

(19)



Europäisches Patentamt  
European Patent Office  
Office européen des brevets



(11)

**EP 0 686 043 B1**

(12)

**EUROPEAN PATENT SPECIFICATION**

(45) Date of publication and mention of the grant of the patent:

**23.06.1999 Bulletin 1999/25**

(21) Application number: **94909760.4**

(22) Date of filing: **23.02.1994**

(51) Int Cl.<sup>6</sup>: **C07H 21/00, A61K 31/70**

(86) International application number:  
**PCT/US94/01913**

(87) International publication number:  
**WO 94/19012 (01.09.1994 Gazette 1994/20)**

(54) **COMPOSITIONS AND METHODS OF APPLICATION OF REACTIVE ANTIVIRAL POLYMERS**

ZUSAMMENSETZUNGEN UND METHODEN ZUR ANWENDUNG VON REAKTIVEN ANTIVIRALEN POLYMEREN

COMPOSITIONS ET PROCEDES D'APPLICATION DE POLYMERES ANTIVIRAUX REACTIFS

(84) Designated Contracting States:  
**AT BE CH DE DK ES FR GB GR IE IT LI LU MC NL  
PT SE**

(30) Priority: **24.02.1993 US 22055**

(43) Date of publication of application:  
**13.12.1995 Bulletin 1995/50**

(73) Proprietor: **WANG, Jui, H.  
Amherst, NY 14226 (US)**

(72) Inventors:  
• **WANG, Jui H.  
Amherst, NY 14226 (US)**  
• **KANG, Insung  
Buffalo, NY 14214 (US)**  
• **RAHAM, Mohammed H.  
Buffalo, NY 14214 (US)**

(74) Representative: **Wablat, Wolfgang, Dr.Dr.  
Patentanwalt,  
Potsdamer Chaussee 48  
14129 Berlin (DE)**

(56) References cited:  
**US-A- 4 966 843                      US-A- 5 091 374**  
**US-A- 5 132 292**

- **NATURE, vol. 357, no. 6373, 7 May 1992 LONDON GB, pages 85-89, E.ARNOLD ET AL. 'Structure of HIV-1 Reverse Transcriptase / DNA Complex at 7A Resolution Showing Active Site Locations.'**
- **JOURNAL OF BIOLOGICAL CHEMISTRY, vol. 263, no. 26, 15 September 1988 MD US, pages 13003-13006, H.CHUAN ET AL. '3'-O-(5-Fluoro-2,4-Dinitrophenyl)ADP Ether and ATP Ether'**

**EP 0 686 043 B1**

Note: Within nine months from the publication of the mention of the grant of the European patent, any person may give notice to the European Patent Office of opposition to the European patent granted. Notice of opposition shall be filed in a written reasoned statement. It shall not be deemed to have been filed until the opposition fee has been paid. (Art. 99(1) European Patent Convention).

**Description**Technical Field

5 **[0001]** The present invention relates to novel compositions useful for, and methods of, treating diseases, such as AIDS, caused by RNA-viruses. The invention also provides a method of making vaccines against such diseases. Further, the composition is usable to inhibit ribonuclease.

Background of Invention

10 **[0002]** In spite of the worldwide effort, the human immunodeficiency virus (HIV) has until now defeated all attempts to develop an effective anti-AIDS drug or vaccine by resorting to hypermutation tactics. Mutation tactics have likewise until now hampered attempts to develop effective vaccines to other RNA-virus caused diseases such as leukemia and influenza.

15 **[0003]** One of the strategies for combating RNA-viruses such as the HIV AIDS (acquired immunodeficiency syndrome) virus is to inactivate the reverse transcriptase of HIV (HIV RT). Unfortunately, the unusually high mutation rate of the virus has hitherto prevented the development of an effective anti-HIV drug or vaccine. The hypermutable HIV can develop resistance rapidly toward all known inhibitors of small molecular weight ( $M_r < 1000$  dalton) such as AZT, ddC, ddI and nevirapine [Larder, et al., Science **243**, 1731 (1989); Larder and Kemp, Science **246**, 1155 (1989); St. Clair, et al., Science **253**, 1557 (1991); Shih, et al., Proc. Natl. Acad. Sci. USA **88**, 9878 (1991)].

20 **[0004]** It is known (Chuan and Wang, J. Biol. Chem. **263**, 13003 (1981)) that the affinity reagents 3'-O-(5-fluoro-2,4-dinitrophenyl) ADP ether and 3'-O-(5-fluoro-2,4-dinitrophenyl) ATP ether are capable of labelling the active site of mitochondrial  $F_1$ -ATPase and of inhibiting the ATPase. However, 3'-O-(5-fluoro-2,4-dinitrophenyl) ADP ether and the corresponding ATP ether require a million-fold higher molar concentration to inhibit the reverse transcriptase of HIV (HIV RT), nor can they inhibit HIV RT or other RNA-viruses in a manner which is mutation-insensitive.

25 **[0005]** It is also known (Fukui and De Clercq, Biochem J **203**, 755-760 and Shannon, Ann. N.Y. Acad. Sci. **284**, 472-507) that anti-viral compounds such as poly(2-fluoro-adenylic acid), poly(2-bromo-adenylic acid), poly(2-iodo-adenylic acid) and 1-(4-fluorobenzoyloxy) adenosine polyadenylic acid, inhibit reverse transcriptase. All of these compounds are derivatized by attaching halogen atoms or other groups to the adenine residues in the polymers. The best of these compounds is  $(fI^2A)_n$  which inhibited murine leukemia RT with an  $IC_{50}$  of 0.04  $\mu$ g/ml. The FDNP-poly[A] of the present invention inhibited the same RT with an  $IC_{50}$  of 0.0017  $\mu$ g/ml (at a 23-fold lower concentration).

Disclosure Of Invention

35 **[0006]** In accordance with an embodiment of the invention a composition of matter is set forth. The composition comprises Y-poly[A] representable by the formula:  $M_n(Y)_m X_i [A]_n$  where:

40  $[A]_n$  = polyadenylic acid (5') with n adenylic acid residues,  
 Y = FDNP, DNP, or partially FDNP and partially DNP,  
 FDNP = 3-fluoro-4,6-dinitrophenyl groups, attached covalently to m of the n 2'-OH groups of  $[A]_n$  via ether linkage,  
 DNP = 2,4-dinitrophenyl groups,  
 attached covalently to m of the n 2'-OH groups of  $[A]_n$  via ether linkage,  
 45 X = an acyl group,  
 i = 0 or 1,  
 M = a cation selected to provide a desired degree of solubility for the composition,

and has one of the generic structures:

50

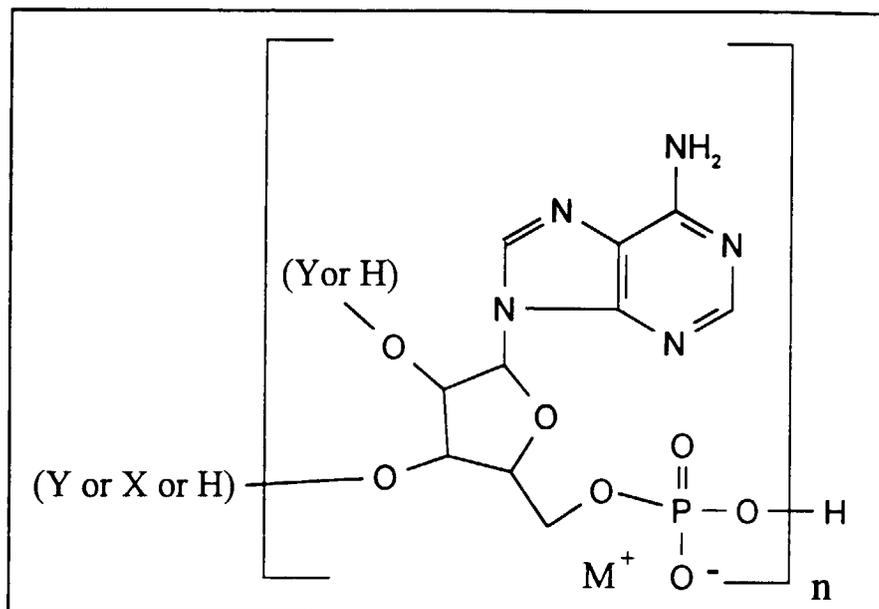
55

5

10

15

20



or

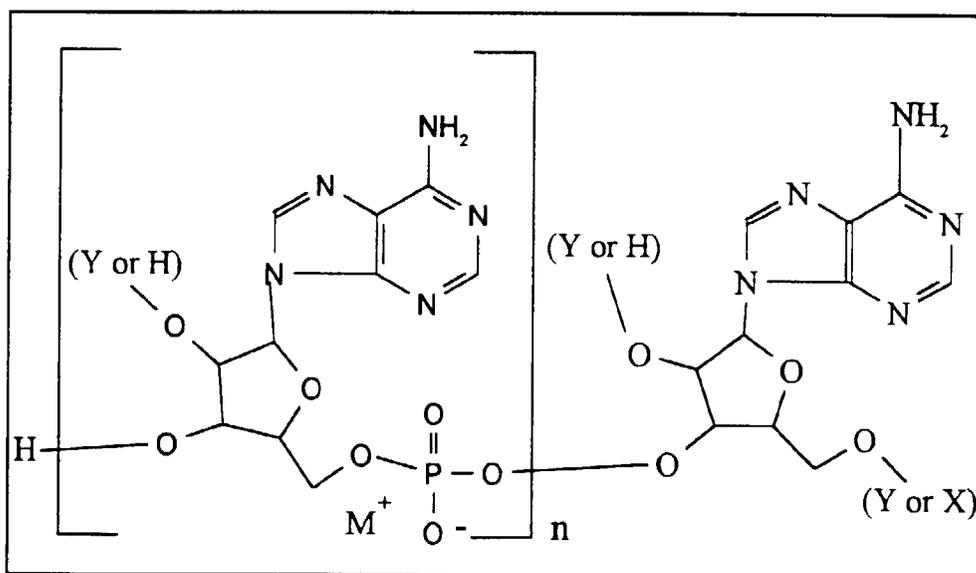
25

30

35

40

45



and having a ratio of Y groups to adenine units of at least about 1:5 and wherein n is sufficiently large so that the Y-poly[A] can substantially completely fill the active site cleft of an RNA-virus RT to effectively inactivate reverse transcriptase and/or can effectively inactivate ribonuclease.

