Office européen des brevets

(12)

EUROPEAN PATENT APPLICATION

(43) Date of publication: 25.09.1996 Bulletin 1996/39

(21) Application number: 95115423.6

(22) Date of filing: 29.09.1995

(51) Int. Cl.⁶: **C12N 15/54**, C12N 9/10, C12N 15/70, C12N 1/21

(84) Designated Contracting States: BE CH DE FR GB IT LI SE

(30) Priority: 14.02.1995 JP 25253/95

(71) Applicant: TOYOTA JIDOSHA KABUSHIKI KAISHA Aichi-ken (JP)

(72) Inventors:

 Ayumi, Koike, c/o Toyota Jidosha K.K. Toyota-shi, Aichi-ken (JP)

 Tokuzo, Nishino, c/o Toyota Jidosha K.K.
 Toyota-shi, Aichi-ken (JP)

 Shusei, Obata, c/o Toyota Jidosha K.K. Toyota-shi, Aichi-ken (JP)

 Shinichi, Ohnuma, c/o Toyota Jidosha K.K. Toyota-shi, Aichi-ken (JP) Takeshi, Nakazawa, c/o Toyota Jidosha K.K. Toyota-shi, Aichi-ken (JP)

(11)

 Kyozo, Ogura, c/o Toyota Jidosha K.K. Toyota-shi, Aichi-ken (JP)

 Tanetoshi, Koyama, c/o Toyota Jidosha K.K. Toyota-shi, Aichi-ken (JP)

(74) Representative: Tiedtke, Harro, Dipl.-Ing. Patentanwaltsbüro
Tiedtke-Bühling-Kinne & Partner
Bavariaring 4
80336 München (DE)

Remarks:

The applicant has subsequently filed a sequence listing and declared, that it includes no new matter.

(54) Mutated farnesyldiphosphate synthase capable of synthesizing geranylgeranyldiphosphate and gene coding therefor

(57) A mutated farnesyldiphosphate synthase capable of synthesizing geranylgeranyldiphosphate and gene coding for said mutated enzyme, wherein the mutated enzyme is modified from a native farnesyldiphosphate synthase by mutation of a gene coding for a native farnesyldiphosphate synthase.

Description

5

10

20

35

45

BACKGROUND OF INVENTION

Field of Invention

The present invention relates to the mutated farnesyldiphosphate synthase capable of synthesizing geranylgeranyldiphosphate and a process for production thereof, as well as genes coding for said mutated enzymes and a process for isolation thereof.

2. Related Art

In nature there are various isoprenoid chain compounds comprising 5 carbon atom-basic structure, isoprene units, and these isoprenoid compounds play important roles for the life of various organisms. It is known that the chain-extension mechanism is catalyzed by a series of prenyltransferases which catalyze a series of catalytic reactions comprising sequential condensation of isopentenyldiphosphate (IPP) having 5 carbon atoms with its isomer dimethylallyldiphosphate (DMAPP). Among the isoprenoid compounds, farnesyldiphosphate (FPP) having 15 carbon atoms is positioned at a branching point in a biosynthesis pathway, from which various physiologically important start to geranylgeranyldiphosphate (GGPP) having 20 carbon atoms, to quinones, squalene, to steroids, farnesylated protein, dolichol etc.

Different prenyltransferases synthesize different isoprenoid compounds having different lengths. However, prenyl-transferases have a common activity to condense an isoprenoid unit to extend the chain, and in fact, amino acids essential for the condensation are being clarified on the basis of homology of amino acid sequences of different prenyl-transferases. However, the mechanism which determines the length of the isoprenoid compound have not yet clarified.

A biosynthesis pathway for geranyldiphosphate (GPP), farnesyldiphosphate (FPP) and geranylgeranyldiphosphate (GGPP) starting from an isoprenoid unit is shown in Fig. 1. In this biosynthesis pathway, the prenyltransferase which synthesizes farnesyldiphosphate is designated "farnesyldiphosphate synthase", and the prenyltransferase which synthesizes geranylgeranyldiphosphate is designated "geranylgeranyldiphosphate synthase".

Farnesyldiphosphate synthases are known in <u>Bacillus thermophils</u> (J. Biochem. <u>113</u>, 355 - 363 (1993)), <u>E. coli</u> (J. Biochem. <u>108</u>, 995 - 1000 (1990)), yeast (J.B.C. <u>265</u>, 19176 - 19184 (1989)), rats (Mol. Cell. Biol. <u>7</u>, 3138 - 3146 (1987)) and in humans (J.B.C. <u>265</u>, 4607 - 4616 (1990)), and their amino acid sequences are also known.

On the other hand, geranylgeranyldiphosphate synthases are known in <u>Rhodopseudomonas capusulata</u> (J. Bacteriol. <u>154</u>, 580 - 590 (1983)), <u>Erwinia uredovora</u> (J. Bacteriol. <u>172</u>, 6704 - 6712 (1990)), <u>Sulfolobus acidocaldarius</u> (J.B.C. <u>269</u>, 14792 - 14797 (1994)) etc.

However, it had not been known that an enzyme having geranylgeranyldiphosphate synthase activity can be obtained by mutation of farnesyldiphosphate synthase.

SUMMARY OF INVENTION

Accordingly, the present invention provides a novel geranylgeranyldiphosphate synthase obtainable by mutating a farnesyldiphosphate synthase and a process for production thereof, as well as gene system therefor and a process for isolation of the gene.

More specifically, the present invention provides a process for production of a gene coding for geranylgeranyldiphosphate synthase comprising the steps of:

- (1) subjecting genes coding for a farnesyldiphosphate synthase to a mutagenesis;
- (2) expressing the genes subjected to the mutagenesis, and
- (3) selecting a gene which provides a geranylgeranyldiphosphate synthase.

The present invention further provides a gene coding for geranylgeranyldiphosphate synthase, an expression vector containing said gene, and a host transformed with said vector.

The present invention also provides a process for production of geranylgeranyldiphosphate synthase comprising expressing said gene, and geranylgeranyldiphosphate synthase obtainable by said process.

From another point of view, the present invention provides a geranylgeranyldiphosphate synthase having an amino acid sequence modified from an amino acid sequence of native farnesyldiphosphate synthase wherein the modification is deletion of one or more amino acids, addition of one or more amino acids, and/or replacement of one or more amino acids with other amino acids.

The present invention still further provides a gene coding for the above-mentioned geranylgeranyldiphosphate synthase, a vector, especially an expression vector comprising said gene, and a host transformed with said vector.

The present invention further provides a process for production of geranylgeranyldiphosphate synthase comprising the steps of cultivation said host, and purification the geranylgeranyldiphosphate synthase from the culture.

The present invention further provides a process for production of geranylgeranyldiphosphate or geranylgeranyol, comprising the steps of acting the present geranylgeranyldiphosphate synthase on isopentenyldiphosphate, dimethylallyldiphosphate, geranyldiphosphate or farnesyldiphosphate as a substrate.

BRIEF EXPLANATION OF DRAWINGS

5

10

20

30

35

Figure 1 represents a biosynthesis pathway for farnesyldiphosphate and geranylgeranyldiphosphate.

Fig. 2 shows the homology of amino acid sequences of farnesyldiphosphate synthase derived from different species. In this Figure, the sequences in the boxes A to E show regions having relatively high homology and which are expected to participate in enzyme activity.

Fig. 3 shows the homology of amino acid sequences of farnesyldiphosphate synthase derived from different species. In this Figure, the sequences in the boxes F and G show regions having relatively high homology and which are expected to participate in enzyme activity.

Fig. 4 shows a native amino acid sequence of farnesyldiphosphate synthase derived from <u>Bacillus</u> stearothermophilus (indicated as W.T), and the mutated points in amino acid sequences of the modified enzymes having geranyldiphosphate synthase activity (No. 1 to No. 4).

Fig. 5 schematically shows a process for construction of the present modified gene.

Fig. 6 is a profile of reversed phase TLC (developer: acetone/water = 9/1) showing products formed by acting the present enzyme on a substrate dimethylallyldiphosphate.

Fig. 7 is a profile of a reversed-phase TLC (developer: acetone/water = 9/1) showing products formed by acting the present enzyme on a substrate geranyldiphosphate.

Fig. 8 is a profile of a reversed phase TLC (developer: acetone/water = 9/1) showing products formed by acting the present enzyme on a substrate (all-E)-farnesyldiphosphate.

Fig. 9 is a profile of a reversed phase TLC (developer: acetone/water = 9/1) showing products formed by acting the present enzyme on a substrate (all-E)-farnesyldiphosphate.

DETAILED DESCRIPTION

Genes of the present invention can be obtained by subjecting a gene coding for a farnesyldiphosphate synthase to mutagenesis, expressing the genes subjected to the mutagenesis, and selecting a gene providing a protein having geranylgeranyldiphosphate synthase activity.

Genes coding for a farnesyldiphosphate synthase used in the present invention may be those of any origin. For example, farnesyldiphosphate synthases of <u>E. coli</u>, yeast, human, rat etc., as well as genes coding therefor are known, and amino acid sequences of these enzymes have high homology as shown in Fig. 2. Therefore, in addition to the gene derived from <u>Bacillus stearothermophilus</u> as described in detail, according to the present invention, any gene coding for an amino acid sequence having a high homology, for example, at least 20% homology with the amino acid sequence of farnesyldiphosphate synthase derived from <u>Bacillus stearothermophilus</u> can be used regardless of its origin. As such gene sources, for example, <u>Bacillus stearothermophilus</u>, <u>E. coli</u>, yeast, humans, rats etc. can be used.

The gene to be mutated is an RNA or DNA coding for a farnesyldiphosphate synthase and sensitive to treatment with a mutagen, and DNA is preferably used for to ease of handling, and especially a single-stranded DNA is preferred due to its high mutation ratio.

A single-stranded DNA can be easily prepared according to a conventional Procedure for preparing a single-stranded DNA, for example, by inserting a double-stranded DNA into a phage, introducing the phage into <u>E. coli</u> cells, culturing the <u>E. coli</u> cells and recovering the phage from the resulting lysate solution; or by introducing a desired double-stranded DNA into host cells, infecting the host cells with helper phage, culturing the host cells and recovering the phage from the resulting lysate solution.

Mutation of a gene can be carried out according to a conventional procedure for artificially mutating a gene. The mutation methods can be a physical method such as irradiation with X-rays, ultraviolet rays, etc., a chemical method such as treatment with a mutagen, a method of cis incorporation by DNA polymerase, a method using synthetic oligonucleotides etc. A chemical method is preferable for ease of operation and a high mutation ratio. As a mutagen, a nitrite, such as sodium nitrite, or the like can be used. To mutate a single-stranded DNA, a nitrite is preferable. Mutagenesis is preferably carried out at a nitrite concentration of 0.01 to 2M, for example, at about 0.1 to 1M, at a temperature of 20 to 30°C, for 10 to 120 minutes.

