heterophylla	DHP_A34_17	gDNAContig	2801	3000
	714	Euphorbia heterophylla	DHP_A34_17	gDNAContig	2801	3000
	715	Euphorbia heterophylla	DHP_A34_18	gDNAContig	2976	3175
	716	Euphorbia heterophylla	DHP_A34_18	gDNAContig	2976	3175
	717	Euphorbia heterophylla	DHP_A34_19	gDNAContig	3151	3350

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	SEQ ID NO	Species	Name   Reference	Type	Start	End
5	718	Euphorbia heterophylla	DHP_A34_19	gDNAContig	3151	3350
10	719	Euphorbia heterophylla	DHP_A34_20	gDNAContig	3326	3525
15	720	Euphorbia heterophylla	DHP_A34_20	gDNAContig	3326	3525
20	721	Euphorbia heterophylla	DHP_A34_21	gDNAContig	3501	3700
25	722	Euphorbia heterophylla	DHP_A34_21	gDNAContig	3501	3700
30	723	Euphorbia heterophylla	DHP_A34_22	gDNAContig	3676	3875
35	724	Euphorbia heterophylla	DHP_A34_22	gDNAContig	3676	3875
40	725	Euphorbia heterophylla	DHP_A34_23	gDNAContig	3851	4050
45	726	Euphorbia heterophylla	DHP_A34_23	gDNAContig	3851	4050
50	727	Euphorbia heterophylla	DHP_A34_24	gDNAContig	4026	4225
55	728	Euphorbia heterophylla	DHP_A34_24	gDNAContig	4026	4225
	729	Euphorbia heterophylla	DHP_A34_25	gDNAContig	4201	4400
	730	Euphorbia heterophylla	DHP_A34_25	gDNAContig	4201	4400
	731	Euphorbia heterophylla	DHP_A34_26	gDNAContig	4376	4575
	732	Euphorbia heterophylla	DHP_A34_26	gDNAContig	4376	4575
	733	Kochia scoparia	DHP_A43_1	gDNAContig	1	200
	734	Kochia scoparia	DHP_A43_1	gDNAContig	1	200
	735	Kochia scoparia	DHP_A43_2	gDNAContig	176	375
	736	Kochia scoparia	DHP_A43_2	gDNAContig	176	375
	737	Kochia scoparia	DHP_A43_3	gDNAContig	351	550
	738	Kochia scoparia	DHP_A43_3	gDNAContig	351	550
	739	Kochia scoparia	DHP_A43_4	gDNAContig	526	725
	740	Kochia scoparia	DHP_A43_4	gDNAContig	526	725
	741	Kochia scoparia	DHP_A43_5	gDNAContig	701	900
	742	Kochia scoparia	DHP_A43_5	gDNAContig	701	900
	743	Kochia scoparia	DHP_A43_6	gDNAContig	876	1075
	744	Kochia scoparia	DHP_A43_6	gDNAContig	876	1075
	745	Kochia scoparia	DHP_A43_7	gDNAContig	1051	1250
	746	Kochia scoparia	DHP_A43_7	gDNAContig	1051	1250
	747	Kochia scoparia	DHP_A43_8	gDNAContig	1226	1425
	748	Kochia scoparia	DHP_A43_8	gDNAContig	1226	1425
	749	Kochia scoparia	DHP_A43_9	gDNAContig	1401	1600
	750	Kochia scoparia	DHP_A43_9	gDNAContig	1401	1600
	751	Kochia scoparia	DHP_A43_10	gDNAContig	1576	1775
	752	Kochia scoparia	DHP_A43_10	gDNAContig	1576	1775
	753	Kochia scoparia	DHP_A43_11	gDNAContig	1751	1950
	754	Kochia scoparia	DHP_A43_11	gDNAContig	1751	1950
	755	Kochia scoparia	DHP_A43_12	gDNAContig	1926	2125

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	SEQ ID NO	Species	Name   Reference	Type	Start	End
5	756	Kochia scoparia	DHP_A43_12	gDNAContig	1926	2125
10	757	Kochia scoparia	DHP_A43_13	gDNAContig	2101	2300
15	758	Kochia scoparia	DHP_A43_13	gDNAContig	2101	2300
20	759	Kochia scoparia	DHP_A43_14	gDNAContig	2276	2475
25	760	Kochia scoparia	DHP_A43_14	gDNAContig	2276	2475
30	761	Kochia scoparia	DHP_A43_15	gDNAContig	2451	2650
35	762	Kochia scoparia	DHP_A43_15	gDNAContig	2451	2650
40	763	Kochia scoparia	DHP_A43_16	gDNAContig	2626	2825
45	764	Kochia scoparia	DHP_A43_16	gDNAContig	2626	2825
50	765	Kochia scoparia	DHP_A43_17	gDNAContig	2801	3000
55	766	Kochia scoparia	DHP_A43_17	gDNAContig	2801	3000
	767	Kochia scoparia	DHP_A43_18	gDNAContig	2976	3175
	768	Kochia scoparia	DHP_A43_18	gDNAContig	2976	3175
	769	Kochia scoparia	DHP_A43_19	gDNAContig	3151	3350
	770	Kochia scoparia	DHP_A43_19	gDNAContig	3151	3350
	771	Kochia scoparia	DHP_A43_20	gDNAContig	3326	3525
	772	Kochia scoparia	DHP_A43_20	gDNAContig	3326	3525
	773	Kochia scoparia	DHP_A43_21	gDNAContig	3501	3700
	774	Kochia scoparia	DHP_A43_21	gDNAContig	3501	3700
	775	Kochia scoparia	DHP_A43_22	gDNAContig	3676	3875
	776	Kochia scoparia	DHP_A43_22	gDNAContig	3676	3875
	777	Kochia scoparia	DHP_A46_1	cDNAContig	1	200
	778	Kochia scoparia	DHP_A46_1	cDNAContig	1	200
	779	Kochia scoparia	DHP_A46_2	cDNAContig	176	375
	780	Kochia scoparia	DHP_A46_2	cDNAContig	176	375
	781	Kochia scoparia	DHP_A46_3	cDNAContig	351	550
	782	Kochia scoparia	DHP_A46_3	cDNAContig	351	550
	783	Kochia scoparia	DHP_A46_4	cDNAContig	526	725
	784	Kochia scoparia	DHP_A46_4	cDNAContig	526	725
	785	Kochia scoparia	DHP_A46_5	cDNAContig	701	900
	786	Kochia scoparia	DHP_A46_5	cDNAContig	701	900
	787	Kochia scoparia	DHP_A46_6	cDNAContig	876	1075
	788	Kochia scoparia	DHP_A46_6	cDNAContig	876	1075
	789	Kochia scoparia	DHP_A46_7	cDNAContig	1051	1250
	790	Kochia scoparia	DHP_A46_7	cDNAContig	1051	1250
	791	Kochia scoparia	DHP_A46_8	cDNAContig	1226	1425
	792	Kochia scoparia	DHP_A46_8	cDNAContig	1226	1425
	793	Kochia scoparia	DHP_A44_1	gDNAContig	1	200

(continued)

	SEQ ID NO	Species	Name   Reference	Type	Start	End
5	794	Kochia scoparia	DHP_A44_1	gDNAContig	1	200
10	795	Kochia scoparia	DHP_A44_2	gDNAContig	176	375
15	796	Kochia scoparia	DHP_A44_2	gDNAContig	176	375
20	797	Kochia scoparia	DHP_A44_3	gDNAContig	351	550
25	798	Kochia scoparia	DHP_A44_3	gDNAContig	351	550
30	799	Kochia scoparia	DHP_A44_4	gDNAContig	526	725
35	800	Kochia scoparia	DHP_A44_4	gDNAContig	526	725
40	801	Kochia scoparia	DHP_A44_5	gDNAContig	701	900
45	802	Kochia scoparia	DHP_A44_5	gDNAContig	701	900
50	803	Kochia scoparia	DHP_A44_6	gDNAContig	876	1075
55	804	Kochia scoparia	DHP_A44_6	gDNAContig	876	1075
	805	Kochia scoparia	DHP_A44_7	gDNAContig	1051	1250
	806	Kochia scoparia	DHP_A44_7	gDNAContig	1051	1250
	807	Kochia scoparia	DHP_A44_8	gDNAContig	1226	1425
	808	Kochia scoparia	DHP_A44_8	gDNAContig	1226	1425
	809	Kochia scoparia	DHP_A44_9	gDNAContig	1401	1600
	810	Kochia scoparia	DHP_A44_9	gDNAContig	1401	1600
	811	Kochia scoparia	DHP_A44_10	gDNAContig	1576	1775
	812	Kochia scoparia	DHP_A44_10	gDNAContig	1576	1775
	813	Kochia scoparia	DHP_A44_11	gDNAContig	1751	1950
	814	Kochia scoparia	DHP_A44_11	gDNAContig	1751	1950
	815	Kochia scoparia	DHP_A44_12	gDNAContig	1926	2125
	816	Kochia scoparia	DHP_A44_12	gDNAContig	1926	2125
	817	Kochia scoparia	DHP_A44_13	gDNAContig	2101	2300
	818	Kochia scoparia	DHP_A44_13	gDNAContig	2101	2300
	819	Kochia scoparia	DHP_A44_14	gDNAContig	2276	2475
	820	Kochia scoparia	DHP_A44_14	gDNAContig	2276	2475
	821	Kochia scoparia	DHP_A44_15	gDNAContig	2451	2650
	822	Kochia scoparia	DHP_A44_15	gDNAContig	2451	2650
	823	Kochia scoparia	DHP_A44_16	gDNAContig	2626	2825
	824	Kochia scoparia	DHP_A44_16	gDNAContig	2626	2825
	825	Kochia scoparia	DHP_A44_17	gDNAContig	2801	3000
	826	Kochia scoparia	DHP_A44_17	gDNAContig	2801	3000
	827	Kochia scoparia	DHP_A44_18	gDNAContig	2976	3175
	828	Kochia scoparia	DHP_A44_18	gDNAContig	2976	3175
	829	Kochia scoparia	DHP_A44_19	gDNAContig	3151	3350
	830	Kochia scoparia	DHP_A44_19	gDNAContig	3151	3350
	831	Lolium multiflorum	DHP_A47_1	gDNAContig	1	200

(continued)

	SEQ ID NO	Species	Name   Reference	Type	Start	End
5	832	Lolium multiflorum	DHP_A47_1	gDNAContig	1	200
10	833	Lolium multiflorum	DHP_A47_2	gDNAContig	176	375
15	834	Lolium multiflorum	DHP_A47_2	gDNAContig	176	375
20	835	Lolium multiflorum	DHP_A47_3	gDNAContig	351	550
25	836	Lolium multiflorum	DHP_A47_3	gDNAContig	351	550
30	837	Lolium multiflorum	DHP_A47_4	gDNAContig	526	725
35	838	Lolium multiflorum	DHP_A47_4	gDNAContig	526	725
40	839	Lolium multiflorum	DHP_A47_5	gDNAContig	701	900
45	840	Lolium multiflorum	DHP_A47_5	gDNAContig	701	900
50	841	Lolium multiflorum	DHP_A47_6	gDNAContig	876	1075
55	842	Lolium multiflorum	DHP_A47_6	gDNAContig	876	1075
	843	Lolium multiflorum	DHP_A47_7	gDNAContig	1051	1250
	844	Lolium multiflorum	DHP_A47_7	gDNAContig	1051	1250
	845	Lolium multiflorum	DHP_A47_8	gDNAContig	1226	1425
	846	Lolium multiflorum	DHP_A47_8	gDNAContig	1226	1425
	847	Lolium multiflorum	DHP_A47_9	gDNAContig	1401	1600
	848	Lolium multiflorum	DHP_A47_9	gDNAContig	1401	1600
	849	Lolium multiflorum	DHP_A47_10	gDNAContig	1576	1775
	850	Lolium multiflorum	DHP_A47_10	gDNAContig	1576	1775
	851	Lolium multiflorum	DHP_A47_11	gDNAContig	1751	1950
	852	Lolium multiflorum	DHP_A47_11	gDNAContig	1751	1950
	853	Lolium multiflorum	DHP_A47_12	gDNAContig	1926	2125
	854	Lolium multiflorum	DHP_A47_12	gDNAContig	1926	2125
	855	Lolium multiflorum	DHP_A47_13	gDNAContig	2101	2300
	856	Lolium multiflorum	DHP_A47_13	gDNAContig	2101	2300
	857	Lolium multiflorum	DHP_A47_14	gDNAContig	2276	2475
	858	Lolium multiflorum	DHP_A47_14	gDNAContig	2276	2475
	859	Lolium multiflorum	DHP_A47_15	gDNAContig	2451	2650
	860	Lolium multiflorum	DHP_A47_15	gDNAContig	2451	2650
	861	Lolium multiflorum	DHP_A47_16	gDNAContig	2626	2825
	862	Lolium multiflorum	DHP_A47_16	gDNAContig	2626	2825
	863	Lolium multiflorum	DHP_A47_17	gDNAContig	2801	3000
	864	Lolium multiflorum	DHP_A47_17	gDNAContig	2801	3000
	865	Lolium multiflorum	DHP_A47_18	gDNAContig	2976	3175
	866	Lolium multiflorum	DHP_A47_18	gDNAContig	2976	3175
	867	Lolium multiflorum	DHP_A47_19	gDNAContig	3151	3350
	868	Lolium multiflorum	DHP_A47_19	gDNAContig	3151	3350
	869	Lolium multiflorum	DHP_A47_20	gDNAContig	3326	3525

(continued)

	SEQ ID NO	Species	Name   Reference	Type	Start	End
5	870	Lolium multiflorum	DHP_A47_20	gDNAContig	3326	3525
10	871	Lolium multiflorum	DHP_A47_21	gDNAContig	3501	3700
15	872	Lolium multiflorum	DHP_A47_21	gDNAContig	3501	3700
20	873	Lolium multiflorum	DHP_A47_22	gDNAContig	3676	3875
25	874	Lolium multiflorum	DHP_A47_22	gDNAContig	3676	3875
30	875	Lolium multiflorum	DHP_A48_1	cDNAContig	1	200
35	876	Lolium multiflorum	DHP_A48_1	cDNAContig	1	200
40	877	Lolium multiflorum	DHP_A48_2	cDNAContig	176	375
45	878	Lolium multiflorum	DHP_A48_2	cDNAContig	176	375
50	879	Lolium multiflorum	DHP_A48_3	cDNAContig	351	550
55	880	Lolium multiflorum	DHP_A48_3	cDNAContig	351	550
	881	Lolium multiflorum	DHP_A48_4	cDNAContig	526	725
	882	Lolium multiflorum	DHP_A48_4	cDNAContig	526	725
	883	Lolium multiflorum	DHP_A48_5	cDNAContig	701	900
	884	Lolium multiflorum	DHP_A48_5	cDNAContig	701	900
	885	Lolium multiflorum	DHP_A48_6	cDNAContig	876	1075
	886	Lolium multiflorum	DHP_A48_6	cDNAContig	876	1075
	887	Lolium multiflorum	DHP_A48_7	cDNAContig	1051	1250
	888	Lolium multiflorum	DHP_A48_7	cDNAContig	1051	1250
	889	Lolium multiflorum	DHP_A48_8	cDNAContig	1226	1425
	890	Lolium multiflorum	DHP_A48_8	cDNAContig	1226	1425
	891	Lolium multiflorum	DHP_A51_1	gDNAContig	1	200
	892	Lolium multiflorum	DHP_A51_1	gDNAContig	1	200
	893	Lolium multiflorum	DHP_A50_1	cDNAContig	1	200
	894	Lolium multiflorum	DHP_A50_1	cDNAContig	1	200
	895	Lolium multiflorum	DHP_A50_2	cDNAContig	176	375
	896	Lolium multiflorum	DHP_A50_2	cDNAContig	176	375
	897	Lolium multiflorum	DHP_A50_3	cDNAContig	351	550
	898	Lolium multiflorum	DHP_A50_3	cDNAContig	351	550
	899	Lolium multiflorum	DHP_A50_4	cDNAContig	526	725
	900	Lolium multiflorum	DHP_A50_4	cDNAContig	526	725
	901	Lolium multiflorum	DHP_A53_1	gDNAContig	1	200
	902	Lolium multiflorum	DHP_A53_1	gDNAContig	1	200
	903	Lolium multiflorum	DHP_A52_1	cDNAContig	1	200
	904	Lolium multiflorum	DHP_A52_1	cDNAContig	1	200
	905	Lolium multiflorum	DHP_A54_1	cDNAContig	1	150
	906	Lolium multiflorum	DHP_A54_1	cDNAContig	1	150

**Table 3**

SEQ ID NO	Gene	# Species	Species
5	907	DHP	5 <i>Amaranthus hybridus, Amaranthus palmeri, Amaranthus rudis, Amaranthus viridis, Kochia scoparia</i>
10	908	DHP	5 <i>Amaranthus hybridus, Amaranthus palmeri, Amaranthus rudis, Amaranthus viridis, Kochia scoparia</i>
15	909	DHP	5 <i>Amaranthus hybridus, Amaranthus palmeri, Amaranthus rudis, Amaranthus viridis, Kochia scoparia</i>
20	910	DHP	5 <i>Amaranthus lividus, Amaranthus palmeri, Amaranthus rudis, Amaranthus viridis, Kochia scoparia</i>
25	911	DHP	5 <i>Amaranthus lividus, Amaranthus palmeri, Amaranthus rudis, Amaranthus viridis, Kochia scoparia</i>
30	912	DHP	5 <i>Amaranthus hybridus, Amaranthus palmeri, Amaranthus rudis, Amaranthus viridis, Kochia scoparia</i>
35	913	DHP	5 <i>Amaranthus hybridus, Amaranthus palmeri, Amaranthus rudis, Amaranthus viridis, Kochia scoparia</i>
40	914	DHP	5 <i>Amaranthus hybridus, Amaranthus palmeri, Amaranthus rudis, Amaranthus viridis, Kochia scoparia</i>
45	915	DHP	5 <i>Amaranthus hybridus, Amaranthus palmeri, Amaranthus rudis, Amaranthus viridis, Kochia scoparia</i>
50	916	DHP	5 <i>Amaranthus hybridus, Amaranthus palmeri, Amaranthus rudis, Amaranthus viridis, Kochia scoparia</i>
55	917	DHP	5 <i>Amaranthus lividus, Amaranthus palmeri, Amaranthus rudis, Amaranthus viridis, Kochia scoparia</i>
	918	DHP	5 <i>Amaranthus hybridus, Amaranthus palmeri, Amaranthus rudis, Amaranthus viridis, Kochia scoparia</i>
	919	DHP	5 <i>Amaranthus hybridus, Amaranthus palmeri, Amaranthus rudis, Amaranthus viridis, Kochia scoparia</i>
	920	DHP	5 <i>Amaranthus hybridus, Amaranthus palmeri, Amaranthus rudis, Amaranthus viridis, Kochia scoparia</i>
	921	DHP	5 <i>Amaranthus hybridus, Amaranthus palmeri, Amaranthus rudis, Amaranthus viridis, Kochia scoparia</i>
	922	DHP	4 <i>Amaranthus lividus, Amaranthus palmeri, Amaranthus rudis, Amaranthus viridis</i>
	923	DHP	4 <i>Amaranthus lividus, Amaranthus palmeri, Amaranthus rudis, Amaranthus viridis</i>
	924	DHP	4 <i>Amaranthus lividus, Amaranthus palmeri, Amaranthus rudis, Amaranthus viridis</i>
	925	DHP	4 <i>Amaranthus lividus, Amaranthus palmeri, Amaranthus rudis, Amaranthus viridis</i>
	926	DHP	4 <i>Amaranthus hybridus, Amaranthus palmeri, Amaranthus rudis, Amaranthus viridis</i>
	927	DHP	4 <i>Amaranthus hybridus, Amaranthus palmeri, Amaranthus rudis, Amaranthus viridis</i>
	928	DHP	4 <i>Amaranthus lividus, Amaranthus palmeri, Amaranthus rudis, Amaranthus viridis</i>

(continued)

	SEQ ID NO	Gene	# Species	Species
5	929	DHP	4	<i>Amaranthus lividus</i> , <i>Amaranthus palmeri</i> , <i>Amaranthus rudis</i> , <i>Amaranthus viridis</i>
10	930	DHP	4	<i>Amaranthus lividus</i> , <i>Amaranthus palmeri</i> , <i>Amaranthus rudis</i> , <i>Amaranthus viridis</i>
15	931	DHP	4	<i>Amaranthus lividus</i> , <i>Amaranthus palmeri</i> , <i>Amaranthus rudis</i> , <i>Amaranthus viridis</i>
20	932	DHP	4	<i>Amaranthus lividus</i> , <i>Amaranthus palmeri</i> , <i>Amaranthus rudis</i> , <i>Amaranthus viridis</i>
25	933	DHP	4	<i>Amaranthus hybridus</i> , <i>Amaranthus palmeri</i> , <i>Amaranthus rudis</i> , <i>Amaranthus viridis</i>
30	934	DHP	4	<i>Amaranthus lividus</i> , <i>Amaranthus palmeri</i> , <i>Amaranthus rudis</i> , <i>Amaranthus viridis</i>
35	935	DHP	4	<i>Amaranthus lividus</i> , <i>Amaranthus palmeri</i> , <i>Amaranthus rudis</i> , <i>Amaranthus viridis</i>
40	936	DHP	4	<i>Amaranthus hybridus</i> , <i>Amaranthus palmeri</i> , <i>Amaranthus rudis</i> , <i>Amaranthus viridis</i>
45	937	DHP	4	<i>Amaranthus lividus</i> , <i>Amaranthus palmeri</i> , <i>Amaranthus rudis</i> , <i>Amaranthus viridis</i>
50	938	DHP	4	<i>Amaranthus lividus</i> , <i>Amaranthus palmeri</i> , <i>Amaranthus rudis</i> , <i>Amaranthus viridis</i>
55	939	DHP	4	<i>Amaranthus lividus</i> , <i>Amaranthus palmeri</i> , <i>Amaranthus rudis</i> , <i>Amaranthus viridis</i>
	940	DHP	4	<i>Amaranthus lividus</i> , <i>Amaranthus palmeri</i> , <i>Amaranthus rudis</i> , <i>Amaranthus viridis</i>
	941	DHP	4	<i>Amaranthus lividus</i> , <i>Amaranthus palmeri</i> , <i>Amaranthus rudis</i> , <i>Amaranthus viridis</i>
	942	DHP	4	<i>Amaranthus hybridus</i> , <i>Amaranthus palmeri</i> , <i>Amaranthus rudis</i> , <i>Amaranthus viridis</i>
	943	DHP	4	<i>Amaranthus lividus</i> , <i>Amaranthus palmeri</i> , <i>Amaranthus rudis</i> , <i>Amaranthus viridis</i>
	944	DHP	4	<i>Amaranthus lividus</i> , <i>Amaranthus palmeri</i> , <i>Amaranthus rudis</i> , <i>Amaranthus viridis</i>
	945	DHP	4	<i>Amaranthus lividus</i> , <i>Amaranthus palmeri</i> , <i>Amaranthus rudis</i> , <i>Amaranthus viridis</i>
	946	DHP	4	<i>Amaranthus lividus</i> , <i>Amaranthus palmeri</i> , <i>Amaranthus rudis</i> , <i>Amaranthus viridis</i>
	947	DHP	4	<i>Amaranthus lividus</i> , <i>Amaranthus palmeri</i> , <i>Amaranthus rudis</i> , <i>Amaranthus viridis</i>
	948	DHP	4	<i>Amaranthus lividus</i> , <i>Amaranthus palmeri</i> , <i>Amaranthus rudis</i> , <i>Amaranthus viridis</i>
	949	DHP	4	<i>Amaranthus lividus</i> , <i>Amaranthus palmeri</i> , <i>Amaranthus rudis</i> , <i>Amaranthus viridis</i>
	950	DHP	4	<i>Amaranthus lividus</i> , <i>Amaranthus palmeri</i> , <i>Amaranthus rudis</i> , <i>Amaranthus viridis</i>

(continued)

	SEQ ID NO	Gene	# Species	Species
5	951	DHP	4	<i>Amaranthus lividus</i> , <i>Amaranthus palmeri</i> , <i>Amaranthus rudis</i> , <i>Amaranthus viridis</i>
10	952	DHP	4	<i>Amaranthus lividus</i> , <i>Amaranthus palmeri</i> , <i>Amaranthus rudis</i> , <i>Amaranthus viridis</i>
15	953	DHP	4	<i>Amaranthus lividus</i> , <i>Amaranthus palmeri</i> , <i>Amaranthus rudis</i> , <i>Amaranthus viridis</i>
20	954	DHP	4	<i>Amaranthus lividus</i> , <i>Amaranthus palmeri</i> , <i>Amaranthus rudis</i> , <i>Amaranthus viridis</i>
25	955	DHP	4	<i>Amaranthus lividus</i> , <i>Amaranthus palmeri</i> , <i>Amaranthus rudis</i> , <i>Amaranthus viridis</i>
30	956	DHP	4	<i>Amaranthus lividus</i> , <i>Amaranthus rudis</i> , <i>Amaranthus viridis</i> , <i>Kochia scoparia</i>
35	957	DHP	4	<i>Amaranthus lividus</i> , <i>Amaranthus palmeri</i> , <i>Amaranthus rudis</i> , <i>Amaranthus viridis</i>
40	958	DHP	4	<i>Amaranthus hybridus</i> , <i>Amaranthus palmeri</i> , <i>Amaranthus rudis</i> , <i>Amaranthus viridis</i>
45	959	DHP	4	<i>Amaranthus palmeri</i> , <i>Amaranthus rudis</i> , <i>Amaranthus viridis</i> , <i>Kochia scoparia</i>
50	960	DHP	4	<i>Amaranthus hybridus</i> , <i>Amaranthus palmeri</i> , <i>Amaranthus rudis</i> , <i>Amaranthus viridis</i>
55	961	DHP	4	<i>Amaranthus lividus</i> , <i>Amaranthus palmeri</i> , <i>Amaranthus rudis</i> , <i>Amaranthus viridis</i>
	962	DHP	4	<i>Amaranthus lividus</i> , <i>Amaranthus palmeri</i> , <i>Amaranthus rudis</i> , <i>Amaranthus viridis</i>
	963	DHP	4	<i>Amaranthus lividus</i> , <i>Amaranthus palmeri</i> , <i>Amaranthus rudis</i> , <i>Amaranthus viridis</i>
	964	DHP	4	<i>Amaranthus hybridus</i> , <i>Amaranthus palmeri</i> , <i>Amaranthus rudis</i> , <i>Amaranthus viridis</i>
	965	DHP	4	<i>Amaranthus hybridus</i> , <i>Amaranthus palmeri</i> , <i>Amaranthus rudis</i> , <i>Amaranthus viridis</i>
	966	DHP	4	<i>Amaranthus hybridus</i> , <i>Amaranthus palmeri</i> , <i>Amaranthus rudis</i> , <i>Amaranthus viridis</i>
	967	DHP	4	<i>Amaranthus lividus</i> , <i>Amaranthus palmeri</i> , <i>Amaranthus rudis</i> , <i>Amaranthus viridis</i>
	968	DHP	4	<i>Amaranthus lividus</i> , <i>Amaranthus palmeri</i> , <i>Amaranthus rudis</i> , <i>Amaranthus viridis</i>
	969	DHP	4	<i>Amaranthus lividus</i> , <i>Amaranthus palmeri</i> , <i>Amaranthus rudis</i> , <i>Amaranthus viridis</i>
	970	DHP	4	<i>Amaranthus hybridus</i> , <i>Amaranthus palmeri</i> , <i>Amaranthus rudis</i> , <i>Amaranthus viridis</i>
	971	DHP	4	<i>Amaranthus lividus</i> , <i>Amaranthus palmeri</i> , <i>Amaranthus rudis</i> , <i>Amaranthus viridis</i>
	972	DHP	4	<i>Amaranthus lividus</i> , <i>Amaranthus palmeri</i> , <i>Amaranthus rudis</i> , <i>Amaranthus viridis</i>
	973	DHP	4	<i>Amaranthus lividus</i> , <i>Amaranthus palmeri</i> , <i>Amaranthus rudis</i> , <i>Amaranthus viridis</i>

(continued)

	SEQ ID NO	Gene	# Species	Species
5	974	DHP	4	<i>Amaranthus lividus</i> , <i>Amaranthus palmeri</i> , <i>Amaranthus rudis</i> , <i>Amaranthus viridis</i>
10	975	DHP	4	<i>Amaranthus lividus</i> , <i>Amaranthus palmeri</i> , <i>Amaranthus rudis</i> , <i>Amaranthus viridis</i>
15	976	DHP	4	<i>Amaranthus lividus</i> , <i>Amaranthus palmeri</i> , <i>Amaranthus rudis</i> , <i>Amaranthus viridis</i>
20	977	DHP	4	<i>Amaranthus lividus</i> , <i>Amaranthus palmeri</i> , <i>Amaranthus rudis</i> , <i>Amaranthus viridis</i>
25	978	DHP	4	<i>Amaranthus lividus</i> , <i>Amaranthus palmeri</i> , <i>Amaranthus rudis</i> , <i>Amaranthus viridis</i>
30	979	DHP	4	<i>Amaranthus lividus</i> , <i>Amaranthus palmeri</i> , <i>Amaranthus rudis</i> , <i>Amaranthus viridis</i>
35	980	DHP	4	<i>Amaranthus lividus</i> , <i>Amaranthus palmeri</i> , <i>Amaranthus rudis</i> , <i>Amaranthus viridis</i>
40	981	DHP	4	<i>Amaranthus hybridus</i> , <i>Amaranthus palmeri</i> , <i>Amaranthus rudis</i> , <i>Amaranthus viridis</i>
45	982	DHP	4	<i>Amaranthus lividus</i> , <i>Amaranthus palmeri</i> , <i>Amaranthus rudis</i> , <i>Amaranthus viridis</i>
50	983	DHP	4	<i>Amaranthus lividus</i> , <i>Amaranthus palmeri</i> , <i>Amaranthus rudis</i> , <i>Amaranthus viridis</i>
55	984	DHP	4	<i>Amaranthus lividus</i> , <i>Amaranthus palmeri</i> , <i>Amaranthus rudis</i> , <i>Amaranthus viridis</i>
	985	DHP	4	<i>Amaranthus hybridus</i> , <i>Amaranthus palmeri</i> , <i>Amaranthus rudis</i> , <i>Amaranthus viridis</i>
	986	DHP	4	<i>Amaranthus lividus</i> , <i>Amaranthus palmeri</i> , <i>Amaranthus rudis</i> , <i>Amaranthus viridis</i>
	987	DHP	4	<i>Amaranthus lividus</i> , <i>Amaranthus palmeri</i> , <i>Amaranthus rudis</i> , <i>Amaranthus viridis</i>
	988	DHP	4	<i>Amaranthus lividus</i> , <i>Amaranthus palmeri</i> , <i>Amaranthus rudis</i> , <i>Amaranthus viridis</i>
	989	DHP	4	<i>Amaranthus lividus</i> , <i>Amaranthus palmeri</i> , <i>Amaranthus rudis</i> , <i>Amaranthus viridis</i>
	990	DHP	4	<i>Amaranthus lividus</i> , <i>Amaranthus palmeri</i> , <i>Amaranthus rudis</i> , <i>Amaranthus viridis</i>
	991	DHP	4	<i>Amaranthus lividus</i> , <i>Amaranthus palmeri</i> , <i>Amaranthus rudis</i> , <i>Amaranthus viridis</i>
	992	DHP	4	<i>Amaranthus lividus</i> , <i>Amaranthus palmeri</i> , <i>Amaranthus rudis</i> , <i>Amaranthus viridis</i>
	993	DHP	4	<i>Amaranthus lividus</i> , <i>Amaranthus palmeri</i> , <i>Amaranthus rudis</i> , <i>Amaranthus viridis</i>
	994	DHP	4	<i>Amaranthus lividus</i> , <i>Amaranthus palmeri</i> , <i>Amaranthus rudis</i> , <i>Amaranthus viridis</i>
	995	DHP	4	<i>Amaranthus lividus</i> , <i>Amaranthus palmeri</i> , <i>Amaranthus rudis</i> , <i>Amaranthus viridis</i>

(continued)

SEQ ID NO	Gene	# Species	Species
5	996	DHP	4 <i>Amaranthus lividus, Amaranthus palmeri, Amaranthus rudis, Amaranthus viridis</i>
10	997	DHP	4 <i>Amaranthus lividus, Amaranthus palmeri, Amaranthus rudis, Amaranthus viridis</i>
15	998	DHP	4 <i>Amaranthus lividus, Amaranthus palmeri, Amaranthus rudis, Amaranthus viridis</i>
20	999	DHP	4 <i>Amaranthus lividus, Amaranthus palmeri, Amaranthus rudis, Amaranthus viridis</i>
25	1000	DHP	4 <i>Amaranthus palmeri, Amaranthus rudis, Amaranthus viridis, Kochia scoparia</i>
30	1001	DHP	4 <i>Amaranthus lividus, Amaranthus palmeri, Amaranthus rudis, Amaranthus viridis</i>
35	1002	DHP	4 <i>Amaranthus lividus, Amaranthus palmeri, Amaranthus rudis, Amaranthus viridis</i>
40	1003	DHP	4 <i>Amaranthus lividus, Amaranthus palmeri, Amaranthus rudis, Amaranthus viridis</i>
45	1004	DHP	4 <i>Amaranthus lividus, Amaranthus palmeri, Amaranthus rudis, Amaranthus viridis</i>
50	1005	DHP	4 <i>Amaranthus lividus, Amaranthus palmeri, Amaranthus rudis, Amaranthus viridis</i>
55	1006	DHP	4 <i>Amaranthus hybridus, Amaranthus palmeri, Amaranthus rudis, Amaranthus viridis</i>
	1007	DHP	4 <i>Amaranthus lividus, Amaranthus palmeri, Amaranthus rudis, Amaranthus viridis</i>
	1008	DHP	4 <i>Amaranthus lividus, Amaranthus palmeri, Amaranthus rudis, Amaranthus viridis</i>
	1009	DHP	4 <i>Amaranthus lividus, Amaranthus palmeri, Amaranthus rudis, Amaranthus viridis</i>
	1010	DHP	4 <i>Amaranthus lividus, Amaranthus palmeri, Amaranthus rudis, Amaranthus viridis</i>
	1011	DHP	4 <i>Amaranthus lividus, Amaranthus palmeri, Amaranthus rudis, Amaranthus viridis</i>
	1012	DHP	4 <i>Amaranthus lividus, Amaranthus palmeri, Amaranthus rudis, Amaranthus viridis</i>
	1013	DHP	4 <i>Amaranthus lividus, Amaranthus palmeri, Amaranthus rudis, Amaranthus viridis</i>
	1014	DHP	4 <i>Amaranthus lividus, Amaranthus palmeri, Amaranthus rudis, Amaranthus viridis</i>
	1015	DHP	4 <i>Amaranthus hybridus, Amaranthus palmeri, Amaranthus rudis, Amaranthus viridis</i>
	1016	DHP	4 <i>Amaranthus hybridus, Amaranthus palmeri, Amaranthus rudis, Amaranthus viridis</i>
	1017	DHP	4 <i>Amaranthus lividus, Amaranthus rudis, Amaranthus viridis, Kochia scoparia</i>
	1018	DHP	4 <i>Amaranthus lividus, Amaranthus palmeri, Amaranthus rudis, Amaranthus viridis</i>

(continued)