50 **[0007]** Another embodiment of the invention comprises a method of treating a mammal having a disease caused by an RNA-virus. The method comprises the administration to the mammal of an effective treatment amount of Y-poly[A].

**[0008]** Still another embodiment of the invention is a method of temporarily protecting a healthy mammal from an RNA-virus infection. The method comprises administering to the mammal an effective preventative amount of Y-poly[A].

55 **[0009]** Another embodiment yet of the invention comprises methods of preparing anti-RNA-virus vaccines and of vaccinating to provide protection against an RNA-virus caused disease. The method comprises introducing to a patient an antigen comprising an RNA-virus which has been irreversibly inactivated by Y-poly[A] wherein Y is at least partially FDNP, whereby the patient produces one or more antibodies to the antigen, the antibodies to the antigen also comprising antibodies to the RNA-virus.

**[0010]** A further embodiment still of the invention comprises a method of sterilizing an RNA-virus in blood aliquots intended for transfusion. The method comprises adding to the blood aliquots an effective amount for sterilization of Y-poly[A] wherein Y is at least partially FDNP.

**[0011]** One more embodiment of the invention comprises a method of removing ribonuclease from a solution comprising passing the solution through an affinity column containing immobilized FDNP-poly[A] or DNP-poly[A].

**[0012]** Ribonuclease can also be irreversibly inhibited by adding an effective amount of Y-poly[A], wherein Y is at least partially FDNP, to a solution of the ribonuclease.

**[0013]** Another useful method in accordance with an embodiment of the invention is to irreversibly inactivating an RNA-virus. The method comprises contacting the RNA-virus with an effective amount for inactivating the RNA-virus of FDNP-poly[A].

#### Best Mode For Carrying Out Invention

**[0014]** A new type of polymeric inhibitor is set forth herein that binds tightly to and substantially completely fills the long binding cleft at the active site of HIV reverse transcriptase (HIV RT). The new inhibitors were synthesized and were discovered to inactivate HIV RT in solution rapidly, and in the case of the Y-poly[A] having at least a portion of the Y being FDNP irreversibly, and to enable susceptible lymphocytes to continue their normal growth after the addition of live HIV to the culture plate. The inhibitors have DNP- and/or FDNP- groups covalently attached to the ribose residues in the polymer. These inhibitors are function-specific and hence have low toxicity, but are not species-specific and hence are indicated to be mutation-insensitive. The compositions and methods of using these novel compounds in five different pharmaceutical and biotechnological applications are disclosed. A group of mutation-insensitive inhibitors has been developed based on the following rationale.

**[0015]** Recent crystallographic studies show that the polymerase site in HIV RT consists of an openended cleft that is long enough to accommodate a segment of RNA 25 to 30 residues in length [K<i>et al., Science 256, 1783 (1992); Arnold, et al., Nature 357, 85 (1992). Presumably, this active site cleft binds polyadenylic acid (5') (hereinafter referred to as "poly[A]") preferentially, because the complex poly[A]-[dT]<sub>12</sub> has been used successfully and quite often as template-promoter in in vitro reverse transcription experiments. Therefore, hydrophobic groups were attached to poly[A] at its 2'-OH positions to improve the binding affinity to RT without hindering its template function. The resulting poly[A] derivative has the capability of binding to HIV RT by substantially completely filling its active site cleft and acts as a multifunctional affinity reagent.

**[0016]** In view of the long-standing practice of labelling nucleophilic groups of proteins with Sanger's reagent 1-fluoro-2,4-dinitrobenzene, electrophilic 3-fluoro-4,6-dinitrophenyl (FDNP) groups were attached to poly[A] at its 2'-OH positions through ether linkages by a previously published procedure for mononucleotides [Chuan and Wang, J. Biol. Chem. 263, 13003 (1981), the disclosure of which is incorporated herein by reference]. These electrophilic FDNP groups serve to irreversibly react with and to bind to nucleophilic groups in the active site cleft thus blocking the action of reverse transcriptase. The structure of the resulting polymer derivative, designated as FDNP-poly[A], is represented by a structure shown below with general formula:  $M_n(\text{FDNP})_m X_i [A]_n$  where:

$[A]_n$  = polyadenylic acid (5') with n adenylic acid residues,

FDNP = 3-fluoro-4,6-dinitrophenyl groups, attached covalently to m of the n 2'-OH groups of  $[A]_n$  via ether linkage,

X = an acyl group,

i = 0 or 1,

M = a cation selected to provide a desired degree of solubility for the composition,

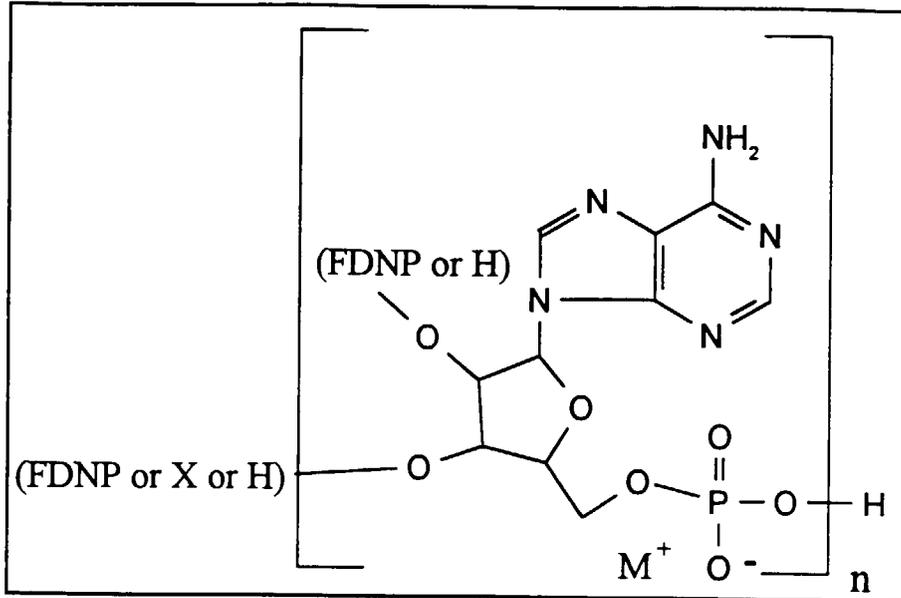
and has one of the generic structures:

5

10

15

20



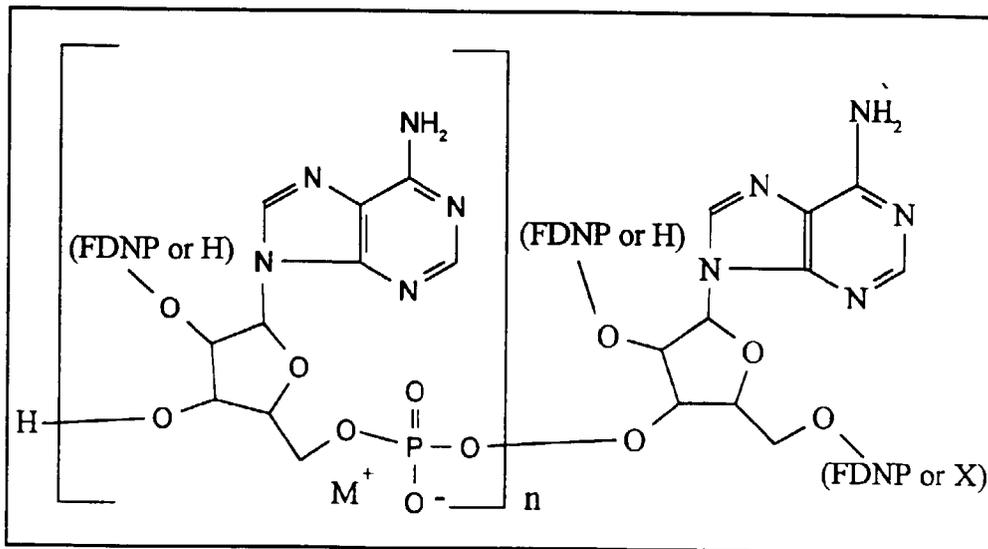
or

25

30

35

40



45

**[0017]** It was further discovered that DNP-poly[A] is also effective in inactivating reverse transcriptases and ribonucleases. Indeed, while the action of DNP-poly[A] is not irreversible it was found that with 1 DNP-group per 1.5 adenine residues, the latter is about twelve times as effective (i.e., is equally effective at one-twelfth the dosage) as is FDNP-poly[A]. The polymer derivative, designated as DNP-poly[A], is represented by a structure shown below with general formula:  $M_n(DNP)_mX_i[A]_n$

50

where:

$[A]_n$  = polyadenylic acid (5') with n adenylic acid residues,

DNP = 2,4-dinitrophenyl groups, attached covalently to m of the n 2'-OH groups of  $[A]_n$  via ether linkage,

X = an acyl group,

55

i = 0 or 1,

M = a cation selected to provide a desired degree of solubility for the composition,

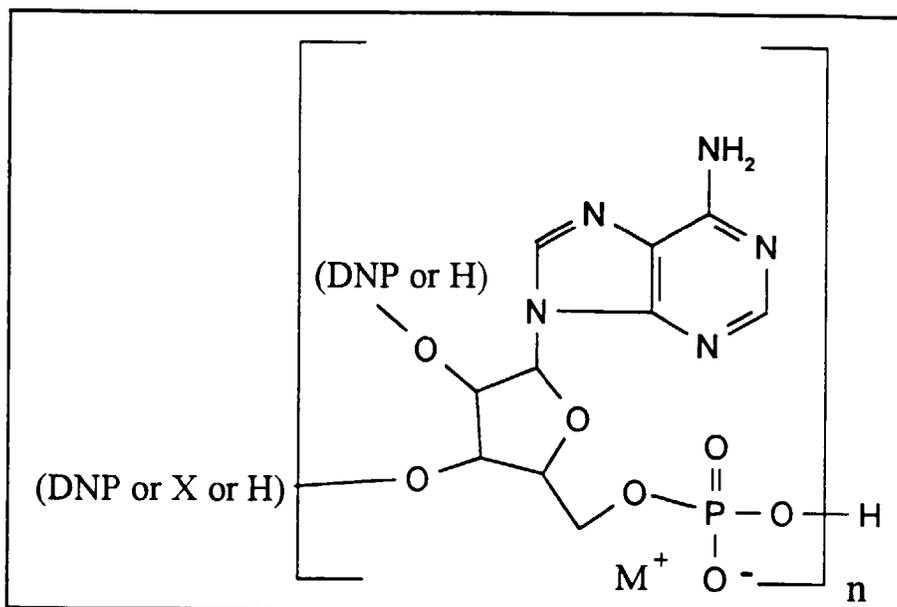
and has one of the generic structures:

