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(54) **METHOD FOR AGGLUTINATING ERYTHROCYTES, METHOD FOR SEPARATING ERYTHROCYTES, AND HEMAGGLUTINATION REAGENT**

(57) Methods of agglutinating and separating erythrocytes, by which erythrocytes can be instantaneously agglutinated into a sufficient size in a blood sample and completely separated from the blood sample; and a hemagglutination reagent are provided. The method of agglutinating erythrocytes according to the present invention includes adding a solution containing a cholic ac-

id-based surfactant and an acid to a blood sample. The method of separating erythrocytes according to the present invention includes separating the erythrocytes agglutinated by the above-described method of the present invention. The hemagglutination reagent according to the present invention contains a cholic acid-based surfactant and an acid.

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Description

TECHNICAL FIELD

5 **[0001]** The present invention relates to a method of agglutinating erythrocytes, a method of separating erythrocytes and a hemagglutination reagent.

BACKGROUND ART

10 **[0002]** Blood is used in diagnostic and therapeutic determinations of various diseases and includes blood cell components such as erythrocyte, leukocyte and platelet, and plasma as a fluid component. The plasma (fluid component) contained in blood, and serum obtained by removing blood cell components and fibrin from blood contain various components required for maintenance of functions in organisms, such as proteins, saccharides and lipids, and are used as analysis samples for biochemical tests for diagnoses and treatments of visceral diseases and the like.

15 **[0003]** However, in cases where blood cell components present in an analysis sample during a biochemical test, blood cell components may disturb the test. In particular, in cases where a biochemical test using colorimetric analysis or the like is carried out, erythrocytes may affect the color and the turbidity of the sample and disturb the test.

20 **[0004]** Thus, in biochemical tests, plasma or serum obtained by previously separating blood cell components such as erythrocyte from a whole blood sample is used as an analysis sample. As a method of separating blood cell components and plasma or serum, a method is used, comprising collecting blood from a patient using a blood collection needle, placing the blood in a blood collection tube, setting the blood collection tube containing the blood in a centrifuge, and centrifuging the blood. However, such a separation method using a centrifuge has problems such as requiring a long time and complicated techniques, and thus a simpler separation method has been demanded.

25 **[0005]** Separation methods other than centrifugation include methods using a special filter such as a blood cell separation membrane, a blood cell separation material, or a hollow fiber. For example, a method of separating erythrocytes from a whole blood sample using a filter comprising a solid phase support particle, a solid phase support matrix membrane, and a hemagglutinin to form plasma has been reported (Patent Document 1).

30 **[0006]** However, when such a filter is used, the dropped blood sample permeates and spreads in the filter, which results in remain of plasma or serum in the filter. Thus, when a small amount of blood sample is used, the amount of plasma or serum after separating erythrocytes is small, which leads to a problem that a sufficient amount of analysis sample for a biochemical test cannot be secured. Furthermore, hemolysis and scattering of blood may sometimes be occurred even when small pressure is applied to the filter during the separation operation.

35 **[0007]** As another method of removing erythrocytes from whole blood, a method comprising contacting a whole blood sample with a solution containing acids selected from the group consisting of acetic acid, citric acid, ascorbic acid, lactic acid, maleic acid, malic acid and malonic acid under conditions effective for agglutination of erythrocytes, and then filtering the sample with a fibrous material is reported (Patent Document 2).

40 **[0008]** Cholic acid-based surfactant, which is one type of surfactants, is an anionic surfactant used for extraction and separation of membrane proteins. For example, the cholic acid-based surfactant is reported to be used as a hemolytic agent when the concentration of hemoglobin is measured using an apparatus equipped with a biosensor (Patent Document 3). However, examples in which a cholic acid-based surfactant is used when erythrocytes are agglutinated have not yet been reported.

PRIOR ART DOCUMENTS

45 PATENT DOCUMENTS

[0009]

50 Patent Document 1: U.S. Patent No. 5981294
 Patent Document 2: U.S. Patent No. 5118428
 Patent Document 3: JP 2014-102143 A

SUMMARY OF THE INVENTION

55 PROBLEMS TO BE SOLVED BY THE INVENTION

[0010] As described above, conventional methods of separating erythrocytes using centrifugation or special filters have problems that they are complex operations requiring a long time or cause hemolysis or blood scattering. In addition,

the method of agglutinating erythrocytes using acids (Patent Document 2) results in incomplete separation of erythrocytes due to the fine size of the agglutinated erythrocytes, which gives inadequate analytical reagent for biochemical tests.