To select a gene coding for a protein having geranylgeranyldiphosphate synthase activity from the genes subjected to the mutagenesis, the gene subjected to the mutagenesis is inserted in an expression vector, the vector is introduced into host cells, the enzyme is expressed, and the expression product is tested for geranylgeranyldiphosphate synthase

activity. Geranylgeranyldiphosphate is converted to phytoene by a phytoene synthase, and the phytoene is converted to lycopene having red color by a phytoene desaturase.

Accordingly, for example, a gene coding for a phytoene synthase and a gene coding for phytoene desaturase are inserted into an expression vector, the vector is introduced into host cells such as <u>E. coli</u> cells, and further an expression plasmid comprising a DNA to be tested is introduced into said host cells, and the double transformed host cells are cultured. If the gene to be tested encodes a geranylgeranyldiphosphate synthase, and the geranylgeranyldiphosphate produced by the gene expression is converted to phytoene and further to lycopene, the cells are red-colored. Accordingly, a desired gene can be selected very easily and efficiently by selecting a red-colored colony.

The present invention provides a protein having geranylgeranyldiphosphate synthase activity, i.e., a geranylgeranyldiphosphate synthase, having an amino acid sequence modified from a native amino acid sequence of a farnesyldiphosphate synthase. Here, the modification of an amino acid sequence means replacement of one or a few amino acids with other amino acids, deletion of one or a few amino acids or addition of one or a few amino acids, or a combination of these modifications. The amino acid replacement is especially preferable. Regarding the number of amino acids to be modified, "a few amino acids" means usually about 15 amino acids, preferably about 10 amino acids, and more preferably about 5 amino acids. Namely, according to the present invention, the number of mutated amino acids is about 1 to 15, preferably about 1 to 10, and more preferably 1 to 5.

To determine the positions of modified amino acids, after the mutagenesis and the selection of a gene coding for a geranylgeranyldiphosphate synthase, a nucleotide sequence of the selected gene is determined, and an amino acid sequence is predicted from the determined nucleotide sequence, the predicted amino acid sequence of the modified enzyme is composed with the corresponding native amino acid sequence. Amino acid sequences thus determined of the modified enzymes are shown in Fig. 4.

In Fig. 4, the row indicated by the symbol W.T shows, by the one-letter expression, a native amino acid sequence of farnesyldiphosphate synthase of <u>Bacillus stearothermophilus</u> origin, and the rows Nos. 1 to 4 show representative amino acid sequences which acquired geranylgeranyldiphosphate synthase activity by amino acid replacement in the amino acid sequence of the farnesyldiphosphate synthase, wherein only the amino acids different from the corresponding amino acids in the native amino acid sequence of the farnesyldiphosphate synthase shown in the line T.W are indicated by the one-letter expression of amino acid.

The modified enzyme No. 1 has two mutations, i.e., the 81st position ($Tyr \rightarrow His$) and 275th position ($Leu \rightarrow Ser$); the modified enzyme No. 2 has two mutations, i.e., 34th position ($Leu \rightarrow Val$) and 59th position ($Arg \rightarrow Gln$); the modified enzyme No. 3 has two mutations, i.e., 157th position ($Val \rightarrow Ala$) and 182nd position ($His \rightarrow Tyr$); and the modified enzyme No. 4 has three mutations, i.e., 81st position ($Tyr \rightarrow His$), 238th position ($Pro \rightarrow Arg$) and 265th position ($Ala \rightarrow Thr$). The amino acid sequences No. 1 to 4 of the above-mentioned modified enzymes and nucleotide sequences coding therefor are shown in SEQ ID NO: 1 to 4, and the native amino acid sequence and a nucleotide sequence coding therefor is shown in SEQ ID NO: 5.

30

35

In the present invention, the amino acid sequence farnesyldiphosphate synthase of <u>Bacillus stearothermophilus</u> origin was used as a specific example. However, as shown in Figs. 2 and 3, farnesyldiphosphate synthases have high homology among a wide spectrum of species covering those derived from the eukaryotes including humans and those derived from prokaryotes including bacteria. Therefore, the present invention can be applied to enzymes derived from various species to obtain novel geranylgeranyldiphosphate synthase.

As shown in Fig. 4, amino acid modification such as replacement occurs on the 34th, 59th, 81st, 157th, 182nd, 239th, 265th, and/or 275th positions of farnesyldiphosphate of <u>Bacillus stearothermophilus</u>. For enzymes from other species, it is expected that replacement at positions corresponding to the above-mentioned positions of the farnesyldiphosphate synthase of <u>Bacillus stearothermophilus</u> origin provides similar effects as that for the modified enzyme derived from <u>Bacillus stearothermophilus</u>. Therefore, the present invention can be applied to any farnesyldiphosphate synthases.

The present invention also relates to genes coding for the various geranylgeranyldiphosphate synthases derived from a farnesyldiphosphate synthase. These genes can be obtained by mutation of a gene coding for a corresponding native amino acid sequence. In addition, once the position of mutated amino acid is determined, a gene coding for the modified enzyme can be obtained by site-specific mutagenesis using a mutagenic primer. In addition, once an entire amino acid sequence is determined, a DNA coding for the amino acid sequence can be chemically synthesized according to a conventional procedure.

Genes coding for farnesyldiphosphate synthases used as starting materials to obtain the present genes have been cloned from various organisms, and therefore they can be used. For example, a gene of <u>Bacillus stearothermophilus</u> origin is described in J. Biochem. <u>113</u>, 355 - 363 (1993), a gene of <u>E. coli</u> origin is described in J. Biochem. <u>108</u>, 995 - 1000 (1990), a gene of yeast origin is described in J.B.C. <u>264</u>, 19176 - 19184 (1989), a gene of rat origin is described in Mol. Cell. Biol. <u>7</u>, 3138 - 3146 (1987), and a gene of human origin is described in J.B.C. <u>265</u>, 4607 - 4614 (1990).

The present invention further provides recombinant vectors, especially expression vectors, comprising the abovementioned gene (DNA), recombinant host transformed with said vector, and a process for production of said enzyme using said recombinant host.

As an example, where <u>E. coli</u> is used as a host, it is known that there are gene expression control mechanisms which regulate transcription of DNA to mRNA, translation of mRNA to protein etc.

As promoter sequences which control the synthesis of mRNA, naturally occurring sequences such as lac, trp, bla, lpp, PL, PR, tet, T3, T7 et al., as well as mutants thereof, such as lacUV5, sequences prepared by fusing naturally occurring promoter sequences, such as tac, tra, etc. are known, and they can be used in the present invention.

As sequences which control the ability to synthesize a protein from mRNA, it is known that a ribosome-binding site (GAGG and similar sequence) and the distance between the ribosome-binding site and the start codon ATG are important. In addition, it is known that a terminator which directs the termination of transcription at the 3'-end (for example, a vector comprising rrnBT1T2 is commercially available from Pharmacia) influences the efficiency of protein synthesis in a recombinant host.

As starting vectors to prepare recombinant vectors of the present invention, those commercially available can be used. Alternatively, various vectors derivatized according to a particular purpose can be used. For example, pBR322, pBR327, pKK223-2, pKK233-2, pTrc99A etc. containing a replicon derived from pMB1; pUC18, pUC19, pUC118, pUC119, pTV118N, pTV119N, pHSG298, pHSG396 etc., which have been modified to increase copy number; pACYC177, pACYC184 etc. containing a replicon derived from p15A; as well as plasmids derived from pSC101, C01E1, R1 or F-factor, may be mentioned.

Further, in addition to plasmids, viral vectors such as λ phage, M13 phage etc., and transposones can be used for introduction of a gene. These vectors are described in Molecular cloning (J. Sambrook, E.F. Fritsch, J. Maniatis, Cold Spring Harbor Laboratory Press); Cloning vector (P.H. Pouwels, B.E. Enger-Valk, W.J. Brammer, Elsevier); and catalogs of manufacturers of vectors.

Especially preferable is pTrc99 (commercially available for Pharmacia) which has an ampicillin resistance gene as a selective maker, Ptrc and lacl^q as a promoter and control gene, an AGGA sequence as a ribosome-binding site and rrnBT1T2 as a terminator, and therefore has a function to control an expression of a geranylgeranyldiphosphate synthase.

Introduction of a DNA coding for geranylgeranyldiphosphate synthase and if necessary DNA fragments having a function to control the expression of said gene into the above-mentioned vectors can be carried out using appropriate restriction enzymes and ligases according to a conventional procedure.

Such a recombinant vector can be used to transform a microorganism such as <u>Escherichia coli</u>, <u>Bacillus</u> etc. Transformation can be carried out according to a conventional procedure, for example by the CaCl₂ method, protoplast method etc. described, for example, in Molecular cloning (J. Sambrook, E.F. Fritsch, T. Maniatis, Cold Spring Harbor Laboratory Press), DNA cloning Vol. I to III (D.M. Glover, IRLPRESS).

Although methods for expression of the present gene in <u>E. coli</u> was described in detail, according to the present invention, a DNA coding for a geranylgeranyldiphosphate synthase is inserted into a conventional expression vector according to a conventional procedure, and the vector is used to transform a host, for example, prokaryotic cells such as various bacterial cells, lower eukaryotic cells for example single cell hosts, for example, yeast cells, or higher eukaryotic cells such as silk-worm. After transformation, the transformant is cultured to produce a geranylgeranyldiphosphate synthase, according to a conventional process.

When a transformant host such as <u>E. coli</u> is cultured, geranylgeranyldiphosphate synthase is intracellularly accumulated. To recover the geranylgeranyldiphosphate from the cultured host cells, the cells are treated physiologically or chemically, for example, with a cell lysating agent to lyze the cells. The cell debris is removed, and the supernatant is subjected to an isolation process conventional for purification of enzymes. The above-mentioned cell-lysing enzyme is preferably lysozyme, and the physical treatment is preferably treatment with ultrasonic radiation. When the supernatant is heated to a temperature of about 55°C, proteins intrinsic to <u>E. coli</u> are insolubilized and removed as an insoluble precipitate. To purify the enzyme, gel-filtration chromatography, ion exchange chromatography, hydrophobic chromatography, reversed chromatography, and affinity chromatography can be used alone or in combination. During the purification and isolation steps, the desired enzyme can be stabilized by addition of a reducing agent such as dithiothreitol, protecting agent against proteases such as PMSF, BSA etc., metal ions such as magnesium, alone or in combination.

The present invention further provides a process for production of geranylgeranyldiphosphate or geranylgeranyol. In this process, isopentenyldiphosphate, dimethylallyldiphosphate, geranyldiphosphate, farnesyldiphosphate may be used as substrates.