5

10

15

20



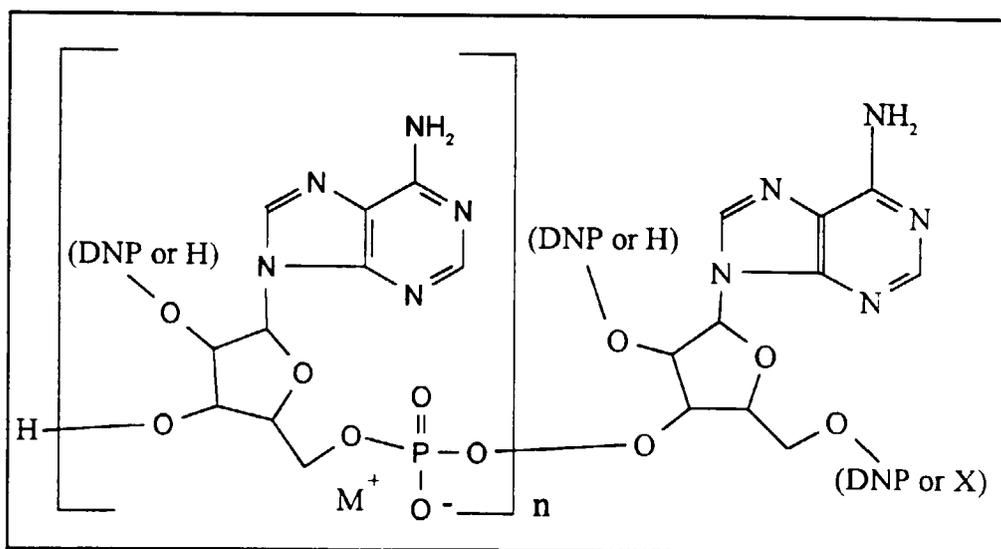
or

25

30

35

40



45 **[0018]** In both cases,  $n$  is sufficiently large so that the Y-poly[A] (where  $Y = \text{DNP}$ , FDNP or where some  $Y$  are DNP and others are FDNP) substantially completely fills the active site cleft of the RNA-virus RT to effectively inactivate the reverse transcriptase and/or can effectively inactivate ribonuclease. Generally it is preferred that  $n$  be relatively large, suitably 20 or greater and preferably 25 or greater. Actual experiments are set forth below for a relatively higher molecular weight Y-poly[A] (h- $M_T$ ),  $n \approx 270$ ,  $m \approx 69$ ,  $M_T \approx 1.1 \times 10^5$  (where  $M_T$  represents molecular weight) and for a relatively lower molecular weight Y-poly[A] (l- $M_T$ ),  $n \approx 28$ ,  $m \approx 9$ ,  $M_T \approx 1.2 \times 10^4$ .

50 **[0019]** FDNP groups are generally attachable to the poly[A] to provide a ratio of FDNP group to [A] of no more than about 1:4. DNP groups are attachable to the poly[A] to provide a considerably higher ratio of about 1:1.5. Mixed FDNP/DNP poly[A] polymers can also be formed. Generally, 1,3-difluoro-2,4-dinitrobenzene (DFDB) will first be reacted with the poly[A] to provide a ratio of FDNP to [A] of from about 1:4 to about 1:10. Then, the product will be reacted with 1-fluoro-2,4-dinitrobenzene (FDNB) so that the resulting composition will have an overall ratio of FDNP plus DNP groups to [A] which is as high as can be attained, generally of greater than 1:4, preferably greater than 1:2.

55 **[0020]** The cation,  $M$ , can be any cation which will provide a sufficient concentration of the composition to accomplish a desired purpose, e.g., inactivate ribonuclease, treat a disease caused by and RNA-virus, produce inactivated (dead)

virus for vaccination, sterilize stored blood, etc. The potassium salt was used in the experiments reported herein but, for example, the ammonium, rubidium, cesium or other salt can be utilized in its place.

[0021] An important factor is the molar ratio of Y groups to [A] (adenine) units. In general, the effectiveness of the individual Y-poly[A] compound increases with increased Y to [A] molar ratio. Accordingly, it is desirable that this ratio be relatively high. Generally, this ratio should be at least about 1:10 with higher ratios, for example 1:5 and above being preferred. The experiments reported herein were carried out with Y-poly[A] compounds having 1:1.5, 1:3.9 and 1:4.8 ratios. Even higher ratios should be desirable.

[0022] We have discovered that at nanomolar concentrations, Y-poly[A] is an effective inhibitor of reverse transcriptases and therefore appears to be an effective anti-RNA-virus treatment (and is also an effective and, when Y is at least partially FDNP, irreversible inhibitor of ribonuclease), but that even at micromolar concentrations it does not inhibit the essential host cell enzymes that were tested *in vitro*, including RNA-polymerase II, cAMP-dependent protein kinase, pyruvate kinase, hexokinase and adenylate kinase. Thus, dosages can be formulated which are effective for irreversibly inhibiting reverse transcriptase in the RNA-viruses but which do not inhibit essential cell enzymes. The association of Y-poly[A] with ribonuclease can be used to inactivate and/or remove ribonuclease from solution. For example, Y-poly[A] wherein at least a portion of Y is FDNP can be added to a solution of ribonuclease to irreversibly inhibit the ribonuclease.

Alternatively and preferably, the Y-poly[A] can be associated with an affinity material, generally with an affinity column, and a solution containing ribonuclease can be passed through the column. The ribonuclease is then held on the column and the solution exiting from the column can be made ribonuclease free. It is preferable when using the column technique that the Y-poly[A] be DNP-poly[A] since the FDNP group of FDNP-poly[A] can be hydrolyzed on the column, whereas a DNP-poly[A] column can be washed and used again. The column material used is not critical. It can be substantially any column material with which the Y-poly[A] can be associated. For example, we have used a cellulose column.

[0023] A long inhibitor molecule such as Y-poly[A] with multiple points of covalent attachment of FDNP groups or non-covalent attachment of DNP groups to many specific amino acid residues at the active site of HIV RT was selected to be a probably mutation-insensitive inhibitor, because it is highly improbable for all these specific amino acid residues to be altered by a single set of random mutations so that the mutant enzyme would no longer be able to bind Y-poly[A] but would still bind the template poly[A]. In order to stay viable, the hypermutable HIV must have a RT that maintains a long active site cleft. It would be very difficult for the viable mutant HIV RT to avoid the binding of such an inhibitor molecule. This inference was confirmed by the observation that in spite of the substantial differences between the primary structures of HIV RT, avian myeloblastosis virus RT (AMV RT) and Moloney murine leukemia virus RT (MLV RT), all three reverse transcriptases were inactivated in 10 minutes or less at 25°C by nanomolar concentrations of FDNP-poly[A].

[0024] Y-poly[A] (h-M<sub>T</sub>) and Y-poly[A] (l-M<sub>T</sub>) were tested at our request as potential anti-HIV drugs *in vitro* with susceptible lymphocytes in the presence of HIV by the Developmental Therapeutics Program of the Division of Cancer Treatment in The National Cancer Institute, (NCI). These reactive polymers were found to be effective in keeping the lymphocytes viable in the presence of HIV (see Example 5 below).

[0025] Since these polymers were designed to inactivate all reverse transcriptases, they can also be utilized for treating other diseases caused by RNA-viruses, e.g., adult T-cell leukemia/lymphoma, hepatitis A, C, D and E, influenza, parainfluenza, infant bronchiolitis and pneumonia, common cold, measles, mumps, etc. Thus, diseases caused by, by way of non-limiting example, the RNA-viruses HTLV (adult T-cell leukemia/lymphoma virus), HAV (Hepatitis A virus), HAC (Hepatitis C virus), HDV (Hepatitis D virus), HAE (Hepatitis E virus), HEV (influenza virus), parainfluenza virus, RSV (respiratory syncytial virus), common cold causing coronavirus and rhinovirus, measles virus and mumps virus can be controlled by use of the present invention. Other RNA-virus caused diseases can likewise be controlled by use of the present invention.

[0026] The invention will be better understood by reference to the following examples.

#### EXAMPLE 1

##### Preparation Of Y-poly[A](h-M<sub>T</sub>)

##### Procedure 1:

[0027] Ten milligrams of the potassium salt of Poly[A] (obtained from Sigma, M<sub>T</sub> ~ 1.1 x 10<sup>5</sup>) were dissolved in 0.5 mL water and 1.6 mL of 0.2 M K<sub>2</sub>CO<sub>3</sub> + 1.5 M KHCO<sub>3</sub> solution (pH 9.2). The stirred solution was mixed with 0.80 mL of an acetone solution containing a tenfold excess of 1,3-difluoro-2,4-dinitrobenzene (DFDB) which was added in three equal aliquots at 12-hour intervals. After the reaction had progressed for 48 hours at room temperature, the excess DFDB was removed by repeated extraction with a 7:1 (v/v) mixture of methylene chloride and dimethylsulfoxide. The

resulting aqueous solution was centrifugally gel-filtered through Sephadex G-25-80 and subsequently lyophilized. The molar ratio of 3-fluoro-4,6-dinitrophenyl to adenine groups in the product was calculated from the observed absorbance at 330 and 259 nm respectively to be 1/3.9, using 3-fluoro-4,6-dinitrophenyl ethyl ether ( $\epsilon_{259} = 3900$ ,  $\epsilon_{330} = 6300$ ) and AMP ( $\epsilon_{259} = 15400$ ) as the standards. Thus the average polymeric inhibitor had 270 adenine residues and 69 2'-O-FDNP groups per molecule. The absorbance of a dilute solution of the FDNP-poly[A] (h-M<sub>T</sub>) did not decrease significantly (<5%) after its pH was lowered from 9 to 5. This observation shows that the fraction of 3-fluoro-4,6-dinitrophenyl groups in the product which had been hydrolyzed to the non-reactive 3-hydroxy-4,6-dinitrophenyl groups during the preparation was less than 5%. Yield of product = 3 mg.