[0011] The present invention has been accomplished in view of the above circumstances, and the purpose of the present invention is to provide a method of agglutinating erythrocytes, a method of separating erythrocytes, and a hemagglutination reagent, which can agglutinate erythrocytes instantaneously into a sufficient size in a blood sample and separate erythrocytes completely from the blood sample.

MEANS FOR SOLVING THE PROBLEMS

[0012] The present inventors intensively studied to find that, in a method of agglutinating erythrocytes in a blood sample, addition of a solution containing a cholic acid-based surfactant together with an acid to the blood sample can provide agglutinated erythrocytes instantaneously in a sufficient size, as compared with conventional methods of agglutinating erythrocytes in which the acid is added alone, thereby completing the present invention.

[0013] Although cholic acid-based surfactants have been known as a hemolytic agent, it is a conventionally unexpected fact that by adding a solution containing a cholic acid-based surfactant in combination with an acid to a blood sample during agglutination of erythrocytes, the erythrocytes can be instantaneously agglutinated into a sufficient size without causing hemolysis.

[0014] That is, the present invention provides a method of agglutinating erythrocytes, comprising adding a solution containing a cholic acid-based surfactant and an acid to a blood sample. The present invention also provides a method of separating erythrocytes from the blood sample, the method comprising separating the erythrocytes agglutinated by the method of the present invention described above. The present invention further provides a hemagglutination reagent containing a cholic acid-based surfactant and an acid. The present invention still further provides use of the reagent of the present invention as a reagent for agglutinating erythrocytes.

EFFECT OF THE INVENTION

[0015] By using the method and the reagent of the present invention, erythrocytes in a blood sample can be instantaneously agglutinated into a sufficient size, which enables complete separation of erythrocytes and easy and convenient separation of erythrocytes from components including serum and plasma, even without using a conventional separation method using long-time centrifugation or a special filter.

BRIEF DESCRIPTION OF THE DRAWINGS

[0016]

Fig. 1 shows a photograph showing results from hemagglutination evaluation tests conducted in Comparative Example 1-1, Comparative Example 1-2 and Example 1.

Fig. 2 shows a photograph showing results from hemagglutination evaluation tests conducted in Comparative Example 2-1, Comparative Example 2-2 and Example 2.

Fig. 3 shows a photograph showing results from hemagglutination evaluation tests conducted in Comparative Example 3-1, Comparative Example 3-2 and Example 3.

Fig. 4 shows a photograph showing results from hemagglutination evaluation tests conducted in Comparative Example 4-1, Comparative Example 4-2 and Example 4.

MODE FOR CARRYING OUT THE INVENTION

[0017] The method of agglutinating erythrocytes according to the present invention is characterized in that a solution containing a cholic acid-based surfactant and an acid is added to a blood sample.

(Blood Sample)

[0018] The blood sample, as used herein, refers to a sample containing erythrocytes, obtained by collecting blood from a human or animal subject. As the blood sample, whole blood collected from subjects may be used as it is or a sample obtained by diluting the collected whole blood with physiologic saline or the like may be used. For ease of operation, whole blood collected from subjects is preferably used as it is. The amount of the blood sample used in the present invention is usually from 10 μ l to 50 ml, preferably from 10 μ l to 1 ml, more preferably from 10 μ l to 100 μ l.

(Cholic Acid-based Surfactant)

[0019] The cholic acid-based surfactant, as used herein, refers to a surfactant having a steroid skeleton in the molecule, specifically, a surfactant having a structure of cholic acid or a derivative of cholic acid. The derivative of cholic acid means any compounds derived from cholic acid, specifically including dehydroxylated compounds such as deoxycholic acid and substituted amide compounds such as taurodeoxycholic acid.