	SEQ ID NO	Gene	# Species	Species
5	1019	DHP	4	<i>Amaranthus lividus</i> , <i>Amaranthus palmeri</i> , <i>Amaranthus rudis</i> , <i>Amaranthus viridis</i>
10	1020	DHP	4	<i>Amaranthus hybridus</i> , <i>Amaranthus palmeri</i> , <i>Amaranthus rudis</i> , <i>Amaranthus viridis</i>
15	1021	DHP	4	<i>Amaranthus lividus</i> , <i>Amaranthus palmeri</i> , <i>Amaranthus rudis</i> , <i>Amaranthus viridis</i>
20	1022	DHP	4	<i>Amaranthus lividus</i> , <i>Amaranthus palmeri</i> , <i>Amaranthus rudis</i> , <i>Amaranthus viridis</i>
25	1023	DHP	4	<i>Amaranthus hybridus</i> , <i>Amaranthus palmeri</i> , <i>Amaranthus rudis</i> , <i>Amaranthus viridis</i>
30	1024	DHP	4	<i>Amaranthus palmeri</i> , <i>Amaranthus rudis</i> , <i>Amaranthus viridis</i> , <i>Kochia scoparia</i>
35	1025	DHP	4	<i>Amaranthus lividus</i> , <i>Amaranthus palmeri</i> , <i>Amaranthus rudis</i> , <i>Amaranthus viridis</i>
40	1026	DHP	4	<i>Amaranthus lividus</i> , <i>Amaranthus palmeri</i> , <i>Amaranthus rudis</i> , <i>Amaranthus viridis</i>
45	1027	DHP	4	<i>Amaranthus lividus</i> , <i>Amaranthus palmeri</i> , <i>Amaranthus rudis</i> , <i>Amaranthus viridis</i>
50	1028	DHP	4	<i>Amaranthus lividus</i> , <i>Amaranthus palmeri</i> , <i>Amaranthus rudis</i> , <i>Amaranthus viridis</i>
55	1029	DHP	4	<i>Amaranthus lividus</i> , <i>Amaranthus palmeri</i> , <i>Amaranthus rudis</i> , <i>Amaranthus viridis</i>
	1030	DHP	4	<i>Amaranthus lividus</i> , <i>Amaranthus palmeri</i> , <i>Amaranthus rudis</i> , <i>Amaranthus viridis</i>
	1031	DHP	4	<i>Amaranthus lividus</i> , <i>Amaranthus palmeri</i> , <i>Amaranthus rudis</i> , <i>Amaranthus viridis</i>
	1032	DHP	4	<i>Amaranthus lividus</i> , <i>Amaranthus palmeri</i> , <i>Amaranthus rudis</i> , <i>Amaranthus viridis</i>
	1033	DHP	4	<i>Amaranthus lividus</i> , <i>Amaranthus palmeri</i> , <i>Amaranthus rudis</i> , <i>Amaranthus viridis</i>
	1034	DHP	4	<i>Amaranthus lividus</i> , <i>Amaranthus palmeri</i> , <i>Amaranthus rudis</i> , <i>Amaranthus viridis</i>
	1035	DHP	4	<i>Amaranthus lividus</i> , <i>Amaranthus palmeri</i> , <i>Amaranthus rudis</i> , <i>Amaranthus viridis</i>
	1036	DHP	4	<i>Amaranthus hybridus</i> , <i>Amaranthus palmeri</i> , <i>Amaranthus rudis</i> , <i>Amaranthus viridis</i>
	1037	DHP	4	<i>Amaranthus lividus</i> , <i>Amaranthus palmeri</i> , <i>Amaranthus rudis</i> , <i>Amaranthus viridis</i>
	1038	DHP	4	<i>Amaranthus hybridus</i> , <i>Amaranthus palmeri</i> , <i>Amaranthus rudis</i> , <i>Amaranthus viridis</i>
	1039	DHP	4	<i>Amaranthus lividus</i> , <i>Amaranthus palmeri</i> , <i>Amaranthus rudis</i> , <i>Amaranthus viridis</i>
	1040	DHP	4	<i>Amaranthus lividus</i> , <i>Amaranthus palmeri</i> , <i>Amaranthus rudis</i> , <i>Amaranthus viridis</i>

(continued)

SEQ ID NO	Gene	# Species	Species
5	1041	DHP	4 <i>Amaranthus lividus, Amaranthus palmeri, Amaranthus rudis, Amaranthus viridis</i>
10	1042	DHP	4 <i>Amaranthus lividus, Amaranthus palmeri, Amaranthus rudis, Amaranthus viridis</i>
15	1043	DHP	4 <i>Amaranthus hybridus, Amaranthus palmeri, Amaranthus rudis, Amaranthus viridis</i>
20	1044	DHP	4 <i>Amaranthus hybridus, Amaranthus palmeri, Amaranthus rudis, Amaranthus viridis</i>
25	1045	DHP	4 <i>Amaranthus lividus, Amaranthus palmeri, Amaranthus rudis, Amaranthus viridis</i>
30	1046	DHP	4 <i>Amaranthus lividus, Amaranthus palmeri, Amaranthus rudis, Amaranthus viridis</i>
35	1047	DHP	4 <i>Amaranthus hybridus, Amaranthus palmeri, Amaranthus rudis, Amaranthus viridis</i>
40	1048	DHP	4 <i>Amaranthus lividus, Amaranthus palmeri, Amaranthus rudis, Amaranthus viridis</i>
45	1049	DHP	4 <i>Amaranthus lividus, Amaranthus palmeri, Amaranthus rudis, Amaranthus viridis</i>
50	1050	DHP	4 <i>Amaranthus lividus, Amaranthus palmeri, Amaranthus rudis, Amaranthus viridis</i>
55	1051	DHP	4 <i>Amaranthus lividus, Amaranthus palmeri, Amaranthus rudis, Amaranthus viridis</i>
	1052	DHP	4 <i>Amaranthus lividus, Amaranthus palmeri, Amaranthus rudis, Amaranthus viridis</i>
	1053	DHP	4 <i>Amaranthus lividus, Amaranthus palmeri, Amaranthus rudis, Amaranthus viridis</i>
	1054	DHP	4 <i>Amaranthus lividus, Amaranthus palmeri, Amaranthus rudis, Amaranthus viridis</i>
	1055	DHP	4 <i>Amaranthus lividus, Amaranthus palmeri, Amaranthus rudis, Amaranthus viridis</i>
	1056	DHP	4 <i>Amaranthus lividus, Amaranthus palmeri, Amaranthus rudis, Amaranthus viridis</i>
	1057	DHP	4 <i>Amaranthus lividus, Amaranthus palmeri, Amaranthus rudis, Amaranthus viridis</i>
	1058	DHP	4 <i>Amaranthus lividus, Amaranthus palmeri, Amaranthus rudis, Amaranthus viridis</i>
	1059	DHP	4 <i>Amaranthus lividus, Amaranthus palmeri, Amaranthus rudis, Amaranthus viridis</i>
	1060	DHP	4 <i>Amaranthus lividus, Amaranthus palmeri, Amaranthus rudis, Amaranthus viridis</i>
	1061	DHP	4 <i>Amaranthus lividus, Amaranthus palmeri, Amaranthus rudis, Amaranthus viridis</i>
	1062	DHP	4 <i>Amaranthus hybridus, Amaranthus palmeri, Amaranthus rudis, Amaranthus viridis</i>

(continued)

	SEQ ID NO	Gene	# Species	Species
5	1063	DHP	4	<i>Amaranthus lividus</i> , <i>Amaranthus palmeri</i> , <i>Amaranthus rudis</i> , <i>Amaranthus viridis</i>
10	1064	DHP	4	<i>Amaranthus lividus</i> , <i>Amaranthus palmeri</i> , <i>Amaranthus rudis</i> , <i>Amaranthus viridis</i>
15	1065	DHP	4	<i>Amaranthus lividus</i> , <i>Amaranthus palmeri</i> , <i>Amaranthus rudis</i> , <i>Amaranthus viridis</i>
20	1066	DHP	4	<i>Amaranthus lividus</i> , <i>Amaranthus palmeri</i> , <i>Amaranthus rudis</i> , <i>Amaranthus viridis</i>
25	1067	DHP	4	<i>Amaranthus lividus</i> , <i>Amaranthus palmeri</i> , <i>Amaranthus rudis</i> , <i>Amaranthus viridis</i>
30	1068	DHP	4	<i>Amaranthus lividus</i> , <i>Amaranthus palmeri</i> , <i>Amaranthus rudis</i> , <i>Amaranthus viridis</i>
35	1069	DHP	4	<i>Amaranthus lividus</i> , <i>Amaranthus palmeri</i> , <i>Amaranthus rudis</i> , <i>Amaranthus viridis</i>
40	1070	DHP	4	<i>Amaranthus lividus</i> , <i>Amaranthus palmeri</i> , <i>Amaranthus rudis</i> , <i>Amaranthus viridis</i>
45	1071	DHP	4	<i>Amaranthus lividus</i> , <i>Amaranthus palmeri</i> , <i>Amaranthus rudis</i> , <i>Amaranthus viridis</i>
50	1072	DHP	4	<i>Amaranthus lividus</i> , <i>Amaranthus palmeri</i> , <i>Amaranthus rudis</i> , <i>Amaranthus viridis</i>
55	1073	DHP	4	<i>Amaranthus lividus</i> , <i>Amaranthus palmeri</i> , <i>Amaranthus rudis</i> , <i>Amaranthus viridis</i>
	1074	DHP	4	<i>Amaranthus lividus</i> , <i>Amaranthus palmeri</i> , <i>Amaranthus rudis</i> , <i>Amaranthus viridis</i>
	1075	DHP	4	<i>Amaranthus lividus</i> , <i>Amaranthus palmeri</i> , <i>Amaranthus rudis</i> , <i>Amaranthus viridis</i>
	1076	DHP	4	<i>Amaranthus lividus</i> , <i>Amaranthus palmeri</i> , <i>Amaranthus rudis</i> , <i>Amaranthus viridis</i>
	1077	DHP	4	<i>Amaranthus lividus</i> , <i>Amaranthus palmeri</i> , <i>Amaranthus rudis</i> , <i>Amaranthus viridis</i>
	1078	DHP	4	<i>Amaranthus lividus</i> , <i>Amaranthus palmeri</i> , <i>Amaranthus rudis</i> , <i>Amaranthus viridis</i>
	1079	DHP	4	<i>Amaranthus lividus</i> , <i>Amaranthus palmeri</i> , <i>Amaranthus rudis</i> , <i>Amaranthus viridis</i>
	1080	DHP	4	<i>Amaranthus lividus</i> , <i>Amaranthus palmeri</i> , <i>Amaranthus rudis</i> , <i>Amaranthus viridis</i>
	1081	DHP	4	<i>Amaranthus lividus</i> , <i>Amaranthus palmeri</i> , <i>Amaranthus rudis</i> , <i>Amaranthus viridis</i>
	1082	DHP	4	<i>Amaranthus lividus</i> , <i>Amaranthus palmeri</i> , <i>Amaranthus rudis</i> , <i>Amaranthus viridis</i>
	1083	DHP	4	<i>Amaranthus lividus</i> , <i>Amaranthus palmeri</i> , <i>Amaranthus rudis</i> , <i>Amaranthus viridis</i>
	1084	DHP	4	<i>Amaranthus lividus</i> , <i>Amaranthus palmeri</i> , <i>Amaranthus rudis</i> , <i>Amaranthus viridis</i>

(continued)

	SEQ ID NO	Gene	# Species	Species
5	1085	DHP	4	<i>Amaranthus lividus</i> , <i>Amaranthus palmeri</i> , <i>Amaranthus rudis</i> , <i>Amaranthus viridis</i>
10	1086	DHP	4	<i>Amaranthus lividus</i> , <i>Amaranthus palmeri</i> , <i>Amaranthus rudis</i> , <i>Amaranthus viridis</i>
15	1087	DHP	4	<i>Amaranthus lividus</i> , <i>Amaranthus palmeri</i> , <i>Amaranthus rudis</i> , <i>Amaranthus viridis</i>
20	1088	DHP	4	<i>Amaranthus lividus</i> , <i>Amaranthus palmeri</i> , <i>Amaranthus rudis</i> , <i>Amaranthus viridis</i>
25	1089	DHP	4	<i>Amaranthus lividus</i> , <i>Amaranthus palmeri</i> , <i>Amaranthus rudis</i> , <i>Amaranthus viridis</i>
30	1090	DHP	4	<i>Amaranthus lividus</i> , <i>Amaranthus palmeri</i> , <i>Amaranthus rudis</i> , <i>Amaranthus viridis</i>
35	1091	DHP	4	<i>Amaranthus palmeri</i> , <i>Amaranthus rudis</i> , <i>Amaranthus viridis</i> , <i>Kochia scoparia</i>
40	1092	DHP	4	<i>Amaranthus lividus</i> , <i>Amaranthus palmeri</i> , <i>Amaranthus rudis</i> , <i>Amaranthus viridis</i>
45	1093	DHP	4	<i>Amaranthus lividus</i> , <i>Amaranthus palmeri</i> , <i>Amaranthus rudis</i> , <i>Amaranthus viridis</i>
50	1094	DHP	4	<i>Amaranthus lividus</i> , <i>Amaranthus palmeri</i> , <i>Amaranthus rudis</i> , <i>Amaranthus viridis</i>
55	1095	DHP	4	<i>Amaranthus lividus</i> , <i>Amaranthus palmeri</i> , <i>Amaranthus rudis</i> , <i>Amaranthus viridis</i>
	1096	DHP	4	<i>Amaranthus lividus</i> , <i>Amaranthus palmeri</i> , <i>Amaranthus rudis</i> , <i>Amaranthus viridis</i>
	1097	DHP	4	<i>Amaranthus lividus</i> , <i>Amaranthus palmeri</i> , <i>Amaranthus rudis</i> , <i>Amaranthus viridis</i>
	1098	DHP	4	<i>Amaranthus lividus</i> , <i>Amaranthus palmeri</i> , <i>Amaranthus rudis</i> , <i>Amaranthus viridis</i>
	1099	DHP	4	<i>Amaranthus lividus</i> , <i>Amaranthus palmeri</i> , <i>Amaranthus rudis</i> , <i>Amaranthus viridis</i>
	1100	DHP	4	<i>Amaranthus lividus</i> , <i>Amaranthus palmeri</i> , <i>Amaranthus rudis</i> , <i>Amaranthus viridis</i>
	1101	DHP	4	<i>Amaranthus hybridus</i> , <i>Amaranthus palmeri</i> , <i>Amaranthus rudis</i> , <i>Amaranthus viridis</i>
	1102	DHP	4	<i>Amaranthus lividus</i> , <i>Amaranthus palmeri</i> , <i>Amaranthus rudis</i> , <i>Amaranthus viridis</i>
	1103	DHP	4	<i>Amaranthus lividus</i> , <i>Amaranthus palmeri</i> , <i>Amaranthus rudis</i> , <i>Amaranthus viridis</i>
	1104	DHP	4	<i>Amaranthus lividus</i> , <i>Amaranthus palmeri</i> , <i>Amaranthus rudis</i> , <i>Amaranthus viridis</i>
	1105	DHP	4	<i>Amaranthus lividus</i> , <i>Amaranthus rudis</i> , <i>Amaranthus viridis</i> , <i>Kochia scoparia</i>
	1106	DHP	4	<i>Amaranthus lividus</i> , <i>Amaranthus palmeri</i> , <i>Amaranthus rudis</i> , <i>Amaranthus viridis</i>
	1107	DHP	4	<i>Amaranthus lividus</i> , <i>Amaranthus palmeri</i> , <i>Amaranthus rudis</i> , <i>Amaranthus viridis</i>

(continued)

	SEQ ID NO	Gene	# Species	Species
5	1108	DHP	4	<i>Amaranthus lividus</i> , <i>Amaranthus palmeri</i> , <i>Amaranthus rudis</i> , <i>Amaranthus viridis</i>
10	1109	DHP	4	<i>Amaranthus lividus</i> , <i>Amaranthus palmeri</i> , <i>Amaranthus rudis</i> , <i>Amaranthus viridis</i>
15	1110	DHP	4	<i>Amaranthus lividus</i> , <i>Amaranthus palmeri</i> , <i>Amaranthus rudis</i> , <i>Amaranthus viridis</i>
20	1111	DHP	4	<i>Amaranthus lividus</i> , <i>Amaranthus palmeri</i> , <i>Amaranthus rudis</i> , <i>Amaranthus viridis</i>
25	1112	DHP	4	<i>Amaranthus lividus</i> , <i>Amaranthus palmeri</i> , <i>Amaranthus rudis</i> , <i>Amaranthus viridis</i>
30	1113	DHP	4	<i>Amaranthus lividus</i> , <i>Amaranthus palmeri</i> , <i>Amaranthus rudis</i> , <i>Amaranthus viridis</i>
35	1114	DHP	4	<i>Amaranthus lividus</i> , <i>Amaranthus palmeri</i> , <i>Amaranthus rudis</i> , <i>Amaranthus viridis</i>
40	1115	DHP	4	<i>Amaranthus lividus</i> , <i>Amaranthus palmeri</i> , <i>Amaranthus rudis</i> , <i>Amaranthus viridis</i>
45	1116	DHP	4	<i>Amaranthus lividus</i> , <i>Amaranthus palmeri</i> , <i>Amaranthus rudis</i> , <i>Amaranthus viridis</i>
50	1117	DHP	4	<i>Amaranthus lividus</i> , <i>Amaranthus palmeri</i> , <i>Amaranthus rudis</i> , <i>Amaranthus viridis</i>
55	1118	DHP	4	<i>Amaranthus lividus</i> , <i>Amaranthus palmeri</i> , <i>Amaranthus rudis</i> , <i>Amaranthus viridis</i>
	1119	DHP	4	<i>Amaranthus hybridus</i> , <i>Amaranthus palmeri</i> , <i>Amaranthus rudis</i> , <i>Amaranthus viridis</i>
	1120	DHP	4	<i>Amaranthus hybridus</i> , <i>Amaranthus palmeri</i> , <i>Amaranthus rudis</i> , <i>Amaranthus viridis</i>
	1121	DHP	4	<i>Amaranthus lividus</i> , <i>Amaranthus palmeri</i> , <i>Amaranthus rudis</i> , <i>Amaranthus viridis</i>
	1122	DHP	4	<i>Amaranthus palmeri</i> , <i>Amaranthus rudis</i> , <i>Amaranthus viridis</i> , <i>Kochia scoparia</i>
	1123	DHP	4	<i>Amaranthus lividus</i> , <i>Amaranthus palmeri</i> , <i>Amaranthus rudis</i> , <i>Amaranthus viridis</i>
	1124	DHP	4	<i>Amaranthus lividus</i> , <i>Amaranthus palmeri</i> , <i>Amaranthus rudis</i> , <i>Amaranthus viridis</i>
	1125	DHP	4	<i>Amaranthus lividus</i> , <i>Amaranthus palmeri</i> , <i>Amaranthus rudis</i> , <i>Amaranthus viridis</i>
	1126	DHP	4	<i>Amaranthus lividus</i> , <i>Amaranthus palmeri</i> , <i>Amaranthus rudis</i> , <i>Amaranthus viridis</i>
	1127	DHP	4	<i>Amaranthus lividus</i> , <i>Amaranthus palmeri</i> , <i>Amaranthus rudis</i> , <i>Amaranthus viridis</i>
	1128	DHP	4	<i>Amaranthus lividus</i> , <i>Amaranthus palmeri</i> , <i>Amaranthus rudis</i> , <i>Amaranthus viridis</i>
	1129	DHP	4	<i>Amaranthus lividus</i> , <i>Amaranthus palmeri</i> , <i>Amaranthus rudis</i> , <i>Amaranthus viridis</i>
	1130	DHP	4	<i>Amaranthus palmeri</i> , <i>Amaranthus rudis</i> , <i>Amaranthus viridis</i> , <i>Kochia scoparia</i>

(continued)

	SEQ ID NO	Gene	# Species	Species
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10	1132	DHP	4	<i>Amaranthus hybridus</i> , <i>Amaranthus palmeri</i> , <i>Amaranthus rudis</i> , <i>Amaranthus viridis</i>
15	1133	DHP	4	<i>Amaranthus hybridus</i> , <i>Amaranthus palmeri</i> , <i>Amaranthus rudis</i> , <i>Amaranthus viridis</i>
20	1134	DHP	4	<i>Amaranthus lividus</i> , <i>Amaranthus palmeri</i> , <i>Amaranthus rudis</i> , <i>Amaranthus viridis</i>
25	1135	DHP	4	<i>Amaranthus lividus</i> , <i>Amaranthus palmeri</i> , <i>Amaranthus rudis</i> , <i>Amaranthus viridis</i>
30	1136	DHP	4	<i>Amaranthus hybridus</i> , <i>Amaranthus palmeri</i> , <i>Amaranthus rudis</i> , <i>Amaranthus viridis</i>
35	1137	DHP	4	<i>Amaranthus lividus</i> , <i>Amaranthus palmeri</i> , <i>Amaranthus rudis</i> , <i>Amaranthus viridis</i>
40	1138	DHP	4	<i>Amaranthus lividus</i> , <i>Amaranthus palmeri</i> , <i>Amaranthus rudis</i> , <i>Amaranthus viridis</i>
45	1139	DHP	4	<i>Amaranthus lividus</i> , <i>Amaranthus palmeri</i> , <i>Amaranthus rudis</i> , <i>Amaranthus viridis</i>
50	1140	DHP	4	<i>Amaranthus hybridus</i> , <i>Amaranthus palmeri</i> , <i>Amaranthus rudis</i> , <i>Amaranthus viridis</i>
55	1141	DHP	4	<i>Amaranthus hybridus</i> , <i>Amaranthus palmeri</i> , <i>Amaranthus rudis</i> , <i>Amaranthus viridis</i>
	1142	DHP	4	<i>Amaranthus lividus</i> , <i>Amaranthus rudis</i> , <i>Amaranthus viridis</i> , <i>Kochia scoparia</i>
	1143	DHP	4	<i>Amaranthus lividus</i> , <i>Amaranthus palmeri</i> , <i>Amaranthus rudis</i> , <i>Amaranthus viridis</i>
	1144	DHP	4	<i>Amaranthus lividus</i> , <i>Amaranthus palmeri</i> , <i>Amaranthus rudis</i> , <i>Amaranthus viridis</i>
	1145	DHP	4	<i>Amaranthus lividus</i> , <i>Amaranthus palmeri</i> , <i>Amaranthus rudis</i> , <i>Amaranthus viridis</i>
	1146	DHP	4	<i>Amaranthus lividus</i> , <i>Amaranthus palmeri</i> , <i>Amaranthus rudis</i> , <i>Amaranthus viridis</i>
	1147	DHP	4	<i>Amaranthus lividus</i> , <i>Amaranthus palmeri</i> , <i>Amaranthus rudis</i> , <i>Amaranthus viridis</i>
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	1152	DHP	4	<i>Amaranthus lividus</i> , <i>Amaranthus palmeri</i> , <i>Amaranthus rudis</i> , <i>Amaranthus viridis</i>

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30	1158	DHP	4	<i>Amaranthus lividus</i> , <i>Amaranthus palmeri</i> , <i>Amaranthus rudis</i> , <i>Amaranthus viridis</i>
35	1159	DHP	4	<i>Amaranthus hybridus</i> , <i>Amaranthus palmeri</i> , <i>Amaranthus rudis</i> , <i>Amaranthus viridis</i>
40	1160	DHP	4	<i>Amaranthus lividus</i> , <i>Amaranthus palmeri</i> , <i>Amaranthus rudis</i> , <i>Amaranthus viridis</i>
45	1161	DHP	4	<i>Amaranthus hybridus</i> , <i>Amaranthus palmeri</i> , <i>Amaranthus rudis</i> , <i>Amaranthus viridis</i>
50	1162	DHP	4	<i>Amaranthus lividus</i> , <i>Amaranthus palmeri</i> , <i>Amaranthus rudis</i> , <i>Amaranthus viridis</i>
55	1163	DHP	4	<i>Amaranthus lividus</i> , <i>Amaranthus palmeri</i> , <i>Amaranthus rudis</i> , <i>Amaranthus viridis</i>
	1164	DHP	4	<i>Amaranthus lividus</i> , <i>Amaranthus palmeri</i> , <i>Amaranthus rudis</i> , <i>Amaranthus viridis</i>
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	1166	DHP	4	<i>Amaranthus lividus</i> , <i>Amaranthus rudis</i> , <i>Amaranthus viridis</i> , <i>Kochia scoparia</i>
	1167	DHP	4	<i>Amaranthus hybridus</i> , <i>Amaranthus palmeri</i> , <i>Amaranthus rudis</i> , <i>Amaranthus viridis</i>
	1168	DHP	4	<i>Amaranthus lividus</i> , <i>Amaranthus palmeri</i> , <i>Amaranthus rudis</i> , <i>Amaranthus viridis</i>
	1169	DHP	4	<i>Amaranthus lividus</i> , <i>Amaranthus palmeri</i> , <i>Amaranthus rudis</i> , <i>Amaranthus viridis</i>
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	1171	DHP	4	<i>Amaranthus lividus</i> , <i>Amaranthus palmeri</i> , <i>Amaranthus rudis</i> , <i>Amaranthus viridis</i>
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	1174	DHP	4	<i>Amaranthus lividus</i> , <i>Amaranthus rudis</i> , <i>Amaranthus viridis</i> , <i>Kochia scoparia</i>
	1175	DHP	4	<i>Amaranthus lividus</i> , <i>Amaranthus palmeri</i> , <i>Amaranthus rudis</i> , <i>Amaranthus viridis</i>

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 Li, Zhaolong  
 Shah, Ronak Hasmukh  
 Tao, Nengbing  
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10 <120> Methods and Compositions for Weed Control

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25 <400> 5

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&lt;211&gt; 1431

&lt;212&gt; DNA

50 &lt;213&gt; Amaranthus palmeri

&lt;400&gt; 6

## EP 2 755 987 B1

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50 &lt;213&gt; Amaranthus palmeri

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&lt;211&gt; 1431

&lt;212&gt; DNA

&lt;213&gt; Amaranthus rudis

&lt;400&gt; 8

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<211> 1059

<212> DNA

<213> Amaranthus rudis

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&lt;211&gt; 792

&lt;212&gt; DNA

&lt;213&gt; Amaranthus rudis

&lt;400&gt; 10

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## EP 2 755 987 B1

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&lt;211&gt; 764

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&lt;213&gt; Amaranthus rudis

&lt;400&gt; 11

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## EP 2 755 987 B1

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&lt;211&gt; 584

&lt;212&gt; DNA

&lt;213&gt; Amaranthus rudis

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&lt;210&gt; 15

&lt;211&gt; 570

&lt;212&gt; DNA

&lt;213&gt; Amaranthus rudis

&lt;400&gt; 15

## EP 2 755 987 B1

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&lt;211&gt; 486

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&lt;213&gt; Ambrosia trifida

&lt;400&gt; 23

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&lt;213&gt; Ambrosia trifida

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&lt;210&gt; 28

&lt;211&gt; 201

&lt;212&gt; DNA

&lt;213&gt; Ambrosia trifida

&lt;400&gt; 28

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&lt;210&gt; 29

&lt;211&gt; 101

&lt;212&gt; DNA

&lt;213&gt; Ambrosia trifida

&lt;400&gt; 29

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&lt;210&gt; 30

&lt;211&gt; 5446

&lt;212&gt; DNA

&lt;213&gt; Conyza canadensis

&lt;400&gt; 30

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&lt;210&gt; 31

&lt;211&gt; 5446

&lt;212&gt; DNA

55 &lt;213&gt; Conyza canadensis

&lt;400&gt; 31

**EP 2 755 987 B1**

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&lt;210&gt; 32

&lt;211&gt; 1401

&lt;212&gt; DNA

55 &lt;213&gt; Conyza canadensis

&lt;400&gt; 32

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&lt;210&gt; 33

&lt;211&gt; 1262

&lt;212&gt; DNA

&lt;213&gt; Conyza canadensis

&lt;400&gt; 33

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EP 2 755 987 B1

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&lt;210&gt; 39

&lt;211&gt; 444

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&lt;213&gt; Euphorbia heterophylla

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&lt;210&gt; 40

&lt;211&gt; 379

&lt;212&gt; DNA

&lt;213&gt; Euphorbia heterophylla

&lt;400&gt; 40

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## EP 2 755 987 B1

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&lt;210&gt; 42

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&lt;213&gt; Digitaria sanguinalis

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&lt;210&gt; 43

&lt;211&gt; 4007

**EP 2 755 987 B1**

<212> DNA

<213> Kochia scoparia

<400> 43

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&lt;213&gt; Kochia scoparia

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 <213> Amaranthus hybridus

50 <400> 56

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 5 tgcaccacct aaaactccag cagttattga agcaacggta gcaggatccc tatcagctgc 180  
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 10 <211> 200  
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 20 ctatttgctt attcctgctt agaagatcat agcattgatc cagactttgg attccatgt 180  
  
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 25 <210> 58  
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 30 <400> 58  
  
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 35 taaagctcta ctttcgagca aactaacggc cttcatcaat atcccaatcg aactttccac 180  
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 40 <210> 59  
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 45 <400> 59  
  
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 50 gtc当地 acaatggaca agaagtcgac cttggaaag cttgatcaag ttacttggac 180  
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&lt;400&gt; 60

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 5 tgcattgg tctctttgac ccatcttgc tcgacacagt ccaaccctca tcgaaccgag 120  
 ccgattgaac catagatcaa ggaacttcaa caattcctca aaggccaat tcgaccatga 180  
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10 <210> 61  
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 <212> DNA  
 <213> Amaranthus lividus

15 &lt;400&gt; 61

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 20 aggtgagct ctgagagcct cactatcccc catgaaagga tatggaaag accatttg 120  
 atggcaccat tgattgattt tattgggtct gatgtagaaa atgacactat ttgtacatgg 180  
 25 cattcattat caaattttt 200

<210> 62  
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 30 <213> Amaranthus lividus

&lt;400&gt; 62

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 35 caatcaatca atgggccat cacaatggt cttccata tccttcatg gggatagt 120  
 aggctctcag agctcacccct aaattccca taaaatagta tgtccaaatgc aattggccta 180  
 40 ggaccataacc ttattccctt 200

<210> 63  
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 45 <213> Amaranthus lividus

&lt;400&gt; 63

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 50 gaagttccct aatcgaaag gatggatga aaagggtttt gcccgttggaa aatcgcttat 120  
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 55 gtgatggcgg aagtttcaa 200

<210> 64  
 <211> 200

&lt;212&gt; DNA

&lt;213&gt; Amaranthus lividus

&lt;400&gt; 64

5                   ttgaaaactt ccgccatcac taaagctatc aggagtcaag ttcaagactc ccattacaga     60  
                  ggtttatga gaccaatccc ataagcgatt tccaacgggc aaaaccctt tcattccatc   120  
 10                cttcccgatt agggaaacttc caccgagttt accccatgct tcaaagattc caccaaaaaa   180  
                  atttgataat gaatgccatg    200

&lt;210&gt; 65

&lt;211&gt; 200

&lt;212&gt; DNA

&lt;213&gt; Amaranthus palmeri

&lt;400&gt; 65

20                cgatatctgtt aaaggagcc ctctcatcct atgcttatca atgactttt catatgccta    60  
                  tcaaacagat cagattggat cgggttcgag ttcaggtcat atataaacat gtcagcaaac   120  
 25                ccttgaccca tacccaaccc atttagttaa ttggcttgaa aatcacaacc ttaattcgac   180  
                  cagcgacaga tcaggttgac    200

&lt;210&gt; 66

&lt;211&gt; 200

&lt;212&gt; DNA

&lt;213&gt; Amaranthus palmeri

&lt;400&gt; 66

30                gtcaacctga tctgtcgctg gtcgaattaa gggtgtgatt ttcaagccaa ttaactaaat   60  
 40                gggttggta tgggtcaagg gtttgctgac atgtttatat atgacctgaa ctcgaacccg   120  
                  atccaatctg atctgtttga taggcatatg aaaaagtcat tgataagcat aggatgagag   180  
                  ggctccctt aacagatccg    200

&lt;210&gt; 67

&lt;211&gt; 200

&lt;212&gt; DNA

&lt;213&gt; Amaranthus palmeri

&lt;400&gt; 67

## EP 2 755 987 B1

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	tttcgattt aagttttta acattgatta aatctaattt tttaggctt aatttaattt	120
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10	<211> 200	
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	<213> Amaranthus palmeri	
	<400> 68	
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	cctgatctgt cgctggtcga	200
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25	<211> 200	
	<212> DNA	
	<213> Amaranthus palmeri	
	<400> 69	
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35	agatctgatc aacccatata attaatttggg tcaaaatctc aactcaaacc catttatttc	180
	gaattttttt cagatcaat	200
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	<211> 200	
	<212> DNA	
	<213> Amaranthus palmeri	
45	<400> 70	
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50	aaaactaaaaa aacccttatata accattttaa tttaatgtgt taatttcgag tttaatcaat	180
	aagtgtttttt taaactcagt	200
55	<210> 71	
	<211> 200	
	<212> DNA	
	<213> Amaranthus palmeri	

&lt;400&gt; 71

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 5 tcaagtttt tcataatgatg ccccttggtt tctttgaata aaattcacca atttaaaatc 120  
 cctgcccttg atccgttgggt attggatattt ggtaaagcac atatggtaaa ttctactcca 180  
 ttgttatctg tagttttgggt 200

10 <210> 72  
 <211> 200  
 <212> DNA  
 <213> Amaranthus palmeri

15 <400> 72

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 20 accaacggat caagggcagg gatTTAAAT tggtaattt tattcaaaga aaacaagggg 120  
 catcatatga aaaaacttga gaggaaaaat tagcgctagt tgatccgacc tgctaatttg 180  
 atctgaattt aattcgaat 200

25 <210> 73  
 <211> 200  
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 <213> Amaranthus palmeri

30 <400> 73

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 35 taatcctcat tactttctgt ttgctgtctt tctctacatt cctactaaat taagacctag 120  
 atgcagtgtt attctgttcg aggttgcgga aaacggtaca atagttgcct aatttgcaat 180  
 40 tcatgtttgt tcttgcgttt 200

45 <210> 74  
 <211> 200  
 <212> DNA  
 <213> Amaranthus palmeri

<400> 74

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 acagaaaagta atgaggattt gtggtgctag gagtaggatt tttgggggt tttagaaccua 180  
 55 aactacagat aacaatggag 200

<210> 75  
 <211> 200

&lt;212&gt; DNA

&lt;213&gt; Amaranthus palmeri

&lt;400&gt; 75

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           ctttcagcct ttcagggagt atttgattt ttctgtAAA ttaccttGCC agatttATTt 120  
 10          ggctatTTG attgtgtGA tttcttGTC cttgtttCC atggggCTAA actccTTTG 180  
           gttaatCTTA aacatttCtt 200

&lt;210&gt; 76

&lt;211&gt; 200

&lt;212&gt; DNA

&lt;213&gt; Amaranthus palmeri

&lt;400&gt; 76

20         aagaaatgtt taagattaAC caaaaggAGT ttAGCCCCAT ggAAACAcAGG gACAAAGAAA 60  
           tacacacaAT caaaatAGCC aaataAAATCT ggCAAGGTAA ttGACAGAA AAATCAAAT 120  
 25         actccCTGAA aggCTGAAAG ttCAACCTTT gttCCAATAG AGTAAACAAT gtagAAAACG 180  
           caagaacAAA catGAATTGC 200

&lt;210&gt; 77

&lt;211&gt; 200

&lt;212&gt; DNA

&lt;213&gt; Amaranthus palmeri

&lt;400&gt; 77

35         ttttggTTAA tcttaAACAT ttcttctaAT tttGAATTGC agtGGTatGA agtGTTGTT 60  
           aataatacaAT caattttCT gaACTGGGTT aatGAAGAGA agcATTGTag acatGAGGAA 120  
 40         tGTGTTCAAG catCTCAAAA caACCCAAATT aatCCCAAT gggatCttCA actGTAATAG 180  
           aggtAAATCA catatatTTCT 200

&lt;210&gt; 78

&lt;211&gt; 200

&lt;212&gt; DNA

&lt;213&gt; Amaranthus palmeri

&lt;400&gt; 78

50         agaatatatATG tgatTTACCT ctattACGT tgaAGATCCC atttGGGATT aatttGGTTG 60  
           ttttGAGATG ctTGAACACA ttCCtCATGT ctacaATGCT tctttCATT AACCCAGTT 120  
 55         agaaaaATTG attGTatATT aaACAAACACT tcataACCAct gcaATTCAAa attGAAGAA 180  
           atGTTAAGA ttaACCAAAA 200

<210> 79  
 <211> 200  
 <212> DNA  
 <213> Amaranthus palmeri

5                   <400> 79

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	ttatgttgtt atcaatgata gaaataagag aagcttgagc attcaaattt atctatcact	120
	cactggtgtt ggacatctaa aatatggacc aagagtttc ttcattttt gtaccagaag	180
	ctgcaaatgt atgattaaag	200

15                   <210> 80  
 <211> 200  
 <212> DNA  
 <213> Amaranthus palmeri

20                   <400> 80

25	cttaaatcat acatttgcag cttctggtag aaaaaatgaa gaaaactctt ggtccatatt	60
	ttagatgtcc acaaccagtg agtgatagat aaatttgaat gctcaagctt ctcttatttc	120
	tatcattgtat aacaacataa taacatcattt aaaataaaac aataaccatt aaatgagaat	180
	atatgtgatt tacctctatt	200

30                   <210> 81  
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 <212> DNA  
 <213> Amaranthus palmeri

35                   <400> 81

40	agaagctgca aatgtatgtat taaaagtttag gttggagttt caataaaatt ttattacccc	60
	gacgcttatta agtggctcac actgaggcggtt ggtttgaggg tcagatgggtt caaccttcag	120
	aagaggttct aagcattttc aacatgttca accgcattttt gtcccgtaaa attggaggct	180
	tcaatattaa attatgttagt	200