EXAMPLES

5

10

20

25

Next, the present invention is explained in more detail by means of examples, though the present invention is not limited thereto.

Example 1. Construction of mutated genes (Fig. 5)

The translation start codon in plasmid pFE15 (Japanese Unexamined Patent Publication (Kokai) No. 5-219761) containing a gene coding for farnesyldiphosphate synthase of <u>Bacillus stearothermophilus</u> origin was changed to ATG to obtain plasmid pEX11 (J. Biochem. 113, 355 - 363 (1993)) for overexpression of farnesyldiphosphate synthase, and the plasmid pEX11 was used in the following Examples. The mutation was carried out according to M. Myers et al. (Science, 229, 242 - 247 (1985)).

First, a farnesyldiphosphate synthase gene present in Ncol-HindIII fragment in pEX11 was removed, and inserted it into plasmid pTV118N (available from Takara Shuzo, Japan) to construct a plasmid, which was then introduced into E. coli cells. The transformed E. coli cells were cultured. With infection of a helper phage M13K07 (available from Takara Shuzo), pTV118N is converted to a single-stranded DNA and preferentially incorporated in phage particles and liberated out of cells. The culture was centrifuged to obtain a supernatant, from which the single-stranded DNA was recovered.

The single-stranded DNA thus recovered was subjected to mutation with sodium nitrite (concentration 1M or 0.2M) to introduce random mutation into the single-stranded DNA, which was then restored to a double-stranded DNA using AMV reverse-transcriptase XL (E.C.2.7.7.7). This farnesyldiphosphate synthase gene fragment was introduced into pTrc99A (available for Pharmacia) and pTV118N, and resulting recombinant plasmids were used to transform <u>E. coli</u> into which a phytoene synthase gene and phytoene desaturase gene had been previously introduced, and red colonies were selected. The principle of the selection is as follows.

The following screening method follows Ohnuma et al. (J. Biol. Chem., <u>269</u>, 14792 - 14797 (1994)). <u>E. coli</u> harboring a plasmid pACYC-IB, into which crtB (phytoene synthase gene) and crtl (phytoene desaturase gene) of a phytopathogen <u>Erwinia uredovora</u> origin had been introduced, was transformed with the mutant plasmid. Note that at present it is believed that <u>E. coli</u> does not have a geranylgeranyldiphosphate synthase. If the mutant plasmid encodes geranylgeranyldiphosphate synthase activity, lycopene having red color is produced in <u>E. coli</u> cells by pACYC-IB resulting in formation of red-colored colonies. However, if the mutant plasmid does not encode geranylgeranyldiphosphate synthase activity, colonies are color-less. In this way, geranylgeranyldiphosphate synthase activity was easily detected by visual observation.

As a result of transformation of the $\underline{E.~coli}$ cells with the mutant plasmid, red colonies were detected. The ratio of positive clones was 1.32×10^{-3} (10 colonies per 7,600 colonies) when the mutation was carried out using 1M NaNO₂, while the ratio of positive clones was 5.98×10^{-5} (one colony per 16,720 colonies) when the mutation was carried out using 0.2M NaNO₂, revealing that the higher the concentration of NaNO₂, the higher the positive ratio.

Among the positive colonies, four colonies were selected, and a nucleotide sequence of an enzyme-coding region in the plasmid was determined, and an amino acid sequence encoded by the nucleotide sequence was determined, for each positive clone. The result is shown in SEQ ID NOs: 1 to 4. In addition, these amino acid sequences were compared with the native amino acid sequence, and positions of the mutation are shown in Fig. 4.

Four mutated enzymes encoded by four mutant genes were further characterized.

Example 2. Production of mutated enzymes

E. coli transformed with the mutant plasmid was cultured in LB medium at 37°C overnight. The culture was centrifuged at 3,000 × G, at 4°C for 5 minutes to collect cells, which were then suspended in a buffer for sonication (50 mM Tris-HCl (pH 7.0), 10 mM 2-mercaptoethanol, 1 mM EDTA). The suspension was subjected to ultrasonic waves to disrupt the cells. The sonicate was centrifuged at 5,000 × g, at a temperature of 4°C for 20 minutes, to obtain a supernatant, which was then heated at 55°C for one hour to inactivate enzymes intrinsic to E. coli to obtain a crude enzyme extract.

50

20

35

To test the enzymatic activity of each mutant enzyme, reactions were carried out in the following reaction mixture.

Table 1

[1-14C]IPP (1 Ci/mol)) 25 nmol Allyl substrate (DMAPP, GPP, FPP) 25 nmol MgCl₂ 5 µmol NH₄CI 50 μmol 2-Mercaptoethanol 50 μmol Tris-HCI buffer (pH 8.5) 50 umol Sample to be tested proper quantity Total 1 ml

Note:

DMAPP: Dimethylallyldiphosphate

GPP: Geranyldiphosphate FPP: Farnesyldiphosphate

The reaction mixture was incubated at 55°C for 30 minutes, and the product was extracted with water-saturated 1butanol, and radioactivity of the extract was counted by a liquid scintillation counter. In addition, the extract (butanol layer) was treated with an acid phosphatase and extracted with pentane. The extract was analyzed by TLC. The TLC analysis showed that the use of dimethylallyldiphosphate and geranyldiphosphate as an allyl substrate provides similar TLC patterns. Note that since the amount of each sample was adjusted so that the radioactivity is approximately same between the samples, the density of the band does not indicate specific activity.

The modified enzymes Nos. 1 and 4 produced an amount of geranylgeranyldiphosphate more than that of farnesyldiphosphate, and therefore it is considered that the modified enzymes Nos. 1 and 4 are suitable for the production of geranylgeranyldiphosphate. On the other hand, the modified enzymes No. 2 and No. 3 provided a small amount of geranylgeranyldiphosphate.

Where (all-E)-farnesyldiphosphate was used as a substrate (primer), (all-E)-geranylgeranyldiphosphate was formed. The results are shown in Figs. 6 to 9.

Specific activity and ratio of product (GGOH/FOH) are shown in Table 2.

Table 2

40			Specific activity* (nmol/min/mg protein)	Ratio of product (GGPP/FPP)
	Wi	ld type	286	0
	No. 1	pTV118N	0.293	18.4
45		pTrc99A	0.253	6.28
	No. 2	pTV118N	110	2.95 × 10 ⁻²
		pTrc99A	83	2.54 × 10 ⁻²
	No. 3	pTV118N	143	1.65 × 10 ⁻¹
50		pTrc99A	19.7	1.73 × 10 ⁻¹
	No. 4	pTV118N	0.262	15.5
		pTrc99A	0.271	8.28

*DMAPP was used as substrate.

7

5

10

15

20

30

35

	SEÇ	QUEN	ICE	LIS	rinc	3											
5	SEÇ) ID	NO.	:	1												
-	SEÇ	QUEN	ICE .	LENG	GTH:	8	94										
	SEÇ	QUEN	CE	TYPI	Ξ:	Nuc	lei	c ac	id								
10	STF	RAND	NES	s:	Dou	ble											
	TOF	OLO	GY:	Lj	inea	r											
	MOI	ECU	LAR	TYE	PE:												
15	sot	JRCE	:]	Baci	llu	s s	tear	coth	erm	oph:	ilus						
	СНА	RAC	TER	ISTI	C:	Mu	tant	(1) 0	f Di	NA c	odi	ng 1	for			
20	far	nes	yld:	iphc	sph	ate	syr	ntha	se								
	SEQ	UEN	CE														
	ATG	GCG	CAG	CTT	TCA	GTT	GAA	CAG	ттт	СТС	AAC	GAG	CAA	AAA	CAG	GCG	48
25	Met	Ala	G1n	Leu	Ser	Va1	Glu	Gln	Phe	Leu	Asn	Glu	Gln	Lys	G1n	Ala	
					5					10					15		
30															CCG		96
	Val	Glu	Thr	Ala 20	Leu	Ser	Arg	Tyr	Ile 25	Glu	Arg	Leu	Glu	Gly 30	Pro	Ala	
	AAG	CTG	AAA		GCG	ATG	GCG	TAC		TTG	GAG	GCC	GGC		AAA	CGA	144
35															Lys		
			35					40				-	45				
40	ATC	CGT	CCG	TTG	CTG	CTT	CTG	TCC	ACC	GTT	CGG	GCG	CTC	GGA	AAA	GAC	192
40	Ile		Pro	Leu	Leu	Leu		Ser	Thr	Val	Arg		Leu	Gly	Lys	Asp	
	CCG	50	GTC	GG4	ጥ ፐር	ccc	55 GTC	CCC	ጥሮር	ccc	ለ ጥጥ	60	ለ ጥር	ል ጥር	CAT	۸۵۵	240
45															His		240
	65			•		70					75					80	
	CAC	TCT	TTG	ATC	CAT	GAT	GAT	TTG	CCG	AGC	ATG	GAC	AAC	GAT	GAT	TTG	288
50	His	Ser	Leu	Ile	His	Asp	Asp	Leu	Pro	Ser	Met	Asp	Asn	Asp	Asp	Leu	
					85					90					95		

	CGG	CGC	GGC	AAG	CCG	ACG	AAC	CAT	` AAA	GTG	TTC	GGC	GAG	GCG	ATG	GCC	336
	Arg	Arg	Gly	Lys	Pro	Thr	Asn	His	Lys	Val	Phe	Gly	G1u	Ala	Met	Ala	
5				100	,				105					110	ı		
	ATC	TTG	GCG	GGG	GAC	GGG	TTG	TTG	ACG	TAC	GCG	TTT	CAA	TTG	ATC	ACC	384
	Ile	Leu	Ala	G1y	Asp	Gly	Leu	Leu	Thr	Tyr	Ala	Phe	G1n	Leu	Ile	Thr	
10			115					120					125				
	GAA	ATC	GAC	GAT	GAG	CGC	ATC	CCT	CCT	TCC	GTC	CGG	CTT	CGG	CTC	ATC	432
4.5	Glu	Ile	Asp	Asp	Glu	Arg	Ile	Pro	Pro	Ser	Val	Arg	Leu	Arg	Leu	Ile	
15		130					135					140					
	GAA	CGG	CTG	GCG	AAA	GCG	GCC	GGT	CCG	GAA	GGG	ATG	GTC	GCC	GGT	CAG	480
20	Glu	Arg	Leu	Ala	Lys	Ala	Ala	Gly	Pro	G1u	Gly	Met	Val	Ala	G1y	Gln	
20	145					150					155					160	
	GCA	GCC	GAT	ATG	GAA	GGA	GAG	GGG	AAA	ACG	CTG	ACG	CTT	TCG	GAG	CTC	528
25	Ala	Ala	Asp	Met	Glu	Gly	Glu	Gly	Lys	Thr	Leu	Thr	Leu	Ser	Glu	Leu	
					165					170					175		
	GAA	TAC	ATT	CAT	CGG	CAT	AAA	ACC	GGG	AAA	ATG	CTG	CAA	TAC	AGC	GTG	576
30	G1u	Tyr	Ile	His	Arg	His	Lys	Thr	Gly	Lys	Met	Leu	Gln	Tyr	Ser	Val	
				180					185					190			
	CAC	GCC	GGC	GCC	TTG	ATC	GGC	GGC	GCT	GAT	GCC	CGG	CAA	ACG	CGG	GAG	624
35	His	Ala	Gly	Ala	Leu	lle	Gly	G1y	Ala	Asp	Ala	Arg	Gln	Thr	Arg	Glu	
			195					200					205				
									GGC								672
40	Leu	Asp	Glu	Phe	Ala	Ala	His	Leu	Gly	Leu	Ala	Phe	Gln	Ile	Arg	Asp	
		210					215					220					
	•								GAA								720
45	Asp	Ile	Leu	Asp			Gly	Ala	Glu	Glu	Lys	Ile	Gly	Lys	Pro	Val	
	225					230					235					240	
	GGC																768.
50	G1y	Ser	Asp			Asn	Asn	Lys	Ala	Thr	Tyr	Pro	Ala	Leu	Leu	Ser	
					245					250					255		