[0020] The cholic acid-based surfactant, as used herein, is preferably selected from the group consisting of anionic surfactants such as cholic acid, sodium cholate, deoxycholic acid, sodium deoxycholate, sodium taurodeoxycholate, chenodeoxycholic acid, sodium chenodeoxycholate, glycocholic acid, sodium glycocholate, glycodeoxycholic acid, sodium glycodeoxycholate, glycolithocholic acid, sodium glycolithocholate, lithocholic acid, sodium lithocholate, taurocholic acid, sodium taurocholate, tauroursodeoxycholic acid, and sodium tauroursodeoxycholate; nonionic surfactants such as polyoxyethylene cholesteryl ether and N,N-Bis(3-D-gluconamidopropyl)cholamide (BIGCHAPS); amphoteric surfactants such as 3-((3-Cholamidopropyl)dimethylammonium)-1-propanesulfonate (CHAPS); and hydrates thereof. These cholic acid-based surfactants may be used alone or in combination of two or more.

[0021] Among them, the cholic acid-based surfactant is preferably anionic cholic acid-based surfactants such as sodium cholate, sodium deoxycholate, sodium taurodeoxycholate and hydrates thereof, particularly preferably sodium taurodeoxycholate and a hydrate thereof, from the viewpoint of rate of agglutination and more complete separation of erythrocytes.

(Acid)

[0022] As the acid used in the present invention, any substance releasing hydrogen ions (H^+) in an aqueous solution may be used. Specific examples of the acid include inorganic acids and organic acids, and salts thereof. Examples of the inorganic acid include hydrochloric acid, hydrobromic acid, sulfuric acid, sulfamic acid, phosphoric acid, and nitric acid. Examples of the organic acid include acetic acid, propionic acid, succinic acid, glycolic acid, stearic acid, lactic acid, malic acid, tartaric acid, citric acid, malonic acid, ascorbic acid, pantoic acid, maleic acid, adipic acid, alginic acid, aspartic acid, hydroxymaleic acid, phenylacetic acid, glutamic acid, benzoic acid, salicylic acid, sulfanilic acid, 2-acetoxybenzoic acid, fumaric acid, benzenesulfonic acid, toluenesulfonic acid, methanesulfonic acid, 2-naphthalenesulfonic acid, ethanedithionylsulfonic acid, oxalic acid, isethionic acid, glucoheptanoic acid, glycerophosphate, hemisulfonic acid, heptanoic acid, hexanoic acid, hydrochloric acid, hydrobromic acid, hydroiodic acid, 2-hydroxyethanesulfonic acid, 2-naphthalenesulfonic acid, pectinic acid, phosphoric acid, sulfuric acid, 3-phenylpropionic acid, picric acid, pivalic acid, thiocyanic acid, p-toluenesulfonic acid, butyric acid, camphoric acid, camphorsulfonic acid, digluconic acid, cyclopentane-propionic acid, disulfuric acid, dodecylsulfuric acid, ethanesulfonic acid, and undecanoic acid. In the present invention, the acids may be used alone or in combination of two or more.

[0023] The organic acids are preferably used as the acid used in the present invention from the viewpoint of agglutination speed and more complete separation of erythrocytes. Among them, preferably organic acids selected from the group consisting of acetic acid, tartaric acid, malonic acid, malic acid and citric acid, more preferably polycarboxylic acids, and particularly preferably citric acid are used.

(Hemagglutination Reagent)

[0024] In the present invention, erythrocytes are agglutinated by adding a solution containing the cholic acid-based surfactant and the acid described above to a blood sample. Such a solution containing the cholic acid-based surfactant and the acid to be added to a blood sample can be used as a reagent for agglutinating erythrocytes (hemagglutination reagent).

[0025] Such a solution containing the cholic acid-based surfactant and the acid (hereinafter referred to as "hemagglutination reagent") can be prepared by a method comprising adding the cholic acid-based surfactant to an aqueous acid solution obtained by diluting the acid with a physiologic saline or the like. The concentration of the acid in the hemagglutination reagent is usually from 1 mM to 1 M, preferably from 5 mM to 500 mM, more preferably from 10 mM to 200 mM, from the viewpoint of agglutination speed and more complete separation of erythrocytes. The pH of the hemagglutination reagent is preferably from 2.0 to 5.0, more preferably from 2.2 to 4.5, still more preferably from 2.5 to 4.0. The pH of the hemagglutination reagent can be measured with a commercially available pH meter by a glass electrode method. The hemagglutination reagent may contain surfactants (for example, surfactants such as Tween 20 and Emulgen A500), reagents and the like, which do not cause hemolysis, in addition to the acid, the cholic acid-based surfactant, and the physiologic saline.