45                   <210> 82  
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 <212> DNA  
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50                   <400> 82

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 gaaaatgctt agaaccttt ctgaagggtt caccatctga ccctcaaacc ccgcctcagt 120  
 5 gtgagccact taatagcgtc gggtaataa aattttattt caactccaac ctaaacttta 180  
 atcatacatt tgcaagttct 200

10 <210> 83  
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15 <400> 83  
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 gaatacataa ttttaaatt gcacagggcc ttcaaagaaa tttcgatt tgctctgccc 120  
 20 ctggcaacct ttggcaaatt tggatagaat tagagaagtt tactttatg tgtgtataat 180  
 atgatgaatg aatgagtgaa 200

25 <210> 84  
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 <213> Amaranthus palmeri

30 <400> 84  
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 aatttgccaa aggttgccag gggcagagca aaatcgaaaa tttctttgaa ggccctgtgc 120  
 35 aatttaaaaa ttatgtattc attaacatat ttatatctt aaacacataa cacatactac 180  
 ataatttaat attgaagcct 200

40 <210> 85  
 <211> 200  
 <212> DNA  
 <213> Amaranthus palmeri

45 <400> 85  
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 50 tgttgtctgt ttcttttctt gggcgagact gtttttggag aaaaacgtta tgttgaatgc 180  
 ttgatttact cagaactgat 200

55 <210> 86  
 <211> 200  
 <212> DNA  
 <213> Amaranthus palmeri

&lt;400&gt; 86

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 5 agaaaagaaa acagacaaca ttagggataa aagatgtctc taattttatt tatatacct 120  
 ttttaacgaa acctcgatag tatataggtc attgactcag aaatgaactt cattgttcac 180  
 tcattcattc atcatattat 200

10 <210> 87  
 <211> 200  
 <212> DNA  
 <213> Amaranthus palmeri

&lt;400&gt; 87

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 20 tttatgttcc gataacaggt ttcaaattcc actcgtatga agtttgatat actgctatgt 120  
 gttttttagt cttcgtgtt tgcttcctt cactcatcac cagtgaatga tgttgaagtt 180  
 tgttccaaga agcaagaagt 200

25 <210> 88  
 <211> 200  
 <212> DNA  
 <213> Amaranthus palmeri

&lt;400&gt; 88

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 35 aaacacgaag ctacaaaaac acatagcagt atatcaaact tcatacgagt ggaatttgaa 120  
 acctgttatac gaaacataaaa caataggaaa acaccatttt tatccatctg tttgcatacag 180  
 ttctgagtaa atcaaggcatt 200

40 <210> 89  
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 <212> DNA  
 <213> Amaranthus palmeri

&lt;400&gt; 89

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 50 tagaaaaattt caaccaagct ctgcaacaaa tgaagaaatt aggcatagac atcacaaggc 120  
 atgggtgttt atatgagacg gAACCTGCAACGGTGTGAAATTAGGCAATGAGCTGCATACGTCAGTTCTTAACT 180  
 ctgctttaag aggctttaca 200

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<213> Amaranthus palmeri

<400> 90

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	cgtctcatat aaacacccat gccttgtat gtctatgcct aatttcttca tttgttgtag	120
10	agcttgggttg aaattttcta atctgtcacc aacattgctc ccgattgcaa ttacgacttc	180
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<210> 91

<211> 200

<212> DNA

<213> Amaranthus palmeri

<400> 91

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	gaaaatttag aaggatatgg gtagaaccaa gggataagg tatggccta ggccaattga	120
25	cttggacata ctatttatg ggaagtttag ggtgagctct gagagcctca ctatccccca	180
	tgaaaggata tggaaagac	200

<210> 92

<211> 200

<212> DNA

<213> Amaranthus palmeri

<400> 92

30	gtctttccca ttcctttca tggggatag tgaggctctc agagctcacc ctaaacttcc	60
	cataaaatag tatgtccaag tcaattggcc taggaccata ctttattccc ttgggtctac	120
35	ccatatcctt ctcattttc tttaataccc ctaataattc atgaggccca agttttgtaa	180
	agctcttaa agcagagtta	200

<210> 93

<211> 200

<212> DNA

<213> Amaranthus palmeri

<400> 93

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	tctgatgtatgaaaatgacac tatttgtacg tggcattcat tatcaaattt ttgggtgga	120
45	atctttgaag catgggtaa actcggtgga agttccctaa tcgggaagga tggaatgaaa	180
	agggtttgc ccgttggaaa	200

<210> 94

<211> 200  
 <212> DNA  
 <213> Amaranthus palmeri

5       <400> 94

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 gtgtcatttt ctacatcaga cccaaatacaa tcaatcaatg gtgccatcac aaatggtctt   180  
 tcccatatcc tttcatgggg    200

15       <210> 95

<211> 200  
 <212> DNA  
 <213> Amaranthus palmeri

20       <400> 95

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 atactgcggt tgctaagggtt cgtcagatga tctcagatgg ggcagatata attgacatcg   180  
 gggcacaatc aaccagaccc    200

30       <210> 96

<211> 200  
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 <213> Amaranthus palmeri

35       <400> 96

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 aaccttagca accgcagtat cgacagattt aaaacttccg ccatcactaa agctatcagg   120  
 agtcaagttc aagactcccc taacagaggt tttatgagac caatcccata agcgatttcc   180  
 aacggggcaaa accctttca    200

45       <210> 97

<211> 200  
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 <213> Amaranthus palmeri

50       <400> 97

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 tgtgaatgat gtctctagtg    200

<210> 98  
 <211> 200  
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5  
 <400> 98

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	aataaaaacgt atccactgac aagattcttc ttcttcctc gatcaaatcc ttgacagctt	120
	ccaatacagg tactacttt tctagcttt cttagccga aattctagtt gccatgggtc	180
	<b>tggttgattg tgcccgatg</b>	200

15  
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20  
 <400> 99

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	gaccttaaag ttccttatat agcaatgcac atgcgaggag atccgacttc aatgcaaaac	120
	tctgagaact tgacctacaa ttagtggaa aagcaagtgg ctttgagtt gagttctagg	180
	<b>gtcatagatg cagaattatc</b>	200

30  
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 <211> 200  
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35  
 <400> 100

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	ttgttaggtca agttctcaga gtttgcatt gaagtcggat ctccctcgcat gtgcattgct	120
	atataaggaa cttaaggc cgcaacaaca ttaaacatct cggaatcgag tttcccacta	180
	<b>gagacatcat tcacaatgtg</b>	200

45  
 <210> 101  
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50  
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 gcatcgatt ttctaagaat acgaatcaa atttgaaat tcttagtgtt ttacaaaaga 120  
 5 tacgggaaga gatacgtaag aagagttgg cggtggctca ttgccccttg ctaattggac 180  
 cttcaagaaa gaggttctg 200

10 <210> 102  
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 20 attcttagaa aatccgatgc cggatcaat aactatcctc caagcaggaa ttcccgataa 180  
 ttctgcatct atgaccctag 200

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 35 agtacataat gtgagggata accttgatgc tgtcaagtta tgtgatgcc tactcgaaa 180  
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40 <210> 104  
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45 <400> 104  
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       tatagttctt acagatgttag aatcattggc tatttgccctt tttctgctta gaagatcata   180  
       gcattgatcc agactttgga   200

10           <210> 106  
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15           <400> 106

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       acttcatcat aatcccaatc gaaccttcca ctagacatca ttcacaagggt ggtacaaaca   180  
       agcagttaat cagttttcc   200

25           <210> 107  
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           <213> Amaranthus palmeri

30           <400> 107

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       tgaagggtgg actgtgtcga gacaagatgg gtcaaagaga ccaatggaca agaagtcgac   180  
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45           <400> 108

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       acaattcctc aaaggccaa ttcgaccatg agtcgcaaca taattctaca tggaatccaa   180  
       agtctggatc aatgctatga   200

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<213> Amaranthus palmeri

<400> 109

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      caaaccatat acggttgtta gttgttactg tcactgtttg agttgaatta gatcatgcaa   180  
10       acactgttagg agtcttgatt   200

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<212> DNA

<213> Amaranthus palmeri

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      caaatgatat gtctaattgtc cgccagagcc tgctgttatac tgcttgtcca agtaacttga   180  
25       tcaagcttgc ccaagggtcga   200

<210> 111

<211> 200

<212> DNA

<213> Amaranthus palmeri

<400> 111

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      tgttgttattg ttgtgtgctt tttggaattt tattcctgca atagaattat tcccgaaaaa   180  
35       tggatttcct ttatatcatc   200

<210> 112

<211> 200

<212> DNA

<213> Amaranthus palmeri

<400> 112

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      taacatcata tcaaagattt cattttgaga agtacagtgt cacatagctt cacataatca   180  
45       agactcctac agtgtttgca   200

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5 <400> 113

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10 tatttatgtc taaacttgtt aaagatgttc tgcttccgcc cttgccacgg gatgtcgaaa 180  
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15 <210> 114

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20 <400> 114

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25 tattaattca gtttgcattc taaagatcaa tgattagaag tacaaaaact cctttgatga 180  
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30 <210> 115

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35 <400> 115

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40 taatgaagtt ttcgagtc aa gtccgagctg ttctataagt ttacgtatct tgaagcttct 180  
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45 <210> 116

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50 <400> 116

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55 tgacttaaaa tcaacgcaaa atcttaaacg agtcaattca agcataggaa tccctcgatt 180

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15	ccaacattaa ctaagctctc gaggcttaat gaggcagggg tagctcaagt ttgttattca	180
	agcaacttac catagtattc	200
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	gagagcttag ttaatgttgg gtttaaactt gctcgactcg aattcgtgat gaactcaaga	120
30	ctttaacgag ttgagtcaag caagcatcaa tgcggttcaa tgtgtcaaaa aataaaggat	180
	aagaagtaag aactaagaag	200
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40	<400> 119	
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	atttcagtag aacgaaagaa gtacattatt aaactggcc attgatgctt gagctagaga	120
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55	<400> 120	

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15 <400> 121

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 20 caagatgtgt tgatgtcatg tgcaatagta attatcaggt gaagtaattc aatatgtac 180  
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30 <400> 122

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 35 gtatgtgagc taggagctgg gcacacatga ttttgtatg cactgctgaa gaataaagga 180  
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45 <400> 123

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 50 cattgacga agatatccag aacacatccc cttgtttct gctagataat caactattct 180  
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 10 gatatgaggt gactggctac 200

&lt;210&gt; 125

&lt;211&gt; 200

&lt;212&gt; DNA

&lt;213&gt; Amaranthus palmeri

&lt;400&gt; 125

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 25 tttgttctcg gcctgaagac gctgcatttc ctcttccatt tgctgcctct caaaccatgt 180  
 atcagcatcc gcccaaactg 200

&lt;210&gt; 126

&lt;211&gt; 200

&lt;212&gt; DNA

&lt;213&gt; Amaranthus palmeri

&lt;400&gt; 126

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 gtcttcaggc cgagaacaaa gcttcaatag aagctgctga agaactgcac agaaagaaac 120  
 40 tggccgagga gcgtgaagga gaaaaagagg acgatcccat ggctagggcc gaagctgagg 180  
 tattacagca gaatcagaat 200

&lt;210&gt; 127

&lt;211&gt; 200

&lt;212&gt; DNA

&lt;213&gt; Amaranthus palmeri

&lt;400&gt; 127

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20 <213> Amaranthus palmeri

<400> 129

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35 <213> Amaranthus palmeri

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50 <213> Amaranthus palmeri

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 10                caaatgaaga aattaggcat agacatcaca aggcatgggt gtttatatga gacggaacct   180  
                  gcatacgtga ctgatcaacc   200

&lt;210&gt; 140

&lt;211&gt; 200

&lt;212&gt; DNA

&lt;213&gt; Amaranthus palmeri

&lt;400&gt; 140

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 25                ttgctcccga ttgcaattac gacttcttgc ttcttggAAC aaacttcaac atcattcact   180  
                  ggtgatgagt gaaggaaagc   200

&lt;210&gt; 141

&lt;211&gt; 200

&lt;212&gt; DNA

&lt;213&gt; Amaranthus palmeri

&lt;400&gt; 141

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 35                ccaagggaat aaggtatggt cctaggccaa ttgacttgga catactattt tatggaaat   180  
                  ttagggtgag ctctgagagc   200

&lt;210&gt; 142

&lt;211&gt; 200

&lt;212&gt; DNA

&lt;213&gt; Amaranthus palmeri

&lt;400&gt; 142

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 45                taattcatga ggcccaagtt ttgtaaagcc tcttaaagca gagttaagaa acttcggttg   180  
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 45 atgatctcag atggggcaga 200

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 5 tcggatctcc tcgcattgtgc attgctatat aaggaacttt aaggccgca acaacattaa 120  
 acatctcgga atcgagtttc ccactagaga catcattcac aatgtgtgcc cccttcttga 180  
 cggcctccga agcaacttta 200

10 <210> 151  
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 <212> DNA  
 <213> Amaranthus palmeri

15 <400> 151

gatgttgta agcaagtggc tttggagttt agttcttaggg tcatacatgc agaattatcg 60  
 20 ggaattcctg ctggaggat agttattgtat cccggatcg gatTTCTAA gaatacgaat 120  
 caaaaatttgg aaattcttag tggtttacaa aagatacggg aagagatagc taagaagagt 180  
 ttggcggtgg ctcattgccc 200

25 <210> 152  
 <211> 200  
 <212> DNA  
 <213> Amaranthus palmeri

30 <400> 152

gggcaatgag ccaccgccaa actcttctta gctatctttt cccgtatctt ttgttaaacca 60  
 35 ctaagaattt ccaaattttt attcgatttcc tttagaaaatc cgatgccggg atcaataact 120  
 atcctccaag caggaattcc cgataattctt gcatttatgc ccctagaact caactccaaa 180  
 gccacttgct tacaaacatc 200

40 <210> 153  
 <211> 200  
 <212> DNA  
 <213> Amaranthus palmeri

45 <400> 153

agagtttggc ggtggctcat tgccccttgc taattggacc ttcaagaagag aggtttctgg 60  
 50 gcgagatttgc tgagcgccct gtacgcgttgc acagggatcc tgctaccatt gcttctataa 120  
 ctgctggagt tttaggtggc gcaaacatttgc taagagtaca taatgtgagg gataaccttgc 180  
 atgctgtcaa gttatgttat 200

55 <210> 154  
 <211> 200  
 <212> DNA

<213> Amaranthus palmeri

<400> 154

5	atcacataac ttgacagcat caaggttatc cctcacatta tgtactctta caatgttgc	60
	accacctaata actccagcag ttatagaagc aatggtagca ggatccctgt cagctgctac	120
10	agggcgctca caaatctcgc ccagaaacct ctttcttcaa ggtccaattt gcaaggggca	180
	atgagccacc gccaaactct	200

<210> 155

<211> 200

<212> DNA

<213> Amaranthus palmeri

<400> 155

20	gcttccttc actttcacc agggAACgtt gttgaagttt gttccaagaa gcaagaagtc	60
	gtaattgcaa tcgggagcaa tgggtgtac agattagaaa atttcaacca agctctgcaa	120
25	caaataaaga aattaggcat agacatcaca aggcatggtt gtttatatga gacggAACct	180
	gcatacgtga ctgatcaacc	200

<210> 156

<211> 200

<212> DNA

<213> Amaranthus palmeri

<400> 156

35	ggttgatcag tcacgtatgc aggttccgtc tcatataaac aaccatgcct tggatgtct	60
	atgcctaatt tcttcatttg ttgcagagct tggttggaaat tttctaatct gtcaccaaca	120
40	ttgctcccga ttgcaattac gacttcttgc ttcttggaaac aaacttcaac aacgttccct	180
	ggtaagagt gaaggaaagc	200

<210> 157

<211> 200

<212> DNA

<213> Amaranthus palmeri

<400> 157

50	aacctgcata cgtgactgat caaccgaagt ttcttaactc tgcttaaga ggctttacaa	60
	gacttggacc tcatgaatta ttagggtat tgaagaaaat tgagaaggat atggtagaa	120
55	ccaaggaaat aaggtatggt cctaggccaa ttgacttggc catactattt tatggaaat	180
	ttagggtagt ctctgagagc	200

<210> 158

<211> 200  
<212> DNA  
<213> Amaranthus palmeri

5 <400> 158

gctctcagag ctcaccctaa acttccata aaatagtatg tccaagtcaa ttggcctagg 60  
accatacctt attcccttgg ttctaccat atccttctca attttcttca ataccctaa 120  
taattcatga ggtccaagtc ttgtaaagcc tcttaaagca gagttaagaa acttcggtt 180  
atcagtcacg tatgcaggtt 200

15 <210> 159

<211> 200  
<212> DNA  
<213> Amaranthus palmeri

20 <400> 159

gaagtttagg gtgagctctg agagcctcac tatccccat gaaaggatat gggaaagacc 60  
atttgtatg gcaccattga ttgattgtat tgggtctgat gtagaaaatg acactattt 120  
tacgtggcat tcattatcaa attttcggg tggaatctt gaagcatggg gtaaactcgg 180  
tggaagctcc ctaatcgga 200

30 <210> 160

<211> 200  
<212> DNA  
<213> Amaranthus palmeri

35 <400> 160

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ttgataatga atgccacgta caaatagtgt cattttctac atcagaccca atacaatcaa 120  
tcaatggtgc catcacaaat ggtcttccc atatccttc atggggata gtgaggctt 180  
cagagctcac cctaaacttc 200

45 <210> 161

<211> 200  
<212> DNA  
<213> Amaranthus palmeri

50 <400> 161

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ctcttatggg attggctca taaaacctct gttatggag tcttgaactt gactcctgat 120  
agcttagtg atggcggaaat tttcaatct gtcaatactg cggttgctaa gttcgtcag 180  
atgatctcag atggggcaga 200

<210> 162  
 <211> 200  
 <212> DNA  
 <213> Amaranthus palmeri

5                   <400> 162

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10	cttccgccat	cactaaagct	atcaggagtc	aagttcaaga	ctcccataac	agaggttta	120
	tgagaccaat	cccataaagag	atttccaacg	ggcaaaaaccc	ttttcattcc	atccttcccg	180
15	attagggagc	ttccaccgag					200

<210> 163  
 <211> 200  
 <212> DNA  
 <213> Amaranthus palmeri

20                   <400> 163

	gtcagatgat	ctcagatggg	gcagatataa	ttgacatcg	ggcacaatca	accagaccca	60
25	tggcaactag	aatttcggct	gaagaagagc	tagcaagagt	agtacctgta	ttagaagctg	120
	tcaaggattt	gatcgaggaa	gaaggaagaa	tcttgtcagt	ggatacgtt	tattctaaag	180
30	ttgctttgga	ggccgtcaag					200

<210> 164  
 <211> 200  
 <212> DNA  
 <213> Amaranthus palmeri

35                   <400> 164

	cttgcggcc	tccaaagcaa	ctttagaata	aaacgtatcc	actgacaaga	ttcttccttc	60
40	ttcctcgatc	aaatccttga	cagttctaa	tacaggtaact	actcttgcta	gctttcttc	120
	agccgaaatt	ctagttgcca	tgggtctgg	tgatttgcc	ccgatgtcaa	ttatatctgc	180
45	cccatctgag	atcatctgac					200

<210> 165  
 <211> 200  
 <212> DNA  
 <213> Amaranthus palmeri

50                   <400> 165

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 taaactcgat tccgagatgt ttaatgttgc tgccggacctt aaagttcctt atatacgaaat 120  
 5 gcacatgcga ggagatccga cttcaatgca aaactctgag aacttgaccc acaatgatgt 180  
 ttgttaagcaa gtggcttcgg 200  
  
 <210> 166  
 10 <211> 200  
 <212> DNA  
 <213> Amaranthus palmeri  
  
 <400> 166  
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 20 acatctcgga atcgagttt ccactagaga catcattcac aatgtgtgcc ccattcttga 180  
 cggcctccaa agcaacttta 200  
  
 <210> 167  
 25 <211> 200  
 <212> DNA  
 <213> Amaranthus palmeri  
  
 <400> 167  
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 ggaattcctg cttggaggat agttattgtt cccggcatcg gatTTCTAA gaatacgaat 120  
 35 caaaaatttgg aaattcttag tggtttacaa aagatacggg aagagatagc taagaagagt 180  
 ttggcggtgg ctcattgccc 200  
  
 <210> 168  
 40 <211> 200  
 <212> DNA  
 <213> Amaranthus palmeri  
  
 <400> 168  
 45 gggcaatgag ccaccgccaa actcttctta gctatctttt cccgttatctt ttgttaaacca 60  
 ctaagaattt ccaaattttt attcgattt ttagaaaatc cgatgccggg atcaataact 120  
 50 atcctccaag caggaattcc cgataattctt gcatctatga ccctagaact caactccgaa 180  
 gccacttgct tacaaacatc 200  
  
 <210> 169  
 55 <211> 200  
 <212> DNA  
 <213> Amaranthus palmeri

&lt;400&gt; 169

agagtttggc ggtggctcat tgcccattgc taattggacc ttcaagaaag aggtttctgg 60  
 5 gcgagatttgc tgagcgccct gtacgcgcgtg acaggatcc tgctaccatt gcttctataa 120  
 ctgctggagt tttaggtggc gcaaacatttga taagagtaca taatgttagg gataaccttg 180  
 atgctgtcaa gttatgttat 200

10 <210> 170  
 <211> 200  
 <212> DNA  
 <213> Amaranthus palmeri

15 <400> 170

atcacataaac ttgacagcat caaggttatc cctaacattta tgtactctta caatgtttgc 60  
 20 accacctaaa actccagcag ttatagaagc aatggtagca ggatccctgt cagctgctac 120  
 agggcgctca caaatctcgc ccagaaacct ctttcttggaa ggtccaattt gcaaggggca 180  
 atgagccacc cccaaactct 200

25 <210> 171  
 <211> 200  
 <212> DNA  
 <213> Amaranthus palmeri

30 <400> 171

ggtgtttatt tgggcattggc cttatgtgtg atgttggtaa attttgtggc ttctgtccact 60  
 35 aatctgatgt atcggatctg ttaaaggag ccctctcatc ctatgcttat caatgacttt 120  
 ttcatatgcc tatcaaacag atcagattgg atcgggttcg agttcagggtc atatataaac 180  
 atgtcagcaa acccttggacc 200

40 <210> 172  
 <211> 200  
 <212> DNA  
 <213> Amaranthus palmeri

45 <400> 172

ggtcaagggt ttgctgacat gtttatatat gacctgaact cgaacccgat ccaatctgat 60  
 50 ctgtttgata ggcataatgaa aaagtcatgg ataagcatag gatgagaggg ctcccttaa 120  
 cagatccgat acatcagatt agtggacgaa gccacaaaaat ttaccaacat cacacataag 180  
 gccatgccccca aataaacacc 200

55 <210> 173  
 <211> 200  
 <212> DNA

<213> Amaranthus palmeri

<400> 173

5	ttaaacatgtc agcaaaccct tgaccatac ccaaccatt tagttaattg gcttgaaaat	60
	cacaacctta attcgaccag cgacagatca gggtgacctg atttcgaccc acttaaccca	120
10	tttataattt tttttcgat ttaagtttt taacattgat taaatcta at ttttaggct	180
	ttaattttaa ttttatgttt	200

<210> 174

<211> 200

15 <212> DNA

<213> Amaranthus palmeri

<400> 174

20	aaacataaaaa taaaattaa agcctaaaaa attagattt atcaatgtt aaaaacttaa	60
	atcgaaaaaa aaattataaa tgggttaagt gggtcgaaat caggtcaacc tgatctgtcg	120
25	ctggtcgaat taagggttgtg attttcaagc caattaacta aatgggttgg gtatgggtca	180
	agggtttgct gacatgttttta	200

<210> 175

<211> 200

30 <212> DNA

<213> Amaranthus palmeri

<400> 175

35	aggcttaat tttaattttt ttttttttgg ttttaattttt atttactatg aaaaataaga	60
	aactattata tgaactgagt ttagcttaca cttattaatt taactcgaaa ttaacacatt	120
40	taattaaatg gttatataagg gtttttttagt tttaaaattt caacctgaac ctaatctatt	180
	taataaaaaag atcagatctg	200

<210> 176

<211> 200

45 <212> DNA

<213> Amaranthus palmeri

<400> 176

50	cagatctgat cttttttatta aatagattag gttcaggttg aaattttaaa actaaaaaac	60
	cctatataac catttaatta aatgtgttaa ttgcagtttta aattaataag tgtaagctaa	120
55	actcagttca tataatagtt tcttattttt catagtaaat acaaattaaa caaaaaaaca	180
	taaaattaaa attaaagcct	200

<210> 177

<211> 200  
<212> DNA  
<213> Amaranthus palmeri

5 <400> 177

ctatttaata aaaagatcag atctgatcaa cccatataat taattgggtc aaaatctcaa 60  
ctcaaacc ca tttat tcga attaaattca gatcaaatta gcagg tcgga tcaacta 120  
ctaattttc ctctcaagtt tttcatatg atgccc ttg tttcttga ataaaattca 180  
ccaatttaaa atccctgccc 200

15 <210> 178

<211> 200  
<212> DNA  
<213> Amaranthus palmeri

20 <400> 178

ggcaggat tttaaattgg tgaattttat tcaaagaaaa caaggcat catagaaaa 60  
aacttgagag gaaaaattag cgctagttga tccgacctgc taatttgc tgaatttaat 120  
tcgaaataaa tgggttgag ttgagattt gacccaatta attatatgg ttgatcagat 180  
ctgatctttt tattaaatag 200

30 <210> 179

<211> 200  
<212> DNA  
<213> Amaranthus palmeri

35 <400> 179

attcaccaat ttaaaatccc tgcccttgat ccgttgtat tggtatttgg taaagcacat 60  
atggtaaatt ctactccatt gttatctgta gtttgggtc taaaccccaa acaaattccta 120  
ctccttagcac cactaatcct cattacttgc tgtttgctgt ctttctctac attcctacta 180  
aattaagacc tagatgcagt 200

45 <210> 180

<211> 200  
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<213> Amaranthus palmeri

50 <400> 180

55

	actgcatcta ggtcttaatt tagtaggaat gtagagaaaag acagcaaaca gaaagtaatg	60
	aggatttagt gtgctaggag taggatttg ttggggtttta gaaccaaaac tacagataac	120
5	aatggagtag aatttaccat atgtgcttta ccaaataccca ataccaacgg atcaagggca	180
	gggattttaa attggtaat	200
	<210> 181	
10	<211> 200	
	<212> DNA	
	<213> Amaranthus palmeri	
	<400> 181	
15	tactaaatata agacctagat gcagtgttat tctgttcgag gttgccgaaa acggtacaat	60
	agttgcctaa tttgcaattc atgtttgttc ttgcgtttc tacattgttt actctattgg	120
20	aacaaagggtt gaactttcag cctttcaggg agtattttga ttttctgtc aaattacctt	180
	gccagattta tttggctatt	200
	<210> 182	
25	<211> 200	
	<212> DNA	
	<213> Amaranthus palmeri	
	<400> 182	
30	aatagccaaa taaatctggc aaggtaattt gacagaaaaa tcaaaatact ccctgaaagg	60
	ctgaaagttc aacctttgtt ccaatagagt aaacaatgta gaaaacgcaa gaacaaacat	120
35	gaattgcaaa ttaggcaact attgtaccgt tttcggcaac ctcgaacaga ataacactgc	180
	atctaggtct taattttagta	200
	<210> 183	
40	<211> 200	
	<212> DNA	
	<213> Amaranthus palmeri	
	<400> 183	
45	accttgccag atttattttgg ctatttgat tgtgtgtatt tctttgtccc ttgtttccat	60
	ggggctaaac tccttttgtt taatcttaaa catttcttct aattttgaat tgcatgttga	120
50	tgaagtgttg tttaatatac aatcaatttt tctgaactgg gttaatgaag agaagcattg	180
	tagacatgag gaatgtgttc	200
	<210> 184	
55	<211> 200	
	<212> DNA	
	<213> Amaranthus palmeri	

&lt;400&gt; 184

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 5 gtatattaaa caacacttca taccactgca attcaaaatt agaagaaaatg tttaagatta 120  
 accaaaagga gtttagcccc atggaaacaa gggacaaaga aatacacaca atcaaaaatag 180  
 ccaaataaaat ctggcaaggt 200

10 <210> 185  
 <211> 200  
 <212> DNA  
 <213> Amaranthus palmeri

15 <400> 185

cattgttagac atgaggaatg tttcaagca tctcaaaaca accaaattaa tcccaaatgg 60  
 20 gatcttcaac tgtaatagag gtaaatcaca tatattctca tttaatgggt attgttttat 120  
 tttaaggatg ttattatgtt gttatcaatg atagaaataa gagaagcttg agcattcaaa 180  
 25 tgtatctatc actcactgg 200

25 <210> 186  
 <211> 200  
 <212> DNA  
 <213> Amaranthus palmeri

30 <400> 186

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 35 aacataataa catccttaaa ataaaaacaat aaccattaaa tgagaatata tgtgatttac 120  
 ctctattaca gttgaagatc ccatttggga ttaatttgggt tgtttgaga tgcttgaaca 180  
 40 cattcctcat gtctacaatg 200

40 <210> 187  
 <211> 200  
 <212> DNA  
 <213> Amaranthus palmeri

45 <400> 187

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 50 catttcttgt accagaattt gcaaatgtat gattaaagtt taggttggag ttgcaataaa 120

atttcattac cccgacgcta ttaagtggct cacactgagg cggggtttga gggtcggatg 180  
 55 gtgcaacctt caggagaggt 200

&lt;210&gt; 188

<211> 200  
<212> DNA  
<213> Amaranthus palmeri

5        <400> 188

          acctctcctg aagggtgcac catccgaccc tcaaaccgg cctcagtgtg agccactaa    60  
          tagcgtcggt gtaatgaaat tttattgcaa ctccaaccta aactttaatc atacatttc    120  
          aaattctggt acaagaaatg aagaaaactc ttggccata ttttagatgt ccacaaccag    180  
          tgagtgatag atacatttg    200

10      <210> 189  
<211> 200  
<212> DNA  
<213> Amaranthus palmeri

15      <400> 189

          ggatggtgca acttcagga gaggttctaa gcattttcaa catgttcaac cgcatgggt    60  
          cccgtaaaat tggaggcttc aatattaaat tatgttagtat gtgttaagtgt tttaaagata    120  
          taaaaatgtt aatgaataca taattttaa aattgcacag ggcttcaaa gaaattttt    180  
          attttgctcc gcccatggca    200

20      <210> 190  
<211> 200  
<212> DNA  
<213> Amaranthus palmeri

25      <400> 190

          tgccatgggc ggagcaaaat caaaaatttc tttgaaagcc ctgtgcaatt ttaaaaatta    60  
          tgtattcatt aacattttta tatctttaaa cacttaacac atactacata atttaatatt    120  
          gaagecctcca atttacggg accctctgctg gttgaacatg ttgaaaatgc tttagaacctc    180  
          tcctgaaggt tgcaccatcc    200

30      <210> 191  
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<212> DNA  
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35      <400> 191

          ttttgatttt gctccgccccca tggcaacctt tggcaaatct ggatagaatt agagaaattt    60  
          acttttatgt atgtataatc tggatgaatga atgaatgaac aatgaagtcc atttctgagt    120

## EP 2 755 987 B1

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 tcttttatcc ctaatgttgt 200

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 <210> 192  
 <211> 200  
 <212> DNA  
 <213> Amaranthus palmeri

10  
 <400> 192

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 15 cgatagtata taggtcattt gactcagaaat gaacttcatt gttcattcat tcattcatca 120  
 gattatacat acataaaaagt aaatttctct aattctatcc agatttgcca aagttgcca 180  
 tgggcggagc aaaatcaaaa 200

20  
 <210> 193  
 <211> 200  
 <212> DNA  
 <213> Amaranthus palmeri

25  
 <400> 193

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 ggtgtttcc tagtgtttat gtttcataaa aggattcaa attccaatcg tatgaagttt 180  
 gaaatactgc tttttgtttt 200

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40  
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 45 ataaacacta ggaaaacacc attttttatac catctgtttg catcagttct gagtaaatca 120  
 agcattcaac ataacgtttt tctccaaaaa cagtctcgcc cagaaaagaa aacagacaac 180  
 attaggata aaagatgtct 200

50  
 <210> 195  
 <211> 200  
 <212> DNA  
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 <400> 195

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 5 cagggAACGT tggtaagtt tggccaaga agcaagaagt cgtaattgca atcgggagca 120  
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 tagacatcac aaggcatggt 200  
 10 <210> 196  
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 20 ttctaattctg tcaccaacat tgctcccgat tgcaattacg acttcttgct tcttggaca 120  
 aacttcaaca acgttccctg gtgaagagtg aaggaaagca aaacacgaag ctacaaaaac 180  
 aaaaagcagt atttcaaact 200  
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 <211> 200  
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 35 tcaaccgaag tttcttaact ctgcttaag aggcttaca agacttggac ctcatgaatt 120  
 attagggta ttgaagaaaa ttgagaagga tatgggtaga accaaggaa taaggatgg 180  
 tcctaggcca attgacttgg 200  
 40 <210> 198  
 <211> 200  
 <212> DNA  
 <213> Amaranthus palmeri  
 45 <400> 198  
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 50 ttttcttcaa taccctaat aattcatgag gtccaaagtct tgtaaagcct cttaaagcag 120  
 agttaagaaa cttcggttga tcagtcacgt atgcaggatc cgtctcatat aaacaaccat 180  
 gccttgtgat gtctatgcct 200  
 55 <210> 199  
 <211> 200  
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 <213> Amaranthus palmeri

&lt;400&gt; 199

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 gagagcctca ctatccccca tgaaaggata tggaaagac catttgtat ggcaccattg   120  
 attgattgtat ttgggtctga tgttagaaaat gacactatTT gtacgtggca ttcattatca   180  
 10          aattttcggtt gtggaatctt   200

&lt;210&gt; 200

&lt;211&gt; 200

&lt;212&gt; DNA

15          &lt;213&gt; Amaranthus palmeri

&lt;400&gt; 200

20          aagattccac ccgaaaaatt tgataatgaa tgccacgtac aaatagtgtc attttctaca   60  
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 tggggatag tgaggctctc agagctcacc ctaaacttcc cataaaatag tatgtccaag   180  
 25          tcaattggcc taggaccata   200

&lt;210&gt; 201

&lt;211&gt; 200

&lt;212&gt; DNA

30          &lt;213&gt; Amaranthus palmeri

&lt;400&gt; 201

35          tatcaaattt ttcgggtgga atctttgaag catgggtaa actcggtgga agctccctaa   60  
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 40          gttttcaatc tgtcaatact   200

&lt;210&gt; 202

&lt;211&gt; 200

&lt;212&gt; DNA

45          &lt;213&gt; Amaranthus palmeri

&lt;400&gt; 202

50          agtattgaca gattgaaaac ttccgccatc actaaagcta tcaggagtca agttcaagac   60  
 tcccataaca gaggttttat gagaccaatc ccataagaga tttccaacgg gcaaaaccct   120  
 tttcattcca tccttccca ttagggagct tccaccgagt ttacccatg cttcaaagat   180  
 55          tccacccgaa aaatttgcata   200

&lt;210&gt; 203

&lt;211&gt; 200

&lt;212&gt; DNA

&lt;213&gt; Amaranthus palmeri

&lt;400&gt; 203

5                   cggaagttt caatctgtca atactgcggc tgctaagggt cgtcagatga tctcagatgg     60  
                  ggcagatata attgacatcg gggcacaatc aaccagaccc atggcaacta gaatttcggc   120  
 10                tgaagaagag ctagcaagag tagtacctgt attagaagct gtcaaggatt tgatcgagga   180  
                  agaaggaaga atcttgcag   200

&lt;210&gt; 204

&lt;211&gt; 200

&lt;212&gt; DNA

&lt;213&gt; Amaranthus palmeri

&lt;400&gt; 204

20                ctgacaagat tcttccttct tcctcgatca aatccttgac agcttctaatt acaggtaacta   60  
                  ctcttgctag ctcttcttca gccgaaattc tagttgccat gggcttggtt gattgtgccc   120  
 25                cgatgtcaat tataatctgcc ccacatctgaga tcacatctgacg aaccttagca acccgagtat   180  
                  tgacagatttggaaaacttccg   200