	CTT	GCC	GGC	GCG	AAG	GAA	AAG	TTG	GCG	TTC	CAT	ATC	GAG	GCG	GCG	CAG	816
5	Leu	Ala	Gly	Ala	Lys	Glu	Lys	Leu	Ala	Phe	His	Ile	Glu	Ala	Ala	Gln	
				260					265					270		-	
							GAC										864
10	Arg	HIS	275	Arg	Asn	Ala	Asp	Val 280	Asp	Gly	Ala	Ala		Ala	Tyr	Ile	
	TGC	GAA		GTC	GCC	GCC	CGC		CAT	ТАА			285				894
							Arg										0,54
15	•	290					295	•									
	SEQ	ID	NO:	: 2													
20	SEQ	UEN	CE I	LENG	TH:	89	94										
	SEQ	UEN	CE 1	YPE	:]	Nuc]	leic	ac.	id								
	STR	AND	NESS	S:	Dou	ble											
25	тор	oro.	Υ:	Lin	ear												
	MOL	ECU	LAR	TYP	E :					÷							
30	SOU	RCE	: E	Baci	llu	s st	ear	oth	ermo	phi	lus						
	СНА	.RAC	reri	STI	C:	Mut	ant	(2) of	DN	A c	odir	ıg f	or			
	far	nes	yldi	pho	sph	ate	sys	tha	se								
35	SEQ	UEN	CE														
	ATG	GCG	CAG	CTT	TCA	GTT	GAA	CAG	TTT	CTC	AAC	GĄG	CAA	AAA	CAG	GCG	48
40	Met	Ala	Gln	Leu	Ser	Va1	Glu	Gln	Phe	Leu	Asn	Glu	Gln	Lys	G1n	Ala	
					5					10					15		
	GTG	GAA	ACA	GCG	CTC	TCC	CGT	TAT	ATA	GAG	CGC	TTA	GAA	GGG	CCG	GCG	96
45	Val	Glu	Thr		Leu	Ser	Arg	Tyr		Glu	Arg	Leu	Glu	Gly	Pro	Ala	
		0.00		20	000	4.50		m. a	25					30			
							GCG										144
50	шys	AUT	ъуs 35	пуз	VIG	rie t	Ala	40	SET	ьeu	GIU	Ата	45	ета	гàг	Arg	
								, 0					7.0				

	ATC	CGT	CCG	TTG	CTG	CTT	CTG	TCC	ACC	GTT	CAG	GCG	CTC	GGC	AAA	GAC	192
_	Ile	Arg	Pro	Leu	Leu	Leu	Leu	Ser	Thr	Val	G1n	Ala	Leu	Gly	Lys	Asp	
5		50					55					60					
	CCG	GCG	GTC	GGA	TTG	ccc	GTC	GCC	TGC	GCG	ATT	GAA	ATG	ATC	CAT	ACG	240
10	Pro	Ala	Val	Gly	Leu	Pro	Val	Ala	Cys	Ala	Ile	Glu	Met	Ile	His	Thr	
70	65					70					75					80	
	TAC	TCT	TTG	ATC	CAT	GAT	GAT	TTG	CCG	AGC	ATG	GAC	AAC	GAT	GAT	TTG	288
15	Tyr	Ser	Leu	Ile	His	Asp	Asp	Leu	Pro	Ser	Met	Asp	Asn	Asp	Asp	Leu	
					85					90					95		
	CGG	CGC	GGC	AAG	CCG	ACG	AAC	CAT	AAA	GTG	TTC	GGC	GAG	GCG	ATG	GCC	336
20	Arg	Arg	Gly	Lys	Pro	Thr	Asn	His	Lys	Val	Phe	Gly	Glu	Ala	Met	Ala	
				100					105					110			
	ATC	TTG	GCG	GGG	GAC	GGG	TTG	TTG	ACG	TAC	GCG	TTT	CAA	TTG	ATC	ACC	384
25	Ile	Leu	Ala	G1y	Asp	Gly	Leu	Leu	Thr	Tyr	Ala	Phe	Gln	Leu	Ile	Thr	
			115					120					125				
	GAA	ATC	GAC	GAT	GAG	CGC	ATC	CCT	CCT	TCC	GTC	CGG	CTT	CGG	CTC	ATC	432
30	Glu	Ile	Asp	Asp	Glu	Arg	Ile	Pro	Pro	Ser	Val	Arg	Leu	Arg	Leu	Ile	
		130					135					140					
	GAA	CGG	CTG	GCG	AAA	GCG	GCC	GGT	CCG	GAA	GGG	ATG	GTC	GCC	GGT	CAG	480
35	Glu	Arg	Leu	Ala	Lys	Ala	Ala	Gly	Pro	Glu	Gly	Met	Val	Ala	Gly	Gln	
	145					150					155					160	
	GCA	GCC	GAT	ATG	GAA	GGA	GAG	GGG	AAA	ACG	CTG	AÇG	CTT	TCG	GAG	CTC	528
40	Ala	Ala	Asp	Met	Glu	Gly	Glu	Gly	Lys	Thr	Leu	Thr	Leu	Ser	Glu	Leu	
					165					170					175		
	GAA	TAC	ATT	CAT	CGG	CAT	AAA	ACC	GGG	AAA	ATG	CTG	CAA	TAC	AGC	GTG	576
45	Glu	Tyr	Ile	His	Arg	His	Lys	Thr	Gly	Lys	Met	Leu	Gln	Tyr	Ser	Val	
				180					185					190			
	CAC	GCC	GGC	GCC	TTG	ATC	GGC	GGC	GCT	GAT	GCC	CGG	CAA	ACG	CGG	GAG	624
50	His	Ala	Gly	Ala	Leu	Ile	Gly	Gly	Ala	Asp	Ala	Arg	Gln	Thr	Arg	Glu	
			195					200					205				

	CTT	GAC	GAA	TTC	GCC	GCC	CAT	CTA	GGC	CTT	GCC	TTT	CAA	ATT	CGC	GAT	672
E	Leu	Asp	Glu	Phe	Ala	Ala	His	Leu	Gly	Leu	Ala	Phe	Gln	Ile	Arg	Asp	
5		210					215					220					
	GAT	ATT	CTC	GAT	TTA	GAA	GGG	GCA	GAA	GAA	AAA	ATC	GGC	AAG	CCG	GTC	720
10	Asp	Ile	Leu	Asp	Ile	G1u	G1y	Ala	Glu	Glu	Lys	Ile	Gly	Lys	Pro	Val	
70	225					230					235					240	
	GGC	AGC	GAC	CAA	AGC	AAC	AAC	AAA	GCG	ACG	TAT	CCA	GCG	TTG	CTG	TCG	768
15	Gly	Ser	Asp	Gln	Ser	Asn	Asn	Lys	Ala	Thr	Tyr	Pro	Ala	Leu	Leu	Ser	
					245					250					255		
	CTT	GCC	GGC	GCG	AAG	GAA	AAG	TTG	GCG	TTC	CAT	ATC	GAG	GCG	GCG	CAG	816
20	Leu	Ala	Gly	Ala	Lys	Glu	Lys	Leu	Ala	Phe	His	Ile	G1u	Ala	Ala	Gln	
				260					265					270			
								GTT									864
25	Arg	His		Arg	Asn	Ala	Asp	Val	Asp	Gly	Ala	Ala		Ala	Tyr	Ile	
			275					280		<u>.</u>			285				
								GAC									894
30	Cys		Leu	Val	Ala	Ala	•	Asp	His	***							
	ano	290		2			295										
	SEQ	ID	NO:	: 3													
35	SEQ	UEN	CE I	ENG	TH:	89	94										
	SEQ	UEN	CE I	YPE	:]	Nuc]	leic	ac:	id								
	STR	AND	NESS	S:	Doul	ble											
40	TOP	OLO.	Y:	Lin	ear												
	MOL	ECU:	LAR	TYP	E:												
45	sou	RCE	: E	Baci	llu	s st	ear	oth	ermo	phi	lus						
	СНА	.RAC'	rer i	STI	C:	Mut	ant	(3) of	DN	A c	odir	ıg f	or			
50	far	nes	yldi	pho	sph	ate	sys	tha	se								
50	SEQ	UEN	CE														