[0026] The concentration of the cholic acid-based surfactant in the hemagglutination reagent is usually from 0.05 to 15.0% by weight, preferably from 0.06 to 13.0% by weight, more preferably from 0.075 to 8.0% by weight, particularly preferably from 0.1 to 6.0% by weight.

[0027] The ratio (molar ratio) of the cholic acid-based surfactant and the acid in the hemagglutination reagent is usually from 1:7 to 1:2000, preferably from 1:8 to 1:1667, more preferably from 1:13 to 1:1333, particularly preferably from 1:17 to 1:1000, from the viewpoint of agglutination speed and more complete separation of erythrocytes. The amount of the hemagglutination reagent added to the blood sample to be used (volume ratio) is usually from 1:4 to 1:80, preferably from 1:8 to 1:20, more preferably from 1:10 to 1:13.

(Method of Agglutinating Erythrocytes)

[0028] In the present invention, when the hemagglutination reagent described above is directly added to a blood sample at room temperature, erythrocytes are instantaneously agglutinated into a sufficient size. In order to allow the agglutination to be progressed more completely, the blood sample is preferably left to stand for 5 seconds to 30 seconds, preferably for about 5 seconds in the state wherein the blood sample contacts the hemagglutination reagent, and then mixed by upside-down mixing several times. By agglutinating erythrocytes in this manner, an agglutinate having a sufficient size can be obtained.

[0029] Since the erythrocytes agglutinated using the method and the reagent of the present invention have a sufficient size, they can be easily and conveniently separated from the blood sample. Examples of the method of separating the agglutinated erythrocytes include a method in which the supernatant is recovered after the agglutinated erythrocytes are precipitated, and a method of separating them by a filtration with a filter paper. Since the serum component and the plasma component from which erythrocytes have been separated off in this manner are substantially free of erythrocytes, they can be used as analysis samples for biochemical tests.

EXAMPLES

[0030] The present invention will now be described in more detail based on Examples. However, the present invention is not limited to these Examples.

(Examples 1 to 4 and Comparative Examples 1-1 to 4-2)

Hemagglutination Depending on the Presence of Cholic Acid-based Surfactant and Salt and pH Range

[0031] To 71 mL of distilled water, 10 mL of 10 w/v% sodium taurodeoxycholate hydrate (TDOC), 10 mL of 1 M citric acid, and 9 mL of 10 w/v% sodium chloride were added to prepare a solution containing 100 mM citric acid, 1% TDOC, and 0.9% sodium chloride as final concentrations (hemagglutination reagent). A solution was also prepared in the similar manner except that TDOC and/or sodium chloride were not added (hemagglutination reagent), and effects of the presence and absence of the cholic acid-based surfactant and the salt and the pH range on agglutination of erythrocytes were compared.

[0032] After adding 60 μ L of a whole blood sample to 400 μ L of the hemagglutination reagent, the rate of agglutination of erythrocytes was visually evaluated. The rate of agglutination was evaluated as +++, ++, +, or \pm in order of rapidity based on the criteria described below. Hemolysis without agglutination was evaluated as -.

+++ : erythrocytes were instantaneously agglutinated into a sufficient size and the agglutinated erythrocytes were precipitated.

++ : erythrocytes started to agglutinate after leaving to stand for a few seconds, and agglutinated erythrocytes were gradually precipitated.

+ : erythrocytes were agglutinated, but the agglutinates were very small and the agglutination was incomplete.

\pm : only very few erythrocytes were agglutinated.

- : erythrocytes were hemolyzed without being agglutinated

[0033] The results are shown in Table 1 below.

[0034] The pH of the hemagglutination reagent was measured using a desktop pH meter (HORIBA) (similarly in the following Examples and Comparative Examples).