#### Procedure 2:

**[0028]** Ten milligrams of the potassium salt of Poly[A] (obtained from Sigma, M<sub>T</sub> ~ 1.1 x 10<sup>5</sup>) were dissolved in 0.5 mL water and 0.1 mL of 0.1 M K<sub>2</sub>CO<sub>3</sub> + 2.0 M KHCO<sub>3</sub> solution (pH 8.8). The stirred solution was mixed with 0.4 mL of an acetone solution containing a five-fold excess of 1,3-difluoro-2,4-dinitrobenzene (DFDB) which was added in four aliquots at 6-hour intervals. During the reaction the pH of the reaction mixture was checked and readjusted to 8.8 by adding K<sub>2</sub>CO<sub>3</sub> and KHCO<sub>3</sub>. The progress of the reaction was monitored by TLC analysis of the reaction mixture as a function of time on a Kodak Cellulose Plate with fluorescent indicator, using (20 mM K<sub>2</sub>HPO<sub>4</sub> + 20 mM KH<sub>2</sub>PO<sub>4</sub>): CH<sub>3</sub>CN = 2:1 (v/v) as the developing solvent. Eventually all of a band at R<sub>f</sub>=0.38 from poly A faded and a new FDNP-poly A band at R<sub>f</sub>=0.77 grew in intensity. After the reaction had progressed until the band at R<sub>f</sub>=0.38 had disappeared, usually after about 24 hours at room temperature, the excess DFDB was removed by repeated extraction with a 7:1 (v/v) mixture of methylene chloride and dimethyl sulfoxide. The resulting aqueous solution was centrifugally gel-filtered through Sephadex G-50-80 or dialyzed and subsequently lyophilized. The molar ratio of 3-fluoro-4,6-dinitrophenyl to adenine groups in the product was calculated from the observed absorbance at 330 and 259 nm respectively to be 1/4.8, using 3-fluoro-4,6-dinitrophenyl ethyl ether ( $\epsilon_{259} = 3900$ ,  $\epsilon_{330} = 6300$ ) and AMP ( $\epsilon_{259} = 15400$ ) as the standards. The FDNP/adenine ratio of 1/4.8 was confirmed by <sup>19</sup>F-NMR and by <sup>31</sup>P-NMR.

**[0029]** While Procedure 1 yielded a product having a desirably higher ratio than did Procedure 2 of FDNP groups to adenine groups it was found that after the 48 hour reaction time about 5% of the 5-fluoro groups had hydrolyzed into OH groups. Hydrolysis was not found to have occurred when the reaction time was limited to 24 hours. The hydrolyzed material was not separable from the product. Accordingly, Procedure 2 is preferred.

**[0030]** DNP-poly A was synthesized similarly to Procedure 2 with 1-fluoro-2,4-dinitrobenzene (FDNB) replacing DFDB. In the synthesis of DNP-poly A, poly A was allowed to react with excess FDNB for 48 hours at room temperature, and the DNP-group to adenine residue molar ratio in the product DNP-poly A was 1/1.5. This ratio was higher than that obtained with the FDNP-poly[A] since in that instance the reaction was terminated after 48 hours to avoid appreciable hydrolysis of the 5-fluoro group.

#### EXAMPLE 2

##### Preparation of FDNP-poly[A] (l-M<sub>T</sub>)

**[0031]** A mixture of ADP and AMP at 28:1 molar ratio was prepared by dissolving 867 mg ADP (K-salt), 25 mg AMP (free acid) and 393 mg K<sub>2</sub>CO<sub>3</sub> in 4 mL water. Its pH was changed from 8.2 to 9.05 by successive additions of solid K<sub>2</sub>CO<sub>3</sub>. The solution was then mixed with 20 μL of 1 M MgCl<sub>2</sub> and 8 units of polynucleotide phosphorylase (obtained from Sigma; derived from E. coli). After the reaction mixture was incubated at 37°C for 48 hours, it was centrifugally gel-filtered through Sephadex G-25-80 to remove all ADP, AMP and oligonucleotides of molecular weight below 5000. Measurement of absorbance at 259 nm showed that the filtrate contained 63 mg of product FDNP-poly[A] (l-M<sub>T</sub>). A second gel-filtration of an aliquot of this FDNP-poly[A] (l-M<sub>T</sub>) solution through Sephadex G-50-80 showed that 99.7% of the oligomers made this way have M<sub>T</sub> < 10000. This FDNP- [A]poly(l-M<sub>T</sub>) was subsequently used to react with FDNB in a similar way as described above to form FDNP-poly[A] (l-M<sub>T</sub>). Molar ratios of FDNP-groups/adenine residues = 1/3.9.

**[0032]** FDNP-poly[A] (l-M<sub>T</sub>) was also synthesized following Procedure 2 above and yielded a product having an FDNP/adenine molar ratio of 1/4.8. DNP-[A]poly(l-M<sub>T</sub>) was prepared similarly and had a DNP/adenine molar ratio of 1/1.5.

#### EXAMPLE 3

##### Irreversible Inhibition of Viral Reverse Transcriptases in Solution by FDNP-poly[A]

**[0033]** Reverse transcriptase samples from HIV (HIV RT) and from Moloney murine leukemia virus (MLV RT) and from avian myeloblastosis virus (AMV RT) were separately incubated with FDNP-poly[A] at 25°C in a Tris-HCl buffer

## EP 0 686 043 B1

(0.1 M, pH 8.2) containing 1 mM MgCl<sub>2</sub> and 125 mM KCl and measured for RT activity. In each experiment, the selected enzyme was incubated for 10 minutes at 25°C in an incubation mixture containing the potassium salt of the inhibitor at a given concentration. After ten minutes the incubated mixture was rapidly injected into an assay solution and further incubated for 10 min at 37°C, then precipitated with trichloroacetic acid, washed, and assayed for the radioactive polynucleotide formed by scintillation counting. The assay mixture contained 125 mM Tris-HCl (pH 8.2), 1 mM MgCl<sub>2</sub>, 125 mM KCl, 250 μM [<sup>3</sup>H]dTTP as the substrate and 23 nM Poly[A] - [dT]<sub>12</sub> as the template.

**[0034]** For each reverse transcriptase the observed percentage of enzyme activity left after 10 minutes preincubation with FDNP-poly[A] at 25°C was plotted as a function of inhibitor concentration, and the concentration of the inhibitor required for 50% inhibition of the enzyme (IC<sub>50</sub>) was determined from the graph. The results are summarized below. Similar results were obtained with DNP-poly[A].

Enzyme	Inhibitor	IC <sub>50</sub> (nM)
HIV RT	FDNP - poly [A] (h-M <sub>T</sub> )	6 pM
HIV RT	FDNP-poly [A] (l-M <sub>T</sub> )	400 nM
MLV RT	FDNP - poly [A] (h-M <sub>T</sub> )	15 pM
AMV RT	FDNP-poly [A] (h-M <sub>T</sub> )	12 pM

**[0035]** The observed IC<sub>50</sub> values show that DNP-poly[A](h-M<sub>T</sub>), and FDNP-poly[A](h-M<sub>T</sub>) and FDNP-poly[A] (l-M<sub>T</sub>) are powerful inhibitors of reverse transcriptase in aqueous solution. Kinetic measurements indicated that both DNP-poly[A] and FDNP-poly[A] compete with the template-primer polyadenylic acid-dodecathymidylic acid (poly[A][dT]<sub>12</sub>) for the same binding site in the HIV RT molecule. Loading the incubated mixture onto a cellulose-oligo[dt] column and subsequent separation by elution chromatography showed that FDNP-poly[A] binds to reverse transcriptase covalently. Therefore its inactivation of the enzyme is irreversible.

**[0036]** The above data also show that in spite of their substantial differences in primary structure, the three viral reverse transcriptases, HIV RT, MLV RT and AMV RT were all inactivated in ten minutes at 25°C by subnanomolar concentrations of FDNP-poly[A]. This is a strong indication that FDNP-poly[A] is mutation-insensitive.

### EXAMPLE 4

#### Inhibition of Different HIV Reverse Transcriptases DNP-poly[A] and FDNP-poly[A]

**[0037]** The inhibition of different mutant HIV RT's by DNP-poly[A] and FDNP-poly[A] was kindly measured gratis by Dr. Joe C. Wu with the RT from four different strains of the human virus: HIV-1 RT (wild type), HIV-2 (wild type), HIV-1<sub>41,215</sub> RT (AZT-resistant mutant, Thr 215 → Tyr, Met 41 → Leu), HIV-1<sub>181c</sub> RT (Nevirapine-resistant mutant), using poly C-[dG]<sub>12-18</sub> as the template and [<sup>3</sup>H]dGTP as the substrate. The assay conditions are the same as in Example 3. The observed approximate IC<sub>50</sub> values are summarized in the Table below:

Inhibitor	IC <sub>50</sub> of the RT (nM)			
	HIV-1 (wild)	HIV-2 (wild)	HIV-1 <sub>41,215</sub>	HIV-1 <sub>181c</sub>
DNP-poly[A]	3.8 ± 0.5	1.6 ± 0.2	2.2 ± 0.3	0.34 ± 0.07
Poly[A]	-----	-----	-----	-----
FDNP-poly[A]	1.5 ± 0.6	1.1 ± 0.2	1.2 ± 0.3	0.65 ± 0.11

**[0038]** It was found in these experiments that both DNP-poly[A] and FDNP-poly[A] are potent inhibitors of all four reverse transcriptases but that poly[A] itself has no inhibitory effect.

### EXAMPLE 5

#### Protection of Susceptible Lymphocytes From HIV by DNP- and FDNP-poly[A] (h-M<sub>T</sub>) and by FDNP-poly[A] (l-M<sub>T</sub>)

**[0039]** FDNP-poly[A] (h-M<sub>T</sub>), DNP-poly[A] (h-M<sub>T</sub>) and FDNP-poly[A] (l-M<sub>T</sub>) were tested as potential anti-AIDS drugs in vitro with susceptible lymphocytes in the presence of HIV by the Developmental Therapeutics Program of Division of Cancer Treatment in the National Cancer Institute (NCI). All three inhibitors were confirmed as being active. The

NCI protocol involves mixing various concentrations of FDNP-poly[A] (h-M<sub>T</sub>), DNP-poly[A] (h-M<sub>T</sub>) or FDNP-poly[A] (l-M<sub>T</sub>) and susceptible T4 lymphocytes (CEM-SS cell line) with or without HIV in microculture plates, incubating the plates for six days, then determining the number of remaining viable cells using a colorimetric endpoint [method described in Weislow, et al., *J. Natl. Cancer Inst.* 81, 577-586 (1989)].