	ATG	GCG	CAG	CTT	TCA	GTT	GAA	CAG	TTT	CTC	AAC	GAG	CAA	AAA	CAG	GCG	48
_	Met	Ala	Gln	Leu	Ser	Val	Glu	Gln	Phe	Leu	Asn	Glu	Gln	Lys	Gln	Ala	
5					5					10					15		
	GTG	GAA	ACA	GCG	CTC	TCC	CGT	TAT	ATA	GAG	CGC	TTA	GAA	GGG	CCG	GCG	96
10	Val	Glu	Thr	Ala	Leu	Ser	Arg	Tyr	Ile	Glu	Arg	Leu	Glu	Gly	Pro	Ala	
10				20					25					30			
	AAG	CTG	AAA	AAG	GCG	ATG	GCG	TAC	TCA	TTG	GAG	GCC	GGC	GGC	AAA	CGA	144
15	Lys	Leu	Lys	Lys	Ala	Met	Ala	Tyr	Ser	Leu	Glu	Ala	Gly	Gly	Lys	Arg	
10			35					40					45				
	ATC	CGT	CCG	TTG	CTG	CTT	CTG	TCC	ACC	GTT	CGG	GCG	CTC	GGA	AAA	GAC	192
20	Ile	Arg	Pro	Leu	Leu	Leu	Leu	Ser	Thr	Val	Arg	Ala	Leu	Gly	Lys	Asp	
		50					55					60					
	CCG	GCG	GTC	GGA	TTG	CCC	GTC	GCC	TGC	GCG	ATT	GAA	ATG	ATC	CAT	ACG	240
25	Pro	Ala	Val	Gly	Leu	Pro	Val	Ala	Cys	Ala	Ile	Glu	Met	Ile	His	Thr	
	65					70					75					80	
	TAC	TCT	TTG	ATC	CAT	GAT	GAT	TTG	CCG	AGC	ATG	GAC	AAC	GAT	GAT	TTG	288
30	Tyr	Ser	Leu	Ile	His	Asp	Asp	Leu	Pro	Ser	Met	Asp	Asn	Asp	Asp	Leu	
					85					90					95		
	CGG	CGC	GGC	AAG	CCG	ACG	AAC	CAT	AAA	GTG	TTC	GGC	GAG	GCG	ATG	GCC	336
35	Arg	Arg	Gly	Lys	Pro	Thr	Asn	His	Lys	Val	Phe	Gly	Glu	Ala	Met	Ala	
				100					105					110			
	ATC	TTG	GCG	GGG	GAC	GGG	TTG	TTG	ACG	TAC	GCG	TTT	CAA	TTG	ATC	ACC	384
40	Ile	Leu	Ala	Gly	Asp	Gly	Leu	Leu	Thr	Tyr	Ala	Phe	Gln	Leu	Ile	Thr	
			115					120					125				
	GAA	ATC	GAC	GAT	GAG	CGC	ATC	CCT	CCT	TCC	GTC	CGG	CTT	CGG	CTC	ATC	432
45	G1u	Ile	Asp	Asp	Glu	Arg	Ile	Pro	Pro	Ser	Val	Arg	Leu	Arg	Leu	Ile	
		130					135					140					
	GAA	CGG	CTG	GCG	AAA	GCG	GCC	GGT	CCG	GAA	GGG	ATG	GCC	GCC	GGT	CAG	480 ⁻
50	Glu	Arg	Leu	Ala	Lys	Ala	Ala	Gly	Pro	Glu	Gly	Met	Ala	Ala	Gly	G1n	
	145					150					155					160	

	GCA	GCC	GAT	ATG	GAA	GGA	GAG	GGG	AAA	ACG	CTG	ACG	CTT	TCG	GAG	CTC	528
	Ala	Ala	Asp	Met	Glu	G1y	Glu	Gly	Lys	Thr	Leu	Thr	Leu	Ser	Glu	Leu	
5					165					170					175		•
	GAA	TAC	ATT	CAT	CGG	TAT	AAA	ACC	GGG	AAA	ATG	CTG	CAA	TAC	AGC	GTG	576
	Glu	Tyr	Ile	His	Arg	Tyr	Lys	Thr	Gly	Lys	Met	Leu	Gln	Tyr	Ser	Val	
10				180					185					190			
	CAC	GCC	GGC	GCC	TTG	ATC	GGC	GGC	GCT	GAT	GCC	CGG	CAA	ACG	CGG	GAG	624
	His	Ala	Gly	Ala	Leu	Ile	Gly	Gly	Ala	Asp	Ala	Arg	Gln	Thr	Arg	Glu	
15			195					200					205				
	CTT	GAC	GAA	TTC	ĢCC	GCC	CAT	CTA	GGC	CTT	GCC	TTT	CAA	ATT	CGC	GAT	672
	Leu	Asp	Glu	Phe	Ala	Ala	His	Leu	Gly	Leu	Ala	Phe	Gln	Ile	Arg	Asp	
20		210					215					220					
	GAT	ATT	CTC	GAT	ATT	GAA	GGG	GCA	GAA	GAA	AAA	ATC	GGC	AAG	CCG	GTC	720
	Asp	Ile	Leu	Asp	Ile	Glu	Gly	Ala	G1u	Glu	Lys	Ile	Gly	Lys	Pro	Val	
25	225					230					235					240	
	GGC	AGC	GAC	CAA	AGC	AAC	AAC	AAA	GCG	ACG	TAT	CCA	GCG	TTG	CTG	TCG	768
	Gly	Ser	Asp	Gln	Ser	Asn	Asn	Lys	Ala	Thr	Tyr	Pro	Ala	Leu	Leu	Ser	
30					245					250					255		
	CTT	GCC	GGC	GCA	AAG	GAA	AAG	TTG	GCG	TTC	CAT	ATC	GAG	GCG	GCG	CAG	816
	Leu	Ala	Gly	Ala	Lys	Glu	Lys	Leu	Ala	Phe	His	Ile	G1u	Ala	Ala	Gln	
35				260					265					270			
55	CGC	CAT	TTA	CGG	AAC	GCC	GAC	GTT	GAC	GGC	GCC	GCG	CTC	GCC	TAT	ATT	864
	Arg	His	Leu	Arg	Asn	Ala	Asp	Val	Asp	Gly	Ala	Ala	Leu	Ala	Tyr	Ile	
			275					280					285				
40	TGC	GAA	CTG	GTC	GCC	GCC	CGC	GAC	CAT	TAA							894
	Cys	Glu	Leu	Val	Ala	Ala	Arg	Asp	His	***							
		290					295										
45	SEQ	ID	NO:	4													
	SEQ	UENC	CE L	ENG	TH:	89	4										
<i>5</i> 0	SEQ	UENC	CE T	YPE	: N	Nucl	eic	aci	.d								
50	STR	ANDI	NESS	:	Doub	ole											
	TOP	OLOY	:	Lin	ear												

MOLECULAR TYPE: SOURCE: Bacillus stearothermophilus CHARACTERISTIC: Mutant (4) of DNA coding for farnesyldiphosphate systhase **SEQUENCE** ATG GCG CAG CTT TCA GTT GAA CAG TTT CTC AAC GAG CAA AAA CAG GCG Met Ala Gln Leu Ser Val Glu Gln Phe Leu Asn Glu Gln Lys Gln Ala GTG GAA ACA GCG CTC TCC CGT TAT ATA GAG CGC TTA GAA GGG CCG GCG Val Glu Thr Ala Leu Ser Arg Tyr Ile Glu Arg Leu Glu Gly Pro Ala AAG CTG AAA AAG GCG ATG GCG TAC TCA TTG GAG GCC GGC GGC AAA CGA Lys Leu Lys Lys Ala Met Ala Tyr Ser Leu Glu Ala Gly Gly Lys Arg ATC CGT CCG TTG CTG CTT CTG TCC ACC GTT CGG GCG CTC GGC AAA GAC Ile Arg Pro Leu Leu Leu Ser Thr Val Arg Ala Leu Gly Lys Asp CCG GCG GTC GGA TTG CCC GTC GCC TGC GCG ATT GAA ATG ATC CAT ACG Pro Ala Val Gly Leu Pro Val Ala Cys Ala Ile Glu Met Ile His Thr CAC TCT TTG ATC CAT GAT GAT TTG CCG AGC ATG GAC AAC GAT GAT TTG His Ser Leu Ile His Asp Asp Leu Pro Ser Met Asp Asn Asp Asp Leu CGG CGC GGC AAG CCG ACG AAC CAT AAA GTG TTC GGC GAG GCG ATG GCC Arg Arg Gly Lys Pro Thr Asn His Lys Val Phe Gly Glu Ala Met Ala ATC TTG GCG GGG GAC GGG TTG TTG ACG TAC GCG TTT CAA TTG ATC ACC Ile Leu Ala Gly Asp Gly Leu Leu Thr Tyr Ala Phe Gln Leu Ile Thr GAA ATC GAC GAT GAG CGC ATC CCT CCT TCC GTC CGG CTT CGG CTC ATC Glu Ile Asp Asp Glu Arg Ile Pro Pro Ser Val Arg Leu Arg Leu Ile

	GAA	CGG	CTG	GCG	AAA	GCG	GCC	GGT	CCG	GAA	GGG	ATG	GTC	GCC	GGT	CAG	480
	Glu	Arg	Leu	Ala	Lys	Ala	Ala	Gly	Pro	Glu	Gly	Met	Val	Ala	Gly	Gln	
5	145					150					155					160	
	GCA	GCC	GAT	ATG	GAA	GGA	GAG	GGG	AAA	ACG	CTG	ACG	CTT	TCG	GAG	CTC	528
	Ala	Ala	Asp	Met	Glu	Gly	Glu	Gly	Lys	Thr	Leu	Thr	Leu	Ser	Glu	Leu	
10					165					170					175		
	GAA	TAC	ATT	CAT	CGG	CAT	AAA	ACC	GGG	AAA	ATG	CTG	CAA	TAC	AGC	GTG	576
	Glu	Tyr	Ile	His	Arg	His	Lys	Thr	Gly	Lys	Met	Leu	Gln	Tyr	Ser	Val	
15				180					185					190			
	CAC	GCC	GGC	GCC	ŢTG	ATC	GGC	GGC	GCT	GAT	GCC	CGG	CAA	ACG	CGG	GAG	624
	His	Ala	Gly	Ala	Leu	Ile	Gly	Gly	Ala	Asp	Ala	Arg	Gln	Thr	Arg	Glu	
20			195					200					205				
	CTT	GAC	GAA	TTC	GCC	GCC	CAT	CTA	GGC	CTT	GCC	TTT	CAA	ATT	CGC	GAT	672
	Leu	Asp	G1u	Phe	Ala	Ala	His	Leu	Gly	Leu	Ala	Phe	Gln	Ile	Arg	Asp	
25		210					215					220					
	GAT	ATT	CTC	GAT	ATT	GAA	GGG	GCA	GAA	GAA	AAA	ATC	GGC	AAG	CGG	GTC	720
	Asp	Ile	Leu	Asp	Ile	Glu	Gly	Ala	Glu	Glu	Lys	Ile	Gly	Lys	Arg	Val	
30	225					230					235					240	
	GGC	AGC	GAC	CAA	AGC	AAC	AAC	AAA	GCG	ACG	TAT	CCA	GCG	TTG	CTG	TCG	768
	Gly	Ser	Asp	Gln	Ser	Asn	Asn	Lys	Ala	Thr	Tyr	Pro	Ala	Leu	Leu	Ser	
35					245					250					255		
	CTT	GCC	GGC	GCG	AAG	GAA	AAG	TTG	ACG	TTC	CAT	ΑŢC	GAG	GCG	GCG	CAG	816
	Leu	Ala	Gly	Ala	Lys	Glu	Lys	Leu	Thr	Phe	His	Ile	Glu	Ala	Ala	Gln	
40				260					265					270			
	CGC	CAT	TTA	CGG	AAC	GCC	GAC	GTT	GAC	GGC	GCC	GCG	CTC	GCC	TAT	ATT	864
	Arg	His	Leu	Arg	Asn	Ala	Asp	Val	Asp	Gly	Ala	Ala	Leu	Ala	Tyr	Ile	
45			275					280					285				
-	TGC	GAA	CTG	GTC	GCC	GCC	CGC	GAC	CAT	TAA							894
	Cys	Glu	Leu	Val	Ala	Ala	Arg	Asp	His	***							
50		290					295								*		
50	SEQ	ID	NO:	5													
	SEQ	UENC	CE L	ENG	rh:	89	4										