&lt;210&gt; 205

&lt;211&gt; 200

&lt;212&gt; DNA

&lt;213&gt; Amaranthus palmeri

&lt;400&gt; 205

30                gaggaagaag gaagaatctt gtcagtggtt acgttttatt ctaaagttgc tttggaggcc   60  
                  gtcaagaagg gggcacacat tgtgaatgtat gtctcttagtg gtaaactcga ttccgagatg   120  
 35                tttaatgttg ttgcggacct taaagttcct tataatagcaa tgcacatgcg aggagatccg   180  
                  acttcaatgc aaaactctga   200

&lt;210&gt; 206

&lt;211&gt; 200

&lt;212&gt; DNA

&lt;213&gt; Amaranthus palmeri

&lt;400&gt; 206

40                tcagagttt gcattgaagt cggatctcct cgcacatgtgc ttgcttatata aggaacttta   60  
                  aggtccgcaa caacattaaa catctcgaa tcgagttac cactagagac atcattcaca   120  
 45                atgtgtgccc ccttcttgac ggcttcggaaa gcaacttttag aataaaacgt atccactgac   180  
                  aagattcttc cttcttcctc   200

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 <212> DNA  
 <213> Amaranthus palmeri

5  
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15  
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20  
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gttttgcatt gaagtcggat	200

30  
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35  
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45  
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50  
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 5 gtccaattag caaggggcaa tgagccaccg ccaaacttctt ctttagctatc tcttcccgta 180  
 tctttgtaa accactaaga 200

10 <210> 211  
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 20 tctagtgaa ggtcgattg ggattatgat gaagtccgtt agtttgctcg aaagaagagc 180  
 tttagccgt aaggttata 200

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 <212> DNA  
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30 <400> 212  
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 caatcgaacc ttccactaga catcattcac aaggtggtaa aaacaagcag ttaatcagtt 120  
 35 tttccgagta tggcatcaca taacttgaca gcatcaaggt tatccctaac attatgtact 180  
 cttacaatgt ttgcaccacc 200

40 <210> 213  
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 <212> DNA  
 <213> Amaranthus palmeri

45 <400> 213  
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 50 cgactcatgg tcgaattggc cctttgagga attgttgaag ttcccttgagt ctatggttca 180  
 atcggctcggtt ttcgtatgt 200

55 <210> 214  
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 <213> Amaranthus palmeri

&lt;400&gt; 214

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 tatgatcttc taagcagaaa taggcaaata gccaatgatt ctacatctgt aagaactata 180  
 accttacggg cttaaagctct 200  
 10

&lt;210&gt; 215

&lt;211&gt; 200

&lt;212&gt; DNA

&lt;213&gt; Amaranthus palmeri

&lt;400&gt; 215

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 20 ccaatggaca agaagtcgac cttgggcaag cttgatcaag ttacttggac gagcagataa 120  
 cagcaggctc tggcgagcat tagacataca tatcatttgt ttttgaagt cagatctgca 180  
 atcaatcagt tgcaatggac 200  
 25

&lt;210&gt; 216

&lt;211&gt; 200

&lt;212&gt; DNA

&lt;213&gt; Amaranthus palmeri

&lt;400&gt; 216

gtccattgca actgatttgat tgcagatctg acttcaaaaa acaaattgata tgtatgtcta 60  
 35 atgctcgcca gagcctgctg ttatctgctc gtccaagtaa cttgatcaag cttgcccaag 120  
 gtcgacttct tgcatttgg tctcttgac ccattttgtc tcgacacagt ccaaccttca 180  
 tcgaaccgag ccgattgaac 200  
 40

&lt;210&gt; 217

&lt;211&gt; 200

&lt;212&gt; DNA

&lt;213&gt; Amaranthus palmeri

&lt;400&gt; 217

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 50 tttgagttga attagatcat gcaaacactg taagagtctt gatgttgc agctatgtga 120  
 tactgtactt ctcaaaatga aatctttgat atgatgttag cttcaagagt tgttttccc 180  
 cccctcacat tctgttctat 200  
 55

&lt;210&gt; 218

&lt;211&gt; 200

&lt;212&gt; DNA

<213> Amaranthus palmeri

<400> 218

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	tcattttgag aagtacagta tcacatagct tcacaaaatc aagactctt cagtgttgc	120
10	atgatctaat tcaactcaaa gagtgacagg aacaactaac aaccgtatat ggtctgtcca	180
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<210> 219

<211> 200

<212> DNA

<213> Amaranthus palmeri

<400> 219

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25	cttctaatac ttgatcttta gatgcaaaac tgaattaata caagtgttct tgatgaaggc	180
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<211> 200

<212> DNA

<213> Amaranthus palmeri

<400> 220

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	taaagatcaa tgattagaag tacaaaaact ccttgatga tataaaggaa taccattttg	120
35	caggaataat tctaattaca ggaataacaat tccaaaaagc acacaacaat acaacataga	180
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<210> 221

<211> 200

<212> DNA

<213> Amaranthus palmeri

<400> 221

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	ccgcccttgc cacggatgt cgggattgac taagggtggt aatcgaggga ttcctatgct	120
45	tgaattgact cgtttaagat tttgcgttga tttaagtca aatgtttgc taagcataaa	180
	ctattcttagc tcgacttgaa	200

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5 <400> 222

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10 acatcccgtg gcaagggcgg aagcagaaca tctttaacaa gtttagacat aaataaaaag 180  
tgagttcttc aataagcctt 200

15 <210> 223

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<213> Amaranthus palmeri

20 <400> 223

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25 ttgaaccgc当地 ttgatgctt当地 ct当地actcaa cccgttaaag tcttgaattc atcacgaatt 180  
cgagtcgagc aagtttaaac 200

30 <210> 224

<211> 200  
<212> DNA  
<213> Amaranthus palmeri

35 <400> 224

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40 ct当地caagata cataaaactta cagaacagct cggacttgac tc当地aaaactt cattattcaa 180  
gtc当地gagctag aatagttat 200

45 <210> 225

<211> 200  
<212> DNA  
<213> Amaranthus palmeri

50 <400> 225

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55 agtc当地agttag tt当地ttgtta attc当地agtag aacgaaagaa gt当地attt当地 aaacttggcc 180  
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<210> 226  
<211> 200  
<212> DNA  
<213> Amaranthus palmeri

5  
<400> 226

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aacaaaacttg agctaccctt gcctcattaa gcctcgagag cttagttaat gttgggtta 180  
aacttgctcg actcgaattc 200

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20  
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<400> 228

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50  
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 10 agtcgtacct gtattagaag 200

&lt;210&gt; 234

&lt;211&gt; 200

&lt;212&gt; DNA

&lt;213&gt; Amaranthus rudis

&lt;400&gt; 234

cttctaatac aggtacgact cttctagct cttttcagc cgaaatccta gttgccatgg 60  
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 ccttagcaac cgca(gtatcg acagattgaa accttccgccc atcactaaag ctatcaggag 180  
 25 tcaagttcaa gactcccatt 200

&lt;210&gt; 235

&lt;211&gt; 200

&lt;212&gt; DNA

&lt;213&gt; Amaranthus rudis

&lt;400&gt; 235

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 attgtgaatg atgtctctag tggaaactc gatcccgaga tggtaatgt tggtgcggac 180  
 40 cttaaagttc cttatatacg 200

&lt;210&gt; 236

&lt;211&gt; 200

&lt;212&gt; DNA

&lt;213&gt; Amaranthus rudis

&lt;400&gt; 236

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 taaaacgtat ccactgacaa gattttct ttttcctcga tcaaatcctt gacagcttct 180  
 55 aatacaggta cgactcttc 200

&lt;210&gt; 237

&lt;211&gt; 200

&lt;212&gt; DNA

&lt;213&gt; Amaranthus rudis

&lt;400&gt; 237

5           cgAACCTTAA AGTTCCATTAT ATAGCAATGC ACATGCGAGG AGATCCGACT TCAATGCAA 60  
           ATTCTGAGAA CTTGACGTAC AATGATGTGT GTAAGCAAGT GGCTTCGGAG TTGAGCTCTA 120  
           GGGTCTGAGA TGCGAGAATTA TCAGGAATTTC CTGCTTGGAG GATGGTTATT GATCCCAGCA 180  
 10           TTGGATTTCTA AGAAATAACG 200

&lt;210&gt; 238

&lt;211&gt; 200

&lt;212&gt; DNA

&lt;213&gt; Amaranthus rudis

&lt;400&gt; 238

20           CGTATTCTTA GAAAATCCAA TGCCGGGATC AATAACCATC CTCCAAGCAG GAATTCCCAGA 60  
           TAATTCTGCA TCTACGACCC TAGAGCTCAA CTCCGAAGGCC ACTTGCTTAC ACACATCATT 120  
           GTACGTCAAG TTCTCAGAAT TTTGCATTGA AGTCGGATCT CCTCGCATGT GCATTGCTAT 180  
 25           ATAAGGAACT TTAAGGTCGG 200

&lt;210&gt; 239

&lt;211&gt; 200

&lt;212&gt; DNA

&lt;213&gt; Amaranthus rudis

&lt;400&gt; 239

30           CGGCATTGGA TTTTCTAAGA ATACGAAGCA AAATTTGGAA ATTCTTAGTG GTTTACAAAAA 60  
           GATACGGCAA GAGATAGCTA AGAAGAGTTT GGCGGTGGCT CATTGCCCT TGCTAATTGG 120  
           ACCTTCAAGA AAGAGGTTTC TGGCGAGAT TTGCAATCGC CCTGTAGCAG CTGATAGGGA 180  
 35           TCCTGCTACC GTTGCTTCTA 200

&lt;210&gt; 240

&lt;211&gt; 200

&lt;212&gt; DNA

&lt;213&gt; Amaranthus rudis

&lt;400&gt; 240

40           TAGAAAGCAAC GGTAGCAGGA TCCCTATCAG CTGCTACAGG GCGATTGCAA ATCTCGCCCA 60  
           GAAACCTCTT TCTTGAAGGT CCAATTAGCA AGGGGCAATG AGCCACCGCC AAACTCTCT 120  
           TAGCTATCTC TTGCCGTATC TTTTGTAAAC CACTAAGAAT TTCCAAATT TGCTTCGTAT 180  
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&lt;210&gt; 241

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<213> Amaranthus rudis

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<213> Amaranthus rudis

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tcttcttcag ccgaaatcct 200

30 <210> 243

<211> 200  
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<213> Amaranthus rudis

35 <400> 243

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aagtggcttc ggagttgagc tctagggtcg tagatgcaga attatcgaa attcctgctt 180  
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45 <210> 244

<211> 200  
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<213> Amaranthus rudis

50 <400> 244

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 10 <211> 200  
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 35 gtaaaccact aagaatttcc aaattttgct tcgtattttttaa agaaaatcca atgcccggat 180  
  
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10 <210> 249  
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15 <400> 249

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 gcaatgttgg tgacagatta gaaaatttca acgaagctct gcaacaaatg aagaaatttag 180  
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25 <210> 250  
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30 <400> 250

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40 ttcaacatcg ttcaactggtg atgagtgaag gaaagcaata caagaagcta caaaaacaca 180  
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45 <210> 251  
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 <212> DNA  
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50 <400> 251

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 attatttaggg gtattgaaga gaatcgagaa ggatatgggc agaaccaagg gaataaggta 180  
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&lt;210&gt; 252

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5       <400> 252

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       taagaaaactt cggctgatca gtcacgtaag caggttccgt ctcgtataaaa caaccatgcc   180  
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15       <210> 253

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20       <400> 253

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       catctcgctc ctaatggacc ccagccccctt aataacgtca atgttgtgcc cacatttctt   180  
       ggagaaccca atgccaggat   200

30       <210> 254

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35       <400> 254

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    180  
       ctgaactata tcctacatta tgtaccctta ctatgttagc accattcaaa attgcagcgg  
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<211> 200

<212> DNA

50       <213> Amaranthus rudis

<400> 255

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 5 caaattttc gggtggaata tttgaagcat gggataaact cggaggaagt tccttaatcg 180  
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15 <400> 256  
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 20 agacccaata caatcaatta atggtgccat cacaatggc cttccata tccttcatg 180  
 ggaaatagtg aggctctcag 200

25 <210> 257  
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30 <400> 257  
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 35 aaggttcaa tctgtcgata ctgcgggtgc taagggtcgt cagatgatct ctgacggggc 180  
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40 <210> 258  
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 50 ccattactga gttttatga gaccaatccc ataaacggtt cccaacgggc aaaaccctt 180  
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55 <210> 259  
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&lt;400&gt; 259

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10 <210> 260  
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15 <400> 260

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25 <210> 261  
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 <213> Amaranthus rudis

30 <400> 261

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40 <210> 262  
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 <213> Amaranthus rudis

45 <400> 262

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 55 atcagtcacg tatgcgggtt 200

<210> 263  
 <211> 200

&lt;212&gt; DNA

&lt;213&gt; Amaranthus rudis

&lt;400&gt; 263

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                   atttgtatgc gcaccattaa ttgattgtat tgggtctgggt gtagaaaatg acactatttgc   120  
 10                tacatggcat tcgtttatcaa atttttcggg tggaatcttt gaagcatggg ataaaactcg   180  
                   tggaagttcc ctaatcgggaa   200

&lt;210&gt; 264

&lt;211&gt; 200

&lt;212&gt; DNA

&lt;213&gt; Amaranthus rudis

&lt;400&gt; 264

20                tcccgattag ggaacttcca ccgagtttat cccatgcttc aaagattcca cccgaaaaat   60  
                   ttgataacga atgccatgtat caaatagtgt cattttctac accagaccca atacaatcaa   120  
 25                ttaatggtgc catcacaaat ggtctttccc atatccttca atggggataa gtgaggctct   180  
                   cagagctcac cctaaacttc   200

&lt;210&gt; 265

&lt;211&gt; 200

&lt;212&gt; DNA

&lt;213&gt; Amaranthus rudis

&lt;400&gt; 265

30                ctcggtgaa gttccctaattt cgaaaaggat ggaatgaaaa gggttttgcc cgttggaaac   60  
                   cgcttatggg attggctca taaaacctct gtaatggag tcttgaacctt gactcctgat   120  
 35                agcttttagtg atggaggaag ttttcaatct gtcgatactg cggttgctaa ggttcgtag   180  
                   atgatatcag atggggccga   200

&lt;210&gt; 266

&lt;211&gt; 200

&lt;212&gt; DNA

&lt;213&gt; Amaranthus rudis

&lt;400&gt; 266

40                tcggccccat ctgatatcat ctgacgaacc ttagcaaccg cagtatcgac agattgaaaa   60  
                   cttcctccat cactaaagct atcaggagtc aagttcaaga ctcccattac agaggttta   120  
 45                tgagaccaat cccataagcg gttcccaacg ggcaaaaccc ttttcattcc atccttcccg   180  
                   attagggaac ttccaccgag   200

<210> 267  
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5                   <400> 267

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10	tggcaactag gatttcggct gaagaagagc tagaaagagt tgtacctgta ttggaagctg	120
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25	ttcctcgatc acatccttaa cagcttccaa tacaggtaca actctttcta gctttcttc	120
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35	<400> 269	
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40	gaaattcgat cctgagatgt tcaatgttgt tgccggacctt aaagttcctt atatacgaaat	120
	gcacatgcga ggagatccga cttcaatgca aaattctgag aacttgacgt acagtgtatgt	180
	ttgttaagcaa gtggcttcgg	200
45	<210> 270 <211> 200 <212> DNA <213> Amaranthus rudis	
50	<400> 270	

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 5 acatctcagg atcgaatttc ccactagaga catcattcac aatgtgtgcc cccttcttga 180  
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 10 <211> 200  
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 <400> 271  
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 20 caaaaatttgg aaattcttac tggtttacaa aagatacggc tagagatagc taagaagagt 180  
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 25 <211> 200  
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 35 attctccaag caggaattcc cgataattct gcatactatga ccctagagct caactccgaa 180  
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 50 ctgcgtggagt ttttaggtggc gcaaacatttgc taagagtaca taatgtgagg gataaccttgc 180  
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 <210> 274  
 55 <211> 200  
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&lt;400&gt; 274

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 agggcgattt caaatctcac ccagaaacct cttttgtaaa ggtccaattt gcaaggggca 180  
 atgagccacc gccaaactct 200

10 <210> 275  
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15 <400> 275

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 20 gtcgatactg cggttgcataa ggttcgtcag atgatctctg acggggcgga cataattgac 120  
 attggggcgca agtcaacaag acccttggca actaggattt cggctgaaga agagctagaa 180  
 25 agagtcgtac ctgttattaga 200

25 <210> 276  
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 <213> Amaranthus rudis

30 <400> 276

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 35 cttgttact gcgc(cc)aat gtcaattatg tccgc(cc)gt cagagatcat ctgacgaacc 120  
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 40 aagttcaaga ctcccattac 200

40 <210> 277  
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45 <400> 277

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 acattgtgaa ttagtgcgttct agtggaaac tcgatcccga gatgttaat gttgtgcgg 180  
 55 accttaaaagt tccttatata 200

<210> 278  
 <211> 200  
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EP 2 755 987 B1

<213> Amaranthus rudis

<400> 278

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      agaaaacatca ttcacaatgt gtgccccatt cttgacggcc tccgaagcaa cttagaata   120  
      aacacgtatcc actgacaaga ttcttccttc ttcctcgatc aaatccttga cagcttctaa 180  
10      tacaggtacg actctttcta   200

<210> 279

<211> 200

15     <212> DNA

<213> Amaranthus rudis

<400> 279

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      aaattctgag aacttgacgt acagtgtatgt ttgtaagcaa gtggcttcgg agttgagctc 120  
      tagggtcata gatgcagaat tatcggaat tcctgcttgg agaatagtta ttgatccgg 180  
25      cattggattt tctaagaata   200

<210> 280

<211> 200

30     <212> DNA

<213> Amaranthus rudis

<400> 280

35      tattcttaga aaatccaatg ccgggatcaa taactattct ccaaggcagga attcccgata 60  
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      acgtcaagtt ctcagaattt tgcattgaag tcggatctcc tcgcattgtgc attgctatat 180  
40      aaggaaacttt aaggccgca   200

<210> 281

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45     <212> DNA

<213> Amaranthus rudis

<400> 281

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      ggacccttcac gaaagagggtt tctgggtgag atttctaattt gccctgttagc agctgatagg 180  
55      gatcctgcta ccgtcgatcc   200

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<211> 200  
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5 <400> 282

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gctatctcta gccgtatctt ttgtaaacca gtaagaattt ccaaattttg ctgcgtattc 180  
tttagaaaatc caatgccggg 200

15 <210> 283

<211> 200  
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<213> Amaranthus rudis

20 <400> 283

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caagaagcaa gaagtcgtaa 200

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<211> 200  
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35 <400> 284

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catcaattctt gagtaaatca 200

45 <210> 285

<211> 200  
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<213> Amaranthus rudis

50 <400> 285

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5  
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ggttccgtct cgtataaaca accatgcctt gtgatgtcta tgcctaattt ctgcatttgt	120
tgcagagctt cggtgaaatt ttctaatctg tcaccaacat tgctcccgat tgcaattacg	180
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15  
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acatactatt ttatggaaag ttttaggtga gctctgagag cctcactatt ccccatgaaa	180
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20  
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 <213> Amaranthus rudis

25  
 <400> 288

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30  
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35  
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	tcgatcccga gatgtttaat	200
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	<210> 291	
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	<400> 291	
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50	tcggatctcc tcgcattgtgc attgctatat aaggaacttt aaggtccgca acaacattaa	180
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 ttggcggtgg ctcattgccc cttgctaatt ggaccttcaa gaaagaggtt tctggcgag 180  
 atttgcaatc gccctgttagc 200

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 20 gggcaatgag ccaccgccaa actcttctta gctatcttgc gccgtatctt ttgtaaacca 120  
 ctaagaattt ccaaattttg ctgcgtatttcc tttagaaaatc caatgccggg atcaataacc 180  
 atcctccaag caggaattcc 200

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 <213> Amaranthus rudis

30 <400> 295  
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 35 ctgctggagt tttaggtggt gcaaacatttgc taagagtaca taatgtgagg gataaccttgc 120  
 atgcagtcaa gttatgttatcc gcccatactca gaaaaacggat ttagccgatttttggattt 180  
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55 <210> 297  
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&lt;213&gt; Amaranthus rudis

&lt;400&gt; 297

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10	tctaagaagc aagaagtcat aattgcaatc gggagcaatg ttggtgacag attagaaaat	180
	tttaaccaag ctctgcaaca	200

&lt;210&gt; 298

&lt;211&gt; 200

15 &lt;212&gt; DNA

&lt;213&gt; Amaranthus rudis

&lt;400&gt; 298

20	tgttcagag ctggtaaaa attttctaat ctgtcaccaa cattgctccc gattgcaatt	60
	atgacttctt gcttcttaga acaaagttca acatcgcca ctggtgatga gtgaaggtac	120
25	gcaaaaaaca aagctgcaaa aacacatagt tgtatttcaa acttcatacg attgggattt	180
	gaaatccttt tatcgaaaca	200

&lt;210&gt; 299

&lt;211&gt; 200

30 &lt;212&gt; DNA

&lt;213&gt; Amaranthus rudis

&lt;400&gt; 299

35	aaaattttaa ccaagctctg caacaaatga agaaattagg catagacatc acaaggcatg	60
	gttgttata tgagacagaa cccgcatacg tgactgatca accgaagttt cttaactctg	120
40	cttaagagg ctttacaaga cttggcctc atgaattatt agggatttg aagaaaattg	180
	agaagaatat ggtagaaacc	200

&lt;210&gt; 300

&lt;211&gt; 200

45 &lt;212&gt; DNA

&lt;213&gt; Amaranthus rudis

&lt;400&gt; 300

50	ggttctaccc atattcttct caattttctt caataccct aataattcat gagggccaaag	60
	tcttgtaaaag cctcttaaag cagatgttaag aaacttcgggt tgatcgtca cgtatgcggg	120
55	ttctgtctca tataaacaac catgccttgt gatgtctatg cctaatttct tcatttgg	180
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&lt;210&gt; 301

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<213> Amaranthus rudis

5        <400> 301

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	aaggatatgg gaaagaccat ttgtgatggc accattaatt gattgtattg ggtctggtgt	180
	agaaaatgac actatttcta	200

15      <210> 302  
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<213> Amaranthus rudis

20      <400> 302

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25	atggtcttcc ccatatcctt tcatggggaa tagtgaggct ctcagagctc accctaaact	120
	tcccataaaa tagtatgtcc aagtcaatttgc gcctaggacc ataccttatt cccttgggtc	180
	tacccatatt cttctcaatt	200

30      <210> 303  
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<213> Amaranthus rudis

35      <400> 303

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40	tttgaagcat gggataaaact cggtggaaatg tccctaatcg ggaaggatgg aatggaaaagg	120
	gttttgcggc ttgggaacctt cttatggat tggtctcata aaacctctgt aatggagtc	180
	ttgaacttga ctccctgatag	200

45      <210> 304  
<211> 200  
<212> DNA  
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50      <400> 304

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	agtttatccc atgcttcaaa gattccaccc gaaaaatttg ataacgaatg ccatgtacaa	180
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                   cacagtcaac aagaccatg gcaacttagga ttccggctga agaagagcta gaaagagttg    180  
 15                tacctgtatt ggaagctgtt    200

<210> 306  
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 <212> DNA  
 20                <213> Amaranthus rufus

25                <400> 306

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 30                catgggtctt gttgactgtg ccccaatgtc aattatgtcg gccccatctg atatcatctg    120  
                   acgaacctta gcaaccgcag tatcgacaga ttgaaaactt cctccatcac taaagctatc    180  
 35                aggagtcaag ttcaagactc    200

<210> 307  
 <211> 200  
 <212> DNA  
 40                <213> Amaranthus viridis

45                <400> 307

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                   atactcgaa aaactgatata actgcttatt tgtaccacct tgtgaatgac gtctagtgaa    180  
 55                aagttcgatt gggatatgtat    200

<210> 308  
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 <212> DNA  
 55                <213> Amaranthus viridis

60                <400> 308

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 5 ttatgtactc ttacaatgtt tgaccacact aaaactccag cagttataga agcaacggta 180  
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 20 gaagatcata gcattgatcc agactttgga ttccatctag aattatgttgcgactcatgg 180  
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 50 gaagtcgacc ttggaaagc ttgatcaagt tacttgacg agcagatagc agcaggactc 180  
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&lt;400&gt; 313

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25           actgggttat taaagagaag cattgttagac atgaggaatg tgttcaagca tctcaaaaca   180  
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30           <210> 314  
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 <213> Amaranthus viridis

&lt;400&gt; 314

35           ccatttggga ttaatttggc tgtttgaga tgcttgaaca cattcctcat gtctacaatg   60  
 cttctcttta ataaccaggc tcagaaaaat tgattgtata ttaaacaaca cttcataccca   120  
 atatgatttt ctggatcaaa aagccaaaga gtacttcttc aaaaatatct tcaaaagttt   180  
 40           caatcttttag gttatacaga   200

45           <210> 315  
 <211> 200  
 <212> DNA  
 <213> Amaranthus viridis

&lt;400&gt; 315

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 55           tgaagaaatt aggcatagac   200

&lt;210&gt; 316

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<213> Amaranthus viridis

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 50 cccgattagg gaacttccac cgagtttacc ccatgctca aagattccac ccaaaaaatt 180  
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&lt;400&gt; 323

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15 <400> 324

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 40 atgcgaggag atccgacttc 200

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 55 tcctcgatca aatccttgac 200

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&lt;212&gt; DNA

&lt;213&gt; Amaranthus viridis

&lt;400&gt; 327

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                  tttgtaagca agtggctacg gagttgagct ctagggtcat agatgcagaa ttatcggaa     120  
 10                ttcctgcttg gaggatagtt attgatcccgcattggatt ttctaagaat acgaatcaa     180  
                  atttggaaat tcttcgtggt   200

&lt;210&gt; 328

&lt;211&gt; 200

&lt;212&gt; DNA

&lt;213&gt; Amaranthus viridis

&lt;400&gt; 328

20               accacgaaga atttccaaat tttgattcgt attcttagaa aatccaatgc cggttatcaat     60  
                  aactatcctc caagcagggaa ttcccgataa ttctgcacatct atgaccctag agctcaactc   120  
 25               cgttagccact tgcttacaaa catcattgta cgtcaagtcc tcagaatttt gcattgaagt   180  
                  cgatatctcct cgcattgtgca   200

&lt;210&gt; 329

&lt;211&gt; 200

&lt;212&gt; DNA

&lt;213&gt; Ambrosia trifida

&lt;400&gt; 329

30               aatgaaggct taagtcaaat gaaaaaatca ggcataaaaa taacaagaca tgcatgttta    60  
                  tacgaaaccg agccagctta cgtgactgat caacctcttt tcctcaattc tgccatcaga   120  
 35               ggcgttacaa agctggcccc acatgagcta ctatggcccc tcaagaaaaat tgaaaaagaa   180  
                  atggccgaa ccaaaggat   200

&lt;210&gt; 330

&lt;211&gt; 200

&lt;212&gt; DNA

&lt;213&gt; Ambrosia trifida

&lt;400&gt; 330

40               atacctttgg ttccggccat ttcttttca attttcttga gggccgatag tagctcatgt    60  
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	tcgagaggcc atttgttatg gctccgttag ttgacttatt ggggtcgaa attgacaatg	180
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15  
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 <213> Ambrosia trifida

20	<400> 332	
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	tctgtgtta ccataaaaca gtatatctaa gtcaatgggt cgccggccgt atctaatacc	180
	tttggttcgg cccatttctt	200

30  
 <210> 333  
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 <213> Ambrosia trifida

35	<400> 333	
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	taatcggtta tgggattggc caaagaaaac ctctgtcatg ggaattttga acttgactcc	180
	cgatagttt agcgacggag	200

45  
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 <213> Ambrosia trifida

50	<400> 334	
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 5 aagattcgcc acctaatttt tcccaagatt caaaaagtcc tcttttgag aaagaatgcc 180  
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&lt;400&gt; 338

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 <213> Ambrosia trifida

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 50 cggaagagct taggggtgtc acgtgcacct ttgttaatcg gtccttcaag aaagagattt 120  
 ttgggggaga tttgtggtcg ggcttctgct gtcgagagag atccagcgac tgggtgtct 180  
 gttacatgtg cgggtttggg 200

55 <210> 342  
 <211> 200  
 <212> DNA

<213> Ambrosia trifida

<400> 342

5	ccaaaaaccg cacatgtAAC agcAGCAACA gTCGCTGGAT ctctctcgac agcagaAGCC	60
	cgaccacaaa tctccccaa aaatctcttt cttGAAGGAC cgattaACAA aggtgcacgt	120
10	gacACCCCTA agctcttccg tgcaatctca ctcctaattc tcttaatcc catcaaAATA	180
	tccaaattat cttcggttt	200

<210> 343

<211> 200

<212> DNA

<213> Ambrosia trifida

<400> 343

20	acctcatcac ctccacAAAT caccaccACC catctccACC ctccaccACC acccacttca	60
	ttctccccAC ccaccgtatC tcAtcAccat catacctggT ggattgcagt ttgactctgt	120
25	ggggTccgct atatctcgTg ttAggaccat gatatctgaa ggggcggaca taattgatct	180
	gggagctcaa tcgacacgtc	200

<210> 344

<211> 200

<212> DNA

<213> Ambrosia trifida

<400> 344

30	gacgtgtcga ttgagctccc agatcaatta tgtccgcccc ttcagatATC atggcctaa	60
	cacgagatat agcggacCCC acagagtCAA actgcaatCC accaggTTG atggtgatGA	120
35	gatacggTgg gtggggagaa tgaagtgggt ggtggTggag ggtggagatG ggtggTggTg	180
	atttgtggag gtgatgagGT	200

<210> 345

<211> 200

<212> DNA

<213> Ambrosia trifida

<400> 345

40	gaccatcca caatgcaAAA cagtgagaAT ttgaagtATA atgatgtttG taaAGAAGTT	60
	ggtgatgagt tgtatgagCG tgtaAGAGCT gcagAGTTat gcggTgtGCC tgcatggagg	120
45	atcattctG acccaggGat cggGTTtca aagAAAACCG aagataATTt ggatATTTG	180
	atgggattaa agagaattAG	200

<210> 346

<211> 200  
<212> DNA  
<213> Ambrosia trifida

5 <400> 346

ctaattctct ttaatcccat caaaatatcc aaattatctt cggtttctt tgaaaacccg 60  
atccctgggt caagaatgat cctccatgca ggcacaccgc ataactctgc agctcttaca 120  
cgctcataca actcatcacc aacttcttta caaacatcat tataacttcaa attctcactg 180  
ttttgcattg tggatgggtc 200

15 <210> 347

<211> 200  
<212> DNA  
<213> Ambrosia trifida

20 <400> 347

gaccatcca caatgcaaaa cagtgagaat ttgaagtata atgatgttg taaagaagtt 60  
ggtgatgagt tgtatgagcg tgtaagagct gcagagttat gcgggtgtgcc tgcattggagg 120  
atcattcttg acccagggat cgggtttca aagaaaacccg aagataattt ggatattttg 180  
atgggattaa agagaattag 200

30 <210> 348

<211> 200  
<212> DNA  
<213> Ambrosia trifida

35 <400> 348

ctaattctct ttaatcccat caaaatatcc aaattatctt cggtttctt tgaaaacccg 60  
atccctgggt caagaatgat cctccatgca ggcacaccgc ataactctgc agctcttaca 120  
cgctcataca actcatcacc aacttcttta caaacatcat tataacttcaa attctcactg 180  
ttttgcattg tggatgggtc 200

45 <210> 349

<211> 200  
<212> DNA  
<213> Ambrosia trifida

50 <400> 349

ttttgatggg attaaagaga attaggagtg agattgccccg gaagagctt ggggtgtcac 60  
gtgcacctt gttaatcggt cttcaagaa agagatttt gggagagatt tgggtcggt 120  
cttctgctgg tgagagagat ccaacaactg ttgctgctgt tacatgtgctg gtttgggtg 180  
gtgctaattgt tgttcgctt 200

<210> 350  
 <211> 200  
 <212> DNA  
 <213> Ambrosia trifida

5  
 <400> 350

aacgcgaaca acattagcac cacccaaaac cgcacatgta acagcagcaa cagtgttgg 60  
 10 atctctctca ccagcagaag cccgaccaca aatctctccc aaaaatctct ttcttgaagg 120  
 accgattaac aaaggtgcac gtgacacccc taagctcttc cgggcaatct cactccta 180  
 15 tctcttaat cccatcaaaa 200

<210> 351  
 <211> 200  
 <212> DNA  
 <213> Ambrosia trifida  
 20  
 <400> 351

gatttgtggt cgggcttctg ctggtgagag agatccaaca actgttgctg ctgttacatg 60  
 25 tgcggtttg ggtggtgcta atgttggtcg cgttcataat gttggagatg atgctgatgc 120  
 tgtaaagctt tgtgattcaa tggtaaccg gtttggaaaga acgtaacagc ttttgaaat 180  
 30 ttgattgtgg gtttggtcc 200

30  
 <210> 352  
 <211> 200  
 <212> DNA  
 <213> Ambrosia trifida  
 35  
 <400> 352

ggacaaaaac ccacaatcaa atttcaaaaa gctgttacgt tcttccaacc cggttcaaca 60  
 40 ttgaatcaca aagctttaca gcatcagcat catctccaac attatgaacg cgaacaacat 120  
 tagcaccacc caaaaccgca catgtaacag cagcaacagt tggatct ctctcaccag 180  
 45 cagaagccccg accacaaatc 200

<210> 353  
 <211> 200  
 <212> DNA  
 <213> Ambrosia trifida  
 50  
 <400> 353

gccttaagtc aaatgaagaa atcaggcata gagataacaa gacatgcatt tttataaaaa 60  
 accgagccag cttatgtAAC tgatcaacct ctTTTCTTA attctgccat cagaggcgTT 120  
 5 acaaAGCTGG GCCCACATGA actactatcg GCCCTCAAGA aaATCGAAAAA agaATTGGGC 180  
 cgaaccaaaAG gtattagata 200

10 <210> 354  
 <211> 200  
 <212> DNA  
 <213> Ambrosia trifida

15 <400> 354  
 tatctaatac cttggttcg gcccattct ttTCGATT tCTTGAGGGC cgatAGTAGT 60  
 tcATGTGGGC ccAGCTTGT AACGCCtCTG ATGGCAGAAAT TAAGAAAAAG AGGTGATCA 120  
 20 gttacataag CTGGCTCGGT tttgtataaa catgcATGTC ttGTTATCTC tatGCCtGAT 180  
 ttcttcattt gacttaaggc 200

25 <210> 355  
 <211> 200  
 <212> DNA  
 <213> Ambrosia trifida

30 <400> 355  
 tgggCCGAAC caaAGGTATT agatacggcc CGCGACCCAT TGACTTAGAT ATACTGTTT 60  
 atggtaaaca cagaattaac tcggaaattc tcactgttcc acatgaaaga atcttcgaga 120  
 35 ggccatttgt tatggctccg ttagttgact tattggggTC ggatattgac aatgatacgg 180  
 ttctatgctg gcattcttc 200

40 <210> 356  
 <211> 200  
 <212> DNA  
 <213> Ambrosia trifida

45 <400> 356  
 gaaAGAATGC cAGCATAGAA CCgtatcATT GTCAATATCC GACCCCAATA AGTCAACTAA 60  
 CGGAGCCATA ACAAAATGGCC TCTCGAAGAT TCTTCATGT GGAACAGTGA GAATTCCGA 120  
 50 GTTAATTCTG TGTtTACCAT AAAACAGTAT ATCTAAGTCA ATGGGTcGCG GGCCTATCT 180  
 aataccTTG GTTCGGCCCA 200

55 <210> 357  
 <211> 101  
 <212> DNA  
 <213> Ambrosia trifida

&lt;400&gt; 357

5           ggattgcagt ttgactctgt ggggtccgct atatctcgta ttaggaccat gatatctgaa   60  
       ggggcggaca taattgatct gggagctcaa tcgacacagtc c                           101

&lt;210&gt; 358

&lt;211&gt; 101

&lt;212&gt; DNA

&lt;213&gt; Ambrosia trifida

&lt;400&gt; 358

15           ggacgtgtcg attgagctcc cagatcaatt atgtccgccc cttagatata catggccta   60  
       acacgagata tagcggaccc cacagagtca aactgcaatc c                           101

&lt;210&gt; 359

&lt;211&gt; 200

&lt;212&gt; DNA

&lt;213&gt; Ambrosia trifida

&lt;400&gt; 359

20           tcgttcttc atttctttc ggatgcttcg atacaagtcc attctcaaga acaagaagta   60  
       gtaattgctt taggttagcaa tgtgggtat aggcttaata acttaatga agccttaagt   120  
       caaatgaaaa aatcaggcat agaaataaca agacatgcat gtttatacga aaccgagcca   180  
       gcttacgtga ctgatcaacc   200

