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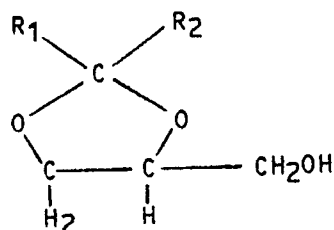
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(54) **Enantioselective preparation of S-2R₁,2R₂-1,3-dioxolane 4-methanol and derivatives thereof.**

(57) The invention relates to the enantioselective conversion of an enantiomer mixture of 2R₁,2R₂-1,3-dioxolane-4-methanol of the formula:

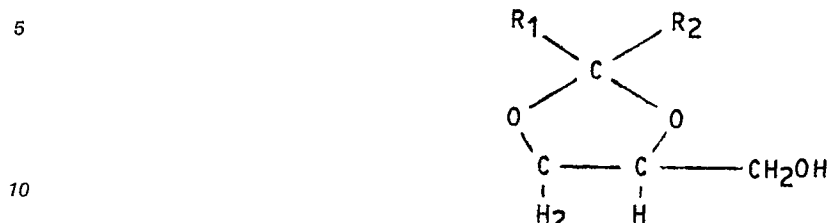


where R₁ and R₂ independently represent hydrogen or an optionally branched alkyl group, or aryl, or R₁ and R₂, together with the carbon atom to which they are attached, represent an optionally substituted carbocyclic ring, the product of said conversion being enriched in one of the enantiomers, which is achieved by subjecting said enantiomer mixture to the action of an enantio-selective enzyme, a 2R₁,2R₂-1,3-dioxolane-4-methanol enriched in S-enantiomer being prepared by the use of a suitable PQQ-dependent alcohol dehydrogenase as enantioselective enzyme.

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ENANTIOSELECTIVE PREPARATION OF S-2R₁,2R₂-1,3-DIOXOLANE-4-METHANOL AND DERIVATIVES THEREOF

This invention relates to the enantioselective conversion of an enantiomer mixture of 2R₁,2R₂-1,3-dioxolane-4-methanol of the formula:

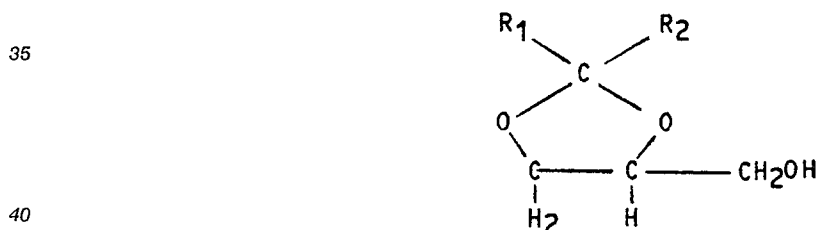


where R₁ and R₂ independently represent hydrogen or an optionally branched alkyl group, an aryl, or R₁ and R₂ together with the carbon atom to which they are attached, represent an optionally substituted carbocyclic ring, the product of said conversion being enriched in one of the enantiomers, which is achieved by subjecting this enantiomer mixture to the action of an enantioselective enzyme.

Such a process is known from EP-A-244.912. According to said patent application a product enriched in R-enantiomer can be obtained from an enantiomer mixture of an above-mentioned compound by subjecting such a mixture to the action of an enzyme that may originate from a great number of micro-organisms. According to said method it is mainly the S-enantiomer of the relevant compounds that is converted into the R-2R₁,2R₂-1,3-dioxolane-4-carboxylic acid, so that also a 2R₁,2R₂-1,3-dioxolane-4-methanol enriched in R-enantiomer remains. In said application it is also stated that the oxidation product, which contains an excess of R-2R₁,2R₂-1,3-dioxolane-4-carboxylic acid, can be reduced to the corresponding alcohol, in which the amount of S-2R₁,2R₂-1,3-dioxolane-4-methanol is predominating.

For many applications, in particular for the use as starting compound for the preparation of pharmaceutical compounds, it is desirable to have S-2R₁,2R₂-1,3-dioxolane-4-methanol at one's disposal. For this purpose it is desirable to be able to prepare a product enriched in S-enantiomer directly from an enantiomer mixture of 2R₁,2R₂-1,3-dioxolane 4-methanol. This can e.g. be achieved by means of an enantioselective enzymatic process.