	SEÇ	JOEN	CE '	LAbr	: :	Nuc.	тетс	: ac	10								
5	STF	RAND	NES	S:	Dou	ble											
	TOF	oro	Y:	Lir	ear												
	MOI	ECU	LAR	TYF	E:												
10	sot	JRCE	: 1	Baci	.llu	s s	tear	oth	ermo	ophi	lus						
	CHA	RAC	TER	ISTI	.C:	DN	A co	din	g fo	or n	nati	ve :	farr	nesy	ldi	phosp	hate
15	syn	tha	se														
	SEÇ	UEN	CE														
	ATG	GCG	CAG	CTT	TCA	GTT	GAA	CAG	TTT	CTC	AAC	GAG	CAA	AAA	CAG	GCG	48
20	Met	Ala	Gln	Leu	Ser	Val	Glu	Gln	Phe	Leu	Asn	Glu	G1n	Lys	Gln	Ala	
					5					10					15		
	GTG	GAA	ACA	GCG	CTC	TCC	CGT	TAT	ATA	GAG	CGC	TTA	GAA	GGG	CCG	GCG	96
25	Val	Glu	Thr	Ala	Leu	Ser	Arg	Tyr	Ile	Glu	Arg	Leu	Glu	Gly	Pro	Ala	
				20					25					30			
	AAG	CTG	AAA	AAG	GCG	ATG	GCG	TAC	TCA	TTG	GAG	GCC	GGC	GGC	AAA	CGA	144
30	Lys	Lys	Lys	Lys	Ala	Met	Ala	Tyr	Ser	Leu	Glu	Ala	Gly	Gly	Lys	Arg	
			35					40					45				
	ATC	CGT	CCG	TTG	CTG	CTT	CTG	TCC	ACC	GTT	CAG	GCG	CTC	GGC	AAA	GAC	192
35	Ile	Arg	Pro	Leu	Leu	Leu	Leu	Ser	Thr	Va1	Gln	Ala	Leu	Gly	Lys	Asp	
		50					55					60					
	CCG	GCG	GTC	GGA	TTG	ccc	GTC	GCC	TGC	G CG	ATT	GAA	ATG	ATC	CAT	ACG	240
40	Pro	Ala	Val	Gly	Leu	Pro	Val	Ala	Cys	Ala	Ile	Glu	Met	Ile	His	Thr	
	65					70					75					80	
	TAC	TCT	TTG	ATC	CAT	GAT	GAT	TTG	CCG	AGC	ATG	GAC	AAC	GAT	GAT	TTG	288
45	Tyr	Ser	Leu	Ile	His	Asp	Asp	Leu	Pro	Ser	Met	Asp	Asn	Asp	Asp	Leu	
					85					90					95		.•
	CGG	CGC	GGC	AAG	CCG	ACG	AAC	CAT	AAA	GTG	TTC	GGC	GAG	GCG	ATG	GCC	336
50	Arg	Arg	Gly	Lys	Pro	Thr	Asn	His	Lys	Va1	Phe	Gly	Glu	Ala	Met	Ala	
				100					105					110			

	ATC	TTG	GCG	GGG	GAC	GGG	TTG	TTG	ACG	TAC	GCG	TTT	CAA	TTG	ATC	ACC	384
	Ile	Leu	Ala	Gly	Asp	Gly	Leu	Leu	Thr	Tyr	Ala	Phe	Gln	Leu	Ile	Thr	
5			115					120					125				
	GAA	ATC	GAC	GAT	GAG	CGC	ATC	CCT	CCT	TCC	GTC	CGG	CTT	CGG	CTC	ATC	432
	Glu	Ile	Asp	Asp	Glu	Arg	Ile	Pro	Pro	Ser	Val	Arg	Leu	Arg	Leu	Ile	
10		130					135					140					
	GAA	CGG	CTG	GCG	AAA	GCG	GCC	GGT	CCG	GAA	GGG	ATG	GTC	GCC	GGT	CAG	480
	G1u	Arg	Leu	Ala	Lys	Ala	Ala	G1y	Pro	G1u	Gly	Met	Val	Ala	Gly	Gln	
15	145					150					155					160	
	GCA	GCC	GAT	ATG	GAA	GGA	GAG	GGG	AAA	ACG	CTG	ACG	CTT	TCG	GAG	CTC	528
	Ala	Ala	Asp	Met	Glu	Gly	Glu	Gly	Lys	Thr	Leu	Thr	Leu	Ser	Glu	Leu	
20					165					170					175		
	GAA	TAC	ATT	CAT	CGG	CAT	AAA	ACC	GGG	AAA	ATG	CTG	CAA	TAC	AGC	GTG	576
	G1u	Tyr	Ile	His	Arg	His	Lys	Thr	Gly	Lys	Met	Leu	Gln	Tyr	Ser	Val	
				180					185					190			
25	CAC	GCC	GGC	GCC	TTG	ATC	GGC	GGC	GCT	GAT	GCC	CGG	CAA	ACG	CGG	GAG	624
	His	Ala	Gly	Ala	Leu	Ile	Gly	Gly	Ala	Asp	Ala	Arg	Gln	Thr	Arg	Glu	
			195					200					205				
30	CTT	GAC	GAA	TTC	GCC	GCC	CAT	CTA	GGC	CTT	GCC	TTT	CAA	ATT	CGC	GAT	672
	Leu	Asp	Glu	Phe	Ala	Ala	His	Leu _.	Gly	Leu	Ala	Phe	Gln	Ile	Arg	Asp	
		210					215					220					
35	GAT	ATT	CTC	GAT	ATT	GAA	GGG	GCA	GAA	GAA	AAA	ATC	GGC	AAG	CCG	GTC	720
	Asp	Ile	Leu	Asp	lle	G1u	G1y	Ala	G1u	Glu	Lys	Ile	Gly	Lys	Pro	Va1	
	225					230					235					240	
40	GGC	AGC	GAC	CAA	AGC	AAC	AAC	AAA	GCG	ACG	TAT	CCA	GCG	TTG	CTG	TCG	768
	G1y	Ser	Asp	G1n	Ser	Asn	Asn	Lys	Ala	Thr	Tyr	Pro	Ala	Leu	Leu	Ser	
					245					250					255		
45	CTT	GCC	GGC	GCG	AAG	GAA	AAG	TTG	GCG	TTC	CAT	ATC	GAG	GCG	GCG	CAG	816.
	Leu	Ala	Gly	Ala	Lys	Glu	Lys	Leu	Ala	Phe	His	Ile	Glu	Ala	Ala	Gln	
				260					265					270			
	CGC	CAT	TTA	CGG	AAC	GCC	GAC	GTT	GAC	GGC	GCC	GCG	CTC	GCC	TAT	ATT	864
50	Arg	His	Leu	Arg	Asn.	Ala	Asp	Val	Asp	Gly	Ala	A1a	Leu	Ala	Tyr	Ile	
			275					280					285				

	TGC	GAA	CTG	GTC	GCC	GCC	CGC	GAC	CAT	TAA AAT	394
5	Cys	G1u	Leu	Val	Ala	Ala	Arg	Asp	His	***	
		290					295				
10											
										ynthesizing geranylgeranyldiphosphate and gene co odified from a native farnesyldiphosphate synthase b	
	tion of a										
15											
20											
25											
30											
35											
40											
45											
50											
55											

SEQUENCE LISTING

5	(1) GENERAL INFORMATION:
10	 (i) APPLICANT: (A) NAME: Toyota Jidosha Kabushiki Kaisha (B) STREET: 1, Toyota-cho (C) CITY: Toyota-shi (D) STATE: Aichi (E) COUNTRY: Japan
15	(F) POSTAL CODE (ZIP): None (ii) TITLE OF INVENTION: MUTATED FARNESYLDIPHOSPHATE SYNTHASE CAPABLE OF SYNTHESIZING GERANYLGERANYLDIPHOSPHATE AND GENE CODING THEREFOR
	(iii) NUMBER OF SEQUENCES: 10
20	 (iv) COMPUTER READABLE FORM: (A) MEDIUM TYPE: Floppy disk (B) COMPUTER: IBM PC compatible (C) OPERATING SYSTEM: PC-DOS/MS-DOS (D) SOFTWARE: PatentIn Release #1.0, Version #1.25 (EPO)
25	(v) CURRENT APPLICATION DATA: APPLICATION NUMBER: EP 95115423.6
	(2) INFORMATION FOR SEQ ID NO: 1:
30	 (i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 894 base pairs (B) TYPE: nucleic acid (C) STRANDEDNESS: double (D) TOPOLOGY: linear
35	<pre>(vi) ORIGINAL SOURCE: (A) ORGANISM: Bacillus stearothermophilus</pre>
40	<pre>(ix) FEATURE:</pre>
45	(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 1:
	ATG GCG CAG CTT TCA GTT GAA CAG TTT CTC AAC GAG CAA AAA CAG GCG Met Ala Gln Leu Ser Val Glu Gln Phe Leu Asn Glu Gln Lys Gln Ala 1 5 10 15
50	GTG GAA ACA GCG CTC TCC CGT TAT ATA GAG CGC TTA GAA GGG CCG GCG Val Glu Thr Ala Leu Ser Arg Tyr Ile Glu Arg Leu Glu Gly Pro Ala 20 25 30
55	

									CGA Arg	144
5							CTC Leu			192
10							ATG Met			240
15							AAC Asn			288
							G A G Glu			336
20							CAA Gln 125			384
25							CTT Leu			432
30							GTC Val			480
							CTT Leu			528
35							CAA Gln			576
40							CAA Gln 205			624
45							C AA Gln			672
							GGC Gly			720
50							GCG Ala			7 68