**[0040]** The test results show that FDNP-poly[A] and DNP-poly[A] are both effective in keeping the lymphocytes viable in the presence of HIV. The observed effective concentrations (EC<sub>50</sub>) are 2.65 ± 0.15 µg/mL or 24 ± 2 nM for FDNP-poly[A] (h-M<sub>T</sub>) and 0.2 ± 0.1 µg/mL or 2 ± 1 nM for DNP-poly[A] [h-M<sub>T</sub>]. Therefore these polymers are potent antiviral agents. The soluble salt of either polymer can be used directly to treat mammals infected by HIV either in an appropriate solution for parenteral administration or in the form of pellets to be taken orally.

EXAMPLE 6

Stability of Y-poly[A] (h-M<sub>T</sub>) in 0.01 M Hydrochloric Acid and in the Presence of Pepsin at 37°C

**[0041]** Human stomach juice contains pepsin and approximately 0.01 M HCl. To simulate the acid conditions inside human stomach, we incubated FDNP-poly[A] in 0.01 M HCl at 37°C, and found that its potency as an irreversible inhibitor of HIV RT decreased only 50% after 4 hours of incubation in 0.01 M HCl. DNP-poly[A] was found to be even more stable under the same conditions. The results for FDNP-poly[A] appear in the following table.

Incubation at 37°C(h)	0	1/2	1	2	4
IC <sub>50</sub> (nM)	27	22	22	21	34

**[0042]** In a separate experiment, FDNP-poly[A] (h-M<sub>T</sub>) (34 nM) was incubated at 37°C in a 0.01 M HCl solution containing 1.0 mg/ml of pepsin. It was found that the potency of the inhibitor was unaffected after 2 hours but decreased to 65% after 4 hours of incubation. These observations suggest that the potency of FDNP-poly[A] against HIV will not be destroyed by stomach juice in 2 hours. Consequently, the solid salt of FDNP-poly[A] may be mixed with proper filler and pressed into pellets or packed in capsules for oral administration.

EXAMPLE 7

Stability of FDNP-poly[A] in Human Blood Serum at 37°C

**[0043]** A frozen sample of normal human serum (Male, Type AB, from Sigma) was thawed and used to dissolve FDNP-poly[A] (h-M<sub>T</sub>) (34 nM). The relative potency of this inhibitor of HIV RT was determined as a function of incubation time at 37°C and listed below:

Incubation time (h)	0	0.5	1.0	2.0	4.0
Relative potency (%)	100	95	91	93	80

**[0044]** Since the potency of FDNP-poly[A] in human serum did not decrease more than 20% in 4 hours at 37°C, the compound can be dissolved in an appropriate solution for parenteral administration.

EXAMPLE 8

Total and Irreversible Inactivation of HIV by FDNP-poly[A] for the Development of Effective Anti-HIV Vaccine

**[0045]** The HIV that has been completely and irreversibly inactivated by FDNP-poly[A] (or Y-poly[A] wherein Y is partially FDNP) is also useful as a complete antigen for the development of an effective anti-AIDS-vaccine.

**[0046]** The world-wide efforts to find an effective vaccine to prevent AIDS have so far been unsuccessful. Vaccines developed with genetically engineered pieces of the AIDS virus do not provide sufficient protection against AIDS mainly because of the extremely high mutation rate of the virus. Recently, encouraging protective effects of a live attenuated HIV vaccine with a deletion in the nef gene has been reported [Daniel, et al., *Science* 258, 1938 (1992)]. This new discovery reminded many researchers of the old practice of using a weakened version of the entire virus to trigger the protective immune response. For example, the live virus can be weakened by treatment with formaldehyde and the weakened virus can then be used as the antigen to trigger immune response. But this is a dangerous procedure because the formation of Schiff base between aldehyde and amino groups is reversible. A reversal of the condensation

reaction would regenerate the fully active virus, which may be fatal to the patient.

**[0047]** Since the inactivation of HIV by Y-poly[A] wherein Y is at least partially FDNP can be 100% effective and irreversible, a truly effective anti-AIDS vaccine is developable for humans by using Y-poly[A] (wherein Y is at least partially FDNP) - inactivated HIV as the antigen and introducing it, e.g., injecting it, into a patient.

#### EXAMPLE 9

##### Sterilization of Possible Traces of Live HIV in Stored Transfusion Blood

**[0048]** The possible presence of even a trace of live HIV in donated blood poses a constant threat to people who receive blood transfusions as well as to people who administer blood transfusions. Very small but adequate amount of Y-poly[A] (wherein Y is at least partially FDNP) can be added to donated blood before storage in blood banks. Since for irreversible inhibitors  $IC_{50}$  decreases with time [Kang and Wang, *J. Biol. Chem.*, in press, May 1994], the added Y-poly[A] inactivates traces of HIV that might be present during storage when used as a safety measure.

**[0049]** In order to test the effectiveness of Y-poly[A], HIV RT (57 nM) was incubated with different concentrations of the inhibitor (FDNP-poly[A] (h-M<sub>7</sub>)) for 10 minutes at 25°C. The percentages of initial enzyme activity left after the incubations are listed below:

Inhibitor concentration (nM)	Enzyme activity (%)
0	100
1.6	89
4.8	72
16.7	54
25	42
33	29
42	15
50	6
55	2
60	0

#### EXAMPLE 10

##### Irreversible Inhibition of Ribonuclease of FDNP-poly[A]

**[0050]** In the biotechnology industry and related research laboratories it is often necessary to isolate large pieces of undamaged RNA. In order to avoid incidental cleavage of the RNA product by traces of exogenous ribonuclease, it is often necessary to preboil all the solution with diethyl pyrocarbonate or to add a ribonuclease inhibitor, generally the commercially available RNasin (obtainable from Promega). Both are expensive procedures for large-scale operation. We discovered that Y-poly[A] (wherein Y is at least partially FDNP) is a potent irreversible inhibitor of ribonuclease. The addition of a small but adequate amount of such Y-poly[A] irreversibly inactivates any trace amount of ribonuclease which might be present in the solutions and containers used in the process. The effectiveness of such Y-poly[A] as a potent inhibitor of ribonuclease is shown by the following experiment.

**[0051]** The catalytic activity of ribonuclease A (RNase A) was determined by monitoring the decrease of absorbance at 300 nm ( $A_{300}$ ) with time due to the catalyzed hydrolysis of cytidine 2':3'-cyclic monophosphate (cCMP) in solution at 25°C. The assay solution contained 1.9 mM cCMP (Na-salt) in 50 mM acetate buffer (pH 5.0). The control reaction in the absence of FDNP-poly[A] was started by injecting 10  $\mu$ l of 54.4  $\mu$ M RNase A stock solution into 2.000 ml of the assay solution, mixing and monitoring the decrease of  $A_{300}$  with time. For inhibition studies the RNase was preincubated with various concentrations of FDNP-poly[A] (h-M<sub>7</sub>) for 1.0 min, and then injected into the cCMP solution to start the assay. The results are summarized below. The total final concentration of RNase was kept constant at 268 nM, the inhibitor concentration at 54  $\mu$ M.

Total preincubation concentration of FDNP-poly[A] ( $\mu$ M)	0	0.5	0.8	1.3	1.7	2.3
% initial RNase activity remaining at 1 minute after mixing	100	89	73	48	25	0

These data show that even at submicromolar concentrations each FDNP-poly[A] molecule can rapidly inactivate many

molecules of exogenous RNase.

#### EXAMPLE 11

##### 5 Effectiveness of Double Helical DNP-Poly A-Poly[dT]

[0052] Double helical DNP-poly A-poly[dT] has been found to be a more potent inhibitor of HIV-1 RT than is DNP-poly A. The double helical DNP-poly[A]poly[dT] was formulated by mixing the two components and heating for 5 minutes at 90°C.

10 [0053] The data listed in the following table were obtained by using poly C-[dG]<sub>12-18</sub> as the template-promoter and [<sup>3</sup>H]TTP as the substrate of HIV-1 RT.

Inhibitor concentration	5 nM	10 nM	20 nM
% inhibition by DNP-poly[A]	44	81	94
% inhibition by DNP-poly[A]-poly[dT]	93	98	100

Clearly the double helical DNP-poly[A]-poly[dT] is a much better inhibitor of HIV-1 RT than is the single-stranded DNP-poly[A].

#### EXAMPLE 12

##### Spontaneous Transport of DNP-poly[A] into Human Cells

25 [0054] A 100- $\mu$ l sample of human adult lymphocytes ( $2 \times 10^6$  cells) was mixed with 100  $\mu$ l RPMI (Roswell Park Memorial Institute) culture medium and 60  $\mu$ l of DNP[<sup>14</sup>C]poly[A] (1.3 mg/ml, with specific radioactivity of 120 cpm/pmole). The mixture was incubated at 37°C and 50  $\mu$ l aliquots were taken out at certain time intervals, filtered, washed and assayed for radioactivity. Each aliquot was spotted on a piece of membrane filter (FA 1.0  $\mu$ m, Millipore), suction-filtered and washed 8 times with aqueous 100 mM KCl solution. Each dried filter with cells was assayed for radioactivity

30 by a liquid scintillation counting system. The data from a typical experiment are summarized below.

Incubation time (h)	0.1	0.5	3	24	48
Counts per min (cpm)	800	900	1300	1800	2200

35 These data show that by either diffusion or endocytosis the hydrophobic DNP[<sup>14</sup>C]poly[A] can be spontaneously transported into human adult lymphocytes. Under the same conditions [<sup>14</sup>C]poly[A] did not get into these lymphocytes at all.