A process has now been found for the enantioselective conversion of an enantiomer mixture of 2R₁,2R₂-1,3-dioxolane-4-methanol of the formula:



where R₁ and R₂ independently represent hydrogen or an optionally branched alkyl group, an aryl, or R₁ and R₂, together with the carbon atom to which they are attached, represent an optionally substituted carbocyclic ring, the product of said conversion being enriched in one of the enantiomers, which is achieved by subjecting the enantiomer mixture to the action of an enantioselective enzyme by - and this characterizes the invention - preparing 2R₁,2R₂-1,3-dioxolane-4-methanol enriched in S-enantiomer while applying a suitable PQQ-dependent alcohol dehydrogenase as enantioselective enzyme.

R₁, R₂ mostly represent H, or an C₁-C₁₀ (cyclo) alkyl, a C₅-C₁₀, preferably C₆, aryl or R₁ and R₂ form together with the C-atom to which they are attached a carbocyclic ring with 4-10, preferably 5 or 6, C-atoms. R₁, R₂ and/or the carbocyclic ring they form together with the C-atom to which they are attached may be substituted with halogen atoms, lower alkyl (C₁-C₄) or lower alkoxy (C₁-C₄).

The so-called PQQ-dependent alcohol dehydrogenase to be used in the present invention is known in itself from, for instance, the dissertation "Quinoprotein (PQQ-containing) Alcohol Dehydrogenase" by J.A.

Duine (PhD thesis, Delft, 19 September 1985) and from B.W. Groen et al. in Biochem. J. 234 (1986) p. 611 ff. PQQ stands for pyrrolo-quinoline-quinone, i.e. 2,7,9 tricarboxy-1H-pyrrolo[2,3f]quinoline-4,5-dione. Suitable PQQ-dependent alcohol dehydrogenases are e.g. Pseudomonas testosteroni or Gluconobacter suboxydans. The PQQ-dependent alcohol dehydrogenase originating from or obtained from Pseudomonas testosteroni is eminently suitable for the subject process. The Pseudomonas testosteroni micro-organism can be cultured by the method described by B.W. Groen et al., loc. cit., but also by any other suitable cultivation method. Such cultivation methods are so commonly known and have been described so frequently in patent specifications and scientific publications as to render it superfluous to describe these cultivation methods in this context. The enzyme preparation as used in the subject invention is not restricted by purity and the like and may be both a crude enzyme solution and a purified enzyme, but it may also consist of (permeabilized and/or immobilized) microbial cells that possess the desired activity, or of a homogenate of cells or whole cells possessing such activity. The enzyme may also be used in immobilized form or in chemically modified form. When some undesired opposite enzyme activity is present in the enzyme preparation used, it is recommendable for this undesired activity to be eliminated or to be suppressed in order to obtain maximum enantioselectivity. The invention is in no way restricted by the form in which the enzyme for the subject invention is used. Within the scope of the invention use may also be made of a PQQ-dependent alcohol dehydrogenase originating from a mutant of Pseudomonas testosteroni or from genetically modified micro-organisms. Preferably use is made of a PQQ-dependent alcohol dehydrogenase originating from or obtained from Pseudomonas testosteroni ATCC 15666, ATCC 15667, NCIB 8893, NCIB 8955 or NCIB 10808. According to Jin Tamaoko et al. in Int. J. Syst. Bact. 37 (1987) p. 52 ff, Pseudomonas testosteroni should be classified as Comamonas testosteroni. The organisms meant are referred to as Pseudomonas testosteroni in this application. Mostly 1 mg - 200 mg cells (dry weight) are used per gram R,S-substrate.