[Table 1]

Example No.	tube No.	pH	NaCl	TDOC	Rate of agglutination	Figure
Comparative Example 1-1	1	2.5	×	×	-	Fig. 1
Comparative Example 1-2	2		○	×	+	
Example 1	3		○	○	+++	
Comparative Example 2-1	4	3.0	×	×	-	Fig. 2
Comparative Example 2-2	5		○	×	+	
Example 2	6		○	○	+++	
Comparative Example 3-1	7	4.0	×	×	-	Fig. 3
Comparative Example 3-2	8		○	×	+	
Example 3	9		○	○	+++	
Comparative Example 4-1	16	2.5	×	○	+++	Fig. 4
Comparative Example 4-2	17	3.0	×	○	+++	
Example 4	18	4.0	×	○	+++	

[0035] As can be seen from the results in Table 1, when the hemagglutination reagent contained the cholic acid-based surfactant and the acid, the rate of agglutination was evaluated as "+++" within a pH range of 2.5 to 5.0 regardless of whether NaCl exists or not, and excellent agglutination effect was observed. On the other hand, when the hemagglutination reagent did not contain the cholic acid-based surfactant and contained only the acid, or only the acid and the salt, the rate of agglutination was evaluated as "-" or "+", which means that hemolysis occurred or that erythrocytes were agglutinated but the agglutinates were very small and the agglutination was incomplete. The results show that the presence of the cholic acid-based surfactant has a great effect on agglutination of erythrocytes.

(Examples 5 to 7)

Hemagglutination Depending on Cholic Acid-based Surfactant and Acid

[0036] Various cholic acid-based surfactants (1% by weight) and citric acid (100 mM) were added to a physiologic saline to prepare hemagglutination reagents (pH3.0) and their abilities to agglutinate erythrocytes were compared.

[0037] After adding 60 μ L of a whole blood sample to 400 μ L of the hemagglutination reagent, the rate of agglutination of erythrocytes was evaluated. The rate of agglutination was evaluated as +++, ++, +, or \pm in order of rapidity based on the criteria described below. Hemolysis without agglutination was evaluated as -. The following criteria were the same as those in Examples 1 to 4 and Comparative Examples 1-1 to 4-2.

+++ : Erythrocytes were instantaneously agglutinated into a sufficient size and the agglutinated erythrocytes were precipitated.

++ : Erythrocytes started to agglutinate after leaving to stand for a few seconds, and agglutinated erythrocytes were gradually precipitated.

+ : Erythrocytes were agglutinated, but the agglutinates were very small and the agglutination was incomplete.

\pm : Only very few erythrocytes were agglutinated.

- : Erythrocytes were hemolyzed without being agglutinated.

[0038] The results are shown in Table 2 below.

[Table 2]

	Surfactant	Rate of agglutination
Example 5	Deoxycholic acid	++
Example 6	Sodium cholate	++
Example 7	Sodium taurodeoxycholate hydrate (TDOC)	+++

[0039] The results in Table 2 shows that when the hemagglutination reagents contained various cholic acid-based surfactants in combination with the acid, the rates of agglutination were evaluated as "++" or "+++" and excellent agglutination effects were observed. In particular, the effect of sodium taurodeoxycholate hydrate (TDOC) on hemagglutination was most significant.

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(Example 8)

Hemagglutination Depending on Concentration of Cholic Acid-based Surfactant

[0040] Sodium taurodeoxycholate hydrate (TDOC) and citric acid (100 mM) were added to a physiologic saline to prepare hemagglutination reagents (pH3.0), and their abilities to agglutinate erythrocytes were compared in the concentration range of TDOC as shown in the table below.

[0041] After adding 60 μ L of a whole blood sample to 400 μ L of the hemagglutination reagent, the rate of agglutination of erythrocytes was evaluated. The rate of agglutination was evaluated as +++, ++, +, or \pm in order of rapidity based on the criteria below. Hemolysis without agglutination was evaluated as -. The following criteria were the same as those in Examples 1 to 7 and Comparative Examples 1-1 to 4-2.

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+++ : Erythrocytes were instantaneously agglutinated into a sufficient size and the agglutinated erythrocytes were precipitated.

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++ : Erythrocytes started to agglutinate after leaving to stand for a few seconds, and agglutinated erythrocytes were gradually precipitated.

+ : Erythrocytes were agglutinated, but the agglutinates were very small and the agglutination was incomplete.

\pm : Only very few erythrocytes were agglutinated.

- : Erythrocytes were hemolyzed without being agglutinated.

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[0042] The results are shown in Table 3 below.

[Table 3]

Concentration of TDOC (wt%)	Rate of agglutination
6.0	+++
1.0	+++
0.5	+++
0.1	+++

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[0043] The results in Table 3 shows that when the concentration of the cholic acid-based surfactant was from 0.1 to 6.0% by weight, the rate of agglutination was evaluated as "+++" and an excellent agglutination effect was observed.