5	CTT Leu	GCC Ala	GGC Gly	GCG Ala 260	AAG Lys	G AA Glu	AAG Lys	TTG Leu	GCG Ala 265	TTC Phe	CAT His	ATC Ile	GAG Glu	GCG Ala 270	GCG Ala	CAG Gln	816
3	CGC A rg	CAT His	TCA Ser 275	CGG Arg	AAC Asn	GCC Ala	GAC Asp	GTT Val 280	GAC Asp	GGC Gly	GCC Ala	GCG Ala	CTC Leu 285	GCC Ala	TAT Tyr	ATT Ile	864
10						GCC Ala				TAA							894
15																	
20																	
25																	
30																	
35																	
40																	
4 5																	
50																	
55																	

	(2)	INF	ORMA'	rion	FOR	SEQ	ID 1	NO: 3	2:							
5			(1	SEQUI A) Li B) T'	ENGTI YPE:	H: 2:	97 ai	mino cid								
		(ii) M O	LECUI	LE T	YPE:	pro	tein								
10		(xi) SE	QUEN	CE DI	ESCR:	IPTI(ON:	SEQ	ID N	0: 2	:				
	Met 1	Ala	Gln	Leu	Ser 5	Val	Glu	Gln	Phe	Leu 10	Asn	Glu	Gln	Lys	Gln 15	Ala
15	Val	Glu	Thr	Ala 20	Leu	Ser	Arg	Tyr	Ile 25	Glu	Arg	Leu	Glu	Gly 30	Pro	Ala
	Lys	Leu	Lys 35	Lys	Ala	Met	Ala	Tyr 40	Ser	Leu	Glu	Ala	Gly 45	Gly	Lys	Arg
20	Ile	Arg 50	Pro	Leu	Leu	Leu	Leu 55	Ser	Thr	Val	Arg	Ala 60	Leu	Gly	Lys	Asp
	Pro 65	Ala	Val	Gly	Leu	Pro 70	Val	Ala	Cys	Ala	Ile 75	Glu	Met	Ile	His	Thr 80
25	His	Ser	Leu	Ile	His 85	Asp	Asp	Leu	Pro	Ser 90	Met	Asp	Asn	Asp	As p 95	Leu
	Arg	Arg	Gly	Lys 100	Pro	Thr	Asn	His	Lys 105	Val	Phe	Gly	Glu	Ala 110	Met	Ala
30	Ile	Leu	Ala 115	Gly	Asp	Gly	Leu	Leu 120	Thr	Tyr	Ala	Phe	Gln 125	Leu	Ile	Thr
35	Glu	Ile 130	Asp	Asp	Glu	Arg	Ile 135	Pro	Pro	Ser	Val	Arg 140	Leu	Arg	Leu	Ile
	Glu 145	Arg	Leu	Ala	Lys	Ala 150	Ala	Gly	Pro	Glu	Gly 155	Met	Val	Ala	Gly	Gln 160
40	Ala	Ala	Asp	Met	Glu 165	-	Glu	_	_			Thr	Leu	Ser	Glu 175	Leu
	Glu	Tyr	Ile	His 180	Arg	His	Lys	Thr	Gly 185	Lys	Met	Leu	Gln	Tyr 190	Ser	Val
45	His	Ala	Gly 195	Ala	Leu	Ile	Gly	Gly 200	Ala	Asp	Ala	Arg	Gln 205	Thr	Arg	Glu
	Leu	Asp 210	Glu	Phe	Ala	Ala	His 215	Leu	Gly	Leu	Ala	Phe 220	Gln	Ile	Arg	Asp
50	As p 225	Ile	Leu	Asp	Ile	Glu 230	Gly	Ala	Glu	Glu	Lys 235	Ile	Gly	Lys	Pro	Val 240
	Gly	Ser	Asp	Gln	Ser 245	Asn	Asn	Lys	Ala	Thr 250	Tyr	Pro	Ala	Leu	Leu 255	Ser

	Leu	Ala	Gly	Ala 260	Lys	Glu	Lys	Leu	Ala 265	Phe	His	Ile	Glu	Ala 270	Ala	Gln
5	Arg	His	Ser 275	Arg	Asn	Ala	Asp	Val 280	Asp	Gly	Ala	Ala	Leu 285	Ala	Tyr	Ile
10	Cys	Glu 290	Leu	Val	Ala	Ala	Arg 295	Asp	His							
15																
20																
25																
30																
35																
40																
45																
50																
55																

	(2)	INF	ORMA'	rion	FOR	SEQ	ID I	, iOi	3:								
5		(i)	(1	QUENCA) LIB) TO	ENGTI YPE: [RAN]	H: 89 nucl	94 ba leic ESS:	ase p acio doub	pair: d	S							
10		(vi)) OR:					illus	s st	earo	ther	noph:	ilus				
15		(ix)	(1	A) NA B) L	AME/I OCAT: THER	ION: INFO	1 CRMA	rion:			ion= hospl					O NA	
••		(xi)) SE	QUEN	CE DI	SCR:	[PTI	ON: S	SEQ :	ID N	0: 3	:					
20		GCG Ala															48
	GTG	GAA	ACA	GCG	CTC	TCC	CGT	TAT	АТА	GAG	CGC	TTA	GAA	GGG	CCG	GCG	96
25	Val	Glu	Thr	Ala 20	Leu	Ser	Arg	Tyr	Ile 25	Glu	Arg	Leu	Glu	Gly 30	Pro	Ala	
		GTG Val														CGA	144
30	цуъ	Vai	д у 5	пур	AIQ	Mec	міа	40	Ser	пеп	Giu	AIG	45	GIY	пуъ	ALG	
		CGT Arg 50															192
35		GCG Ala															240
40	_	TCT Ser	_			_	_	_		_		_	_	_	_	_	288
45		CGC A rg															336
		TTG Leu															384
50		ATC Ile 130															432

5				GCG Ala 150						480
				GGA Gly						528
10	_			CAT His						576
15				ATC Ile						624
20				GCC Ala						672
25				GAA Glu 230						720
25				AAC Asn						768
30				GAA Glu						816
35				GCC Ala						864
40				GCC Ala		TAA				894

	(2)	INF	ORMA'	1.TON	FOR	SEQ	1D .	NO:	4:							
5			(1	SEQUI A) LI B) T	ENGT:	H: 2 ami	97 a	mino cid								
		(ii) M O	LECU	LE T	YPE:	pro	tein								
10		(xi) SE	QUEN	CE D	ESCR	IPTI	ON:	SEQ	ID N	0: 4	:				
	Met 1	Ala	Gln	Leu	Ser 5	Val	Glu	Gln	Phe	Leu 10	Asn	Glu	Gln	Lys	Gln 15	Ala
15	Val	Glu	Thr	Ala 20	Leu	Ser	Arg	Tyr	Ile 25	Glu	Arg	Leu	Glu	Gly 30	Pro	Ala
	Lys	Val	Lys 35	Lys	Ala	Met	Ala	Tyr 40	Ser	Leu	Glu	Ala	Gly 45	Gly	Lys	Arg
20	Ile	Arg 50	Pro	Leu	Leu	Leu	Leu 55	Ser	Thr	Val	Gln	Ala 60	Leu	Gly	Lys	Asp
	Pro 65	Ala	Val	Gly	Leu	Pro 70	Val	Ala	Cys	Ala	Ile 7 5	Glu	Met	Ile	His	Thr 80
25	Tyr	Ser	Leu	Ile	His 85	Asp	Asp	Leu	Pro	Ser 90	Met	Asp	Asn	Asp	As p 95	Leu
	Arg	Arg	Gly	Lys 100	Pro	Thr	Asn	His	Lys 105	Val	Phe	Gly	Glu	Ala 110	Met	Ala
30	Ile	Leu	Ala 115	Gly	Asp	Gly	Leu	Leu 120	Thr	Tyr	Ala	Phe	Gln 125	Leu	Ile	Thr
	Glu	Ile 130	Asp	Asp	Glu	Arg	Ile 135	Pro	Pro	Ser	Val	Arg 140	Leu	Arg	Leu	Ile
35	Glu 145	Arg	Leu	Ala	Lys	Ala 150	Ala	Gly	Pro	Glu	Gly 155	Met	Val	Ala	Gly	Gln 160
40	Ala	Ala	Asp	Met	Glu 165	Gly	Glu	Gly	Lys	Thr 170	Leu	Thr	Leu	Ser	Glu 175	Leu
40	Glu	Tyr	Ile	His 180	Arg	His	Lys	Thr	Gly 185	Lys	Met	Leu	Gln	Tyr 190	Ser	Val
45	His	Ala	Gl y 195	Ala	Leu	Ile	Gly	Gly 200	Ala	Asp	Ala	Arg	Gln 205	Thr	Arg	Glu
	Leu	Asp 210	Glu	Phe	Ala	Ala	His 215	Leu	Gly	Leu	Ala	Phe 220	Gln	Ile	Arg	Asp
50	Asp 225	Ile	Leu	Asp	Ile	Glu 230	Gly	Ala	Glu	Glu	Lys 235	Ile	Gly	Lys	Pro	Val 240
	Gly	Ser	Asp	Gln	Ser 245	Asn	Asn	Lys	Ala	Thr 250	Tyr	Pro	Ala	Leu	Leu 255	Ser

	Leu	Ala	Gly	Ala 260	Lys	Glu	Lys	Leu	Ala 265	Phe	His	Ile	Glu	Ala 270	Ala	Gln
5	Arg	His	Leu 275	Arg	Asn	Ala	Asp	Val 280	Asp	Gly	Ala	Ala	Leu 285	Ala	Tyr	Ile
10	Cys	Glu 290	Leu	Val	Ala	Ala	Arg 295	Asp	His							
15																
20																
25																
30																
35																
40																
4 5																
50																
55																

	(2)	INF	ORMA	TION	FOR	SEQ	ID	NO:	5:								
5		(i	(. ()	A) L B) T C) S	CE C ENGT YPE: TRAN	H: 8 nuc DEDN	94 b leic ESS:	ase aci dou	pair d	S							
10		(vi			AL S			illu	s st	earo	ther	moph	ilus				
15		(ix	() (1	B) L	AME/ OCAT THER	ION:	1 ORMA	TION		unct ldip					DNA		
		(xi)) SE	QUEN	CE D	ESCR	IPTI	ON:	SEQ	ID N	O: 5	:					
20		GCG Ala														4	8 8
25		G AA Glu														9	96
30		CTG Leu													CGA Arg	14	: 4
		CGT Arg 50														19	12
35		GCG Ala														. 24	:0
40		TCT Ser														28	8
45		CGC Arg														33	6
		TTG Leu														38	4
50		ATC Ile 130														43	2

5				AAA Lys							480
	_	_		GAA Glu 165							528
10				CGG A rg							576
15				TTG Leu							624
20				GCC Ala							672
25				ATT Ile							720
23				AGC Ser 245							768
30				AAG Lys							816
35				AAC Asn							864
40				GCC Ala			TAA				894