As is apparent from the publication by B.W. Groen et al. (loc. cit.), the PQQ-dependent alcohol dehydrogenase, such as can be obtained, for instance, from Pseudomonas testosteroni, is not active in all cases with regard to the oxidation of some alcohols: for activity a sufficient amount of PQQ must be present. If not enough PQQ is present in the cultivation of the micro-organism, then it is desirable for oxidation purposes to add it in sufficient quantities. Mostly the amount of PQQ is 0.5-2 equivalents with respect to the enzyme. For optimal results a minimum quantity of 1 equivalent is used. Instead of PQQ also PQQ analogues, such as monoesters of PQQ (at the 2-position), 4-hydroxy-PQ (PQ standing for pyrroloquinoline) or 5-hydroxy-PQ after oxidation, 3-methyl-PQQ, 3-ethyl-PQQ, 3-propyl-PQQ, N-methyl-PQQ, N-ethyl-PQQ, 8-methyl-PQQ or the PQQ-acetone adduct and other aldehyde and ketone adducts, as described by J.A. Jongejan, B.W. Groen and J.A. Duine in "PQQ and Quinoproteins" (J.A. Jongejans and J.A. Duine, eds.), Kluwer Academic Publishers, Dordrecht, 1989, pp. 205-216), may be used.

Enantioselective enzymatic conversions are known by themselves. The enantioselectivity resides in a difference in conversion rate between the enantiomers in question. Reaction products are obtained that are enriched in one of the enantiomers. For practical purposes it generally is desirable for one of the enantiomers to be obtained in a large excess. This is achieved by terminating the conversion at a certain degree of conversion. For enantioselective enzymatic hydrolyses this is described by Qu-Ming Gu et al. in Tetrahedron Letters 27 (1986), 5203 ff., and more in general by Ghing Shih Chen et al. in J. Am. Chem. Soc. 104 (1982), 7294 ff. The general doctrine of enantioselective enzymatic conversions described in these publications also applies to the subject process.

According to the subject process the PQQ-dependent alcohol dehydrogenase oxidizes R-enantiomer of 2R₁,2R₂-1,3-dioxolane-4-methanol at a more rapid rate to form the S-enantiomer of the corresponding 2R₁,2R₂-1,3-dioxolane-4-carboxylic acid, as a result of which also a reaction product enriched in S-2R₁,2R₂-1,3-dioxolane-4-methanol is obtained. The conversion rates of the enantiomers appear to differ considerably, as a result of which at 50% conversion of a racemic mixture already an enantiomer excess of more than 90% of the 2R₁,2R₂-1,3-dioxolane-4-methanol can be obtained, while an enantiomer excess of more than 95% can already be reached at a conversion between 50 and 55 %.

The cells used in the reaction may be separated from the reaction mixture and reused in a subsequent conversion, without considerable loss of activity.

According to the subject process, preferably, the S-enantiomer is prepared from 2,2-dimethyl-1,3-dioxolane-4-methanol. An enantiomer excess of S-2,2-dimethyl-1,3-dioxolane-4-methanol of at least 95% can be obtained.

The S-2,2-dimethyl-1,3-dioxolane-4-methanol is an important starting product for the preparation of pharmaceuticals, crop protection and/or agricultural pest control agents, as is also reported in a publication by J. Jurczak et al. in Tetrahedron 42 (1986) 447 ff. The R- and S-enantiomers of 2,2-dimethyl-1,3-dioxolane-4-methanol are important chiral C₃ synthons in organic synthesis. Examples of syntheses of more

complex structures from such C_3 synthons are β -receptor blocking agents and sn-glyceryl-phosphoryl choline (GPC).

Racemic 2,2-dimethyl-1,3-dioxolane-4-methanol can be prepared in the way known for the preparation of acetals by acid-catalyzed coupling of glycerol and acetone. For compounds of the formula presented in the above, in which R_1 and R_2 do not both represent a methyl group, glycerol is to be coupled with the aldehyde or ketone in question.

pH and temperature at which the process of the present invention is performed are not critical. Mostly the pH is between 5 and 9 and the temperature between 15 and 50 °C.

The invention will be further elucidated by the following examples, without being restricted by these examples. In these examples the analyses of the reaction products, notably with regard to the determination of the enantiomeric excess (ee), were performed using the method described below (TAGIT method). It is possible, if so desired, to calculate the selectivity value for the reaction with the aid of the formulae of Chen et al. (J. Am. Chem. Soc. 104, 7294 ff. (1982)). For conversion determination use was made of column separation over an Aminex HPX-87H HPLC column with 0.01 N H_2SO_4 as mobile solvent and glycerol detection on the basis of the refractive index.