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[0044] In Examples 1 to 4 described above, after the agglutinated erythrocytes were precipitated, the supernatant was separated and added dropwise to an immunochromatographic assay kit (Quick Navi® -Ebola). When the movement on a membrane was observed, the membrane was not colored red. These results also supported the fact that the above-mentioned separated supernatant did not substantially contain erythrocytes and is suitable as an analysis sample for biochemical tests.

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Claims

1. A method of agglutinating erythrocytes, the method comprising adding a solution containing a cholic acid-based surfactant and an acid to a blood sample.

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2. The method according to claim 1, wherein the solution has a pH of from 2.5 to 4.0.

3. The method according to claim 1 or 2, wherein said cholic acid-based surfactant is sodium taurodeoxycholate hydrate.

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4. A method of separating erythrocytes from a blood sample, the method comprising separating erythrocytes agglutinated by the method according to any one of claims 1 to 3.

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5. A hemagglutination reagent, which contains a cholic acid-based surfactant and an acid.
6. The reagent according to claim 5, wherein said reagent has a pH of from 2.5 to 4.0.
7. The reagent according to claim 5 or 6, wherein said cholic acid-based surfactant is sodium taurodeoxycholate hydrate.
8. Use of the reagent according to any one of claims 5 to 7 as a reagent for agglutinating erythrocytes.

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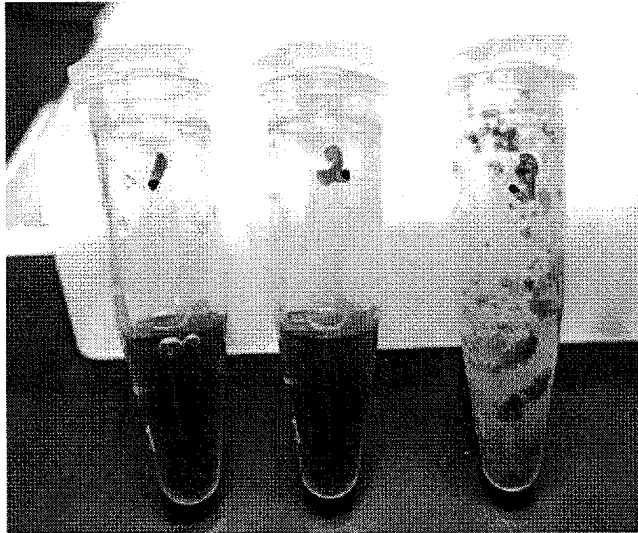