&lt;210&gt; 360

&lt;211&gt; 200

&lt;212&gt; DNA

&lt;213&gt; Ambrosia trifida

&lt;400&gt; 360

25           ggttgatcag tcacgtaagc tggctcggtt tcgtataaac atgcatagtct tgttatttct   60  
       atgcctgatt tttcatttg acttaaggct tcattaaagt tattaaggct atcacccaca   120  
       ttgctaccta aagcaattac tacttcttgt tcttgagaat ggacttgtat cgaagcatcc   180  
       gaagagaaaat gaaagaacga   200

&lt;210&gt; 361

&lt;211&gt; 200

&lt;212&gt; DNA

&lt;213&gt; Ambrosia trifida

&lt;400&gt; 361

	agccagctta cgtgactgat caacctttt tcctcaattc tgccatcaga ggcgttacaa	60
	agctggccc acatgagcta ctatcgccc tcaagaaaat tgaaaaagaa atggccgaa	120
5	ccaaaggat tagatacggc ccgcgaccca ttgacttaga tatactgtt tatggtaaac	180
	acagagttaa ctcggaaatt	200
	<210> 362	
10	<211> 200	
	<212> DNA	
	<213> Ambrosia trifida	
	<400> 362	
15	aatttccgag ttaactctgt gttaccata aaacagtata tctaagtcaa tgggtcgccc	60
	gccgtatcta ataccttggttccat ttcttttca attttcttga gggccgatag	120
20	tagctcatgt gggcccagct ttgttaacgccc tctgatggca gaattgagga aaagaggttg	180
	atcagtcacg taagctggct	200
	<210> 363	
25	<211> 200	
	<212> DNA	
	<213> Ambrosia trifida	
	<400> 363	
30	taaacacaga gttaactcgaaattctcac tgttccacat gaaagaatct tcgagaggcc	60
	atttgttatg gctccgttag ttgacttatt ggggtcgaa attgacaatg atacggttct	120
35	atgctggcat tctttctcaa aaagaggact ttttgaatct tggaaaaat tagtgtggca	180
	atctttgata ggcaaagatg	200
	<210> 364	
40	<211> 200	
	<212> DNA	
	<213> Ambrosia trifida	
	<400> 364	
45	catcttgcc tatcaaagat tcgccccatcattttccca agattcaaaa agtcctttt	60
	ttgagaaaaga atgccagcat agaaccgtat cattgtcaat ttccgacccc aataagtcaa	120
50	ctaacggagc cataacaaat ggcctctcga agattcttc atgtgaaaca gtgagaattt	180
	ccgagttAAC tctgtgttta	200
	<210> 365	
55	<211> 200	
	<212> DNA	
	<213> Ambrosia trifida	

&lt;400&gt; 365

ggcgaatctt tgataggcaa agatgggtta agaagagttt taccagtcaa taatcggtta 60  
 5 tgggatttgtt caaaagaaaac ctctgtcatg ggaattttga acttgactcc cgatagtttt 120  
 agcgacggag gaaagtttga cactgtgggg tccgctatat ctcgtgttag ggccatgata 180  
 tctgaagggg cagacataat 200

10 <210> 366  
 <211> 200  
 <212> DNA  
 <213> Ambrosia trifida

15 <400> 366

attatgtctg ccccttcaga tatcatggcc ctaacacgag atatacgcca ccccacagtg 60  
 20 tcaaaacttgc ctcgcgtcgct aaaactatcg ggagtcaagt tcaaaattcc catgacagag 120  
 gttttctttg accaatcccc taaccgatta ttgactggta aaactcttct taaccatct 180  
 ttgccttatca aagattcgcc 200

25 <210> 367  
 <211> 200  
 <212> DNA  
 <213> Ambrosia trifida

30 <400> 367

tgatatctga aggggcagac ataattgatc tgggagctca atcgacacgt ccaatggcaa 60  
 35 ccaagatttc agtccaagaa gaactagata ggttaatccc tggctcgaa aagattctt 120  
 aattaccgcga aattgaagga aaactgctct ctatcgacac gttttactca gaagttgctt 180  
 tagaagcgat taagaaaggg 200

40 <210> 368  
 <211> 200  
 <212> DNA  
 <213> Ambrosia trifida

45 <400> 368

ccctttctta atcgcttcta aagcaacttc tgagtaaaac gtgtcgatag agaggcgttt 60  
 50 tccttcaatt tcgggttaatt caagaatctt ttccgagaaca gggattaacc tatctagttc 120  
 ttcttgact gaaatcttgg ttgccattgg acgtgtcgat tgagctccca gatcaattat 180  
 gtctgcccct tcagatata 200

55 <210> 369  
 <211> 200  
 <212> DNA

<213> Ambrosia trifida

<400> 369

5	tgcttagaa gcgattaaga aaggggctca tataatcaat gatgtatcg gtggaaatct	60
	agattctgat atgttcggg tagttctaa tcgtatgtt ccatatattg ctatgcacat	120
	gagagggac ccatccacaa tgcaaaacag tgagaatttg aagtatgtg atgttgtaa	180
10	agaagtttgt gatgagttgt	200

<210> 370

<211> 200

<212> DNA

<213> Ambrosia trifida

<400> 370

20	acaactcatc accaacttct ttacaaacat catcatactt caaattctca ctgtttgca	60
	ttgtggatgg gtcccctctc atgtcatag caatatatgg aacatcaaga ttagcaacta	120
	cccgaaacat atcagaatct agattccac ccgatacatc attgattata tgagcccctt	180
25	tcttaatcgc ttctaaagca	200

<210> 371

<211> 200

<212> DNA

<213> Ambrosia trifida

<400> 371

35	tgtaaagaag ttgggtatga gttgtatgag cgtgtaaagag ctgcagaggat atgcgggttg	60
	cctgcatttgc ggatcattct tgaccaggg atcgggtttt caaagaaaac cgaagataat	120
	ttggatattt tgatggatt aaagagaatt aggagtgaga ttgcacggaa gagcttaggg	180
40	gtgtcacgtt cacctttgtt	200

<210> 372

<211> 200

<212> DNA

<213> Ambrosia trifida

<400> 372

50	aacaaaggtg cacgtgacac ccctaagctc ttccgtgcaa tctcactcct aattctcttt	60
	aatcccatca aaatatccaa attatctcg gttttcttg aaaacccgat ccctgggtca	120
	agaatgatcc tccatgcagg cacaccgcat aactctgcag ctcttacacg ctcatacaac	180
55	tcatcaccaa cttctttaca	200

<210> 373

<211> 200  
 <212> DNA  
 <213> Ambrosia trifida

5       <400> 373

ttcgtgtatc tcatgatctc tgtataatgt taaacttatt atagttcaat ataagtcact    60  
 gtgtgatcta atttcgtata caattacttt atttgtgttgc attctgatcc taaatattgc    120  
 taatagtttgc tgcttaacta tatatccgt taaacatcac atttacttta tccctagaag    180  
 atttggcaa gttttacaa    200

15       <210> 374

<211> 200  
 <212> DNA  
 <213> Ambrosia trifida

20       <400> 374

ttgtaaaaac ttgcccaaattt cttctaggaa taaaagtaaat gtgatgttta acgggatata    60  
 tagttaagca caaactatta gcaatattta ggatcagaat caacacaaat aaagtaattt    120  
 tatacgaaat tagatcacac agtgacttat attgaactat aataagtttta acattataca    180  
 gagatcatga gatacacgaa    200

30       <210> 375

<211> 200  
 <212> DNA  
 <213> Ambrosia trifida

35       <400> 375

agaagatttgc ggcaagtttt tacaaatttta tggcgacgtt attgtatattt tttaaatcat    60  
 tttcgtgcag catcttttc gttctttcat ttctcttcgg atgcttcgat acaagtccat    120  
 tctcaagaac aagaagtagt aattgcttta ggtacaatg tgggtgatacg gcttaataac    180  
 tttaatgaag ccttaagtca    200

45       <210> 376

<211> 200  
 <212> DNA  
 <213> Ambrosia trifida

50       <400> 376

tgacttaagg cttcattaaa gttttaagc ctatcaccca cattgctacc taaagcaatt    60  
 actacttctt gttcttgaga atggacttgtt atcgaagcat ccgaagagaa atgaaagaac    120  
 gaaaaagatg ctgcacgaaa atgatttaaa aaattacaat aacgtcgcca taaatttgc    180  
 aaaacttgcc caaatcttct    200

<210> 377  
 <211> 200  
 <212> DNA  
 <213> Conyza canadensis

5  
 <400> 377

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ctttttctca actctgccgt tagagccact acaaagctt g gccctcatga gctactgtcc	120
attctcaaga aaattgaaaa gaaaatgggt cgaaccaaag ggcttaggta cggcccacga	180
cccattgacc tagatatatt	200

10  
 <210> 378  
 <211> 200  
 <212> DNA  
 <213> Conyza canadensis

15  
 <400> 378

aatatatcta ggtcaatggg tcgtggccg tacctaagcc ctttggttcg acccatttcc	60
ttttcaattt tcttgagaat ggacagtgc tcatgagggc caagcttgc agtggctcta	120
acggcagagt tgagaaaaag aggctggtcc gtcacataag ctggttcagt ttctataga	180
caagcatgcc ttgttatttc	200

20  
 <210> 379  
 <211> 200  
 <212> DNA  
 <213> Conyza canadensis

25  
 <400> 379

cacgacccat tgacctagat atattgttct atggtaatg caaagttaac tctgatattc	60
taactgttcc tcatgaaaga atctttgaga ggccattcgt tatggctcca ttagttgact	120
tgttaggatc agatgtagac aatgatacgg ttctatgctg gcattcttt tcaaaaaatg	180
ggcttttgg atcttggaa	200

30  
 <210> 380  
 <211> 200  
 <212> DNA  
 <213> Conyza canadensis

35  
 <400> 380

ttcccaagat ccaaaaagcc catttttga aaaagaatgc cagcatagaa ccgtatcatt 60  
 gtctacatct gatcctaaca agtcaactaa tggagccata acgaatggcc tctcaaagat 120  
 5 tctttcatga ggaacagtta gaatatcaga gttaactttg catttaccat agaacaatat 180  
 atctaggta atgggtcgta 200  
  
 <210> 381  
 10 <211> 200  
 <212> DNA  
 <213> Conyza canadensis  
  
 <400> 381  
 15 aaatgggctt ttggatctt gggaaacatt aggtggtaa tcttcatacg gaaaagatgg 60  
 tttaagaagg gtttacctg tcaatgatcg tttatggat tggtaaaga aaacttcgt 120  
 20 catgggtatt ttgaatataa ctccgtatag ttttagtgat ggagggaagt ttgattccat 180  
 ggggtccgct ttatctcgta 200  
  
 <210> 382  
 25 <211> 200  
 <212> DNA  
 <213> Conyza canadensis  
  
 <400> 382  
 30 cacgagataa agcggacccc atggaatcaa acttcctcc atcactaaaa ctatcaggag 60  
 ttatattcaa aatacccatg acggaagttt tcttgacca atcccataaa cgatcattga 120  
 35 caggtaaaac ccttcttaaa ccatctttc ctatggaaga ttcaccaccc aatgtttccc 180  
 aagatccaaa aagcccat 200  
  
 <210> 383  
 40 <211> 200  
 <212> DNA  
 <213> Conyza canadensis  
  
 <400> 383  
 45 tccatgggt ccgtttatc tcgtgttcag accatgatat ctgaagggt tgacataatt 60  
 gatctaggag ctcaatccac acgcccattg gcgaccaaga tctcagtcga agaggaacta 120  
 50 gataggctaa tacccgttct tgaaaagatt cttgaattac ctgaaattga aggaaagttg 180  
 ttgtctgtgg acacattta 200  
  
 <210> 384  
 55 <211> 200  
 <212> DNA  
 <213> Conyza canadensis

&lt;400&gt; 384

taaaaatgtgt ccacagacaa caacttcct tcaattcag gtaattcaag aatctttca 60  
 5 agaacgggta ttagcctatc tagttcctct tcgactgaga tcttggtcgc cattggcgt 120  
 gtggatttagag ctcctagatc aattatgtca accccttcag atatcatggt ctgaacacga 180  
 gataaaagcg 10 accccatgga 200

&lt;210&gt; 385

&lt;211&gt; 200

&lt;212&gt; DNA

&lt;213&gt; Conyza canadensis

&lt;400&gt; 385

agttgttg 15 tc tgtggacaca ttttactcgg aagttgcttc agaggcaatc aagaaagggg 60  
 20 ctcatatgg 200 caatgatgta tcgggtggaa tgtagattc tgatatgctt catgttgtgg 120  
 ctgatctaaa tttccat 25 atcactatgc acatgagagg ggaccatcc acaatgcaaa 180  
 acagtgagaa tttgaagtat 200

&lt;210&gt; 386

&lt;211&gt; 200

&lt;212&gt; DNA

&lt;213&gt; Conyza canadensis

&lt;400&gt; 386

atacttcaa 30 ttctca 200 ctgttgttgg 60 gcatgggtcc cctctcatgt gcata 200  
 35 atatggaaca tttagatcag ccacaacatg aagcatatca gaatctaaca ttccacccga 120  
 tacatcattg accatatgag cccctttctt gattgcctct gaagcaactt ccgagtaaaa 180  
 tttgtccaca gacaacaact 200

&lt;210&gt; 387

&lt;211&gt; 200

&lt;212&gt; DNA

&lt;213&gt; Conyza canadensis

&lt;400&gt; 387

gcaaaacagt 45 gagaatttga agtatgatga tggtgtcaaa gaagttgggg aggaattgta 60  
 50 tgaacgtgta aggaatgcag agttatgtgg tggtcccgca tggaggatgg ttcttgatcc 120  
 agggatcggg tttcaaaga aaaccgaaga taatttgag atattgatgg gactaaagag 180  
 gtttagaagt gagattggac 200

&lt;210&gt; 388

&lt;211&gt; 200

&lt;212&gt; DNA

<213> Conyza canadensis

<400> 388

5 gtccaatctc acttctaaac ctcttagtc ccatcaatat ctccaaatta tcttcggttt 60  
tcttgaaaa cccgatccct ggatcaagaa ccatacctcca tgcgaaaaca ccacataact 120  
ctgcattcct tacacgttca tacaattcct ccccaacttc tttgacaaca tcatacatact 180  
10 tcaaattctc actgttttgc 200

<210> 389

<211> 200

15 <212> DNA

<213> *Conyza canadensis*

<400> 389

20 aagagggtta gaagtgagat tggacaaaag agcttagggg tgtctcggtc acctttgtta 60  
atcggacctt caagaaaaag gttttgggt gagatttgtg gtccggccttc tgctgttag 120  
25 agagatccag ggactgttagc tgctgttacc agtgcgattt tgggtgggtgc caatatcggtt 180  
agggttcata acattggaca 200

<210> 390

<211> 200

30 <212> DNA

## 2.1.2 DNA

<400> 390

35 tgtccaatgt tatgaacctt aacgatatttgcaccaccca aaatcgcaact ggtaacagca 60  
gctacagtcc ctggatctctt ctaaacagca gaaggccgac cacaatctc accaaaaaac 120  
cttttcttg aaggtccgat taacaaaggt gcacgagaca cccctaagct ctttgtcca 180  
40 atctcaatttcaaaacctttt 200

<210> 391

<211> 200

45 <212> DNA

<212> BNA

<400> 391

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        ctattaccca cattgctacc taaagcaatt actacttctt gttctggaga atgcacttgt 180  
        attgaagtat ccgaggacga 200

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        ggtagccccc acgaccatt gacctagata tattgttcta tggtaaatgc aaagtttaact 180  
        ctgatattct aactgttccct 200

30       <210> 394

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35       <400> 394

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 40      aatgggtcgt gggccgtacc taagcccttt ggtagacccc atttcctttt caatttctt 120  
        gagaatggac agtagctcat gagggccaag cttttagtg gctctaacgg cagagtttag 180  
        aaaaagaggc tggccgtca 200

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        aaaagatggt ttaagaaggg 200

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35  
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30 <400> 401  
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15 <400> 404

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 atccctggat caagaaccat cctccatgcg ggaacaccac ataactctgc attccttaca 180  
 25 cgttcataca attcctcccc 200

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 30 <213> Conyza canadensis

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 cgcaatctaa catctggaa ttgtacaaag tttattgaag atagtgaagg cagattcgac 180  
 40 tgccacacctac acatataaga 200

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 45 <213> Conyza canadensis

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 55 gctccaacta gaagaagaaa 200

<210> 407  
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&lt;213&gt; Conyza canadensis

&lt;400&gt; 407

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                   acatcatttgc atgttcgtc aaaaacaata ccatttctta tcttgctca atggatatga 120  
 10                aatctgccaa tacaagatta ttcatttgac ctcaatctag catctgatgt ctattcaagg 180  
                   tatcgttat aatgaagatt    200

&lt;210&gt; 408

&lt;211&gt; 200

&lt;212&gt; DNA

&lt;213&gt; Conyza canadensis

&lt;400&gt; 408

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                   taatcttgta ttggcagatt tcatatccat tgagcaaaga taagaaatgg tatttttttt 120  
 25                gacgaacatg caaatgatgt tttatattgt gagttaagta tagaagttaa tggtttctta 180  
                   tatgtgttagg tggcagtgcga    200

&lt;210&gt; 409

&lt;211&gt; 200

&lt;212&gt; DNA

&lt;213&gt; Conyza canadensis

&lt;400&gt; 409

30                caaggtatcg tttataatga agattattgt acttcaaaat actttcttgt gtagtggtat 60  
 40                caaattttga gttaaagtttta ctccatttga cctcaaaagc caaataatga gcaacaaaca 120  
                   acacaatttc acctccctta taccctgcga ctatatttta tcttcaaatc aatccatttc 180  
                   catgtttatc ctctccatac    200

&lt;210&gt; 410

&lt;211&gt; 200

&lt;212&gt; DNA

&lt;213&gt; Conyza canadensis

&lt;400&gt; 410

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 5 taactttaac tcaaaatttg ataccactac acaagaaaagt attttgaagt acaataatct 180  
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 20 atatattctt tatttacaac tataaatagc atatgggtat gttggtat agatatacat 180  
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 35 ctgtaaacctt agtttagttt agtttggttt ttccctaaa acttattatg tagctgtatg 180  
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40 <210> 413  
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 50 aaccccccctt tctatgggtt 200

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10	tacacaggta atatatgttt cctttttctt ccccatttac acacatataa tacacatcta	180
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	tccaaattt caaaaccga aataatgtac gacattcggt gcacaaggta caaacagaaa	120
25	aaactacttt tgtatTTAG atctcaagaa aggagtggcc ttttggtaa taacataactt	180
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<400> 416

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40	tcggtttgg aaaatttggaa ttgtaaaaga gataaatgtaa aagccctcat cttaaaccc	180
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<210> 417

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 35  
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 45 taagaatctt tttataggca 200

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 20 gcattacagg gatgctctac agtttgcata cacaagtccc agtggtagct ctattcaatt 180  
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30 <400> 423  
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 35 ccataaccct agggcaacac gagcctttag attatgaatg ggcacaccca attgtttaga 180  
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 50 ttcaaacaca aacttcatcc taattcttat caattctaac attcgcgact ccctattgta 180  
 tcattcagta atgagcataa 200

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10	tgacggacca gcctctttt ctcaactctg ccgttagagc cactacaag cttggccctc	180
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10 ataggaaaag atgguttaag aagggttta cctgtcaatg atcgtttatg ggattggta 180  
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 20 gtgcacctt gttaatcgga ccttcaagaa aaaggtttt gggtagatt tgtggtcggc 180  
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 35 cctcttagt cccatcaata tctccaaatt atttcggtt ttcttgaaa acccgatccc 180  
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 50 ttgtgattca atgttggatc gagctggtag atcttagtag cttaaagtta tttgacatct 180  
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 tttactttt cagttttct tggtagttt tgttttgaa aatttgctag atcagattaa 180  
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10	taaaaggagt ttacaatacc actaaatgaa actcgagcg tcccaaaaaa gacacgaaga	180
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25	accacaagaa accaaaactg tcttgcat ttctctctt tccaggtaaa aactcaata	180
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40	tgtcttagact cttgcttca ttttagaaag cataacttgc actctgtgga atcgaattat	180
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55	ttttctgtgc aattcctctg caacttgctt tgatgcttgg tcttctttt gaaggcgttg	180
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10 acaccaaaga tgatcctatg gctatagctg aagctgaatc attgaagcag aataatagtt 180  
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25 taagaaaaaa aatgttagtga tgatttaact caaaacaact cacacagctc caaaatggat 180  
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40 ttgtttggc tctaaattac agtttggcc gatgcggata catggttga acgacagcaa 180  
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50 <400> 455

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aaaaaacgat catggcaatg tattacacaa tctaaagcaa ccactatgaa taataaagat 120  
55 aaatcttcc gagagtccac atgccttgcatttccataag ataggcttt tcagaaatca 180  
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	cattgccatg	atcgaaaa	tttccccata	attagaagct	tctatctatt	aggctatcct	180
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20                  <400> 457

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	aatatcaagt	aaccacatta	tgaattgact	tgtttaatca	aagttcatct	tacttctca	180
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30                  <210> 458  
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35                  <400> 458

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	tactatgg	ac	ac	aaataaagat	ttacttaata	acatgcatta	gactgat	ttt	agtgttat	180
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50                  <400> 459

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 20 tcttcctaca gctaggAGTA attatatgtt agaacaactg atgtgttctg tatttgcata 180  
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 10 tttgttaatt ggaaagttt 200

&lt;210&gt; 464

&lt;211&gt; 200

&lt;212&gt; DNA

15 &lt;213&gt; Conyza canadensis

&lt;400&gt; 464

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 ccataagtac atctgatgaa ccaatttagac ttgtcacaaa tatcaaaccg agagtcggac 180  
 25 atgtgatgct tttagttgaa 200

&lt;210&gt; 465

&lt;211&gt; 200

&lt;212&gt; DNA

30 &lt;213&gt; Conyza canadensis

&lt;400&gt; 465

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&lt;210&gt; 466

&lt;211&gt; 200

&lt;212&gt; DNA

45 &lt;213&gt; Conyza canadensis

&lt;400&gt; 466

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&lt;210&gt; 467

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&lt;212&gt; DNA

&lt;213&gt; Conyza canadensis

&lt;400&gt; 467

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 10          aaatctgccca atacaagatt attcatttga cctcaatcta gcatctgatg tctattcaag 180  
           gtatcgutta taatgaagat   200

&lt;210&gt; 468

&lt;211&gt; 200

&lt;212&gt; DNA

&lt;213&gt; Conyza canadensis

&lt;400&gt; 468

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           acgaacatgc aaatgatgtt ttatattgtg agttaagtat agaagttaat ggtttcttat 180  
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&lt;211&gt; 200

&lt;212&gt; DNA

&lt;213&gt; Conyza canadensis

&lt;400&gt; 469

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           aacacaattt cacccctt ataccctgctg actatatttt atcttcaaat caatccattt 180  
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&lt;211&gt; 200

&lt;212&gt; DNA

&lt;213&gt; Conyza canadensis

&lt;400&gt; 470

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 50          aaggaggatg aaattgtgtt gtttggct cattatttgg cttttgaggt caaatggagt 120  
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	gagtgaaact gggaaacccct ttttaattg tggtaattt tgtaatttc ttgaaggtaa	180
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 15 aacaatttta tgtacaaattt 200

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<213> Digitaria sanguinalis

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25	ttcatacacctc aaatttgtct ttgctttctt attgaccagg tgtgatggag tgatgcttgc	180
	gtgaaatatac ctacaaggct	200
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	<213> Digitaria sanguinalis	
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40	cattagacgtc aaataaacctt ctcaatgtaa ggagaggaga tgccccactg gaggcaagca	180
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	<210> 545	
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55	atgaactact ttggagggtt cctccatcg cattcgttt caggtactgt tcaaatcttt	180
	tctttcaaa tcccctatca	200

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5                   <400> 546

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                 aggagcatcg ttgaacatat gataattaag aaagcaaatg cttcaaataa attgttagcct   180  
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20                <400> 547

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                 cctagacatg ctattccatt cagggctcgt tcgtttacgc gatgttcgct tgagggatgt   180  
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 35                <213> Digitaria sanguinalis

35                <400> 548

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                 actgtttaga aatgcaaagg ggtgagttag aagcttagag ctgctttga tcgcatgata   180  
 45                ggggatttga aaagaaaaga   200

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 50                <213> Digitaria sanguinalis

50                <400> 549

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30 <400> 551  
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 35 agatataactt ctgtacggca attcccagat tgataactgag gctctaattt tgccacatga 180  
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 <213> Digitaria sanguinalis

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 50 gtcctatatac cttctcaatt tccttagtt tcttaagcaa ctcgtgaggt cccagcctag 180  
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 tttgaattat ggaataaaact tgggggtgaa tctataattg gaacagaaaag tattaaaagg 180  
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&lt;400&gt; 554

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 tcgacactat cattgccaga tgacactaga aggtcaacaa gaggtgctag aacaaatggc 180  
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 <213> Digitaria sanguinalis

&lt;400&gt; 555

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 ctgctatttc tcaggctaaa ttattaattt cagaaggtgc agatatcata gatattggtg 180  
 ctcaatctac caggcctttt 200

40 <210> 556  
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 <212> DNA  
 <213> Digitaria sanguinalis

&lt;400&gt; 556

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<213> Digitaria sanguinalis

<400> 557

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10	agtggataca ttctatgcag aagttgcatg tgaagctgtg aaaagaggag ttcacatgat	180
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<210> 558

<211> 200

<212> DNA

<213> Digitaria sanguinalis

<400> 558

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25	catccagaac aggaaccaat cttcaattt cttcggttgg agataatctc tttgcaaaag	180
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<210> 559

<211> 200

<212> DNA

<213> Digitaria sanguinalis

<400> 559

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	ctcaaagttc cgtatgttgc aatgcacatg aggggagatc catcaacaat gcaaagtgaa	120
40	caaaaatttac agtatgacga tgtctgcaag gaagttgctt cagagctata tgcacaggtg	180
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<210> 560

<211> 200

<212> DNA

<213> Digitaria sanguinalis

<400> 560

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	tcgtcatact gtaaattttg ttcaacttgc attgttcatg gatctcccatt catgtgcatt	120
55	gcaacatacg gaactttgag ttcaagcagca actttaagaa ttttgggtc aagctgtcca	180
	ccagatacat cattgatcat	200

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5            <400> 561

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ggagggagat gagtaaaatg agtattggtg cttcacatgt gccaatattt ctgggaccct	180
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<213> Digitaria sanguinalis

15          <400> 562

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ggatttcttg gagaacccaa tgcctggatc tagaactatc ctccacaatg gaatcccaga	180
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<213> Digitaria sanguinalis

25          <400> 563

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acataatgca ggatacagtt cagatgctgc aaagttttgt gatgcattga ataagagaag	180
aagaatggaa gactgaacca	200

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35          <400> 564

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cagctgcaac agtagcaaca tctctctcaa ctggattttc acgattgcattt atttcaccctt	180
aaaatctttt ccttagagggt	200

<210> 565  
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5                   <400> 565

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	gagaaaaatgg tcatgcagga tagttactct gctgctaat gggattctca tattacatca	120
	tttggagt attgttttg taataaataa accagggatg acgttttcc tgtgtcatct	180
	cctatctagt ttcttagatac	200

15                  <210> 566  
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 <213> Digitaria sanguinalis

20                  <400> 566

25	gtatctagaa actagatagg agatgacaca ggaaaaacgt catccctggt ttatttatta	60
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	tcctgcataa ccattttctc gaataaaatc agagctcggt atcttttga tcagctggtt	180
	cagtcttcca ttcttcttct	200

30                  <210> 567  
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 <212> DNA  
 <213> Digitaria sanguinalis

35                  <400> 567

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	aagagtagtc aaggaataag gttacagaat cagttagtaa ttacagattc ttaccttttg	120
	tggcctatt ttagatgatt ttccagaaca ttttcttcc aatttaataa tatcagaagg	180
	gcctaatttg tgtgtatctt	200

45                  <210> 568  
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 <213> Digitaria sanguinalis

50                  <400> 568

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 5 cttattcctt gactacttt tccaagtgtat ctgagttgaa ccatgcttcA ttgaagtatC 180  
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 20 aaaagctcAG gcgtgaACat cactaggcat gcctgtctat atgagactgc ccctgcttat 180  
 gtgaccgatC agccacggTT 200

25 <210> 570  
 <211> 200  
 <212> DNA  
 <213> Digitaria sanguinalis

30 <400> 570  
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 35 cccacgttgc ttcccataGC aatcacaATC tcttggTCAG ttgaACatCC ctcaAGcgAA 180  
 catcgcgtaa acgaacgagc 200

40 <210> 571  
 <211> 200  
 <212> DNA  
 <213> Digitaria sanguinalis

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 50 gaataaggta tggcccaAGA ccaatcgatC tagatataCT tctgtacggc aattcccaga 180  
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55 <210> 572  
 <211> 200  
 <212> DNA  
 <213> Digitaria sanguinalis

&lt;400&gt; 572

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 cttaagcaac tcgtgaggtc ccagcctagt cgtacacctga atggcagagt tcaggaaccg 180  
 10 tggctgatcg gtcacataag 200

&lt;210&gt; 573

&lt;211&gt; 200

&lt;212&gt; DNA

&lt;213&gt; Digitaria sanguinalis

&lt;400&gt; 573

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 gcactctctt tcaaagtgcg gtgggtgttt ctttgaatta tggaataaac ttgggggtga 180  
 25 atctataatt ggaacagaaaa 200

&lt;210&gt; 574

&lt;211&gt; 200

&lt;212&gt; DNA

&lt;213&gt; Digitaria sanguinalis

&lt;400&gt; 574

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 35 tgcactttga aagagagtgc cagcttgttt cgacactatc attgccagat gcacctagaa 120  
 ggtcaacaag aggtgctaga acaaatggcc tctcctggat gcgttcatgt ggcacaatta 180  
 40 gagcctcagt atcaatctgg 200

&lt;210&gt; 575

&lt;211&gt; 200

&lt;212&gt; DNA

&lt;213&gt; Digitaria sanguinalis

&lt;400&gt; 575

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 50 tttgatttgt gtgagagaac cctcgtcatg ggggtcctta atctaacacc agacagctt 120  
 agtcatggag gtaagttca acaagtggaa gctgctattt ctcaggctaa attattaatt 180  
 55 tcagaaggta cagatatcat 200

&lt;210&gt; 576

&lt;211&gt; 200

&lt;212&gt; DNA

<213> Digitaria sanguinalis

<400> 576

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	tgaaacctac ctccatcact aaagctgtct ggtgttagat taaggacccc catgacgagg	120
10	gttctctcac accaatcaaa caaatgattc ccaacaggtta atacccttt aatactttct	180
	gttccaatta tagattcacc	200

<210> 577

<211> 200

<212> DNA

<213> Digitaria sanguinalis

<400> 577

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25	aaattcctga aatggaggga aagttgctct cagtggatac attctatgca gaagttgcat	180
	gtgaagctgt gaaaagagga	200

<210> 578

<211> 200

<212> DNA

<213> Digitaria sanguinalis

<400> 578

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40	ttcggttggaa gataatctct ttgcaaaagg cctggtagat tgacccacaa tatctatgat	180
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<210> 579

<211> 200

<212> DNA

<213> Digitaria sanguinalis

<400> 579

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	tgaccccaaa attctaaag ttgctgctga actcaaagtt ccgtatgttgc aatgcacat	120
55	gaggggagat ccatcaacaa tgcaaagtga acaaaattta cagttatgacg atgtctgcaa	180
	ggaagttgct tcagagctat	200

<210> 580

<211> 200  
<212> DNA  
<213> Digitaria sanguinalis

5        <400> 580

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          10      cttaagaat tttgggtca agctgtccac cagatacatc attgatcatg tgaactcctc    180  
                  tttcacagc ttcacatgca    200

15      <210> 581  
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<212> DNA  
<213> Digitaria sanguinalis

20      <400> 581

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          ccattgtgga ggatagttct agatccaggc attgggtct ccaagaaatc caaacataac    120  
          25      cttgaagtaa ttatggatt ggaatccatt aggagggaga ttagtaaaat gagtatttgt    180  
                  gcttcacatg tgccaatatt    200

30      <210> 582  
<211> 200  
<212> DNA  
<213> Digitaria sanguinalis

35      <400> 582

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          aatcccataa ttacttcaag gttatgtttg gatttcttgg agaacccaa gcctggatct    120  
          40      agaactatcc tccacaatgg aatcccagat aactctgctt ctctcacctg tgcatatacg    180  
                  tctgaagcaa cttccttgca    200

45      <210> 583  
<211> 200  
<212> DNA  
<213> Digitaria sanguinalis

50      <400> 583

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          tatgcaatcg tgaaaatcca gttgagagag atgttgctac tgttgcagct gtgacggcgg    120  
          55      gtatTTGAA tggtgctaAC atagtaAGGG tacataatgc aggatacagt tcagatgctg    180  
                  caaagtttg tgatgcattg    200

<210> 584  
<211> 200  
<212> DNA  
<213> Digitaria sanguinalis

5  
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tggattttca cgattgcata tttcacccaa aaatctttc ctagagggtc ccagtaatat 180  
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15  
<210> 585  
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<212> DNA  
<213> Euphorbia heterophylla  
20  
<400> 585

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25 atttaaacca tgttttaaaa atagtgttat gaaaaactcc gcttgggcgc cgtctagata 120  
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&lt;211&gt; 200

&lt;212&gt; DNA

&lt;213&gt; Euphorbia heterophylla

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&lt;211&gt; 200

&lt;212&gt; DNA

&lt;213&gt; Euphorbia heterophylla

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&lt;213&gt; Euphorbia heterophylla

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 tttttgttg tataaatata 200

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30 <400> 634  
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45 <400> 635  
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 50 aaccaAGcat tgcaattaat gaaaAGttcc ggcattaaca taactagaca tggttgttG 180  
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55 <210> 636  
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&lt;400&gt; 636

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 5 attaattgca atgcttggtt gaaattatga actctatctc ccacattgct tcctaattgca 120  
 attactactt cttgctcttg agaatgaact tccaccgaac tatttggttc ggtatggaga 180  
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10 <210> 637  
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15 <400> 637

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25 <210> 638  
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30 <400> 638

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40 <210> 639  
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45 <400> 639

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<211> 200

15           <212> DNA

<213> Euphorbia heterophylla

<400> 641

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30           <212> DNA

<213> Euphorbia heterophylla

<400> 642

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       caatccata aatggttcc gatcggttta actctttca ttccgtcctt cccgatcagg   180  
 40           cttcgcgc ccaactttt   200

<210> 643

<211> 200

45           <212> DNA

<213> Euphorbia heterophylla

<400> 643

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       aagcggttgtt gaaaatcccc gagatgaccg aaaagctcat atcggtcgac acatttactt   180  
 55           cggaagtgcgc ctttagaagcg   200

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5 <400> 644

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ggaaatcatc gagcgaacct 200

15 <210> 645

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20 <400> 645

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taatgtttgt aaagaggttg 200

30 <210> 646

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35 <400> 646

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45 <210> 647

<211> 200  
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50 <400> 647

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acgaatcaga atctggaaat cctgatggga attccaagga tccggccga gattgggagg 180  
aaaagcgtgg gtatttctcg 200

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5                   <400> 648

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	caggctggaa ttccggacat ctcggcttct ttaaccctcg aaaacaactc aaaggcaacc	180
	tctttacaaa cattatcgta	200

15                   <210> 649  
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 <213> Euphorbia heterophylla

20                   <400> 649

25	ggagggaaaag cgtgggtatt tctcgtgggc cgattcttat tgggccttcg agaaagaggt	60
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	ctgttactac tggattctg ggagggcaa atattgttag agttcataat gtagggata	180
	atgtggatgc tgtgaagcta	200

30                   <210> 650  
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35                   <400> 650

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	ttcgcaaatt tcgccccaaa acctctttct cgaaggccca ataagaatcg gcccacgaga	180
	aatacccacg ctttccttc	200

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50                   <400> 651

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 10 <211> 200  
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 45 <400> 654  
  
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&lt;400&gt; 655

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15 <400> 656

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 25 actttccata gaaaagtata 200

30 <210> 657  
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 40 cgctcgatga tttccgaaagg 200

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 55 ctttcggcgc ccaactttc 200

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&lt;213&gt; Euphorbia heterophylla