All analyses were always performed using samples taken, at different points of time, from the reaction mixtures of the experiments, the amounts being 10-50 ml for conversion determination and 10-50 ml for ee determination, depending on the conversion.

In ee determination the samples are extracted with two times 20 ml dichloromethane, following which 200 μ l of the residue obtained after extract evaporation is dissolved in 5 ml diethylether. Subsequently 0.35 g tosylchloride and about 0.5 g potassium hydroxide powder are added to the ether solution, and the resulting solution is stirred at room temperature for 15 minutes. The tosyl addition product of the 2 R_1 ,2 R_2 -1,3-dioxolane-4-methanol is then isolated by extraction with diethylether and water, after which the ether fraction is washed with water and evaporated. The tosyl-2 R_1 ,2 R_2 -1,3-dioxolane-4-methanol obtained is then dissolved in 2.0 ml n-butylamine and heated for 1 hour at 100 °C. The reaction product is subsequently isolated by extraction with diethylether and water, after which the ether fraction is washed four times with water and evaporated. This residue is dissolved in 1.0 ml acetonitrile, after which 2.5 μ l of the resulting solution is mixed with 50 μ l acetonitrile and 1 mg 2,3,4,6-tetra-O-acetyl- β -glucopyranosyl-isothiocyanate (TAGIT). Five minutes after mixing the ee is determined using a reversed phase (C_{18}) column with 60/40 methanol/water as eluent solvent.

This method for determination of the enantiomer excess has been found to be very suitable. It gives an excellent separation of the R- and S-enantiomers of 2 R_1 ,2 R_2 -1,3 dioxolane-4-methanol (in derivatized form), as is illustrated in figures I.1, I.2 and I.3, which are added as specimen chromatograms of this separation. Figure I.1 relates to the racemate, for I.2 use has been made of the commercially available S-enantiomer (Janssen Chimica), and I.3 relates to the product obtained according to the invention after 56% conversion.

Example I

Pseudomonas testosteroni (American Type Culture Collection No. 15667) was cultured in a fermentor containing 20 l of mineral salts medium (composition per litre of demineralized water: 15.4 g $K_2HPO_4 \cdot 3H_2O$, 4.52 g KH_2PO_4 , 0.5 g $MgCl_2 \cdot 6H_2O$, 3 g $(NH_4)_2SO_4$, 15 mg $CaCl_2$, 15 mg $FeSO_4 \cdot 7H_2O$ and the trace elements Cu, B, Mn, Zn, Mo), to which, after sterilization, 100 ml ethanol had been added. The pH was 7.0. The fermentor contents were kept at 20 °C and stirred at a speed of 350 rpm. The aeration rate was 3.5 l per minute. After 80 hours, cultivation the cells were harvested by centrifuging. A portion of these cells (wet weight 11 g) was suspended in 100 ml tris/HCl buffer (50 mM, pH 7.5), to which 5.3 μ mol PQQ had been added. After the cells had been saturated with PQQ, 1.8 l of the same buffer and 20 g R,S-2,2-dimethyl-1,3-dioxolane-4-methanol were added. During incubation the cell suspension was aerated (1.0 l air per minute) and stirred (400 rpm), while the pH was kept at 7.5 by titration with 0.2 N NaOH. The temperature was kept at 28 °C.

During this oxidation at regular intervals samples were taken, which were analyzed in the way described above. The results are summarized in Table 1.

Table 1

Sample	Conversion	ee (S-enantiomer)
	(%)	(%)
1	15	17
2	32	45
3	45	75
4	56	99

Thus with respect to Sample 4, there was 56% conversion of 2,2-dimethyl-1,3-dioxolane-4-methanol and the enantiomeric excess (ee) of the S-enantiomer of 2,2-dimethyl-1,3 dioxolane-4-methanol was 99%.

Example II

Using the purification method described in Biochem. J. (1986) 234 , 611 ff., PQQ-dependent alcohol dehydrogenase was recovered from *Pseudomonas testosteroni* ATCC 15667, cultured as described in Example I.