#### EXAMPLE 13

##### Spontaneous Transport of DNP-poly[A] into Murine Leukemia Virus

40 [0055] Murine Moloney leukemia virus and DNP-poly[A] were used in a model experiment for demonstrating the spontaneous transport of the inhibitor into a retrovirus. In each experiment, 5  $\mu$ l of the virus suspension was incubated on ice with 37.5 nM DNP-poly[A]. After a certain time, the mixture was centrifuged for 10 min at 100,000 g. The supernatant was carefully removed and the pellet (practically invisible) was washed with 200  $\mu$ l Tris buffer (50 mM Tris-HCl, pH 7.8). The pellet was then homogenized in the assay buffer (50 mM Tris-HCl, pH 7.8, 60 mM KCl, 2 mM DTT and 0.6 mM Mn-acetate) and incubated with 0.02 U poly[A]-oligo[dT], 1.5 mM [<sup>3</sup>H]dTTP and 0.1% Triton-100 for 60 min at 37°C. The reaction was stopped with 10% TCA and the catalytic activity (A) of the reverse transcriptase was calculated

50 from the cpm of the precipitated radioactive product [<sup>3</sup>H]poly[dT]. The experimental data is summarized in the following table where A<sup>o</sup> represent the initial catalytic activity before incubation with the inhibitor.

Incubation time (h)	0	0.16	0.5	1.0	24	48
A/A <sup>o</sup>	1	0.54	0.50	0.45	0.25	0.16

55 These data show that DNP-poly[A] can penetrate the envelop of murine leukemia virus and inhibit the reverse transcriptase inside. They indicate that FDNP-poly[A] can also penetrate the envelop of HIV and produce a permanently

inactivated HIV suitable for use as an ideal antigen for an anti-AIDS vaccine. Under these same conditions [<sup>3</sup>H]poly [dT] did not get into the virus at all.

Industrial Applicability

5 [0056] The present invention provides a composition of matter which is useful in the treatment of RNA-virus caused diseases such as AIDS, and in the sterilization of HIV in stored transfusion blood, and which can provide vaccines against RNA-virus caused diseases.

10 [0057] While the invention has been described in connection with specific embodiments thereof, it will be understood that it is capable of further modification, and this application is intended to cover any variations, uses, or adaptations of the invention, as may be applied to the essential features hereinbefore set forth, and as fall within the scope of the invention and the limits of the appended claims.

15 **Claims**

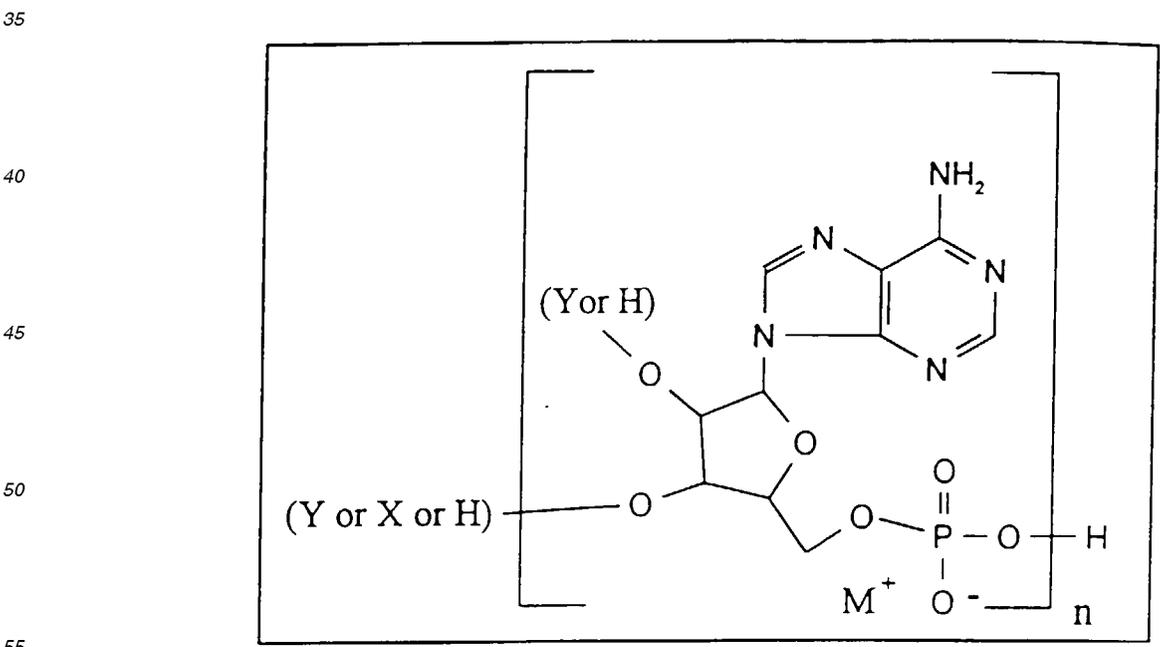
1. A composition of matter comprising Y-poly[A] representable by the formula:

20 
$$M_n(Y)_m X_i [A]_n$$

where

- 25 [A]<sub>n</sub> = polyadenylic acid (5') with n adenylic acid residues,  
 Y = FDNP, DNP or partially FDNP and partially DNP,  
 FDNP = 3-fluoro-4,6-dinitrophenyl groups, attached covalently to m of the n 2'-OH groups of [A]<sub>n</sub> via ether linkage,  
 DNP = 2,4-dinitrophenyl groups, attached covalently to m of the n 2'-OH groups of [A]<sub>n</sub> via ether linkage  
 X = an acyl group,  
 30 i = 0 or 1,  
 M = a cation selected to provide a desired degree of solubility for the composition,

and has one of the generic structures:



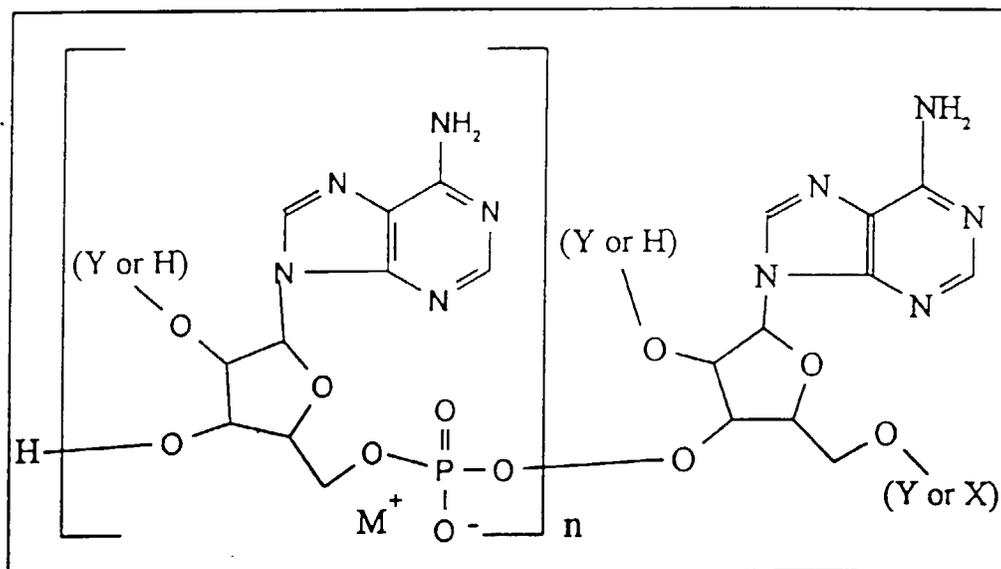
or

5

10

15

20



25

and having a ratio of Y groups to adenine units of at least about 1:5 and wherein n is sufficiently large so that the Y-poly[A] can substantially completely fill the active site cleft of an RNA-virus RT to effectively inactivate reverse transcriptase and/or can effectively inactivate ribonuclease.

30

2. A composition of matter as set forth in claim 1, wherein n is at least about 20.
3. Use of a composition of matter according to claim 1 or 2 for the preparation of pharmaceutical compositions for the treatment of a disease caused by an RNA-virus.
4. Use according to claim 3, wherein the RNA-virus is HIV, HTLV, HAV, HCV, influenza-viruses, parainfluenza viruses, RSV, coronavirus or rhinovirus, measles virus or mumps virus and the disease is, respectively, acquired immuno-deficiency syndrome (AIDS), adult T-cell leukemia/lymphoma, hepatitis A, hepatitis C, influenza, parainfluenza, infant bronchiolitis and infant pneumonia, common cold, measles or mumps.
5. Use of a composition of matter according to claim 1 or 2 for the preparation of pharmaceutical compositions for the temporarily protection from a RNA-virus infection.
6. Use according to claim 5, wherein the RNA-virus infection is caused by HIV, HTLV, HAV, HCV, influenza-viruses, parainfluenza viruses, RSV, coronavirus or rhinovirus, measles virus or mumps virus and the disease is, respectively, acquired immuno-deficiency syndrome (AIDS), adult T-cell leukemia/lymphoma, hepatitis A, hepatitis C, influenza, parainfluenza, infant bronchiolitis and infant pneumonia, common cold, measles or mumps.
7. Use of composition of matter according to claim 1 or 2 for the preparation of pharmaceutical compositions for vaccinating against an anti-RNA-virus caused disease, and where the vaccinating is performed by:

50

introducing to a patient an antigen comprising an RNA-virus which has been inactivated by Y-poly [A] wherein Y is at least partially FDNP whereby the patient produces one or more antibodies to the antigen, the antibodies to the antigen also comprising antibodies to the RNA-virus.

55

8. Use according to claim 7 wherein the RNA-virus is HIV, HTLV, HAV, HCV, influenza-viruses, parainfluenza viruses, RSV, coronavirus or rhinovirus, measles virus or mumps virus and the disease is, respectively, acquired immuno-deficiency syndrome (AIDS), adult T-cell leukemia/lymphoma, hepatitis A, hepatitis C, influenza, parainfluenza, infant bronchiolitis and infant pneumonia, common cold, measles or mumps.
9. A method of sterilizing an RNA-virus in blood aliquots intended for transfusion comprising adding to the blood aliquots an effective amount for sterilization of Y-poly [A] wherein Y is at least partially FDNP.

10. A method as set forth in claim 9, wherein the RNA-virus is HIV and the disease is acquired immuno-deficiency syndrome (AIDS).

5 11. A method as set forth in claim 9, wherein the RNA-virus is HIV, HTLV, HAV, HCV, influenza-viruses, parainfluenza viruses, RSV, coronavirus or rhinovirus, measles virus or mumps virus and the disease is, respectively, acquired immuno-deficiency syndrome (AIDS), adult T-cell leukemia/lymphoma, hepatitis A, hepatitis C, influenza, parainfluenza, infant bronchiolitis and infant pneumonia, common cold, measles or mumps.