Fig.1

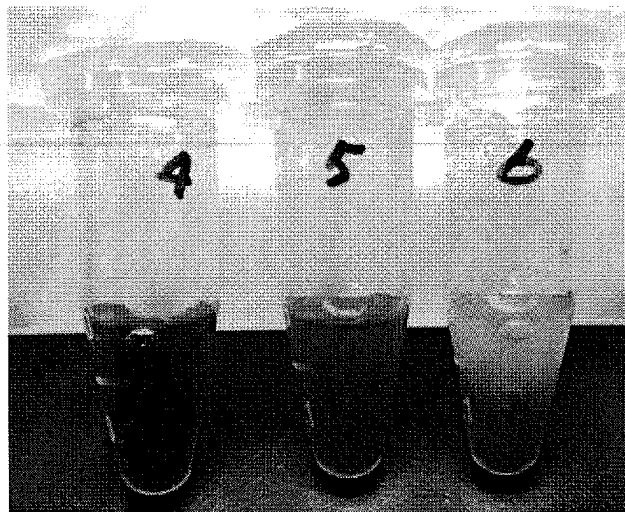


Fig.2

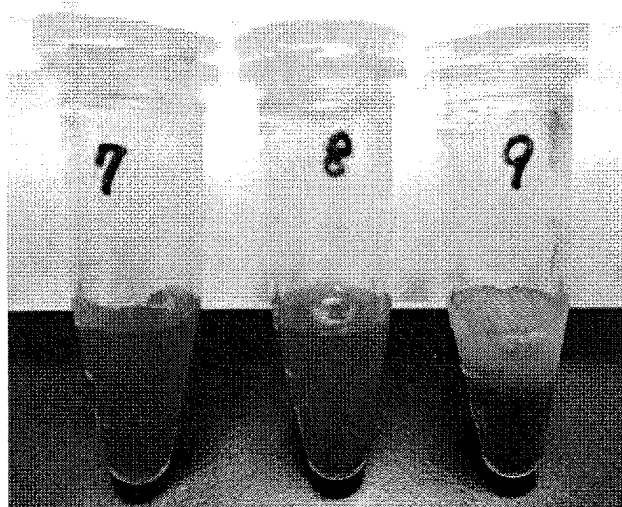


Fig.3

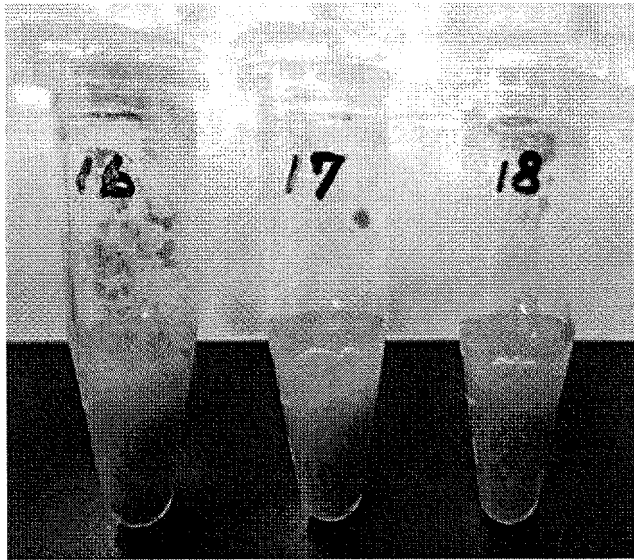


Fig.4

INTERNATIONAL SEARCH REPORT

International application No.
PCT/JP2017/036087

5	<p>A. CLASSIFICATION OF SUBJECT MATTER Int.Cl. G01N33/48 (2006.01) i</p> <p>According to International Patent Classification (IPC) or to both national classification and IPC</p>													
10	<p>B. FIELDS SEARCHED</p> <p>Minimum documentation searched (classification system followed by classification symbols) Int.Cl. G01N33/48-33/98</p>													
15	<p>Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched</p> <table border="0"> <tr> <td>Published examined utility model applications of Japan</td> <td>1922-1996</td> </tr> <tr> <td>Published unexamined utility model applications of Japan</td> <td>1971-2017</td> </tr> <tr> <td>Registered utility model specifications of Japan</td> <td>1996-2017</td> </tr> <tr> <td>Published registered utility model specifications of Japan</td> <td>1994-2017</td> </tr> </table>		Published examined utility model applications of Japan	1922-1996	Published unexamined utility model applications of Japan	1971-2017	Registered utility model specifications of Japan	1996-2017	Published registered utility model specifications of Japan	1994-2017				
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20	<p>Electronic data base consulted during the international search (name of data base and, where practicable, search terms used) JSTPlus/JMEDPlus/JST7580 (JDreamIII) CPlus/MEDLINE/EMBASE/BIOSIS (STN)</p>													
25	<p>C. DOCUMENTS CONSIDERED TO BE RELEVANT</p> <table border="1"> <thead> <tr> <th>Category*</th> <th>Citation of document, with indication, where appropriate, of the relevant passages</th> <th>Relevant to claim No.</th> </tr> </thead> <tbody> <tr> <td>A</td> <td>WO 2013/147307 A1 (SEKISUI MEDICAL CO., LTD.) 03 October 2013, paragraphs [0013]-[0014], [0037]-[0043] & US 2015/0086974 A1, paragraphs [0045]-[0057], [0133]-[0153] & EP 2833142 A1</td> <td>1-8</td> </tr> <tr> <td>A</td> <td>JP 2005-114359 A (MATSUSHITA ELECTRIC INDUSTRIAL CO., LTD.) 28 April 2005, claims, paragraphs [0029]-[0035] & US 2005/0145490 A1, claims, paragraphs [0039]-[0051]</td> <td>1-8</td> </tr> </tbody> </table>		Category*	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.	A	WO 2013/147307 A1 (SEKISUI MEDICAL CO., LTD.) 03 October 2013, paragraphs [0013]-[0014], [0037]-[0043] & US 2015/0086974 A1, paragraphs [0045]-[0057], [0133]-[0153] & EP 2833142 A1	1-8	A	JP 2005-114359 A (MATSUSHITA ELECTRIC INDUSTRIAL CO., LTD.) 28 April 2005, claims, paragraphs [0029]-[0035] & US 2005/0145490 A1, claims, paragraphs [0039]-[0051]	1-8			
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A	WO 2013/147307 A1 (SEKISUI MEDICAL CO., LTD.) 03 October 2013, paragraphs [0013]-[0014], [0037]-[0043] & US 2015/0086974 A1, paragraphs [0045]-[0057], [0133]-[0153] & EP 2833142 A1	1-8												
A	JP 2005-114359 A (MATSUSHITA ELECTRIC INDUSTRIAL CO., LTD.) 28 April 2005, claims, paragraphs [0029]-[0035] & US 2005/0145490 A1, claims, paragraphs [0039]-[0051]	1-8												
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40	<p><input checked="" type="checkbox"/> Further documents are listed in the continuation of Box C. <input type="checkbox"/> See patent family annex.</p>													