The isolated enzyme was dissolved in 20 ml potassium phosphate buffer (pH 8.0; 50 mM) and saturated with 100 nmol PQQ. Subsequently 1.01 mmol potassium ferricyanide and 0.5 mmol R,S-2,2-dimethyl-1,3-dioxolane-4-methanol were added. The pH of the solution was kept at 8.0 by titration with 0.1 N NaOH. After reduction of ferricyanide the reaction mixture was analyzed. There was 51% conversion of S-2,2-dimethyl-1,3-dioxolane-4-methanol yielding the S-2,2-dimethyl-1,3-dioxolane-4-methanol with ee value of 99 %.

Example III

The alcohol dehydrogenase recovered from *Pseudomonas testosteroni* ATCC 15667 (see Example II) was dissolved in 20 ml potassium phosphate buffer (pH 8.0; 50 mM) and saturated with 100 nmol PQQ, and subsequently excess potassium ferricyanide and 0.5 mmol 2,2-pentylene-1,3-dioxolane-4-methanol were added. After oxidation of about 50% of the substrate the reaction was stopped and the reaction mixture analyzed. Conversion appeared to be 54% and the enantiomer excess of the S-enantiomer was 99%.

Example IV

Example III was repeated with 2,2-butylene-1,3-dioxolane-4-methanol instead of 2,2-pentylene-1,3-dioxolane-4-methanol. The conversion was 53% and the enantiomer excess of the S-enantiomer 99%.

Example V

A 2.0 l Erlenmeyer flask, filled with 500 ml mineral salts medium (see Example I), 2.5 ml ethanol and 2 μ mol PQQ, was inoculated with *Pseudomonas testosteroni* ATCC 15667. Both PQQ (filter sterilized) and ethanol had been added after sterilization by heating of the medium. The cultivation temperature was 28 °C, and shaking took place at 200 rpm. After 36 hours the cells were harvested by centrifuging, after which they were suspended in 100 ml potassium phosphate buffer (pH 7.0; 50 mM). After addition of 1.0 g R,S-2,2-dimethyl-1,3-dioxolane-4-methanol, the reaction medium was incubated at 28 °C while being shaken (200 rpm). After 24 hours the reaction medium was analyzed. Conversion was 54%, and the ee value for the S-enantiomer 96%.

Example VI

Using the method of Example V, *Pseudomonas testosteroni* ATCC 15666, NCIB 8893 and NCIB 10808 were respectively cultured, and each time the PQQ-dependent alcohol dehydrogenase obtained was used for enantiospecific oxidation of R,S-2,2-dimethyl-1,3-dioxolane-4-methanol as described in example IV, with similar results as regards the enantioselectivity.

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Example VII

Using the purification method described in Agric. Biol. Chem. 42 (1978) 2045-2056, purified, membrane bound, PQQ-dependent alcohol dehydrogenase recovered from *Gluconobacter suboxydans* ATCC 621, was dissolved in 20 ml potassium phosphate buffer (pH = 7.0; 50 mM) and saturated with 100 nmol PQQ. Subsequently excess potassium ferricyanide and 0.5 mmol R,S-2,2-dimethyl-1,3-dioxolane-4-methanol were added. After oxidation of about 60% of the substrate, the reaction was stopped. There was 64% conversion to S-2,2-dimethyl-1,3-dioxolane-4-methanol with enantiomeric 96%.

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Example VIII

Pseudomonas testosteroni ATCC 15667 was cultured in a fermentor containing 5 l of a medium containing mineral salts as described in Example I (pH = 7), to which, after sterilization, 25 ml ethanol, as a Carbon-source, was added. At the end of the exponential growth phase, the pH was increased to 7.5. Subsequently 150 mmol R,S-2,2-dimethyl-1,3-dioxolane-4-methanol and 2.5 mg PQQ were added while the pH was kept at 7.5 by titration with 0.2 N NaOH. After oxidation of 57 % of the substrate added, the cells were separated off with a centrifuge, and the remaining substrate was isolated by extraction. The enantiomeric excess of the S-enantiomer of 2,2-dimethyl-1,3-dioxolane-4-methanol was $\geq 99.5\%$.