10 12. A method of irreversibly inactivating ribonuclease in solution comprising adding to the solution an effective amount for inactivating the ribonuclease of Y-poly [A] wherein Y is at least partially FDNP.

15 13. A method of removing ribonuclease from a solution containing ribonuclease comprising contacting the solution with an affinity material containing immobilized FDNP-poly[A] or DNP-poly[A] and then separating the solution from the affinity material.

14. A method of irreversibly inactivating an RNA-virus comprising contacting the RNA-virus with an effective amount for inactivating the RNA-virus of Y-poly[A] wherein Y is at least partially FDNP.

20 **Patentansprüche**

1. Y-Poly[A]-enthaltende Stoffzusammensetzung gekennzeichnet durch die Formel:

25 
$$M_n(Y)_m X_i [A]_n$$

worin

- 30 [A]<sub>n</sub> = Polyadenylsäure (5') mit n Adenylsäureresten,  
 Y = FDNP, DNP oder partiell FDNP und partiell DNP,  
 FDNP = 3-Fluor-4,6-dinitrophenylgruppen, über Etherbrücken kovalent mit m der n 2'-OH-Gruppen von [A]<sub>n</sub> verknüpft,  
 DNP = 2,4-Dinitrophenylgruppen, über Etherbrücken kovalent mit m der n 2'-OH-Gruppen von [A]<sub>n</sub> verknüpft,  
 X = Acylgruppe,  
 35 i = 0 oder 1,  
 M = ein Kation ist, ausgesucht um einen gewünschten Löslichkeitsgrad der Zusammensetzung zu erreichen,

und eine dieser generischen Strukturen besitzt:

40

45

50

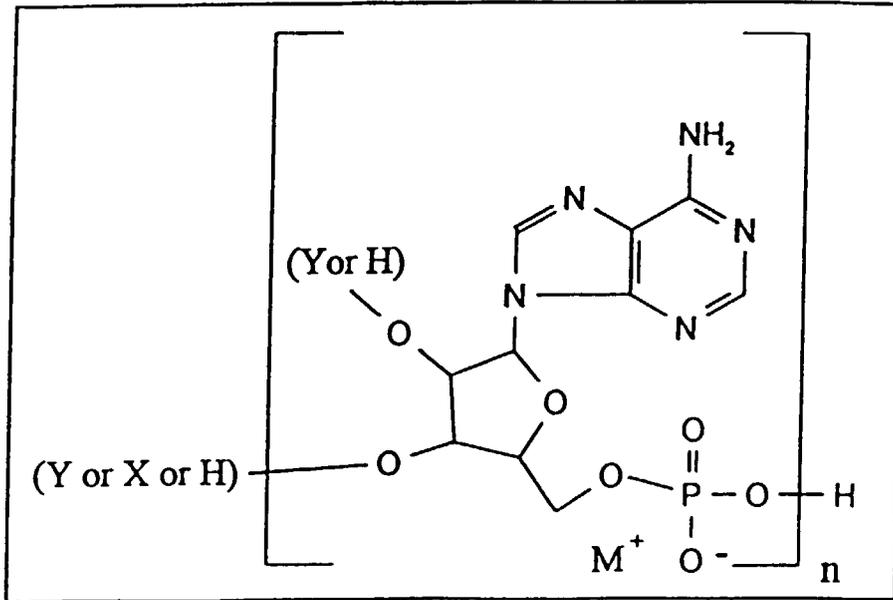
55

5

10

15

20



oder

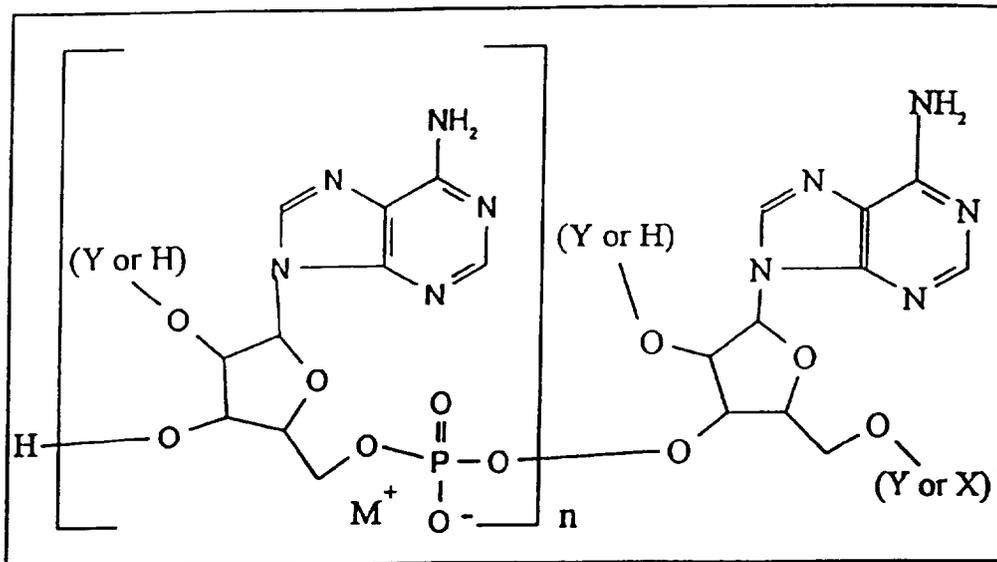
25

30

35

40

45



und worin das Verhältnis von Y-Gruppen zu Adenineinheiten wenigstens 1:5 beträgt und worin n hinreichend groß ist, daß das y-Poly[A] im wesentlichen vollständig das aktive Zentrum eines RNA-Virus RT ausfüllen kann, um wirkungsvoll die reverse Transcriptase und/oder wirkungsvoll die Ribonuclease zu inaktivieren.

50

2. Stoffzusammensetzung nach Anspruch 1, dadurch gekennzeichnet, daß n wenigstens 20 beträgt.
3. Verwendung der Stoffzusammensetzung nach Anspruch 1 oder 2 zur Herstellung pharmazeutischer Zusammensetzungen zur Behandlung von Krankheiten verursacht durch RNA-Viren.
4. Verwendung nach Anspruch 3, dadurch gekennzeichnet, daß das RNA-Virus ein HIV, HTLV, HAV, HCV, Influenzaviren, Parainfluenzaviren, RSV, Coronavirus oder Rhinovirus, Masern-Virus oder Mumps-Virus ist und die Erkrankung Immunschwächesyndrom (AIDS), T-Zellen-Leukämie/Lymphoma bei Erwachsenen, Hepatitis A, Hepa-

55

titis C, Influenza, Parainfluenza, Säuglings-Bronchiolitis und Säuglings-Lungenentzündung, Erkältung, Masern oder Mumps ist.

5 5. Verwendung der Stoffzusammensetzung nach Anspruch 1 oder 2 zur Herstellung pharmazeutischer Zusammensetzungen zum zeitweiligen Schutz vor einer RNA-Virusinfektion.

10 6. Verwendung nach Anspruch 5, dadurch gekennzeichnet, daß die RNA-Virusinfektion durch HIV, HTLV, HAV, HCV, Influenzaviren, Parainfluenzaviren, RSV, Coronavirus oder Rhinovirus, Masern-Virus oder Mumps-Virus verursacht wurde, und die Erkrankung Immunschwächesyndrom (AIDS), T-Zellen-Leukämie/Lymphoma bei Erwachsenen, Hepatitis A, Hepatitis C, Influenza, Parainfluenza, Säuglings-Bronchiolitis und Säuglings-Lungenentzündung, Erkältung, Masern oder Mumps ist.

15 7. Verwendung der Stoffzusammensetzung nach Anspruch 1 oder 2 zur Herstellung pharmazeutischer Zusammensetzungen zur Impfung gegen Erkrankungen verursacht durch Anti-RNA-Viren, wobei die Impfung wie folgt durchgeführt wird:

20 der Patient erhält eine Injektion eines Antigens, welches einen RNA-Virus enthält, der durch Y-Poly[A] inaktiviert wurde, wobei Y wenigstens partiell FDNP bedeutet, wodurch der Patient einen oder mehrere Antikörper gegen das Antigen bildet, wobei die Antikörper zum Antigen auch Antikörper zum RNA-Virus enthalten.

25 8. Verwendung nach Anspruch 7, dadurch gekennzeichnet, daß das RNA-Virus ein HIV, HTLV, HAV, HCV, Influenzaviren, Parainfluenzaviren, RSV, Coronavirus oder Rhinovirus, Masern-Virus oder Mumps-Virus ist und die Erkrankung Immunschwächesyndrom (AIDS), T-Zellen-Leukämie/Lymphoma bei Erwachsenen, Hepatitis A, Hepatitis C, Influenza, Parainfluenza, Säuglings-Bronchiolitis und Säuglings-Lungenentzündung, Erkältung, Masern oder Mumps ist.

30 9. Verfahren zur Sterilisierung von RNA-Viren in zur Transfusion vorgesehener Blut-Aliquots, dadurch gekennzeichnet, daß zu den Blut-Aliquots eine für die Sterilisierung wirkungsvolle Menge von Y-Poly[A] hinzugefügt wird, wobei Y wenigstens partiell FDNP bedeutet.

35 10. Verfahren nach Anspruch 9, dadurch gekennzeichnet, daß das RNA-Virus HIV ist und die Erkrankung Immunschwächesyndrom (AIDS) ist.

40 11. Verfahren nach Anspruch 9, dadurch gekennzeichnet, daß das RNA-Virus ein HIV, HTLV, HAV, HCV, Influenzaviren, Parainfluenzaviren, RSV, Coronavirus oder Rhinovirus, Masern-Virus oder Mumps-Virus ist und die Erkrankung Immunschwächesyndrom (AIDS), T-Zellen-Leukämie/Lymphoma bei Erwachsenen, Hepatitis A, Hepatitis C, Influenza, Parainfluenza, Säuglings-Bronchiolitis und Säuglings-Lungenentzündung, Erkältung, Masern oder Mumps ist.

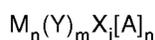
45 12. Verfahren zur irreversiblen Inaktivierung der Ribonuclease in Lösung, dadurch gekennzeichnet, daß zur Lösung eine zur Inaktivierung der Ribonuclease wirkungsvolle Menge von Y-Poly[A] hinzugefügt wird, wobei Y wenigstens partiell FDNP bedeutet.