&lt;400&gt; 659

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 10                aagcggttgt gaaaataacct gagatgaccg gaaagctcat atcggtcgac acatttact   180  
                   cggaagtgcgc cttagaagcg    200

&lt;210&gt; 660

&lt;211&gt; 200

&lt;212&gt; DNA

&lt;213&gt; Euphorbia heterophylla

&lt;400&gt; 660

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 25                tatcctttta gccatagggc gtgtcgattg agcaccaaaa tcgacaatat ccgccccttc   180  
                   ggaaatcatc gagcgaacct    200

&lt;210&gt; 661

&lt;211&gt; 200

&lt;212&gt; DNA

&lt;213&gt; Euphorbia heterophylla

&lt;400&gt; 661

30                ttactcgaa gtcgccttag aagcggtcca aagaggagca aatctcgtaa atgacgtatc   60  
 40                tggggggcaa ttagatccaa aaatgacgga aaatgtcgct aatctcgagg taccatatat   120  
                   cttaatgcac atgagggggg acccccaccac aatgcagaac actcagaatc taaaatacga   180  
                   taatgtttgt aaagaggttg    200

&lt;210&gt; 662

&lt;211&gt; 200

&lt;212&gt; DNA

&lt;213&gt; Euphorbia heterophylla

&lt;400&gt; 662

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 10 <211> 200  
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<213> Euphorbia heterophylla

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15 <212> DNA

<213> Euphorbia heterophylla

<400> 667

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30 <212> DNA

<213> Euphorbia heterophylla

<400> 668

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45 <212> DNA

<213> Euphorbia heterophylla

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35 <400> 672

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50 <213> Euphorbia heterophylla  
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## EP 2 755 987 B1

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15 <400> 678

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10	ggtatcacaa acacaaagac acccttaaag ctcctatgtt tgtggagagt tgtaagtcga	180
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<210> 682

<211> 200

<212> DNA

<213> Euphorbia heterophylla

<400> 682

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25	tttgatattt gatattttcg gtttattaaat actttttcta ttttattaaatc tggatcattt	180
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<210> 683

<211> 200

<212> DNA

<213> Euphorbia heterophylla

<400> 683

35	gtcgactaat ttcaagattgc aaaagttaaa aatggccaac ttttttagct atgaggtag	60
	attagatacg agaatgcctt ggtggaaagc tggtcactgt ttcttcattc gcgtggcaat	120
40	taacccggca taatcgatac ccaattcacg gacaagggtca aaggtatcag cttaactcaa	180
	cccttcgctt tgatactaaa	200

<210> 684

<211> 200

<212> DNA

<213> Euphorbia heterophylla

<400> 684

50	tttagtatca aagcgaaggg tttagtaaag ctgatacatt tgaccttgc cgtgaattgg	60
	gtatcgatta tgccgggta attgccacgc gaatgaagaa acagtgacca gctttcacc	120
55	aaggcattct cgtatctaatt ctaacctcat agctaaaaaa gttggccatt tttaactttt	180
	gcaatctgaa attagtcgac	200

<210> 685

<211> 200  
<212> DNA  
<213> Euphorbia heterophylla

5        <400> 685

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10	tacttcattt	tattgtctat	gcacataagg	tgttcgacaa	aaagcctgaa	tgaaaactgtg	120
	agtgctggta	cgcggttaca	atactcctga	ttatgtccaa	tgattatttgc	ggtactttaa	180
	taccctaatt	tgtgaagttc					200

15      <210> 686  
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<212> DNA  
<213> Euphorbia heterophylla

20      <400> 686

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25	tgtaaccgcg	taccagcact	cacagttca	ttcaggcttt	ttgtcgaaca	gcttatgtgc	120
	atagacaata	aatgaagta	acgaaataaaa	gcataatcag	aataagaatc	agcaatttag	180
	tatcaaagcg	aagggtttag					200

30      <210> 687  
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<212> DNA  
<213> Euphorbia heterophylla

35      <400> 687

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40	tcattttatg	aatccccaaag	tctttgaagt	acagctgatt	aaaatgcaag	tactcataag	120
	ttgtcattgt	atgtttgaag	tccttacatt	tttggatttt	tacagaattt	gtactgaaga	180
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45      <210> 688  
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<213> Euphorbia heterophylla

50      <400> 688

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	cttggggatt	cataaaatga	taaataaccg	gggaagaaac	taccctttcg	ctagtgaact	180
	tcacaaatta	gggttattaaa					200

<210> 689  
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 <212> DNA  
 <213> Euphorbia heterophylla

5                   <400> 689

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10	cggctggtgt ccactaaaca agggttcaat aatgccatca accgtttgg aggtaatata	120
	gttatgccta tgatgctta agtcgcttt catcgatttgc ttacattacc aggctaagat	180
	tctgcaattt aagttagaaa	200

15                   <210> 690  
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 <213> Euphorbia heterophylla

20                   <400> 690

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25	taaagcatca taggcataaac tatattacct ccaaaacgggt tgatggcatt attgaaccct	120
	tgttagtgg acaccagccg cttgaggaga agcatattag gaacaaggaa gcctaactag	180
	gtatgaattt atgcttcttc	200

30                   <210> 691  
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 <213> Euphorbia heterophylla

35                   <400> 691

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40	tttgcaccc tggggccggc tcaagattgc ttcccaggt tgctgtcttgc acatcacgtc	120
	ctaatccaaa accctaaggat gatgggttaa tgggtccctt cgcaattata tatcattcac	180
	ttaactcatt aacttccatg	200

45                   <210> 692  
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50                   <400> 692

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 5 accggcccca gggtgacaaa acaaaatcca tactttcaa gacaagactc gtcgatttct 180  
 aacttcaatt gcagaatctt 200

10 <210> 693  
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 <212> DNA  
 <213> Euphorbia heterophylla

15 <400> 693  
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 20 aagagccaaa atctttgac caagacccaa atatgttgc ttcatgttgc agcattgtca 180  
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25 <210> 694  
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 <213> Euphorbia heterophylla

30 <400> 694  
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 35 aaggtcaaaa tctacgttta tacagtgttgc gggcaagtgt ggagttgatt atccacatgg 180  
 aagttaatga gttaagtgaa 200

40 <210> 695  
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45 <400> 695  
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 50 cattgcaatt aatgaaaagt tccggcatta acataactag acatggttgt ttgtaccaga 180  
 cagcacctgc ttatgtcact 200

55 <210> 696  
 <211> 200  
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 <213> Euphorbia heterophylla

&lt;400&gt; 696

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 5 acttttcatt aattgcaatg ctgggtgaa attatgaact ctatctccca cattgcttcc 120  
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 atggagaaaag gatgatgaca 200

10 <210> 697  
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15 <400> 697

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 gggccgtacc aaagggatta ggtacggacc aaggccaatc gattggata tactttcta 180  
 25 tggaaaagttc cgattaaatt 200

25 <210> 698  
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 <212> DNA  
 <213> Euphorbia heterophylla

30 <400> 698

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 35 taatcccttt ggtacggccc aagtcctttt cgatctgctt taaaactcct aataactcgt 120  
 ggggtccgag tttcgtgaaa gctcgaactg ctgagttgag aaagtgaggt tggtcagtga 180  
 40 cataaggcagg tgctgtctgg 200

40 <210> 699  
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45 <400> 699

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 50 gagagaccct ttgtgatggc cccattggtg gatttactcg gctcgaaat cgagaatgac 120  
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 55 ggcgaaaagcc tgatcgaa 200

<210> 700  
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<213> Euphorbia heterophylla

<400> 700

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       gccatcacaa agggtctctc ccatattctc tcatgaggaa caataagtgt atcggatta     180  
 10        atccggaact ttccatagaa    200

<210> 701

<211> 200

15        <212> DNA

<213> Euphorbia heterophylla

<400> 701

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       gtttcagcga tggggcaag ttgacaacta ttgattctat agtttctaag gttcgctcga    180  
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<210> 702

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30        <212> DNA

<213> Euphorbia heterophylla

<400> 702

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       ttccgaccaa tccctataat ggtttccgat cggggtaact ctttcattt cgtcattccc    180  
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<210> 703

<211> 200

45        <212> DNA

<213> Euphorbia heterophylla

<400> 703

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       ggctaaaagg atatccccgc aagaggaaat ggataggcta atccctgtat tggaaagcggt    120  
       tgtgaaaata cccgagatga ccgaaaagct catatcggtc gacacattt actcgaaat    180  
 55        cgcccttagaa gcggtccaaa    200

<210> 704

<211> 200  
<212> DNA  
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5 <400> 704

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10 gcggggatat ccttttagcc atagggcgtg tcgattgagc accaaaatcg acaatatccg 180  
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15 <210> 705

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20 <400> 705

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25 cacatgaggg gggaccccac cacaatgcag aacactcaga atctaaaata cgataatgtt 180  
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30 <210> 706

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35 <400> 706

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40 gtcatttttg gatctaatttgc ccccccagat acgtcattta cgagatttgc tcctctttgg 180  
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45 <210> 707

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50 <400> 707

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gaattccagc ctggcgaatg tttattgacc ctggaattgg gtttcgaag aacacgaatc 120  
55 agaatctgga aatcctgatg ggaattccaa ggatccgggc cgagattggg aggaaaagcg 180  
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5  
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 cattcgccag gctggaattc cgacatctc ggcttctta accctcgaaa acaactcaaa 180  
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15  
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 20  
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 25 cgaaatttgc gaacgcccctg aagcaactga aagagaccgg gcaactgttg cttctgttac 120  
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 tgctgtgaag ctatgtgact 200

30  
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 35  
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 40 cccctcccaag aatcccagta gtaacagaag caacagttgc cgggtctctt tcagttgctt 120  
 cagggcggttc gcaaatttgc cccaaaaacc tctttctcga aggcccaata agaatcgcc 180  
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 50  
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 10 <211> 200  
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 25 <211> 200  
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 35 aattagaagt tatattgaag ggtatattagg ataaataaaa gtccaaacca caaaaaaaaaa 180  
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&lt;400&gt; 715

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 10 cgaatcattt tggacttttc 200

&lt;210&gt; 716

&lt;211&gt; 200

&lt;212&gt; DNA

15 &lt;213&gt; Euphorbia heterophylla

&lt;400&gt; 716

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 25 atagttttac ggttttttt 200

&lt;210&gt; 717

&lt;211&gt; 200

&lt;212&gt; DNA

30 &lt;213&gt; Euphorbia heterophylla

&lt;400&gt; 717

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 aatttgttat aactccacca agatttaatt tttccaatc tgaaaatgtc ttatgttata 180  
 40 gtaagaagtg agatgagttt 200

&lt;210&gt; 718

&lt;211&gt; 200

&lt;212&gt; DNA

45 &lt;213&gt; Euphorbia heterophylla

&lt;400&gt; 718

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 55 gtccaaaatg attcgaaac 200

&lt;210&gt; 719

&lt;211&gt; 200

&lt;212&gt; DNA

&lt;213&gt; Euphorbia heterophylla

&lt;400&gt; 719

5           ttatagtaag aagttagatg agtttatgtc tttgtaaaa gatttgccc aattaatgca   60  
          aaaattttgt gacaagacaa gaggggtgaa agagacaagc aacaagagaa agcagtgaaa   120  
 10          ggggcaaatg ttaggaaact gtttagctaga aaagggtaga gggtggccta cttagccat   180  
          ttgagcatta aaaaagcaca   200

15          &lt;210&gt; 720

&lt;211&gt; 200

&lt;212&gt; DNA

&lt;213&gt; Euphorbia heterophylla

20          &lt;400&gt; 720

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          agtttcctaa catttgc(cc) tttcactgct ttctcttggtt gcttgcctt ttcacccctc   120  
          ttgtcttgc acaaaatttt tgcatatt gggcaaaatc tttcacaaa gacataaact   180  
          catctcactt cttaactataa   200

30          &lt;210&gt; 721

&lt;211&gt; 200

&lt;212&gt; DNA

&lt;213&gt; Euphorbia heterophylla

35          &lt;400&gt; 721

40           ccaaatttgag cattaaaaaa gcacatggct atgtatTTTt gacAAAGTC CCTATTGTTG   60  
          gcagatgaac taagcaacca tatggttcat taactaacta actagttatt gcaaatttga   120  
          ctgttccaat tcataaagg actctcccc tagaccattc aacacttggg ttctagccct   180  
          ccctctcacc ctcagacaac   200

45          &lt;210&gt; 722

&lt;211&gt; 200

&lt;212&gt; DNA

&lt;213&gt; Euphorbia heterophylla

50          &lt;400&gt; 722

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          cctttgatga attgaaacag tcaaatttgc aataactagt tagttgtta atgaaccata   120  
          tggttgctta gttcatctgc caacaatagg gactttggtc taaaatacat agccatgtgc   180  
          tttttaatg ctcaaattgg   200

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 <213> Euphorbia heterophylla

5  
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 15 catcaaacta gggttctaaa 200

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 <212> DNA  
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 30 acaagtgtag tcctaacaaa aaaggggacc aatgatgaaa atgaggttca caagggttgt 180  
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<400> 725

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 45 ggtggattag attgaaaacg actattttt gttgtataaa tatatctaaa aagtatata 180  
 attacattct ttattggaat 200

<210> 726  
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 <212> DNA  
 <213> Euphorbia heterophylla

<400> 726

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 5 ttgaatttag aatgatctcg caagttcaat ccgaaccaga gtactgaagt atgggttag 180  
 aaccctagtt tgatgttgc 200

10 <210> 727  
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 <212> DNA  
 <213> Euphorbia heterophylla

15 <400> 727

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25 ttcattaatt ctgtttcaa ttgtcaattt aaattttcat aatattttc accattttt 180  
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30 <210> 728  
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 <213> Euphorbia heterophylla

35 <400> 728

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 ttgaaaacag aattaatgaa acttttaat gaactattga aactagacga agggagtagc 120  
 ttatcgaact ttcaaaatgg ctacaacaaa gtcaaattct tatatatattt gatgaattcc 180  
 aataaagaat gtaatttat 200

45 <210> 729  
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 <212> DNA  
 <213> Euphorbia heterophylla

50 <400> 729

55 tttttttcc aatttgcct tgtatttattt gagaaagaga ttatgttatg atgaaagtat 60  
 aagttgctaa tcaagaagaa agagattta atgtaattat ttaaataata aattcctacc 120  
 attttaattt gttgacaattt gttataatg attccgttgc atcgacaact aaaactagat 180  
 agaaggatca aaatataaaa 200

<210> 730  
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<213> Euphorbia heterophylla

<400> 730

5           tttatattt tgatccttct atctagttt agttgtcgat gcaacggaat cattataacc   60  
       aattgtcaca caattaaaat ggttagaatt tattattaa ataattacat taaaatctct   120  
       ttcttcttga ttagcaactt atacttcat cataacataa tctcttctc aataaataca   180  
 10           agggcaaatt ggaaaaaaa   200

<210> 731

<211> 200

<212> DNA

<213> Euphorbia heterophylla

<400> 731

20           tagatagaag gatcaaataa taaaattta acataattaa tacatataat atataaatgt   60  
       ataatagata caattattat tcctcgaatt aaaattggct aattatataa taatataatc   120  
 25           caacctataa aaactttga gctttaagc ttgagatgag ctgtaaaa atgattgtac   180  
       aagcccgacc atgaaagaga   200

<210> 732

<211> 200

<212> DNA

<213> Euphorbia heterophylla

<400> 732

35           tctttcat ggtcggtt gtacaatcat tttacaaag ctcatctcaa gctaaaagc   60  
       tcaaaagttt ttatagggtt gattatatta ttatataatt agccaatttt aattcgagga   120  
 40           ataataattt tatctattat acatttat attatatgtt ttaattatgt taaaatttt   180  
       tattttgatc cttctatcta   200

<210> 733

<211> 200

<212> DNA

<213> Kochia scoparia

<400> 733

50           gaatgtgctt aggccgcgtc aatcagccag tttgatttg aaggaaattt ccaagtatga   60  
       tagaggtaaa gcacagttgt ttcattttt gactatataat ttaatttggg aatctgtact   120  
 55           gtttcatttt gtaactttat tttgatcctg agttattgaa tctgtcatgt ctcattttga   180  
       gacggtattt tgaatctgag   200

<210> 734  
 <211> 200  
 <212> DNA  
 <213> Kochia scoparia

5  
 <400> 734

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 ataaaagttac aaaatgaaac agtacagatt cccaaattaa atatatagtc aaaaaatgaa 120  
 acaactgtgc tttacctcta tcatacttgg aaattccctt ccaaatcaa ctggctgatt 180  
 gcagccgcct aagcacattc 200

15  
 <210> 735  
 <211> 200  
 <212> DNA  
 <213> Kochia scoparia  
 20  
 <400> 735

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 25 tataaatata gggtttaagt tgatgatctg gatgcttatca gagctctatt atgcaggatt 120  
 acaaatcata atttcattgt gaaatttact tctttatgtg tattcacttg aagattcata 180  
 30 attttgatgt attcataaaat 200

35  
 <210> 736  
 <211> 200  
 <212> DNA  
 <213> Kochia scoparia

<400> 736

40 atttatgaat acatcaaaat tatgaatctt caagtgaata cacataaaga agtaaatttc 60  
 acaatgaaat tatgatttgt aatcctgcat aatagagctc tgatagcatc cagatcatca 120  
 acttaaaccc tatatttata aatacctatg gcctattcat aatttcatat ccagactcag 180  
 45 attcaaaaata ccgtctcaaa 200

50  
 <210> 737  
 <211> 200  
 <212> DNA  
 <213> Kochia scoparia

<400> 737

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 5 gattgatgtc atgattctaa aatgccttta cctaattcaa atggatttt aagattacgt 180  
 atgtacttt gcagcttcaa 200

10 <210> 738  
 <211> 200  
 <212> DNA  
 <213> Kochia scoparia

15 <400> 738  
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 20 ttttccccct tatactttca gcacatgctg actctttctg ttttagggcac atcaaattt 180  
 tgaatacatc aaaatttatga 200

25 <210> 739  
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 <212> DNA  
 <213> Kochia scoparia

30 <400> 739  
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 35 agattagata actttaacca agctctgcag ctaatgaaga aattaggagt aaatatcaca 180  
 aggcatggct gcttatacga 200

40 <210> 740  
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 <212> DNA  
 <213> Kochia scoparia

45 <400> 740  
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 50 tctttagaac aaacctcaat agtagttct ggtgatgagt gcaaaaaagc aagacttgaa 180  
 gctgcaaaag tacatacgt 200

55 <210> 741  
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 <212> DNA  
 <213> Kochia scoparia

&lt;400&gt; 741

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 5 ttcttaactc tgcagtgagg ggcttcacaa aacttggcc tctagaacta ttagggatgc 120  
 tgaagaaaat tgagaaggat atgggtcgaa ctaatgaaat aagatatggc cctaggccaa 180  
 ttgacttaga cattctttt 200

10 <210> 742  
 <211> 200  
 <212> DNA  
 <213> Kochia scoparia

15 <400> 742  
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 20 atccttctca atttcttca gcatccctaa tagttctaga ggcccaagtt ttgtgaagcc 120  
 cctcaactgca gagttaagaa actttggttg atcagtcaca taggcaggat ctgtctcgta 180  
 taagcagcca tgccttgtga 200

25 <210> 743  
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 <212> DNA  
 <213> Kochia scoparia

30 <400> 743  
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 35 agttccgcat gaaaggatat gggaaagacc ctttgtgatg gcaccactaa ttgatataat 120  
 tggatcggat gtagaaaaatg acactgttgc tgcattggcat tcattatcaa aattttctgg 180  
 tggactattt gaagcgtggg 200

40 <210> 744  
 <211> 200  
 <212> DNA  
 <213> Kochia scoparia

45 <400> 744  
 cccacgcttc aaatagtcca ccagaaaatt ttgataatga atgccatgca gcaacagtgt 60  
 50 cattttctac atccgatcca attatatcaa tttagtggtgc catcacaaag ggtctttccc 120  
 atatccttc atgcggaact gtaagttct cagagcttac cctaaacttc ccataaaaaaa 180  
 gaatgtctaa gtcaattggc 200

55 <210> 745  
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 <212> DNA

<213> Kochia scoparia

<400> 745

5	tctggtggac tatttgaagc gtgggataaaa cttggtggag attcactcat tgggaaggac	60
	ggaatgagta gggtttgcc agttggaaac cacttgtggg attggtcgtg caaaacctct	120
10	gtaatggag tcttgaattt gactccggac agcttttagtg atggaggaaa gtttctacct	180
	gtagaaaaatg cagttctca	200

<210> 746

<211> 200

<212> DNA

<213> Kochia scoparia

<400> 746

20	tgagaaaactg cattttctac aggtagaaac tttcctccat cactaaagct gtccggagtc	60
	aaattcaaga ctcccattac agaggtttg cacgaccaat cccacaagtg gttcccaact	120
25	ggcaaaaccc tactcattcc gtccttccca atgagtgaat ctccaccaag tttatcccac	180
	gcttcaaata gtccaccaga	200

<210> 747

<211> 200

<212> DNA

<213> Kochia scoparia

<400> 747

35	tacctgtaga aaatgcagtt tctcaggttc gtcaaatgat ctcagaaggt gctgacataa	60
	ttgatattgg agcgcaatca acaaggccca tggcaactag gatttctgct gaggaagagc	120
40	tggaaagact agtccctgtt tttagaaggtg tcaaggatgt tatcgaggaa gaaggaagaa	180
	tgttatcagt ggatacattt	200

<210> 748

<211> 200

<212> DNA

<213> Kochia scoparia

<400> 748

50	aaatgtatcc actgataaca ttcttccttc ttcctcgata acatccttga caccttctaa	60
	aacaggact agtctttcca gctttcctc agcagaaaatc ctagttgcca tgggccttgt	120
55	tgattgcgtt ccaatatcaa ttatgtcagc accttctgag atcatttgac gaacctgaga	180
	aactgcattt tctacaggtt	200

<210> 749

<211> 200  
<212> DNA  
<213> Kochia scoparia

5 <400> 749

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10 tgcagggcctt aaagtgcctt atatagcaat gcacatgcga ggtgatcctt cttcaatgca 180  
aaatgctgat aatttgacat 200

15 <210> 750

<211> 200  
<212> DNA  
<213> Kochia scoparia

20 <400> 750

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aaggcacttt aagccctgca acaacattga acatctcagg atctagcttt ccactcgaaa 120  
25 catcattcac catatgtgct cccttattga cagttcaga agcaactttt gagtaaaatg 180  
tatccactga taacatttt 200

30 <210> 751

<211> 200  
<212> DNA  
<213> Kochia scoparia

35 <400> 751

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agctctagga ttacagatgc agaattatca ggtattcctg cttggaggat agttattgtat 120  
40 cctggcattt gattttccaa gaatacaaag caaaatttgg aaattctcac agggttgaaa 180  
agaattcggc aagaaatagc 200

45 <210> 752

<211> 200  
<212> DNA  
<213> Kochia scoparia

50 <400> 752

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ttggaaaatc caatgccagg atcaataact atcctccaag caggaatacc tgataattct 120  
55 gcatctgtaa tccttagagct caactctaaa gccacccct tacaaacatc attgtatgtc 180  
aaattatcag cattttgcat 200

<210> 753  
 <211> 200  
 <212> DNA  
 <213> Kochia scoparia

5  
 <400> 753

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 10 tgattggacc ttcaagaaag aggttttgg gtgagatctg caatcgtcct gtagcatctg 120  
 atagagatcc ggcaactgtt gcttctatca ccgctggagt tttgggttgt gcaaacattg 180  
 taagagtaca taatgttcgg 200

15  
 <210> 754  
 <211> 200  
 <212> DNA  
 <213> Kochia scoparia  
 20  
 <400> 754

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 25 aacagttgcc ggatctctat cagatgctac aggacgattg cagatctcac ccaaaaacct 120  
 ctttcttcaa ggtccaatca gtaagggacc atgagccaca gccaaactct ttttgctat 180  
 ttcttgccga attctttca 200

30  
 <210> 755  
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 <212> DNA  
 <213> Kochia scoparia  
 35  
 <400> 755

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 40 gcttgaatat cagtataact gaagattctt agattcttag attcttagat tctaagagtt 120  
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 attttgttgc ttatgatttt 200

45  
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 <212> DNA  
 <213> Kochia scoparia  
 50  
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aaaatcataa gcaacaaaat tacaatcata ttattcatat ctgccgaagg cacacaacaa 60  
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 5 agttatactg atattcaagc attgcacatcac atagcttgac accatcaagg ctatccgaa 180  
 cattatgtac tcttacaatg 200

10 <210> 757  
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 <212> DNA  
 <213> Kochia scoparia

15 <400> 757  
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 aaaaatgtac ttgaatttca taaatggtga aataaagttt gtcggtgctt catgcaatac 120  
 20 gactttcttca gtttgcattt ctctatcata ggggctgtaa gcagattagc ccagtcaatt 180  
 cacatttagat ttagacggcc 200

25 <210> 758  
 <211> 200  
 <212> DNA  
 <213> Kochia scoparia

30 <400> 758  
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 caatgcaaac tagaaaagtc gtattgcattg aagcaccgac aaactttatt tcaccattta 120  
 35 tgaaattcaa gtacattttt taatctaaat tctatgataa aagaaaggta ggaagaaaat 180  
 cataagcaac aaaattacaa 200

40 <210> 759  
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 <212> DNA  
 <213> Kochia scoparia

45 <400> 759  
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 atacatttaa actatatgtat ctctaaagagc aaaatacaaa ataatacaat gtgtttcatg 120  
 50 aagatgtcca atcatttctc cctcaaaaaa agatcgtgat gaacaatcag gtactttgat 180  
 tttgttgtag aacctcagct 200

55 <210> 760  
 <211> 200  
 <212> DNA  
 <213> Kochia scoparia

&lt;400&gt; 760

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 5 gagaatgtat tggacatctt catgaaacac attgtattat ttgttatttt gctcttagag 120  
 atcatatagt ttaaatgtat aaacagcggtt agcaacattt tgctgtttt gcaccggccg 180  
 tctaaatcta atgtgaattt 200

10 <210> 761  
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 <212> DNA  
 <213> Kochia scoparia

15 <400> 761

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 20 tcgcctcctca gccaatttct ttctgtgcat ctccctgtca gcttctattt aagctttgtg 120  
 ctcagcctgg agacgctgca tttcctttt catctgctgt ctttcgaacc atgtatcagc 180  
 atccgccccaa actgcagaaaa 200

25 <210> 762  
 <211> 200  
 <212> DNA  
 <213> Kochia scoparia

30 <400> 762

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 35 tgcagcgtct ccaggctgag cacaaagctt caatagaagc tgcagaggag atgcacagaaa 120  
 agaaaattggc tgaggagcga gaaggagaaa aagatgatcc catggctagg gctgaagctg 180  
 agtttctaca acaaaatcaa 200

40 <210> 763  
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 <212> DNA  
 <213> Kochia scoparia

45 <400> 763

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 50 atggcaaatt ggcaataaga tggtgcaagt aacaagctca aatgatccac catagccaaa 120  
 ttccttaacca ttggactgtg tcataactc atatcagcta gttagaaacat tctcaccata 180  
 acccttaacta ttcgaggata 200

55 <210> 764  
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<213> Kochia scoparia

<400> 764

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	cacagtccaa tggtaggaa tttggctatg gtggatcatt tgagcttggt acttgcacca	120
10	tcttattgcc aatttgcacat gctgcacact ccgtatgatt aacttggta aattatttct	180
	gcagtttggg cgatgctga	200

<210> 765

<211> 200

<212> DNA

<213> Kochia scoparia

<400> 765

20	ccataaccct aactattcga ggataagtga taccattcaa caatagtaaa gatcaaatca	60
	ggtattacta gacctatcaa agtaatacgg agtagttgtt attgttacag agttaggaag	120
25	aatcctattt tcccatttct cctatcaatc ctaaactttt gtgactgaac tatcgacata	180
	tagtgtgaag aacatgaatg	200

<210> 766

<211> 200

<212> DNA

<213> Kochia scoparia

<400> 766

35	cattcatgtt ctgcacacta tatgtcgata gttcagtcac aaaagtttag gattgatagg	60
	agaaaatggga aaataggatt ctgcctaact ctgtaacaat aacaactact ccgtattact	120
40	ttgataggc tagtaatacc tgatttgatc tttactattt ttgaatggta tcacttatcc	180
	tcgaatagtt agggttatgg	200

<210> 767

<211> 200

<212> DNA

<213> Kochia scoparia

<400> 767

50	acatatagtg tgaagaacat gaatggcaga agtcacaact ctgattcctg agtcctgact	60
	ataaaactatt ggatgactgt gatacatcat tagtcctta agcattgagt tactctattc	120
55	tacatagtta aattatatacg attccaagat ccgaatccat gaattatcaa ttaccataaa	180
	ttgatcttct tcctttgagt	200

<210> 768

<211> 200  
<212> DNA  
<213> Kochia scoparia

5 <400> 768

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ctatataatt taactatgt aaatagagta actcaatgct taaaggacta atgatgtatc 120  
10 acagtcatcc aatagtttat agtcaggact caggaatcag agttgtgact tctgccattc 180  
atgttcttca cactatatgt 200

15 <210> 769

<211> 200  
<212> DNA  
<213> Kochia scoparia

20 <400> 769

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atctagaagc aatagacact tatcaatgat acagcaatag acgaattcat ctacccaagg 120  
25 cagcatcaac aacagctgcc gtttgcgata atcgaaacta tcatagtgtc cttttcacta 180  
ttagtgcaga tgttatctag 200

30 <210> 770

<211> 200  
<212> DNA  
<213> Kochia scoparia

35 <400> 770

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40 agtgtctatt gtttctagat ttaagacata tttcgagta ggaactgttg aaaaaactca 180  
aaggaagaag atcaatttat 200

45 <210> 771

<211> 200  
<212> DNA  
<213> Kochia scoparia

50 <400> 771

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tcttattctg tactgcataa tcctctcaca aaaaattcgc caatgaatga caaatttagct 120  
55 atccagaatt gcagattgat tgaatcaaat tgatcatggc agcagcacaa ttgcagcagt 180  
tatcaattga agcacaatat 200

<210> 772  
 <211> 200  
 <212> DNA  
 <213> Kochia scoparia

5  
 <400> 772

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ttatgcagta cagaataaga ttttccagga atgtagaaac atagaggaaa agaccctaga	180
taacatctgc actaatagtgc	200

10  
 <210> 773  
 <211> 200  
 <212> DNA  
 <213> Kochia scoparia

15  
 <400> 773

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ccaatcaata actccacaat atagtatcaa tctcttaactc agaaactaaa cagaagcaaa	120
aacccattca atcaaagcaa cagtaaaattt agggttgaa tttagaaagaa agagaggaa	180
gagaaacata ctgggttgcg	200

20  
 <210> 774  
 <211> 200  
 <212> DNA  
 <213> Kochia scoparia

25  
 <400> 774

cgcaacccag tatgtttctc ttccctctct ttctttctaa ttcaaaccct aattttactg	60
ttgccttgat tgaatgggtt tttgccttgc ttttagttct gagtttagaga ttgataactat	120
attgtggagt tattgattgg aatttctgtg ctttgattgt tatgatttag aggtaatatt	180
gtgcctcaat tgataactgc	200

30  
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 <211> 200  
 <212> DNA  
 <213> Kochia scoparia

35  
 <400> 775

## EP 2 755 987 B1

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	ggaagagcgt aattgacagt gagaacacga ccgtaaagct cagcaccatc catgttatcc	120
5	atggcagaag aagcgtcgta tctctccaag aaagtgacga aaccgaaaga acgatgcttg	180
	tttgtggcca aatcaagagg	200
	<210> 776	
10	<211> 200	
	<212> DNA	
	<213> Kochia scoparia	
	<400> 776	
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	gacgacgctt cttctgccat ggataaacatg gatggtgctg agctttacgg tcgtgttctc	120
20	actgtcaatt acgctcttcc cgagaagatt aagggtggtg aacagggatg ggctgcgcaa	180
	cccagtatgt ttctcttccc	200
	<210> 777	
25	<211> 200	
	<212> DNA	
	<213> Kochia scoparia	
	<400> 777	
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	aaagagcatg aagttgtaat tgctctaggg agcaatgtag gagacagatt agataacttt	120
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10 <210> 780  
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15 <400> 780

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 25 agatctgtc tcgtataaagc 200

25 <210> 781  
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30 <400> 781

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 40 gtgggataaa cttgggtggag 200

40 <210> 782  
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45 <400> 782

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 55 acttccccata aaaaagaatg 200

<210> 783  
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&lt;212&gt; DNA

&lt;213&gt; Kochia scoparia

&lt;400&gt; 783

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 10                aatttgactc cggacagctt tagtcatggaa ggaaagtttc tacctgtaga aaatgcagtt   180  
                  tctcagggttc gtcaaatgtat   200

&lt;210&gt; 784

&lt;211&gt; 200

&lt;212&gt; DNA

&lt;213&gt; Kochia scoparia

&lt;400&gt; 784

20               atcatttgac gaacctgaga aactgcattt tctacaggtt gaaactttcc tccatcacta   60  
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 25               aagtggttcc caactggcaa aaccctactc attccgtcct tcccaatgag tgaatctcca   180  
                  ccaagtttat cccacgcttc   200

&lt;210&gt; 785

&lt;211&gt; 200

&lt;212&gt; DNA

&lt;213&gt; Kochia scoparia

&lt;400&gt; 785

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                  aatcaacaag gccccatggca actaggattt ctgctgagga agagctggaa agactagtcc   120  
 40               ctgtttttaga aggtgtcaag gatgttatcg aggaagaagg aagaatgtta tcagtggata   180  
                  cattttactc aaaagttgtat   200

&lt;210&gt; 786

&lt;211&gt; 200

&lt;212&gt; DNA

&lt;213&gt; Kochia scoparia

&lt;400&gt; 786

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55

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10 <400> 787

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20 <210> 788

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<213> Kochia scoparia

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<213> Kochia scoparia

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10     agtttctggt gatgagtgc aaaaagcaag acttgaagct gcaaaagtac atacgtaatc  180
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25     gcctctagaa ctattaggaa tgctgaagaa aattgagaag gatatgggtc gaactaatgg  180
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30 gtgatggagg aaagtttcta 200

<210> 800  
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35 <213> Kochia scoparia

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45 ttttgataat gaatgccatg 200

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50 <213> Kochia scoparia

<400> 801

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30 <400> 807

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40 <210> 808  
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	gcatcacata gcttgacacc atcaaggcta tcccgaacat tatgtactct tacaatgttt	180
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<210> 811

<211> 200

<212> DNA

<213> Kochia scoparia

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	tttgcggtg cttcatgcaa tacgactttt ctagttgca ttgctctatc ataggggctg	180
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          aaaataatac aatgtgttgc atgaagatgt ccaatcattt ctccctcaaa aaaagatcgt    180  
          gatgaacaat caggtacttt    200

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15      <400> 814

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          cagcgtagc aacattatgc tggggggca ccggccgtct aaatctaattt tgaatttgc    180  
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 <213> Kochia scoparia

15 <400> 825

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 20 tatgtttcta cattcctgga aaatcttatt ctgtactgca taatcctctc aaaaaaaatt 120  
 cgccaatgaa tgacaaatta gctatccaga attgcagatt gattgaatca aattgatcat 180  
 25 ggcagcagca caattgcagc 200

25 <210> 826  
 <211> 200  
 <212> DNA  
 <213> Kochia scoparia

30 <400> 826

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 35 taatttgtca ttcattggcg aattttttgt gagaggatta tgcagtacag aataagattt 120  
 tccaggaatg tagaaacata gaggtttaga ccctagataa catctgcact aatagtgaaa 180  
 40 aggacactat gatagttcg 200

40 <210> 827  
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 <212> DNA  
 <213> Kochia scoparia

45 <400> 827

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 50 ataacaatca aagcacagaa attccaatca ataactccac aatatagtat caatctctaa 120  
 ctcagaaact aaacagaagc aaaaacccat tcaatcaaag caacagtaaa attagggttt 180  
 55 gaatttagaaa gaaagagagg 200

<210> 828  
 <211> 200  
 <212> DNA

<213> Kochia scoparia

<400> 828

5	cctcttttc tttctaattc aaaccctaatttactgttg ctggattga atgggaaaa	60
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	ttctgtgctt tgattgttat gattgagagg taatattgtt cttcaatttga taactgctgc	180
10	aatttgtgctg ctgccatgtat	200