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Example IX

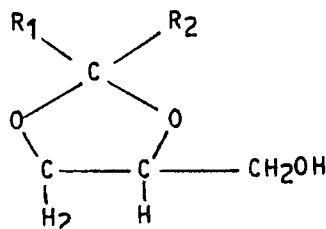
The cells remaining from example VIII were reused. To this end the cells were suspended in 1.0 l tris/HCl buffer (50 mM, pH 7.5), to which 20 g R,S-2,2-dimethyl-1,3-dioxolane-4-methanol were added. After oxidation of 54% of the racemate, the S-enantiomer of R,S-2,2-dimethyl-1,3-dioxolane-4-methanol remained with an enantiomeric excess of 97%.

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Claims

1. Process for the enantioselective conversion of an enantiomer mixture of 2R₁,2R₂-1,3-dioxolane-4-methanol of the formula:

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where R₁ and R₂ independently represent hydrogen or an optionally branched alkyl group, an aryl, or R₁ and R₂, together with the carbon atom to which they are attached, represent an optionally substituted carbocyclic ring, the product of said conversion being enriched in one of the enantiomers, which is achieved by subjecting said enantiomer mixture to the action of an enantioselective enzyme, characterized in that a 2R₁,2R₂-1,3-dioxolane-4-methanol enriched in S-enantiomer is prepared by applying a suitable PQQ-dependent alcohol dehydrogenase as enantioselective enzyme.

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2. Process according to claim 1, characterized in that S-2R₁, 2R₂-1,3-dioxolane-4-methanol is prepared with an enantiomer excess of at least 95%.

3. Process according to either of claims 1 and 2, characterized in that 2,2-dimethyl-1,3-dioxolane-4-

methanol enriched in S-enantiomer is prepared.

4. Process according to any one of claims 1-3, characterized in that as enzyme use is made of a PQQ-dependent alcohol dehydrogenase originating from or obtained from Pseudomonas testosteroni .

5. Process according to claim 4, characterized in that as enzyme use is made of a PQQ-dependent alcohol
5 dehydrogenase originating from or obtained from Pseudomonas testosteroni ATCC 15666, 15667, NCIB 8893, NCIB 8955 and/or NCIB 10808.

6. Process according to any one of claims 1-3, characterized in that as enzyme use is made of a PQQ-dependent alcohol dehydrogenase originating from or obtained from Gluconobacter suboxydans .

7. Process according to claim 6, characterized in that as enzyme use is made of a PQQ-dependent alcohol
10 dehydrogenase originating from or obtained from Gluconobacter suboxydans ATCC 621.

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Absorption (260 n m)