45	<table border="0"> <tr> <td>* Special categories of cited documents:</td> <td>"T" later document published after the international filing date or priority date and not in conflict with the application but cited to understand the principle or theory underlying the invention</td> </tr> <tr> <td>"A" document defining the general state of the art which is not considered to be of particular relevance</td> <td>"X" document of particular relevance; the claimed invention cannot be considered novel or cannot be considered to involve an inventive step when the document is taken alone</td> </tr> <tr> <td>"E" earlier application or patent but published on or after the international filing date</td> <td>"Y" document of particular relevance; the claimed invention cannot be considered to involve an inventive step when the document is combined with one or more other such documents, such combination being obvious to a person skilled in the art</td> </tr> <tr> <td>"L" document which may throw doubts on priority claim(s) or which is cited to establish the publication date of another citation or other special reason (as specified)</td> <td>"&" document member of the same patent family</td> </tr> <tr> <td>"O" document referring to an oral disclosure, use, exhibition or other means</td> <td></td> </tr> <tr> <td>"P" document published prior to the international filing date but later than the priority date claimed</td> <td></td> </tr> </table>		* Special categories of cited documents:	"T" later document published after the international filing date or priority date and not in conflict with the application but cited to understand the principle or theory underlying the invention	"A" document defining the general state of the art which is not considered to be of particular relevance	"X" document of particular relevance; the claimed invention cannot be considered novel or cannot be considered to involve an inventive step when the document is taken alone	"E" earlier application or patent but published on or after the international filing date	"Y" document of particular relevance; the claimed invention cannot be considered to involve an inventive step when the document is combined with one or more other such documents, such combination being obvious to a person skilled in the art	"L" document which may throw doubts on priority claim(s) or which is cited to establish the publication date of another citation or other special reason (as specified)	"&" document member of the same patent family	"O" document referring to an oral disclosure, use, exhibition or other means		"P" document published prior to the international filing date but later than the priority date claimed	
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50	<p>Date of the actual completion of the international search 20 December 2017 (20.12.2017)</p>	<p>Date of mailing of the international search report 09 January 2018 (09.01.2018)</p>												
55	<p>Name and mailing address of the ISA/ Japan Patent Office 3-4-3, Kasumigaseki, Chiyoda-ku, Tokyo 100-8915, Japan</p>	<p>Authorized officer Telephone No.</p>												

INTERNATIONAL SEARCH REPORT

International application No. PCT/JP2017/036087
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C (Continuation). DOCUMENTS CONSIDERED TO BE RELEVANT		
Category*	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
A	WO 2006/109685 A1 (NIPPON KAYAKU CO., LTD.) 19 October 2006, claims, paragraphs [0019]-[0037] & JP 4671441 B2	1-8
A	US 2015/0204843 A1 (QIAGEN GMBH) 23 July 2015, claims, paragraphs [0150]-[0153] & WO 2014/033208 A1 & EP 2890980 A1	1-8

REFERENCES CITED IN THE DESCRIPTION

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Patent documents cited in the description

- US 5981294 A [0009]
- US 5118428 A [0009]
- JP 2014102143 A [0009]