<210> 829

<211> 200

<212> DNA

<213> Kochia scoparia

<400> 829

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	gttcaccacc cttaatcttc tcgggaagag cgtaatttgc acgtgagaaca cgaccgtaaa	120
	gctcagcacc atccatgtta tccatggcag aagaagcgctc gtctctctcc aagaaagtga	180
25	cgaaaccgaa agaacgtatgc	200

<210> 830

<211> 200

<212> DNA

<213> Kochia scoparia

<400> 830

35	gcatcgttctt ttcgggttcg tcactttctt ggagagagac gacgcttctt ctgcatttggaa	60
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	gaagattaag ggtggtaaac agggatgggc tgcgcaccc agtatgtttc tcttccctct	180
40	ctttctttctt aattcaaacc	200

<210> 831

<211> 200

<212> DNA

<213> Lolium multiflorum

<400> 831

50	acgaaagctt tgaactctca agtggtgcaa catagataga ttatatatata ttcgaatata	60
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	ttcctcgatg gccccgcctc atcgctgggg cggcgacgct tccggctcg cacccctcg	180
55	tccggaggaag ttgggtcctc	200

<210> 832

<211> 200  
<212> DNA  
<213> Lolium multiflorum

5 <400> 832

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gaggcggggc catcgaggaa gaagaagtag tttaagttt tttaaagtt tttatatgtt 120  
10 attttatgt tcattcaaat tatattcgaa atatatataa tctatctatg ttgcaccact 180  
tgagagttca aagctttcgt 200

15 <210> 833

<211> 200  
<212> DNA  
<213> Lolium multiflorum

20 <400> 833

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cctcctcttc ctggccctct tcgtcggtct gtccttggc ctcttgcgtcc tcctccttgt 120  
25 cgtcatccca tgcgtaatcc tcctcgtagt catccattc gtccattgtg gccggcgcccc 180  
ggaaatcgtc gctgctggac 200

30 <210> 834

<211> 200  
<212> DNA  
<213> Lolium multiflorum

35 <400> 834

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ggaggacgca tggatgacg acgaggagga ggacgaagag gccaaaggagc aggccgacga 120  
40 agaggccgag gaagaggagg aggacgacga agaggctgag gacactgacg aggaggagga 180  
cccaacttcc tccgacgagg 200

45 <210> 835

<211> 200  
<212> DNA  
<213> Lolium multiflorum

50 <400> 835

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cggcttcccc tcgtcgatgg tggatggatgg ccaagagaga tcgtcgatgg ttgacggagg 120  
55 atggcgatgg gagaacagat gaatggcgatgg ggccgtcgatgg aaccgtatat gtaggtctct 180  
cgacgaagag agcggcggtt 200

<210> 836  
 <211> 200  
 <212> DNA  
 <213> Lolium multiflorum

5  
 <400> 836

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 10 atctgttctc tcctcaccat cctccgtcaa ccatgagcga tctctttgg ccatcgacaa 120  
 ccgacagcga gggaaagccg cctggatggc gccattggtg ggataaagct gcatcggtcca 180  
 15 gcagcgacga ttcccccccg 200

<210> 837  
 <211> 200  
 <212> DNA  
 <213> Lolium multiflorum  
 20  
 <400> 837

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 25 tctcgcatcg aggccacttc ggccgttccc gacgagtcgt ttgggtcttc cagaccactt 120  
 tgcgacggtt caatgcgtcg agggcaactcc gacgattgcc cttcccggtg actgcgcgt 180  
 30 cgctatgcac gcggtgtgtg 200

30  
 <210> 838  
 <211> 200  
 <212> DNA  
 <213> Lolium multiflorum  
 35  
 <400> 838

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 40 cgacgcattt aaccgtcgca aagtggtctg gagagccaa acgactcgct gggAACGGCC 120  
 gaagtggcct cgatgcgaga accgctgtaa tggagcacga actccgcggaa agagcaaccg 180  
 45 ccgcctcttt cgtcgagaga 200

<210> 839  
 <211> 200  
 <212> DNA  
 <213> Lolium multiflorum  
 50  
 <400> 839

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	gccgtcgcta tgcacgcggt gtgtgcgcaa ccatgggctc gcggctggga aaatgggcct	60
	ccccaggcca caaaatacat ccatccggcg ataaactaacg ccggatttcg gcctgggagc	120
5	cccaacggct gggatgctct tagcccgtgg aacacattcc cgaatcggtt ccgacggctc	180
	catctctctc gccctgatct	200
	<210> 840	
10	<211> 200	
	<212> DNA	
	<213> Lolium multiflorum	
	<400> 840	
15	agatcagggc gagagagatg agaccgtcgg taccgattcg ggaatgtttt ccacgggcta	60
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20	atgtatttg tggcctgggg aggcccattt tcccagccgc gagcccatgg ttgcgcacac	180
	accgcgtgca tagcgacggc	200
	<210> 841	
25	<211> 200	
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	<213> Lolium multiflorum	
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35	cttctcgctc cgtcctccct ctccaaccga gcggcacgag gtagaccggg agaccccaac	180
	tccgtcttag cccagtggcg	200
40	<210> 842	
	<211> 200	
	<212> DNA	
	<213> Lolium multiflorum	
45	<400> 842	
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50	gcgcgggtgg ttgcgcgcgc acgccgtcgg cccttccaga tcgcggaaaga gagatagatc	180
	agggcgagag agatgagacc	200
55	<210> 843	
	<211> 200	
	<212> DNA	
	<213> Lolium multiflorum	

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<400> 843

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gcttgtttga ttacgatttc tagggttctt ggcattgtcg gtagttgcag atttgtcttc 180  
tgttgggtgt ttgtttccat 200

10 <210> 844  
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<213> Lolium multiflorum

15 <400> 844  
  
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20 gaaatcgtaa tcaaacaagc caagaccaag aaactgggtt aaggaggaag agtatgtgca 120  
tacaagtgcg aagcagacgg agatggagag ggccgcggag atgacgcctc acgggcgcca 180  
25 ctgggctagg acggagttgg 200

25 <210> 845  
<211> 200  
<212> DNA  
<213> Lolium multiflorum

30 <400> 845  
  
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35 ttgtttgttt ccatgccatt gcttgtatcc atcaatttggaa aaggaaagaa agggctacag 120  
ggatattagg aattagatca gtccttccta gctgatttc tctgacaggt tgtagtctc 180  
40 gcgcatgtgg tggacttctt 200

45 <210> 846  
<211> 200  
<212> DNA  
<213> Lolium multiflorum

<400> 846  
  
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50 tgatctaatt cctaataatcc ctgttagccct ttcttccctt tccaattgtat ggataacaagc 120  
aatggcatgg aaacaaacaa ctacataata cgaaacgatg gatgcagcca aggccatgga 180  
55 aacaaacacc caacagaaga 200  
  
<210> 847  
<211> 200

&lt;212&gt; DNA

&lt;213&gt; Lolium multiflorum

&lt;400&gt; 847

5                   gtctcgcgca tgtggtggac ttcttgtgct tttggctagc tccaaagtctg atgtgttagct   60  
                  gctttctgag atttccaca taaacgttgc tatggctagt aagttgtggc taactaaatc   120  
 10                cttgggcaca agctagtaat ctgaatggca tatccgcccc catgccttta attgttgct   180  
                  aacgatgggc taggttctct   200

&lt;210&gt; 848

&lt;211&gt; 200

&lt;212&gt; DNA

&lt;213&gt; Lolium multiflorum

&lt;400&gt; 848

20                agagaaccta gcccattcggtt aggcaacaat taaaggcatg ggggcggata tgccattcag   60  
                  attactagct tgtgccaag gattttagtta gccacaactt actagccata gcaacgttta   120  
 25                tgtggaaaat ctcagaaagc agctacacat cagacttgga gctagccaaa agcacaagaa   180  
                  gtccaccaca tgcgcgagac   200

&lt;210&gt; 849

&lt;211&gt; 200

&lt;212&gt; DNA

&lt;213&gt; Lolium multiflorum

&lt;400&gt; 849

35                tgccataacga tgggctaggt tctcttattt attcattttt gttaaaagaa gaacttgggc   60  
                  atgctaattc agcaaaatag aaaaaatatg tgttgtttct gtatatagag gttcttggtt   120  
 40                ttcccttgcc gattctgacc gatttgccggg tttttttttt ttttgtctt cctcatcgtc   180  
                  aggcaagaag agcaagatgt   200

&lt;210&gt; 850

&lt;211&gt; 200

&lt;212&gt; DNA

&lt;213&gt; Lolium multiflorum

&lt;400&gt; 850

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acatattgct cttcttgccc gacgatgagg aaagacaaaa aaaaaaaaaa cccgcaaatc 60  
 ggtcagaatc ggcaaggaa aaccaagaac ctcttatatac agaaacaaca catattttt 120  
 5 ctatttgct gaatttagcat gcccaagttc ttcttttaac aaaaatgaat aaataagaga 180  
 acctagccca tcgttaggca 200

10 <210> 851  
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 <212> DNA  
 <213> Lolium multiflorum

15 <400> 851  
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 20 caagggctac tactgcggag ggctagcgtc tcctaattgcc ttctcaggtt gcttgccaa 180  
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25 <210> 852  
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 <212> DNA  
 <213> Lolium multiflorum

30 <400> 852  
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 35 gcatcttaga gcataggagc caggtctagt tctgctgctg cttaaatcct ctgtcacatc 180  
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40 <210> 853  
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 <213> Lolium multiflorum

45 <400> 853  
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 caaagacagt cctaactcgt ttcccctctg aacataccag cattggatct tcggcatcct 120  
 50 ctcagctcct caagaaaacc accaggccac gctaacctgt tcagggtccg ttcatacacg 180  
 caatgcgccc tcaccaacga 200

55 <210> 854  
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<213> Lolium multiflorum

<400> 854

5           tcgttggta gggcgattg cgttatgaa cggaccctga acaggttagc gtggcctggt     60  
       ggtttcttg aggagctgag agatgccga agatccaatg ctgttatgtt cagagggaa     120  
       acgagttagg actgtcttg gttggaatgt gaaagaggtg gaaaagagat gttccctagt     180  
 10           tattattggg gaactttggg   200

<210> 855

<211> 200

15           <212> DNA

<213> Lolium multiflorum

<400> 855

20           acacgcaatg cgccctcacc aacgattccg ccgaccagga ggtcgtgatt gccctggca     60  
       gcaacgtggg agacagggtc agcatgttcg acagggcgct gcggctgatg aggagctcgg   120  
       gcatcaaggt caccaggcac gcctgcctgt acgagaccgc cccagcctac gtgaccgacc   180  
 25           agccgcgggtt cctcaactct   200

<210> 856

<211> 200

30           <212> DNA

<213> Lolium multiflorum

<400> 856

35           agagttgagg aaccgcggct ggtcggtcac gtaggctggg gcggctctgt acaggcaggc   60  
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       gaccctgtct cccacgttgc tgcccaggc aatcacgacc tcctggtcgg cggaatcggt   180  
 40           ggtgagggcg cattgcgtgt   200

<210> 857

<211> 200

45           <212> DNA

<213> Lolium multiflorum

<400> 857

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       gttgctcagg aagctcaagg agatcgagaa ggatatacgtt cgtaccgcgg gggtaaggta   120  
       cggcccgagg ccgatcgatc tggacattct tctgtatggg gactcccgaa tcaagaccga   180  
 55           gtctctgatt gtgccgcgt   200

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<210> 858  
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<212> DNA  
<213> *Lolium multiflorum*

5 <400> 858

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ccttgagacctt cctgagcaac tcgtgaggcc ccagcttcgt cgtcccccta acggcagagt 180  
tgaggaaaccq caqcttaqtcaq 200

15 <210> 859  
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<212> DNA  
<212> *Lolium multiflorum*

<400> 859

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qqaacagaag qcattaaaaag 200

30 <210> 860  
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<212> *Lolium multiflorum*

35

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aacccgcccac tgcacttcga gagagagtgc cagttttct ccataccgtc ctcagccgac 120  
gaacctcagga ggtcaacaag aggcgccaag acgaacggtc tctcatggat gcgttcatgc 180

ggcacaaatca gag  
45  
<210> 861  
<211> 200  
<212> DNA

50

## EP 2 755 987 B1

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 5 ctgacataat tgatatttgt 200  
 <210> 862  
 <211> 200  
 <212> DNA  
 10 <213> Lolium multiflorum  
 <400> 862

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 <211> 200  
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 25 <213> Lolium multiflorum  
 <400> 863

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 gatggaaggg aagttgcttt cgtagacac gttctacgac caagtcgctg ctgaagctgt 180  
 30 35 aaaaagagga gccaccatga 200  
 <210> 864  
 <211> 200  
 <212> DNA  
 40 <213> Lolium multiflorum  
 <400> 864

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 tctcaagctc ttcttctgac gatagcctcc ttgcaaaggg cctggtggac tggcaccaa 180  
 45 50 tatcaattat gtcagcacct 200  
 <210> 865  
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 55 <213> Lolium multiflorum  
 <400> 865

## EP 2 755 987 B1

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 5 ccatcaacta tgcagaatga acagaatcta cagtatgtatc atgtctgcaa agaagttgct 180  
 tctgagttat acgccccggtt 200

10 <210> 866  
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 <213> Lolium multiflorum

15 <400> 866  
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 20 tcagcaacaa cttaagaat acttgggtca agctgccac cagatacatc attgatcatg 180  
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25 <210> 867  
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 <212> DNA  
 <213> Lolium multiflorum

30 <400> 867  
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 35 tcgcgggttt ggaatccatt agagaagaga tgggtaaaat gaggcttagt gcttcacatg 180  
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40 <210> 868  
 <211> 200  
 <212> DNA  
 <213> Lolium multiflorum

45 <400> 868  
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 50 gccagggtca agaataatcc tccacaaagg aattccagaa agctctgctg ctctcaaccg 180  
 gggtataac tcagaacgaa 200

55 <210> 869  
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 <212> DNA  
 <213> Lolium multiflorum

&lt;400&gt; 869

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 t gatgcatta ctcaagtaca 200

10 <210> 870  
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 <213> Lolium multiflorum

15 <400> 870

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 ctctctgaac tggatcggcg cgactacata tttcacctag gaaacttttc ctcgagggtc 180  
 25 caagtaatat tggcacatgt 200

25 <210> 871  
 <211> 200  
 <212> DNA  
 <213> Lolium multiflorum

30 <400> 871

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 35 agccatacca gttgccagtt ttgtgcaagg gaatgctgat gtggagctca aaccatatgg 120  
 ggtgtatcat cacttaataa gatcaaagca tgcgtgaata actattacag tgaacaagag 180  
 40 atatacagtt tttcgtgcg 200

40 <210> 872  
 <211> 200  
 <212> DNA  
 <213> Lolium multiflorum

45 <400> 872

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 50 ttatataagt atgatacacc ccatatggtt tgagctccac atcagcattc ccttgcacaa 120  
 aactggcaac tggtatggct tctatccggg ctgtatactt tctattattt tcttctgtac 180  
 55 ttgagtaatg catcagagac 200

<210> 873  
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 <212> DNA

&lt;213&gt; Lolium multiflorum

&lt;400&gt; 873

5           aagagatata cagtttttc gtgcgcctcc tccttcgttag aatcttcaaa gaagcaaagg     60  
      taacacagac ctaggcctct ggggttggta tatgatgtaa ccagtttgt gagggtttaa     120  
      taatttatcta atagtgagac ctaagagtaa gaccttata atatactttg cttggaattt     180  
 10           cctacttgag agcatcattt   200

&lt;210&gt; 874

&lt;211&gt; 200

&lt;212&gt; DNA

&lt;213&gt; Lolium multiflorum

&lt;400&gt; 874

20           aatgtatgct ctcaagttagg aaattccaag caaagtatat tataaaggtc ttactcttag     60  
      gtctcactat tagataatta ttaaaccctc acaaaaactgg ttacatcata taccaacccc     120  
      agaggcctag gtctgtgtta cctttgcttc tttgaagatt ctacgaagga ggaggcgcac     180  
 25           gaaaaaaactg tataatctctt   200

&lt;210&gt; 875

&lt;211&gt; 200

&lt;212&gt; DNA

&lt;213&gt; Lolium multiflorum

&lt;400&gt; 875

30           agctcctcaa gaaaaccacc aggccacgct aacctgttca gggtccgttc atacacgcaa     60  
      tgcgccctca ccaacgattc cgccgaccag gaggtcgtga ttgccctggg cagcaacgtg     120  
      ggagacaggg tcagcatgtt cgacagggcg ctgcggctga tgaggagctc gggcatcaag     180  
 35           gtcaccaggg acgcctgcct   200

&lt;210&gt; 876

&lt;211&gt; 200

&lt;212&gt; DNA

&lt;213&gt; Lolium multiflorum

&lt;400&gt; 876

40           aggcaggcgt gcctggtgac cttgatgccc gagctcctca tcagccgcag cgccctgtcg     60  
      aacatgctga ccctgtctcc cacgttgctg cccagggcaa tcacgacctc ctggtcggcg     120  
      gaatcggttgg tgagggcgca ttgcgtgtat gaacggaccc tgaacaggtt agcgtggcct     180  
 45           ggtggttttc ttgaggagct   200

&lt;210&gt; 877

<211> 200  
<212> DNA  
<213> Lolium multiflorum

5        <400> 877

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10	cgcggttcct caactctgcc gttagggca cgacgaagct ggggcctcac gagttgctca	120
	ggaagctcaa ggagatcgag aaggatata gacgtaccgc cggggtaagg tacggccgaa	180
	ggccgatcga tctggacatt	200
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	<211> 200	
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	<213> Lolium multiflorum	
20	<400> 878	
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25	ctcgatctcc ttgagcttcc tgagcaactc gtgaggcccc agcttacgtcg tgccctaacc	120
	ggcagagttg aggaaccgcg gctggtcggt cacgtaggct ggggcggct cgtacaggca	180
	ggcgtgcctg gtgacacctga	200
30	<210> 879	
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35	<400> 879	
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40	tctgattgtg ccgcattaaac gcatccatga gagaccgttc gtcttggcgc ctattgttga	120
	cctcctgggt tcgtcggtc aggacggtat ggagaaaagc tggactctc tctcgaaatgt	180
	cagtggcggg ttctttgatt	200
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	<213> Lolium multiflorum	
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55	cagccgacga acccaggagg tcaacaagag ggcggcaagac gaacggtctc tcatggatgc	120
	gttcatgcgg cacaatcaga gactcggtct tgatccggga gtccccatac agaagaatgt	180
	ccagatcgat cggcctcggg	200

<210> 881  
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 <212> DNA  
 <213> Lolium multiflorum

5  
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acagaaggca taaaaagggt catgtctgtt ggaaatacgc	tgttgactg gctcagagg	120
acccttgtta tgggggtgct taacctcacg ccagacagct	tcagcgacgg aggttaagttt	180
caagaagtgg aagctgccat		200

15  
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20  
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agcaccccca taacaagggt cctctgacgc cagtccaaaca	gcgtatttcc aacagacatg	120
acccttttaa tgccttctgt tccaaacaaca gattcaccgc	cgagcttgtt ccacaaatca	180
aagaacccgc cactgcactt		200

30  
 <210> 883  
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 <212> DNA  
 <213> Lolium multiflorum

35  
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acataattga tattggtgcc cagtcacca ggccctttgc aaggaggcta	tccgcagaag	120
aagagcttga gaggctggtc cctgttctgg atgctatcat	gaaactccca gagatggaag	180
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45  
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50  
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 5 ggcaccaata tcaattatgt cagcacccctc tgatatcaac aacctggcct gggaaatggc 180  
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10 <210> 885  
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15 <400> 885  
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 20 agttgttgct gaactgggag ttccgtatgt taccatgcac atgagaggcg atccatcaac 180  
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25 <210> 886  
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 <213> Lolium multiflorum

30 <400> 886  
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 35 tgatcatggc ggctcctttt tttacagott cagcagcgac ttgcgcgttag aacgtgtcta 180  
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40 <210> 887  
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45 <400> 887  
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 50 cttgaccctg gcattgggtt ttctaaagaaa tccacacaga atattgaagt aatcgccgggt 180  
 ttggaaatcca ttagagaaga 200

55 <210> 888  
 <211> 200  
 <212> DNA  
 <213> Lolium multiflorum

&lt;400&gt; 888

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 5 aacccaatgc cagggtaaag aataatcctc cacaaaggaa ttccagaaag ctctgctgct 120  
 ctcaaccggg cgtataactc agaagcaact tctttcaga catcatcata ctgttagattc 180  
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10 <210> 889  
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15 <400> 889

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 20 caatattact tggaccctcg aggaaaagtt tccttaggtga aatatgtagt cgcggccatc 120  
 cagttcagag agatgctgct actgcttcgt ccgttacaat tgcgatcttgc aatggcgcta 180  
 atatagtttagt ggtccataat 200

25 <210> 890  
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 <212> DNA  
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30 <400> 890

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40 <210> 891  
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45 <400> 891

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55 <210> 892  
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10	ccgggaatcc ccatacagaa gaatgtccag atcgatcggc ctcggaccgt accttatccc	180
	ggcggtgcgc cctatatcct	200

<210> 893

<211> 200

<212> DNA

<213> Lolium multiflorum

<400> 893

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25	gccatttctc aggccaggct gttaattca gaaggtgcag gcataattga tattggcgct	180
	cagtccacca ggccctttgc	200

<210> 894

<211> 200

<212> DNA

<213> Lolium multiflorum

<400> 894

30	gcaaagggcc tggtggactg agcgccaata tcaattatgc ctgcacccctc tgaaattaac	60
	agcctggcct gagaaatggc agcttccacc tcttggaaact tacctccatc gctaaagctg	120
35	tctggcgtga ggttaagcac cccataaacc agggtcctct cccgccaatc caacagcgta	180
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<210> 895

<211> 200

<212> DNA

<213> Lolium multiflorum

<400> 895

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	tggtccctgt tcttaatgct atcatgaaac tccccagat ggaagggaag ttgcttcag	120
45	tagatacggtt ctacgcgcaa gtcgcggctg aagccgtgaa aagaggggcc accatgatca	180
	atgatgtatc tggtgccag	200

<210> 896

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5 <400> 896

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10 agcattaaga acagggacca gcctctcaag ctcttcttct gcagatagcc accttgcaaa 180  
gggcctggtg gactgagcgc 200

15 <210> 897

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25 gaatttacag tatgtatgatg tctgcaaaga agttgcttct gagttatact ctcggtttag 180  
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35 <400> 898

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50 <400> 899

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55 gaagagatgg gtaaaatgag tcttaggtgct tcacatgtgc caatattact tggaccctcc 180  
aggaaaacgtt tcttagggca 200

<210> 900  
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 gtggatttct tggaaaaccc aatgccaggg tcaagaataa tcctccacaa aggaattcca 180  
 15 gacaactctg ctgctctcaa 200

<210> 901  
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20  
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 ccaagagatc gtgattgcc tggcagcaa cgtggagac agggtcagca cgttcgacag 120  
 30 ggcgctgcgg ctgatgagga gctcggcat caggatcacc aggcacgcct gcctgtacga 180  
 gaccgcccct gcttacgtga 200

35  
 <210> 902  
 <211> 200  
 <212> DNA  
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35  
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 45 gggcaatcac gatctcttgg tcggcggaat cgttggtgac cgccgattgc atgtacgaac 180  
 ggaccctgaa caggttagcg 200

45  
 <210> 903  
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 <212> DNA  
 <213> Lolium multiflorum

50  
 <400> 903

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	<212> DNA	
	<213> Lolium multiflorum	
	<400> 904	
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	<211> 150	
	<212> DNA	
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35	tgccctgtacg agaccgcccc tgcttacgtg	150
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40	<212> DNA	
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<210> 924

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<212> DNA

<213> Unknown

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<220>

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<210> 925

<211> 21

<212> DNA

20 <213> Unknown

<220>

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25

<400> 925

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<210> 926

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30 <212> DNA

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<220>

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50

<210> 928

<211> 21

<212> DNA

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20      <210> 936  
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### Claims

- 15 1. A method of plant control comprising: treating a plant with a composition comprising a double-stranded RNA (dsRNA) polynucleotide and a transfer agent, wherein said dsRNA polynucleotide is identical or complementary to an RNA sequence of a 7,8-dihydropteroate synthase (DHPS) gene sequence, wherein said DHPS gene sequence is selected from the group consisting of SEQ ID NOs:1, 2, 5, 9, 11, 18, 19, and a polynucleotide fragment thereof with at least 18 contiguous nucleotides from said DHPS gene sequence, wherein said transfer agent is an organosilicone composition or an organosilicone compound contained therein and conditions the surface of said plant for permeation by said dsRNA polynucleotide, and whereby said plant's growth, development, or reproductive ability is suppressed or delayed or said plant is more sensitive to a DHPS inhibitor herbicide or a mitosis inhibitor herbicide as a result of said dsRNA polynucleotide containing composition relative to an untreated plant.
- 25 2. The method as claimed in claim 1, wherein
  - (i) said plant is selected from the group consisting of *Amaranthus palmeri*, *Amaranthus rudis*, *Amaranthus hybridus*, *Amaranthus lividus*, *Amaranthus viridis*, *Ambrosia trifida*, *Conyza canadensis*, and *Euphorbia heterophylla*; ; or
  - 30 (ii) said composition further comprises said DHPS inhibitor herbicide or said mitosis inhibitor herbicide and said treating comprises external application to said plant with said composition, preferably said composition further comprises one or more co-herbicides different from said DHPS inhibitor herbicide or said mitosis inhibitor herbicide, wherein said one or more co-herbicides are selected from the group consisting of amide herbicides, arsenical herbicides, benzothiazole herbicides, benzoylcyclohexanedione herbicides, benzofuranyl alkylsulfonate herbicides, cyclohexene oxime herbicides, cyclopropylisoxazole herbicides, dicarboximide herbicides, dinitroaniline herbicides, dinitrophenol herbicides, diphenyl ether herbicides, dithiocarbamate herbicides, glycine herbicides, halogenated aliphatic herbicides, imidazolinone herbicides, inorganic herbicides, nitrile herbicides, organophosphorus herbicides, oxadiazolone herbicides, oxazole herbicides, phenoxy herbicides, phenylenediamine herbicides, pyrazole herbicides, pyridazine herbicides, pyridazinone herbicides, pyridine herbicides, 35 pyrimidinediamine herbicides, pyrimidinyloxybenzylamine herbicides, quaternary ammonium herbicides, thiocarbamate herbicides, thiocarbonate herbicides, thiourea herbicides, triazine herbicides, triazinone herbicides, triazole herbicides, triazolone herbicides, triazolopyrimidine herbicides, uracil herbicides, and urea herbicides.
- 45 3. The method as claimed in claim 1, wherein said composition comprises any combination of two or more of said dsRNA polynucleotides and said treating comprises external application to said plant with said composition.
4. A composition comprising a dsRNA polynucleotide and a transfer agent, wherein said dsRNA polynucleotide is identical or complementary to an RNA sequence of a DHPS gene sequence, wherein said DHPS gene sequence is selected from the group consisting of SEQ ID NOs:1, 2, 5, 9, 11, 18, 19, and a polynucleotide fragment thereof with at least 18 contiguous nucleotides from said DHPS gene sequence, wherein said transfer agent is an organosilicone composition or an organosilicone compound contained therein and conditions the surface of a plant for permeation by said dsRNA polynucleotide, and whereby said plant treated with said composition has its growth, development, or reproductive ability regulated, suppressed or delayed or said plant is more sensitive to a DHPS inhibitor herbicide or a mitosis inhibitor herbicide as a result of said dsRNA polynucleotide containing composition relative to an untreated plant.
- 55 5. The composition of claim 4, wherein

(i) said composition further comprises said DHPS inhibitor herbicide or said mitosis inhibitor herbicide, preferably said DHPS inhibitor herbicide is selected from the group consisting of carbamates and asulam, or said mitosis inhibitor herbicide is selected from the group consisting of dinitroaniline herbicides, or  
 5 (ii) said composition further comprises a co-herbicide, wherein said co-herbicide is selected from the group consisting of amide herbicides, arsenical herbicides, benzothiazole herbicides, benzoylcyclohexanedione herbicides, benzofuranyl alkylsulfonate herbicides, cyclohexene oxime herbicides, cyclopropylisoxazole herbicides, dicarboximide herbicides, dinitroaniline herbicides, dinitrophenol herbicides, diphenyl ether herbicides, dithiocarbamate herbicides, glycine herbicides, halogenated aliphatic herbicides, imidazolinone herbicides, inorganic herbicides, nitrile herbicides, organophosphorus herbicides, oxadiazolone herbicides, oxazole herbicides, phenoxy herbicides, phenylenediamine herbicides, pyrazole herbicides, pyridazine herbicides, pyridazinone herbicides, pyridine herbicides, pyrimidinediamine herbicides, pyrimidinyloxybenzylamine herbicides, quaternary ammonium herbicides, thiocarbamate herbicides, thiocarbonate herbicides, thiourea herbicides, triazine herbicides, triazinone herbicides, triazole herbicides, triazolone herbicides, triazolopyrimidine herbicides, uracil herbicides, and urea herbicides.

- 15 6. A method of reducing expression of a DHPS gene in a plant comprising: external application to said plant of a composition comprising a dsRNA polynucleotide and a transfer agent, wherein said dsRNA polynucleotide is identical or complementary to an RNA sequence of a DHPS gene sequence, wherein said DHPS gene sequence is selected from the group consisting of SEQ ID NOs:1, 2, 5, 9, 11, 18, 19, and a polynucleotide fragment thereof with at least 20 18 contiguous nucleotides from said DHPS gene sequence, wherein said transfer agent is an organosilicone composition or an organosilicone compound contained therein and conditions the surface of said plant for permeation by said dsRNA polynucleotide, and whereby said expression of said DHPS gene is reduced relative to an untreated plant.
- 25 7. A method of identifying dsRNA polynucleotides useful in modulating DHPS gene expression when externally treating a plant comprising: a) providing a plurality of dsRNA polynucleotides that comprise a region identical or complementary to a polynucleotide fragment of at least 18 contiguous nucleotides in length that is identical or complementary to a DHPS gene sequence selected from the group consisting of SEQ ID NOs:1, 2, 5, 9, 11, 18, and 19; b) externally treating said plant with a composition comprising one or more of said dsRNA polynucleotides and a transfer agent, wherein said transfer agent is an organosilicone composition or an organosilicone compound contained therein and conditions the surface of said plant for permeation by said one or more of said dsRNA polynucleotides; c) analyzing said plant, or extract for modulation of DHPS gene expression, and d) said plant treated with said composition has its growth, development or reproductive ability regulated, suppressed, or delayed or said plant is more sensitive to a DHPS inhibitor herbicide or a mitosis inhibitor herbicide as a result of said composition, relative to an untreated plant.
- 35 8. The method of claim 7, wherein  
 40 (i) said plant is selected from the group consisting of *Amaranthus palmeri*, *Amaranthus rudis*, *Amaranthus hybridus*, *Amaranthus lividus*, *Amaranthus viridis*, *Ambrosia trifida*, *Conyza canadensis*, and *Euphorbia heterophylla*;  
 45 (ii) said DHPS gene expression is reduced in said plant relative to a plant not treated with said composition.  
 50 9. An agricultural chemical composition comprising an admixture of a dsRNA polynucleotide, a transfer agent, a DHPS inhibitor herbicide or a mitosis inhibitor herbicide, and a co-herbicide, wherein said dsRNA polynucleotide is identical or complementary to a portion of an RNA sequence of a DHPS gene sequence, wherein said DHPS gene sequence is selected from the group consisting of SEQ ID NOs:1, 2, 5, 9, 11, 18, 19, and a polynucleotide fragment thereof with at least 18 contiguous nucleotides from said DHPS gene sequence, wherein said transfer agent is an organosilicone composition or an organosilicone compound contained therein and conditions a surface of a plant for permeation by said dsRNA polynucleotide, and whereby said plant treated with said composition has its growth, development, or reproductive ability regulated, suppressed, or delayed or said plant is more sensitive to said DHPS inhibitor herbicide or said mitosis inhibitor herbicide as a result of said dsRNA polynucleotide containing composition, relative to an untreated plant.  
 55 10. The agricultural chemical composition of claim 9, wherein said co-herbicide is selected from the group consisting of amide herbicides, arsenical herbicides, benzothiazole herbicides, benzoylcyclohexanedione herbicides, benzofuranyl alkylsulfonate herbicides, cyclohexene oxime herbicides, cyclopropylisoxazole herbicides, dicarboximide herbicides, dinitroaniline herbicides, dinitrophenol herbicides, diphenyl ether herbicides, dithiocarbamate herbicides, glycine herbicides, halogenated aliphatic herbicides, imidazolinone herbicides, inorganic herbicides, nitrile herbicides,

cides, organophosphorus herbicides, oxadiazolone herbicides, oxazole herbicides, phenoxy herbicides, phenylenediamine herbicides, pyrazole herbicides, pyridazine herbicides, pyridazinone herbicides, pyridine herbicides, pyrimidinediamine herbicides, pyrimidinyloxybenzylamine herbicides, quaternary ammonium herbicides, thiocarbamate herbicides, thiocarbonate herbicides, thiourea herbicides, triazine herbicides, triazinone herbicides, triazole herbicides, triazolone herbicides, triazolopyrimidine herbicides, uracil herbicides, and urea herbicides.

- 5           11. An agricultural chemical composition comprising an admixture of a dsRNA polynucleotide, a transfer agent, a DHPS inhibitor herbicide, and a pesticide, wherein said dsRNA polynucleotide is identical or complementary to a portion of an RNA sequence of a DHPS gene sequence, wherein said DHPS gene sequence is selected from the group consisting of SEQ ID NOS:1, 2, 5, 9, 11, 18, 19, and a polynucleotide fragment thereof with at least 18 contiguous nucleotides from said DHPS gene sequence, wherein said transfer agent is an organosilicone composition or an organosilicone compound contained therein and conditions a surface of a plant for permeation by said dsRNA polynucleotide, whereby said plant treated with said composition has its growth, development, or reproductive ability regulated, suppressed, or delayed or said plant is more sensitive to said DHPS inhibitor herbicide or a mitosis inhibitor herbicide as a result of said dsRNA polynucleotide containing composition relative to an untreated plant.
- 10          12. The agricultural chemical composition of claim 11, wherein said pesticide is selected from the group consisting of insecticides, fungicides, nematicides, bactericides, acaricides, growth regulators, chemosterilants, semiochemicals, repellents, attractants, pheromones, feeding stimulants, and biopesticides.
- 15          13. A composition comprising a dsRNA polynucleotide and a transfer agent according to claim 4, wherein said dsRNA polynucleotide is identical or complementary to an RNA sequence of a DHPS gene sequence, wherein said DHPS gene sequence comprises SEQ ID NOS:1176, 1186, a complement thereof, and a polynucleotide fragment thereof with at least 18 contiguous nucleotides from said DHPS gene sequence.