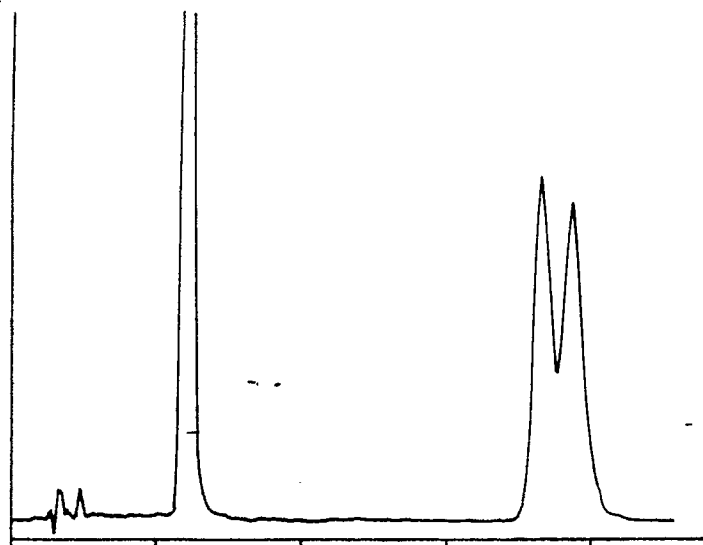


Fig. I.1

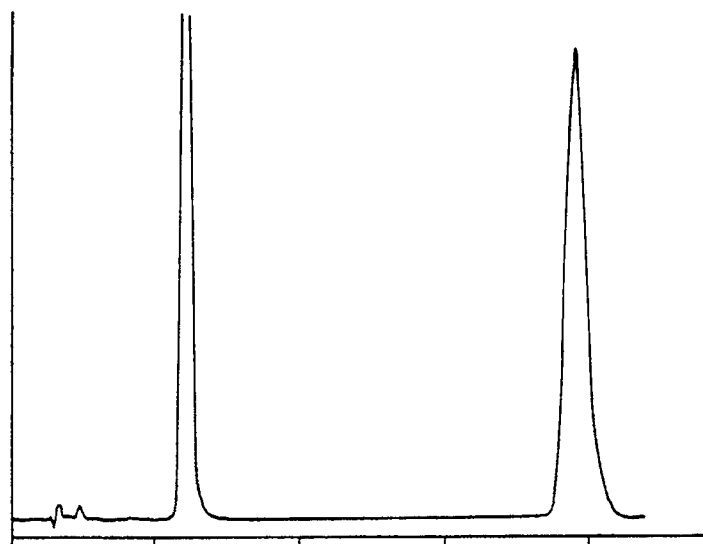


Fig. I.2

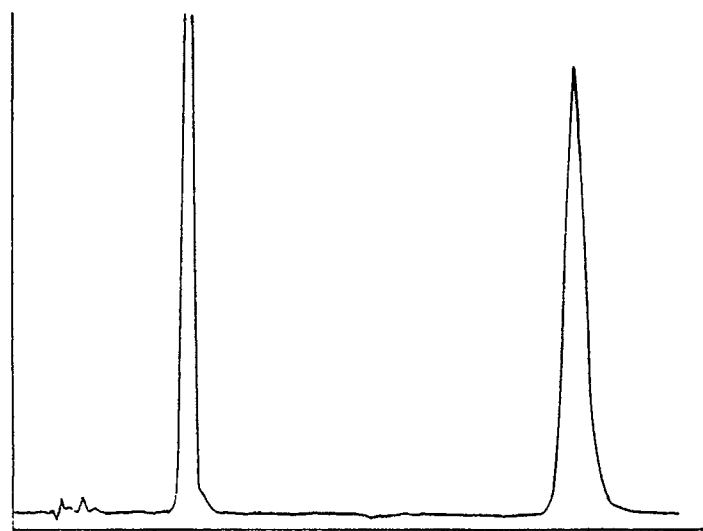


Fig. I.3

0 10 20 30 40
 —————> Time (min.)



European
Patent Office

EUROPEAN SEARCH REPORT

Application Number

EP 90 20 1651

DOCUMENTS CONSIDERED TO BE RELEVANT			
Category	Citation of document with indication, where appropriate, of relevant passages	Relevant to claim	CLASSIFICATION OF THE APPLICATION (Int. Cl.5)
D,A	EP-A-0 244 912 (GIST BROCADES) * Claims * - - -	1	C 12 P 41/00 C 12 P 17/04
A	WO-A-8 503 704 (AMERICAN HOSPITAL SUPPLY) * Claims * - - -	1	
A	CHEMISTRY LETTERS, 1984, pages 401-404, The Chemical Society of Japan; T. MUKAIYAMA et al.: "Highly efficient asymmetric synthesis of (S)-2,2-dimethyl-1,3-dioxolane-4-methanol from glycerol" * Whole document * - - -	1	
A	CHEMICAL ABSTRACTS, vol. 104, no. 21, 26th May 1986, page 256, abstract no. 182157c, Columbus, Ohio, US; B.W. GROEN et al.: "Quinohemoprotein alcohol dehydrogenase apoenzyme from Pseudomonas testosteroni", & BIOCHEM. J. 1986, 234(3), 611-15 - - - - -	1	
The present search report has been drawn up for all claims			TECHNICAL FIELDS SEARCHED (Int. Cl.5) C 12 P
Place of search The Hague		Date of completion of search 23 November 90	Examiner DELANGHE L.L.M.
CATEGORY OF CITED DOCUMENTS X: particularly relevant if taken alone Y: particularly relevant if combined with another document of the same category A: technological background O: non-written disclosure P: intermediate document T: theory or principle underlying the invention E: earlier patent document, but published on, or after the filing date D: document cited in the application L: document cited for other reasons ----- &: member of the same patent family, corresponding document			