50 13. Verfahren zur Entfernung von Ribonuclease aus einer Ribonuclease-enthaltenden Lösung, dadurch gekennzeichnet, daß die Lösung mit einem affinen Material, welches immobilisiertes FDNP-Poly[A] oder DNP-Poly[A] enthält, in Kontakt gebracht wird, und danach die Lösung vom affinen Material getrennt wird.

55 14. Verfahren zur irreversiblen Inaktivierung eines RNA-Virus, dadurch gekennzeichnet, daß das RNA-Virus mit einer zur Inaktivierung des RNA-Virus wirkungsvollen Menge von Y-Poly[A] in Kontakt gebracht wird, wobei Y wenigstens partiell FDNP bedeutet.

**Revendications**

55 1. Composition comprenant Y-poly[A] pouvant être représenté par la formule :



dans laquelle

[A]<sub>n</sub> = acide polyadénylique (5') avec n résidus d'acide adénylique,

Y = FDNP, DNP ou partiellement FDNP ou partiellement DNP,

5 FDNP = groupes 3-fluoro-4,6-dinitrophényle, attachés de manière covalente à m des n groupes 2'-OH de [A]<sub>n</sub> via une liaison éther,

DNP = groupes 2,4-dinitrophényle, attachés de manière covalente à m des n groupes 2'-OH de [A]<sub>n</sub> via une liaison éther,

X = un groupe acyle,

10 i = 0 ou 1,

M = un cation choisi pour fournir un degré désiré de solubilité à la composition,

et a une des structures génériques :

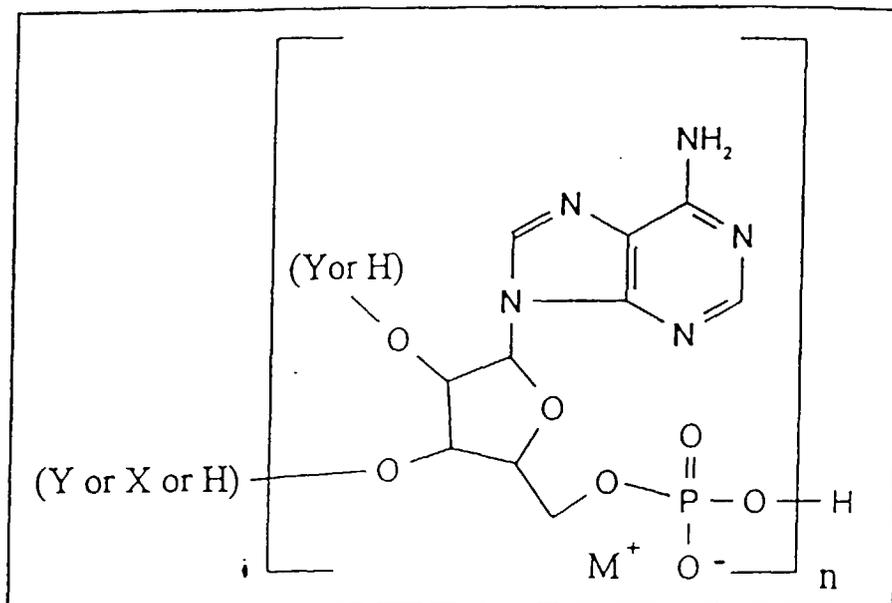
15

20

25

30

35



ou

40

45

50

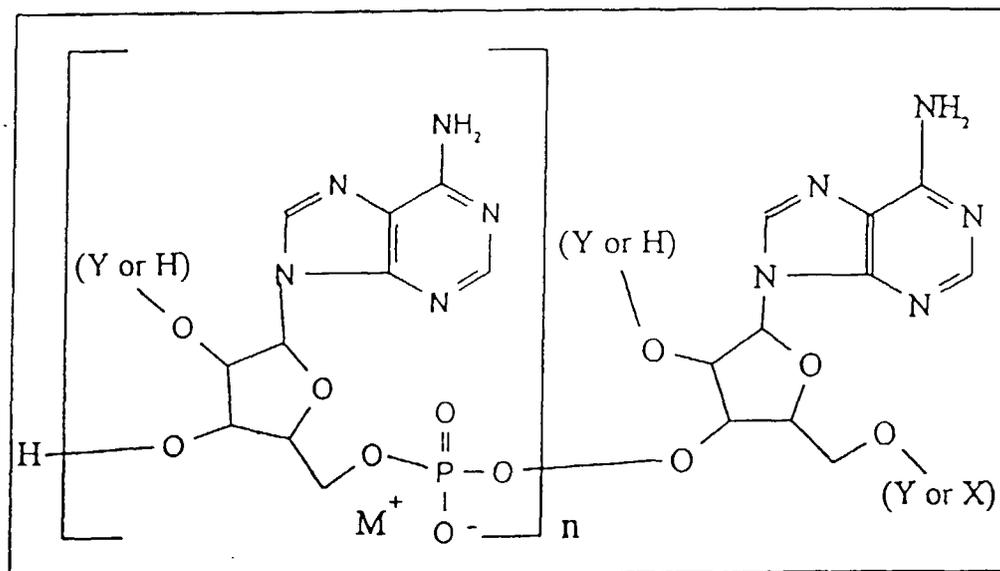
55

5

10

15

20



25

et ayant un rapport des groupes Y sur les unités adénine d'au moins environ 1:5 et dans laquelle n est suffisamment grand de manière à ce que le Y-poly[A] puisse essentiellement remplir complètement la crevasse du site actif d'une transcriptase inverse (RT) d'un virus à ARN pour inactiver de manière efficace la transcriptase inverse et/ou puisse inactiver de manière efficace une ribonucléase.

30

2. Composition selon la revendication 1 dans laquelle n est au moins environ 20.
3. Utilisation d'une composition selon la revendication 1 ou 2 pour la préparation de compositions pharmaceutiques pour le traitement d'une maladie causée par un virus à ARN.
4. Utilisation selon la revendication 3 dans laquelle le virus à ARN est le VIH, le HTLV, le HAV, le HCV, les virus de la grippe, les virus para-influenza, le RSV, le coronavirus ou le rhinovirus, le virus de la rougeole ou le virus des oreillons et la maladie est, respectivement le syndrome de l'immuno-déficience acquise (SIDA), les leucémies/lymphomes à cellules T chez l'adulte, l'hépatite A, l'hépatite C, la grippe, la para-influenza, la bronchiolite infantile et la pneumonie infantile, le rhume commun, la rougeole ou les oreillons.
5. Utilisation d'une composition selon la revendication 1 ou 2 pour la préparation de compositions pharmaceutiques pour la protection temporaire contre une infection par un virus à ARN.
6. Utilisation selon la revendication 5, dans laquelle l'infection par le virus à ARN est causée par le VIH, le HTLV, le HAV, le HCV, les virus de la grippe, les virus para-influenza, le RSV, le coronavirus ou le rhinovirus, le virus de la rougeole ou le virus des oreillons et la maladie est, respectivement le syndrome de l'immuno-déficience acquise (SIDA), les leucémies/lymphomes à cellules T chez l'adulte, l'hépatite A, l'hépatite C, la grippe, la para-influenza, la bronchiolite infantile et la pneumonie infantile, le rhume commun, la rougeole ou les oreillons.
7. Utilisation d'une composition selon la revendication 1 ou 2 pour la préparation de compositions pharmaceutiques pour la vaccination contre une maladie causée par un virus à ARN et dans laquelle la vaccination est effectuée par :
 

l'introduction à un patient d'un antigène comprenant un virus à ARN qui a été inactivé par le Y-poly[A], Y étant au moins partiellement FDNP ce en quoi le patient produit un ou plusieurs anticorps contre l'antigène, les anticorps contre l'antigène comprenant aussi des anticorps contre le virus à ARN.
8. Utilisation selon la revendication 7, dans laquelle le virus à ARN est le VIH, le HTLV, le HAV, le HCV, les virus de la grippe, les virus para-influenza, le RCV, le coronavirus ou le rhinovirus, le virus de la rougeole ou le virus des oreillons et la maladie est, respectivement le syndrome de l'immuno-déficience acquise (SIDA), les leucémies/lymphomes à cellules T chez l'adulte, l'hépatite A, l'hépatite C, la grippe, la para-influenza, la bronchiolite infantile et la pneumonie infantile, le rhume commun, la rougeole ou les oreillons.

50

55

## EP 0 686 043 B1

9. Procédé de stérilisation d'un virus à ARN dans des fractions aliquotes de sang destinées à la transfusion comprenant l'ajout aux fractions aliquotes de sang d'une quantité efficace pour la stérilisation de Y-poly[A], Y étant au moins partiellement FNNDP.

5 10. Procédé selon la revendication 9, dans lequel le virus à ARN est le VIH et la maladie est le syndrome de l'immuno-déficience acquise (SIDA).

10 11. Procédé selon la revendication 9, dans lequel le virus à ARN est le VIH, le HTLV, le HAV, le HCV, les virus de la grippe, les virus parainfluenza, le RCV, le coronavirus ou le rhinovirus, le virus de la rougeole ou le virus des oreillons et la maladie est, respectivement le syndrome de l'immuno-déficience acquise (SIDA), les leucémies/lymphomes à cellules T chez l'adulte, l'hépatite A, l'hépatite C, la grippe, la para-influenza, la bronchiolite infantile et la pneumonie infantile, le rhume commun, la rougeole ou les oreillons.

15 12. Procédé pour inactiver de manière irréversible une ribonucléase en solution comprenant l'ajout à la solution d'une quantité efficace pour inactiver la ribonucléase de Y-poly[A], Y étant au moins partiellement FNNDP.

20 13. Procédé pour éliminer une ribonucléase d'une solution contenant une ribonucléase comprenant l'étape consistant à mettre en contact la solution avec un matériau affiné contenant le FNNDP-poly[A] ou le DNP-poly[A] immobilisé et puis séparer la solution à partir du matériau affiné.

25 14. Procédé pour inactiver de manière irréversible un virus à ARN comprenant l'étape consistant à mettre en contact le virus à ARN avec une quantité efficace pour l'inactivation du virus à ARN de Y-poly[A], Y étant au moins partiellement FNNDP.

30

35

40

45

50

55