### Patentansprüche

- 30          1. Verfahren zur Kontrolle von Pflanzen, Schritte umfassend, bei denen man: eine Pflanze mit einer Zusammensetzung behandelt, die ein doppelsträngiges RNA (dsRNA)-Polynukleotid und ein Übertragungsmittel umfasst, wobei das dsRNA-Polynukleotid identisch mit oder komplementär zu einer RNA-Sequenz einer 7,8-Dihydropteroatsynthase (DHPS)-Gensequenz ist, wobei die DHPS-Gensequenz aus der Gruppe ausgewählt wurde, die aus SEQ ID Nummern:1, 2, 5, 9, 11, 18, 19 und einem Polynukleotidfragment davon mit mindestens 18 aufeinanderfolgenden Nukleotiden der DHPS-Gensequenz besteht, wobei das Übertragungsmittel eine siliciumorganische Zusammensetzung oder eine darin enthaltene siliciumorganische Verbindung ist, welche die Oberfläche der Pflanze für das Eindringen des dsRNA-Polynukleotids konditioniert, und wobei das Wachstum, die Entwicklung oder die Fortpflanzungsfähigkeit der Pflanze unterdrückt oder verzögert wird oder die Pflanze im Vergleich zu einer unbehandelten Pflanze durch die Zusammensetzung, welche das dsRNA-Polynukleotid enthält, auf ein DHPS-Inhibitor-Herbizid oder ein Mitoseinhibitor-Herbizid empfindlicher reagiert.
- 35          2. Verfahren gemäß Anspruch 1, bei dem
- 40           (i) die Pflanze aus der Gruppe bestehend aus *Amaranthus palmeri*, *Amaranthus rudis*, *Amaranthus hybridus*, *Amaranthus lividus*, *Amaranthus viridis*, *Ambrosia trifida*, *Conyza canadensis* und *Euphorbia heterophylla* ausgewählt wurde; oder
- 45           (ii) die Zusammensetzung ferner das DHPS-Inhibitor-Herbizid oder das Mitoseinhibitor-Herbizid umfasst und die Behandlung eine äußere Verabreichung der Zusammensetzung auf die Pflanze umfasst, wobei die Zusammensetzung vorzugsweise ferner ein oder mehrere Co-Herbizide umfasst, die sich von dem DHPS-Inhibitor-Herbizid oder dem Mitoseinhibitor-Herbizid unterscheiden, und wobei das eine oder die mehreren Co-Herbizide aus der Gruppe bestehend aus Amidherbiziden, Arsenherbiziden, Benzothiazolherbiziden, Benzoylcyclohexandionherbiziden, Benzofuranylalkylsulfonat-Herbiziden, Cyclohexandion-Herbiziden, Cyclopropylisoxazol-Herbiziden, Dicarboximid-Herbiziden, Dinitroanilin-Herbiziden, Dinitrophenol-Herbiziden, Diphenyletherherbiziden, Dithiocarbamatherbiziden, Glycerinherbiziden, halogenierten aliphatischen Herbiziden, Imidazolinonherbiziden, anorganischen Herbiziden, Nitrilherbiziden, phosphororganischen Herbiziden, Oxadiazolon-Herbiziden, Oxazol-Herbiziden, Phenoxy-Herbiziden, Phenylendiamin-Herbiziden, Pyrazol-Herbiziden, Pyridazin-Herbiziden, Pyridazinon-Herbiziden, Pyridin-Herbiziden, Pyrimidindiamin-Herbiziden, Pyrimidinyloxybenzylamin-Herbiziden, quaternären Ammonium-Herbiziden, Thiocarbamat-Herbiziden, Thiocarbonat-Herbiziden, Thioharnstoff-Herbiziden, Triazinon-Herbiziden, Triazol-Herbiziden, Triazolon-Herbiziden, Triazolopyrimidin-Herbiziden.

ziden, Uracil-Herbiziden und Harnstoff-Herbiziden ausgewählt wurden.

3. Verfahren gemäß Anspruch 1, wobei die Zusammensetzung eine beliebige Kombination von zwei oder mehr der dsRNA-Polynukleotide umfasst und die Behandlung eine äußere Verabreichung der Zusammensetzung auf die Pflanze umfasst.
4. Zusammensetzung, die ein dsRNA-Polynukleotid und ein Übertragungsmittel umfasst, wobei das dsRNA-Polynukleotid identisch mit oder komplementär zu einer RNA-Sequenz einer DHPS-Gensequenz ist, wobei die DHPS-Gensequenz aus der Gruppe bestehend aus SEQ ID Nummern:1, 2, 5, 9, 11, 18, 19 und einem Polynukleotidfragment davon mit mindestens 18 aufeinanderfolgenden Nukleotiden der DHPS-Gensequenz ausgewählt wurde, wobei das Übertragungsmittel eine siliciumorganische Zusammensetzung oder eine darin enthaltene siliciumorganische Verbindung ist, welche die Oberfläche der Pflanze für das Eindringen des dsRNA-Polynukleotids konditioniert, und wobei das Wachstum, die Entwicklung oder die Fortpflanzungsfähigkeit der mit der Zusammensetzung behandelten Pflanze reguliert, unterdrückt oder verzögert wird oder die Pflanze im Vergleich zu einer unbehandelten Pflanze durch die Zusammensetzung, welche das dsRNA-Polynukleotid enthält, auf ein DHPS-Inhibitor-Herbizid oder ein Mitoseinhibitor-Herbizid empfindlicher reagiert.
5. Zusammensetzung gemäß Anspruch 4, in der
  - (i) die Zusammensetzung ferner das DHPS-Inhibitor-Herbizid oder das Mitoseinhibitor-Herbizid umfasst, wobei das DHPS-Inhibitor-Herbizid vorzugsweise aus der Gruppe bestehend aus Carbamaten und Asulam ausgewählt wurde oder das Mitoseinhibitor-Herbizid aus der Gruppe bestehend aus Dinitroanilin-Herbiziden ausgewählt wurde, oder
  - (ii) die Zusammensetzung ferner ein Co-Herbizid umfasst, wobei das Co-Herbizid aus der Gruppe bestehend aus Amidherbiziden, Arsenherbiziden, Benzothiazolherbiziden, Benzoylcyclohexandionherbiziden, Benzofuranylalkylsulfonat-Herbiziden, Cyclohexandion-Herbiziden, Cyclopropylisoxazol-Herbiziden, Dicarboximid-Herbiziden, Dinitroanilin-Herbiziden, Dinitrophenol-Herbiziden, Diphenyletherherbiziden, Dithiocarbamatherbiziden, Glycerinherbiziden, halogenierten aliphatischen Herbiziden, Imidazolinonherbiziden, anorganischen Herbiziden, Nitrilherbiziden, phosphororganischen Herbiziden, Oxadiazolon-Herbiziden, Oxazol-Herbiziden, Phenoxy-Herbiziden, Phenylendiamin-Herbiziden, Pyrazol-Herbiziden, Pyridazin-Herbiziden, Pyridazinon-Herbiziden, Pyridin-Herbiziden, Pyrimidindiamin-Herbiziden, Pyrimidinyloxybenzylamin-Herbiziden, quaternären Ammonium-Herbiziden, Thiocarbamat-Herbiziden, Thiocarbonat-Herbiziden, Thioharnstoff-Herbiziden, Triazinon-Herbiziden, Triazol-Herbiziden, Triazolon-Herbiziden, Triazolopyrimidin-Herbiziden, Uracil-Herbiziden und Harnstoff-Herbiziden ausgewählt wurde.
6. Verfahren zur Verringerung der Expression eines DHPS-Gens in einer Pflanze, Schritte umfassend, bei denen man: eine Zusammensetzung, die ein dsRNA-Polynukleotid und ein Übertragungsmittel umfasst, äußerlich auf die Pflanze aufträgt, wobei das dsRNA-Polynukleotid identisch mit oder komplementär zu einer RNA-Sequenz einer DHPS-Gensequenz ist, wobei die DHPS-Gensequenz aus der Gruppe bestehend aus SEQ ID Nummern:1, 2, 5, 9, 11, 18, 19 und einem Polynukleotidfragment davon mit mindestens 18 aufeinanderfolgenden Nukleotiden der DHPS-Gensequenz ausgewählt wurde, wobei das Übertragungsmittel eine siliciumorganische Zusammensetzung oder eine darin enthaltene siliciumorganische Verbindung ist, welche die Oberfläche der Pflanze für das Eindringen des dsRNA-Polynukleotids konditioniert, und wobei die Expression des DHPS-Gens im Vergleich zu einer unbehandelten Pflanze reduziert wird.
7. Verfahren zur Identifizierung von dsRNA-Polynukleotiden, welche zur Veränderung der DHPS-Genexpression bei äußerlicher Auftragung auf eine Pflanze nützlich sind, Schritte umfassend, bei denen man: a) eine Vielzahl an dsRNA-Polynukleotiden bereitstellt, welche einen Bereich umfassen, der identisch mit oder komplementär zu einem Polynukleotidfragment mit einer Länge von mindestens 18 aufeinanderfolgenden Nukleotiden ist, welcher identisch mit oder komplementär zu einer DHPS-Gensequenz ist, die aus der Gruppe bestehend aus SEQ ID Nummern:1, 2, 5, 9, 11, 18 und 19 ausgewählt wurde; b) eine Zusammensetzung äußerlich auf die Pflanze aufträgt, wobei die Zusammensetzung eines oder mehrere der dsRNA-Polynukleotide und ein Übertragungsmittel umfasst, wobei das Übertragungsmittel eine siliciumorganische Zusammensetzung oder eine darin enthaltene siliciumorganische Verbindung ist, welche die Oberfläche der Pflanze für das Eindringen der einen oder mehreren dsRNA-Polynukleotide konditioniert; c) die Pflanze oder einen Extrakt davon auf die Veränderung der DHPS-Genexpression hin analysiert, und d) wobei das Wachstum, die Entwicklung oder die Fortpflanzungsfähigkeit der mit der Zusammensetzung behandelten Pflanze reguliert, unterdrückt oder verzögert wird oder die Pflanze im Vergleich zu einer unbehandelten Pflanze durch die Zusammensetzung, welche das dsRNA-Polynukleotid enthält, auf ein DHPS-Inhibitor-Herbizid

oder ein Mitoseinhibitor-Herbizid empfindlicher reagiert.

8. Verfahren gemäß Anspruch 7, bei dem

- 5           (i) die Pflanze aus der Gruppe bestehend aus *Amaranthus palmeri*, *Amaranthus rufus*, *Amaranthus hybridus*, *Amaranthus lividus*, *Amaranthus viridis*, *Ambrosia trifida*, *Conyza canadensis* und *Euphorbia heterophylla* ausgewählt wurde;
- 10          (ii) die DHPS-Genexpression in der Pflanze im Vergleich zu einer nicht mit der Zusammensetzung behandelten Pflanze reduziert ist.

10          9. Landwirtschaftliche chemische Zusammensetzung, die eine Mischung aus einem dsRNA-Polynukleotid, einem Übertragungsmittel, einem DHPS-Inhibitor-Herbizid oder einem Mitoseinhibitor-Herbizid und einem Co-Herbizid umfasst, wobei das dsRNA-Polynukleotid identisch mit oder komplementär zu einem Teil einer RNA-Sequenz einer DHPS-Gensequenz ist, wobei die DHPS-Gensequenz aus der Gruppe bestehend aus SEQ ID Nummern:1, 2, 5, 9, 11, 18, 19 und einem Polynukleotidfragment davon mit mindestens 18 aufeinanderfolgenden Nukleotiden der DHPS-Gensequenz ausgewählt wurde, wobei das Übertragungsmittel eine siliciumorganische Zusammensetzung oder eine darin enthaltene siliciumorganische Verbindung ist, welche die Oberfläche der Pflanze für das Eindringen des dsRNA-Polynukleotids konditioniert, und wobei das Wachstum, die Entwicklung oder die Fortpflanzungsfähigkeit der mit der Zusammensetzung behandelten Pflanze reguliert, unterdrückt oder verzögert wird oder die Pflanze im Vergleich zu einer unbehandelten Pflanze durch die Zusammensetzung, welche das dsRNA-Polynukleotid enthält, auf ein DHPS-Inhibitor-Herbizid oder ein Mitoseinhibitor-Herbizid empfindlicher reagiert.

15          10. Landwirtschaftliche chemische Zusammensetzung gemäß Anspruch 9, wobei das Co-Herbizid aus der Gruppe bestehend aus Amidherbiziden, Arsenherbiziden, Benzothiazolherbiziden, Benzoylcyclohexandionherbiziden, Benzofuranylalkylsulfonat-Herbiziden, Cyclohexandion-Herbiziden, Cyclopropylisoxazol-Herbiziden, Dicarboximid-Herbiziden, Dinitroanilin-Herbiziden, Dinitrophenoxy-Herbiziden, Diphenyletherherbiziden, Dithiocarbamatherbiziden, Glycinherbiziden, halogenierten aliphatischen Herbiziden, Imidazolinonherbiziden, anorganischen Herbiziden, Nitrilherbiziden, phosphororganischen Herbiziden, Oxadiazolon-Herbiziden, Oxazol-Herbiziden, Phenoxy-Herbiziden, Phenylendiamin-Herbiziden, Pyrazol-Herbiziden, Pyridazin-Herbiziden, Pyridazinon-Herbiziden, Pyridin-Herbiziden, Pyrimidindiamin-Herbiziden, Pyrimidinyloxybenzylamin-Herbiziden, quaternären Ammonium-Herbiziden, Thiocarbamat-Herbiziden, Thiocarbonat-Herbiziden, Thioharnstoff-Herbiziden, Triazinon-Herbiziden, Triazol-Herbiziden, Triazolon-Herbiziden, Triazolopyrimidin-Herbiziden, Uracil-Herbiziden und Harnstoff-Herbiziden ausgewählt wurde.

20          11. Landwirtschaftliche chemische Zusammensetzung, die eine Mischung aus einem dsRNA-Polynukleotid, einem Übertragungsmittel, einem DHPS-Inhibitor-Herbizid und einem Pestizid umfasst, wobei das dsRNA-Polynukleotid identisch mit oder komplementär zu einem Teil einer RNA-Sequenz einer DHPS-Gensequenz ist, wobei die DHPS-Gensequenz aus der Gruppe bestehend aus SEQ ID Nummern:1, 2, 5, 9, 11, 18, 19 und einem Polynukleotidfragment davon mit mindestens 18 aufeinanderfolgenden Nukleotiden der DHPS-Gensequenz ausgewählt wurde, wobei das Übertragungsmittel eine siliciumorganische Zusammensetzung oder eine darin enthaltene siliciumorganische Verbindung ist, welche die Oberfläche der Pflanze für das Eindringen des dsRNA-Polynukleotids konditioniert, und wobei das Wachstum, die Entwicklung oder die Fortpflanzungsfähigkeit der mit der Zusammensetzung behandelten Pflanze reguliert, unterdrückt oder verzögert wird oder die Pflanze im Vergleich zu einer unbehandelten Pflanze durch die Zusammensetzung, welche das dsRNA-Polynukleotid enthält, auf ein DHPS-Inhibitor-Herbizid oder ein Mitoseinhibitor-Herbizid empfindlicher reagiert.

25          12. Landwirtschaftliche chemische Zusammensetzung gemäß Anspruch 11, bei der das Pestizid aus der Gruppe bestehend aus Insektiziden, Fungiziden, Nematiziden, Bakteriziden, Akariziden, Wachstumsregulatoren, chemische Sterilisationsmittel, Botenstoffe, Repellent, Lockstoffen, Pheromonen, Futterstimulanzien und Biopestiziden ausgewählt wurde.

30          13. Zusammensetzung, die ein dsRNA-Polynukleotid und ein Übertragungsmittel gemäß Anspruch 4 umfasst, wobei das dsRNA-Polynukleotid identisch mit oder komplementär zu einer RNA-Sequenz einer DHPS-Gensequenz ist, wobei die DHPS-Gensequenz die SEQ ID Nummern: 1176, 1186, ein Komplement davon und ein Polynukleotidfragment davon mit mindestens 18 aufeinanderfolgenden Nukleotiden der DHPS-Gensequenz umfasst.

**Revendications**

1. Procédé de lutte contre des végétaux, qui comporte le traitement d'un végétal avec une composition comprenant un polynucléotide d'ARN double brin (ARNdb) et un agent de transfert, étant entendu que ledit polynucléotide d'ARNdb est identique ou complémentaire à une séquence d'ARN correspondant à une séquence de gène de 7,8-dihydroptéroate synthase (DHPS), ladite séquence de gène de DHPS étant choisie dans l'ensemble constitué par les séquences SEQ ID NO : 1, SEQ ID NO : 2, SEQ ID NO : 5, SEQ ID NO : 9, SEQ ID NO : 11, SEQ ID NO : 18 et SEQ ID NO : 19, et un fragment polynucléotidique de celles-ci comportant au moins 18 nucléotides contigus provenant de ladite séquence de gène de DHPS, et ledit agent de transfert est une composition à base d'organosilicone ou un composé de type organosilicone contenu en son sein, et prépare la surface dudit végétal à la perméation dudit polynucléotide d'ARNdb, moyennant quoi la croissance, le développement ou la capacité de reproduction dudit végétal est supprimé(e) ou retardé(e), ou bien ledit végétal, sous l'effet de ladite composition contenant ledit polynucléotide d'ARNdb, est plus sensible qu'un végétal non traité à un herbicide inhibiteur de DHPS ou un herbicide inhibiteur de mitose.

15 2. Procédé selon la revendication 1, dans lequel :

(i) ledit végétal est choisi dans le groupe constitué par les végétaux *Amaranthus palmeri*, *Amaranthus rudis*, *Amaranthus hybridus*, *Amaranthus lividus*, *Amaranthus viridis*, *Ambrosia trifida*, *Conyza canadensis* et *Euphorbia heterophylla*,

(ii) ou ladite composition contient également ledit herbicide inhibiteur de DHPS ou ledit herbicide inhibiteur de mitose et ledit traitement comporte l'application externe de ladite composition sur ledit végétal et, de préférence, ladite composition comprend en outre un ou plusieurs co-herbicide(s) différent(s) dudit herbicide inhibiteur de DHPS ou dudit herbicide inhibiteur de mitose, étant entendu que le(s)dit(s) co-herbicide(s) est ou sont choisi(s) dans l'ensemble constitué par les herbicides de type amide, herbicides arsenicaux, herbicides de type benzothiazole, herbicides de type benzoyl-cyclohexanedione, herbicides de type alkylsulfonate de benzofuranyle, herbicides de type cyclohexène-oxime, herbicides de type cyclopropyl-isoxazole, herbicides de type dicarboximide, herbicides de type dinitroaniline, herbicides de type dinitrophénol, herbicides de type éther diphenyle, herbicides de type dithiocarbamate, herbicides de type glycine, herbicides aliphatiques halogénés, herbicides de type imidazolinone, herbicides minéraux, herbicides de type nitrile, herbicides organophosphorés, herbicides de type oxadiazolone, herbicides de type oxazole, herbicides phénoxylques, herbicides de type phénylénediamine, herbicides de type pyrazole, herbicides de type pyridazine, herbicides de type pyridazinone, herbicides de type pyridine, herbicides de type pyrimidine-diamine, herbicides de type pyrimidinyloxy-benzylamine, herbicides de type ammonium quaternaire, herbicides de type thiocarbamate, herbicides de type thiocarbonate, herbicides de type thiourée, herbicides de type triazine, herbicides de type triazinone, herbicides de type triazole, herbicides de type triazolone, herbicides de type triazolo-pyrimidine, herbicides de type uracile et herbicides de type urée.

40 3. Procédé selon la revendication 1, dans lequel ladite composition comprend n'importe quelle combinaison d'au moins deux desdits polynucléotides d'ARNdb et ledit traitement comporte l'application externe de ladite composition sur ledit végétal.

45 4. Composition comprenant un polynucléotide d'ARNdb et un agent de transfert, dans laquelle ledit polynucléotide d'ARNdb est identique ou complémentaire à une séquence d'ARN correspondant à une séquence de gène de DHPS, ladite séquence de gène de DHPS étant choisie dans l'ensemble constitué par les séquences SEQ ID NO : 1, SEQ ID NO : 2, SEQ ID NO : 5, SEQ ID NO : 9, SEQ ID NO : 11, SEQ ID NO : 18 et SEQ ID NO : 19, et un fragment polynucléotidique de celles-ci comportant au moins 18 nucléotides contigus provenant de ladite séquence de gène de DHPS, et ledit agent de transfert est une composition à base d'organosilicone ou un composé de type organosilicone contenu en son sein, et prépare la surface d'un végétal à la perméation dudit polynucléotide d'ARNdb, moyennant quoi ledit végétal traité par ladite composition voit sa croissance, son développement ou sa capacité de reproduction régulé(e), supprimé(e) ou retardé(e), ou bien ledit végétal, sous l'effet de ladite composition contenant ledit polynucléotide d'ARNdb, est plus sensible qu'un végétal non traité à un herbicide inhibiteur de DHPS ou un herbicide inhibiteur de mitose.

55 5. Composition selon la revendication 4, dans laquelle :

(i) ladite composition contient également ledit herbicide inhibiteur de DHPS ou ledit herbicide inhibiteur de mitose et, de préférence, ledit herbicide inhibiteur de DHPS est choisi dans l'ensemble constitué par les car-

bamates et l'asulame, ou ledit herbicide inhibiteur de mitose est choisi parmi les herbicides de type dinitroaniline, (ii) ou ladite composition comprend en outre un co-herbicide, ledit co-herbicide étant choisi dans l'ensemble constitué par les herbicides de type amide, herbicides arsenicaux, herbicides de type benzothiazole, herbicides de type benzoyl-cyclohexanedione, herbicides de type alkylsulfonate de benzofuranyle, herbicides de type cyclohexène-oxime, herbicides de type cyclopropyl-isoxazole, herbicides de type dicarboximide, herbicides de type dinitroaniline, herbicides de type dinitrophénol, herbicides de type éther diphenylique, herbicides de type dithiocarbamate, herbicides de type glycine, herbicides aliphatiques halogénés, herbicides de type imidazolnone, herbicides minéraux, herbicides de type nitrile, herbicides organophosphorés, herbicides de type oxa-diazolone, herbicides de type oxazole, herbicides phénoxyliques, herbicides de type phénylène-diamine, herbicides de type pyrazole, herbicides de type pyridazine, herbicides de type pyridazinone, herbicides de type pyridine, herbicides de type pyrimidine-diamine, herbicides de type pyrimidinyloxy-benzylamine, herbicides de type ammonium quaternaire, herbicides de type thiocarbamate, herbicides de type thiocarbonate, herbicides de type thiourée, herbicides de type triazine, herbicides de type triazinone, herbicides de type triazole, herbicides de type triazolone, herbicides de type triazolo-pyrimidine, herbicides de type uracile et herbicides de type urée.

15 6. Procédé de réduction de l'expression d'un gène de DHPS dans un végétal, qui comporte l'application externe sur ledit végétal d'une composition comprenant un polynucléotide d'ARNdb et un agent de transfert, étant entendu que ledit polynucléotide d'ARNdb est identique ou complémentaire à une séquence d'ARN correspondant à une séquence de gène de DHPS, ladite séquence de gène de DHPS étant choisie dans l'ensemble constitué par les séquences SEQ ID NO : 1, SEQ ID NO : 2, SEQ ID NO : 5, SEQ ID NO : 9, SEQ ID NO : 11, SEQ ID NO : 18 et SEQ ID NO : 19, et un fragment polynucléotidique de celles-ci comportant au moins 18 nucléotides contigus provenant de ladite séquence de gène de DHPS, et ledit agent de transfert est une composition à base d'organosilicone ou un composé de type organosilicone contenu en son sein, et prépare la surface dudit végétal à la perméation dudit polynucléotide d'ARNdb, moyennant quoi ladite expression dudit gène de DHPS est moins importante que dans un végétal non traité.

20 7. Procédé d'identification de polynucléotides d'ARNdb utilisables pour moduler l'expression du gène de DHPS en traitant un végétal par voie externe, qui comprend le fait de :

25 a) se procurer plusieurs polynucléotides d'ARNdb qui comportent une région identique ou complémentaire à un fragment polynucléotidique d'une longueur d'au moins 18 nucléotides contigus, qui est lui-même identique ou complémentaire à une séquence de gène de DHPS choisie dans l'ensemble constitué par les séquences SEQ ID NO : 1, SEQ ID NO : 2, SEQ ID NO : 5, SEQ ID NO : 9, SEQ ID NO : 11, SEQ ID NO : 18 et SEQ ID NO : 19, b) traiter par voie externe ledit végétal avec une composition comprenant un ou plusieurs desdits polynucléotides d'ARNdb et un agent de transfert, ledit agent de transfert étant une composition à base d'organosilicone ou un composé de type organosilicone contenu en son sein, et préparant la surface dudit végétal à la perméation dudit ou desdits polynucléotide(s) d'ARNdb, c) analyser la modulation de l'expression du gène de DHPS dans ledit végétal ou extrait végétal, d) et vérifier si ledit végétal traité par ladite composition voit sa croissance, son développement ou sa capacité de reproduction régulé(e), supprimé(e) ou retardé(e), ou bien si ledit végétal, sous l'effet de ladite composition, est plus sensible qu'un végétal non traité à un herbicide inhibiteur de DHPS ou un herbicide inhibiteur de mitose.

30 8. Procédé selon la revendication 7, dans lequel :

35 (i) ledit végétal est choisi dans le groupe constitué par les végétaux *Amaranthus palmeri*, *Amaranthus rudis*, *Amaranthus hybridus*, *Amaranthus lividus*, *Amaranthus viridis*, *Ambrosia trifida*, *Conyza canadensis* et *Euphorbia heterophylla*, (ii) ladite expression du gène de DHPS dans ledit végétal est moins importante que dans un végétal non traité.

40 9. Composition agrochimique comprenant un mélange constitué par un polynucléotide d'ARNdb, un agent de transfert, un herbicide inhibiteur de DHPS ou un herbicide inhibiteur de mitose, et un co-herbicide, dans laquelle ledit polynucléotide d'ARNdb est identique ou complémentaire à une portion d'une séquence d'ARN correspondant à une séquence de gène de DHPS, ladite séquence de gène de DHPS étant choisie dans l'ensemble constitué par les séquences SEQ ID NO : 1, SEQ ID NO : 2, SEQ ID NO : 5, SEQ ID NO : 9, SEQ ID NO : 11, SEQ ID NO : 18 et SEQ ID NO : 19, et un fragment polynucléotidique de celles-ci comportant au moins 18 nucléotides contigus provenant de ladite séquence de gène de DHPS, et ledit agent de transfert est une composition à base d'organosilicone ou un composé de type organosilicone contenu en son sein, et prépare la surface d'un végétal à la perméation dudit polynucléotide d'ARNdb, moyennant quoi ledit végétal traité par ladite composition voit sa croissance, son déve-

loppement ou sa capacité de reproduction régulé(e), supprimé(e) ou retardé(e), ou bien ledit végétal, sous l'effet de ladite composition contenant ledit polynucléotide d'ARNdb, est plus sensible qu'un végétal non traité audit herbicide inhibiteur de DHPS ou audit herbicide inhibiteur de mitose.

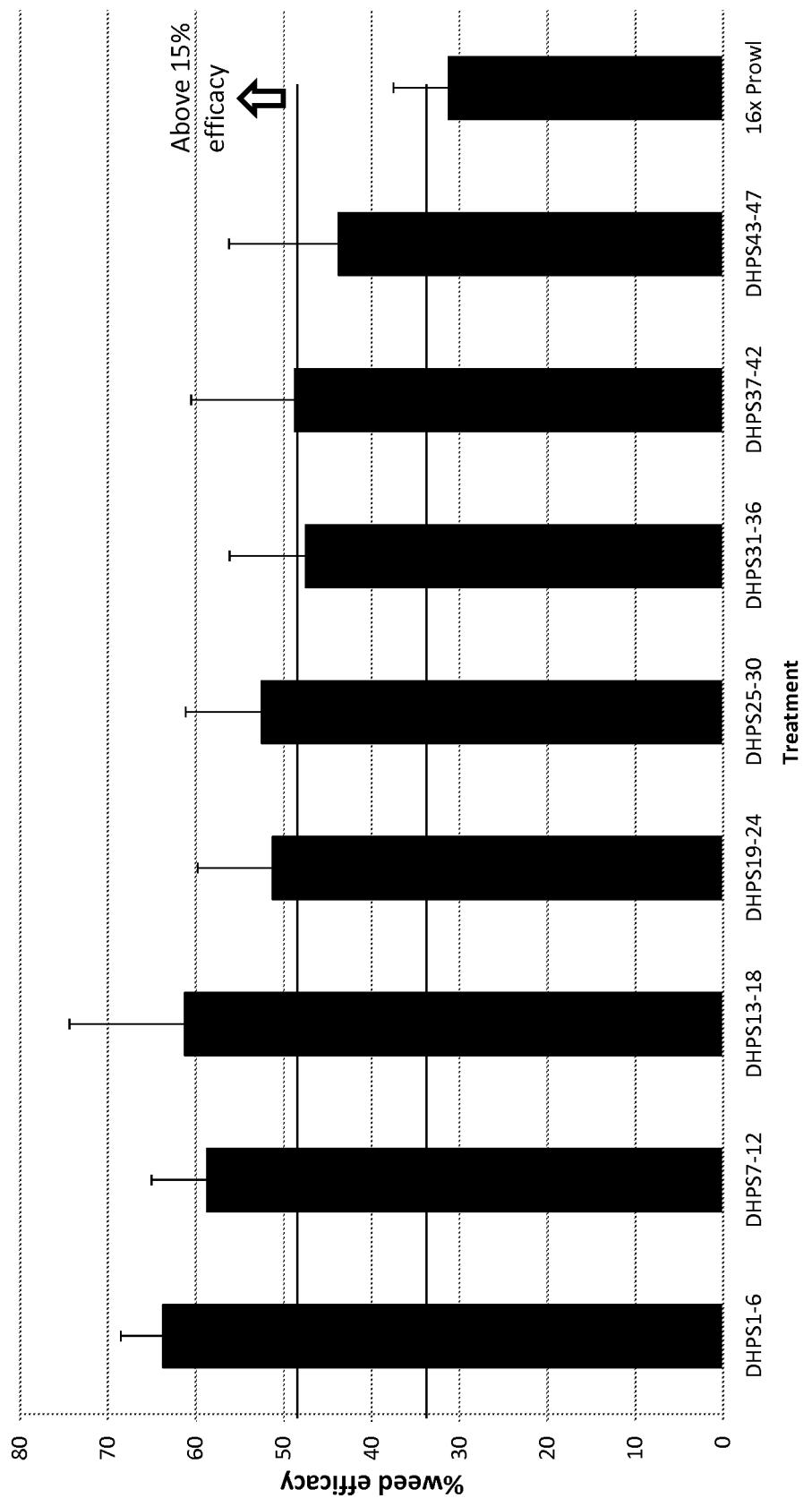
- 5      **10.** Composition agrochimique selon la revendication 9, dans laquelle ledit co-herbicide est choisi dans l'ensemble constitué par les herbicides de type amide, herbicides arsenicaux, herbicides de type benzothiazole, herbicides de type benzoyl-cyclohexanedione, herbicides de type alkylsulfonate de benzofuranyle, herbicides de type cyclohexène-oxime, herbicides de type cyclopropyl-isoxazole, herbicides de type dicarboximide, herbicides de type dinitroaniline, herbicides de type dinitrophénol, herbicides de type éther diphénylique, herbicides de type dithiocarbamate, herbicides de type glycine, herbicides aliphatiques halogénés, herbicides de type imidazolinone, herbicides minéraux, herbicides de type nitrile, herbicides organophosphorés, herbicides de type oxadiazolone, herbicides de type oxazole, herbicides phénoxyliques, herbicides de type phénylène-diamine, herbicides de type pyrazole, herbicides de type pyridazine, herbicides de type pyridazinone, herbicides de type pyridine, herbicides de type pyrimidine-diamine, herbicides de type pyrimidinyloxy-benzylamine, herbicides de type ammonium quaternaire, herbicides de type thiocarbamate, herbicides de type thiocarbonate, herbicides de type thiourée, herbicides de type triazine, herbicides de type triazinone, herbicides de type triazole, herbicides de type triazolone, herbicides de type triazolo-pyrimidine, herbicides de type uracile et herbicides de type urée.
- 10     **11.** Composition agrochimique comprenant un mélange constitué par un polynucléotide d'ARNdb, un agent de transfert, un herbicide inhibiteur de DHPS et un pesticide, dans laquelle ledit polynucléotide d'ARNdb est identique ou complémentaire à une portion d'une séquence d'ARN correspondant à une séquence de gène de DHPS, ladite séquence de gène de DHPS étant choisie dans l'ensemble constitué par les séquences SEQ ID NO : 1, SEQ ID NO : 2, SEQ ID NO : 5, SEQ ID NO : 9, SEQ ID NO : 11, SEQ ID NO : 18 et SEQ ID NO : 19, et un fragment polynucléotidique de celles-ci comportant au moins 18 nucléotides contigus provenant de ladite séquence de gène de DHPS, et ledit agent de transfert est une composition à base d'organosilicone ou un composé de type organosilicone contenu en son sein, et prépare la surface d'un végétal à la perméation dudit polynucléotide d'ARNdb, moyennant quoi ledit végétal traité par ladite composition voit sa croissance, son développement ou sa capacité de reproduction régulé(e), supprimé(e) ou retardé(e), ou bien ledit végétal, sous l'effet de ladite composition contenant ledit polynucléotide d'ARNdb, est plus sensible qu'un végétal non traité audit herbicide inhibiteur de DHPS ou à un herbicide inhibiteur de mitose.
- 15     **12.** Composition agrochimique selon la revendication 11, dans laquelle ledit pesticide est choisi dans le groupe constitué par les insecticides, fongicides, nématicides, bactéricides, acaricides, régulateurs de croissance, chimiostérilisants, substances sémiochimiques, répulsifs, attractifs, phéromones, phagostimulants et biopesticides.
- 20     **13.** Composition comprenant un polynucléotide d'ARNdb et un agent de transfert selon la revendication 4, dans laquelle ledit polynucléotide d'ARNdb est identique ou complémentaire à une séquence d'ARN correspondant à une séquence de gène de DHPS, ladite séquence de gène de DHPS comprenant les séquences SEQ ID NO : 1176 et SEQ ID NO : 1186, un complément de celles-ci et un fragment polynucléotidique de celles-ci comportant au moins 18 nucléotides contigus provenant de ladite séquence de gène de DHPS.

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**Treatment with oligonucleotide pools followed by Prowl  
(pendimethalin)**



**Figure 1**

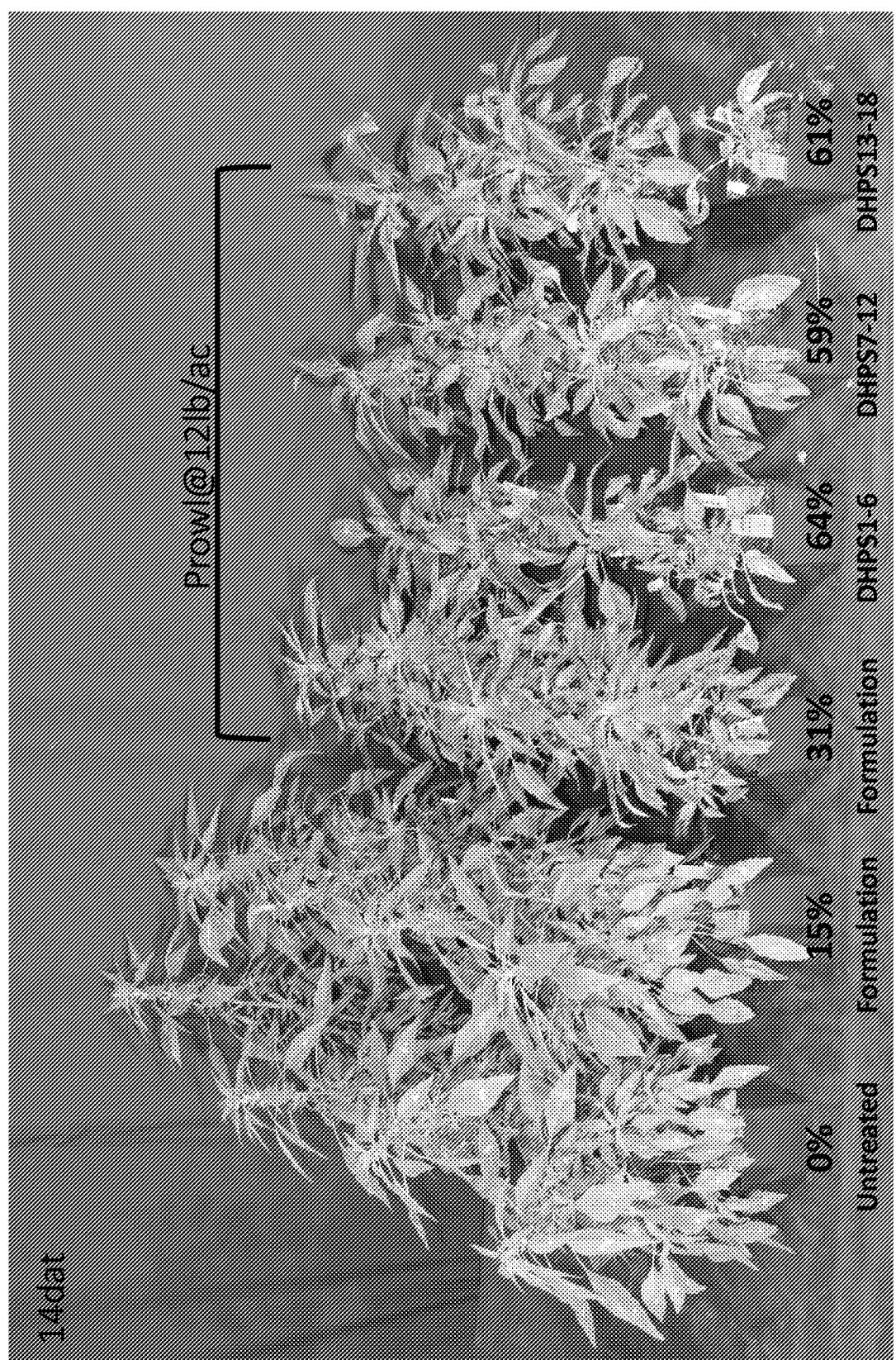


Figure 2

## REFERENCES CITED IN THE DESCRIPTION

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