	(2)	INF	ORMA'	TION	FOR	SEQ	ID I	NO:	6:							
5			(1	A) Li B) T	ENCE ENGTI YPE:	H: 2:	97 ai	mino cid								
		(ii) MOI	LECUI	LE T	YPE:	pro	tein								
10		(xi) SE(QUEN	CE DI	ESCR:	IPTI	ON:	SEQ :	ID N	0: 6	:				
	Met 1	Ala	Gln	Leu	Ser 5	Val	Glu	Gln	Phe	Leu 10	Asn	Glu	Gln	Lys	Gln 15	Ala
15	Val	Glu	Thr	Ala 20	Leu	Ser	Arg	Tyr	Ile 25	Glu	Arg	Leu	Glu	Gly 30	Pro	Ala
	Lys	Leu	Lys 35	Lys	Ala	Met	Ala	Tyr 40	Ser	Leu	Glu	Ala	Gly 45	Gly	Lys	Arg
20	Ile	Arg 50	Pro	Leu	Leu	Leu	Leu 55	Ser	Thr	Val	Arg	Ala 60	Leu	Gly	Lys	Asp
	Pro 65	Ala	Val	Gly	Leu	Pro 70	Val	Ala	Cys	Ala	Ile 75	Glu	Met	Ile	His	Thr 80
25	Tyr	Ser	Leu	Ile	His 85	Asp	Asp	Leu	Pro	Ser 90	Met	Asp	Asn	Asp	Asp 95	Leu
	Arg	Arg	Gly	Lys 100	Pro	Thr	Asn	His	Lys 105	Val	Phe	Gly	Glu	Ala 110	Met	Ala
30	Ile	Leu	Ala 115	Gly	Asp	Gly	Leu	Leu 120	Thr	Tyr	Ala	Phe	Gln 125	Leu	Ile	Thr
	Glu	Ile 130	Asp	Asp	Glu	Arg	Ile 135	Pro	Pro	Ser	Val	Arg 140	Leu	Arg	Leu	Ile
35	Glu 145	Arg	Leu	Ala	Lys	Ala 150	Ala	Gly	Pro	Glu	Gly 155	Met	Ala	Ala	Gly	Gln 160
40	Ala	Ala	Asp		Glu 165	Gly	Glu	Gly	-	Thr 170		Thr	Leu	Ser	Glu 175	
40	Glu	Tyr	Ile	His 180	Arg	Tyr	Lys	Thr	Gly 185	Lys	Met	Leu	Gln	Tyr 190	Ser	Val
45	His	Ala	Gly 195	Ala	Leu	Ile	Gly	Gly 200	Ala	Asp	Ala	Arg	Gln 205	Thr	Arg	Glu
	Leu	Asp 210	Glu	Phe	Ala	Ala	His 215	Leu	Gly	Leu	Ala	Phe 220	Gln	Ile	Arg	Asp
50	Asp 225	Ile	Leu	Asp	Ile	Glu 230	Gly	Ala	Glu	Glu	Lys 235	Ile	Gly	Lys	Pro	V al 240
	Gly	Ser	Asp	Gln	Ser 245	Asn	Asn	Lys	Ala	Thr 250	Tyr	Pro	Ala	Leu	Leu 255	Ser
55																

	Leu	Ala	Gly	Ala 260	Lys	Gļu	Lys	Leu	Ala 265	Phe	His	Ile	Glu	Ala 270	Ala	Gln
5	Arg	His	Leu 275	Arg	Asn	Ala	Asp	Val 280	Asp	Gly	Ala	Ala	Leu 285	Ala	Tyr	Ile
10	Cys	Glu 290	Leu	Val	Ala	Ala	Arg 295	Asp	His							
15																
20																
25																
30																
35																
40																
4 5																
50																
55																

	(2)	INF	ORMA	TION	FOR	SEQ	ID :	NO:	7:							-	
5	 (i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 894 base pairs (B) TYPE: nucleic acid (C) STRANDEDNESS: double (D) TOPOLOGY: linear (vi) ORIGINAL SOURCE: 																
10		(vi					E: Bac	illu	s st	earo	ther	moph	ilus				
15	<pre>(ix) FEATURE:</pre>																
20		(xi)	SE	QUEN	CE D	ESCR	IPTI(ON:	SEQ :	ID N	0: 7	:					
20		GCG Ala															48
25		GAA Glu															96
30		CTG Leu														CGA Arg	144
		CGT Arg 50															192
35		GCG Ala															240
40		TCT Ser															288
4 5		CGC Arg													_		336
		TTG Leu															384
50		ATC Ile 130															432

5			AAA Lys							480
			GAA Glu 165							528
10			CGG A rg							576
15			TTG Leu							624
20			GCC Ala							672
25			ATT Ile							720
			AGC Ser 245							768
30			AAG Lys							816
35			AAC Asn							864
40			GCC Ala			TAA				894

	(2)	INF	OR MA T	rion	FOR	SEQ	ID I	4O: 8	3:							
5			(I	SEQUI A) Li B) T' C) T(ENGTI	H: 29	97 ar	mino cid								
		(ii)	MOI	LECUI	LE TY	PE:	prot	ein								
10		(xi)	SEÇ	QUENC	CE DI	ESCR	PTI	ON: S	SEQ :	ID NO	D: 8	:				
	Met 1	Ala	Gln	Leu	Ser 5	Val	Glu	Gln	Phe	Leu 10	Asn	Glu	Gln	Lys	Gln 15	Ala
15	Val	Glu	Thr	Ala 20	Leu	Ser	Arg	Tyr	Ile 25	Glu	Arg	Leu	Glu	Gly 30	Pro	Ala
	Lys	Leu	Lys 35	Lys	Ala	Met	Ala	Tyr 40	Ser	Leu	Glu	Ala	Gly 45	Gly	Lys	Arg
20	Ile	Arg 50	Pro	Leu	Leu	Leu	Leu 55	Ser	Thr	Val	Arg	Ala 60	Leu	Gly	Lys	Asp
	Pro 65	Ala	Val	Gly	Leu	Pro 70	Val	Ala	Cys	Ala	Ile 75	Glu	Met	Ile	His	Thr 80
25	His	Ser	Leu	Ile	His 85	Asp	Asp	Leu	Pro	Ser 90	Met	Asp	Asn	Asp	Asp 95	Leu
	Arg	Arg	Gly	Lys 100	Pro	Thr	Asn	His	Lys 105	Val	Phe	Gly	Glu	Ala 110	Met	Ala
30	Ile	Leu	Ala 115	Gly	Asp	Gly	Leu	Leu 120	Thr	Tyr	Ala	Phe	Gln 125	Leu	Ile	Thr
	Glu	Ile 130	Asp	Asp	Glu	Arg	Ile 135	Pro	Pro	Ser	Val	Arg 140	Leu	Arg	Leu	Ile
35	Glu 145	Arg	Leu	Ala	Lys	Ala 150	Ala	Gly	Pro	Glu	Gly 155	Met	Val	Ala	Gly	Gln 160
	Ala	Ala	Asp	Met	Glu 165	Gly	Glu	Gly	Lys	Thr 170	Leu	Thr	Leu	Ser	Glu 175	Leu
40	Glu	Tyr	Ile	His 180	Arg	His	Lys	Thr	Gly 185	Lys	Met	Leu	Gln	Tyr 190	Ser	Val
45	His	Ala	45	Ala	Leu	Ile	Gly	Gly 200	Ala	Asp	Ala	Arg	Gln 205	Thr	Arg	Glu
 -	Leu	Asp 210	Glu	Phe	Ala	Ala	His 215	Leu	Gly	Leu	Ala	Phe 220	Gln	Ile	Arg	Asp
50	Asp 225	Ile	Leu	Asp	Ile	Glu 230	Gly	Ala	Glu	Glu	Lys 235	Ile	Gly	Lys	Arg	Val 240
	Gly	Ser	Asp	Gln	Ser 245	Asn	Asn	Lys	Ala	Thr 250	Tyr	Pro	Ala	Leu	Leu 255	Ser

	Leu	Ala	Gly	Ala 260	Lys	Glu	Lys	Leu	Thr 265	Phe	His	Ile	Glu	Ala 270	Ala	Gln
5	Arg	His	Leu 275	Arg	Asn	Ala	Asp	Val 280	Asp	Gly	Ala	Ala	Leu 285	Ala	Tyr	Ile
10	Cys	Glu 290	Leu	Val	Ala	Ala	Arg 295	Asp	His							
15																
20																
25																
30																
35																
40																
45																
50																
55																

	(2)	INF	ORMA	TION	FOR	SEQ	ID.	NO:	9:									
5		(i	(. (A) L B) T C) S	CE CI ENGTI YPE: TRANI	H: 8 nuc DEDN	94 b leic ESS:	ase aci dou	pair d	S								
10		(vi			AL SO			illu	s st	earo	ther	moph	ilus					
15		(ix	(. ()	B) L	AME/1 OCAT: THER	ION:	1 ORMA	TION	: /fi phato				A co	ding	for	native	÷	
		(xi) SE	QUEN	CE DI	ESCR:	IPTI	ON:	SEQ :	ID N	O: 9	:						
20														AAA Lys				48
25														GGG Gly 30				96
30														GGC Gly				144
														GGC Gly				192
35														ATC Ile				240
40														GAT Asp				288
45														GCG Ala 110				336
														TTG Leu				384
50														CGG Arg				432

5	CGG Arg									480
	GCC Ala									528
10	 TAC Tyr		 -						_	576
15	GCC Ala									624
20	 GAC Asp 210		 	_						672
05	ATT Ile									720
25	AGC Ser									768
30	GCC Ala									816
35	CAT His									864
40	GAA Glu 290					TAA				894

	(2)	INF	ORMA'	TION	FOR	SEQ	ID I	NO:	10:							
5			(1	A) Li B) T	ENCE ENGT YPE: OPOL	H: 2	97 ai	mino cid								
		(ii)) M O:	LECU:	LE T	YPE:	pro	tein								
10		(xi) SE	QUEN	CE D	ESCR	IPTI	ON:	SEQ	ID N	0: 1	0:				
	Met 1	Ala	Gln	Leu	Ser 5	Val	Glu	Gln	Phe	Leu 10	Asn	Glu	Gln	Lys	Gln 15	Ala
15	Val	Glu	Thr	Ala 20	Leu	Ser	Arg	Tyr	Ile 25	Glu	Arg	Leu	Glu	Gly 30	Pro	Ala
	Lys	Lys	Lys 35	Lys	Ala	Met	Ala	Tyr 40	Ser	Leu	Glu	Ala	Gly 45	Gly	Lys	Arg
20	Ile	Arg 50	Pro	Leu	Leu	Leu	Leu 55	Ser	Thr	Val	Gln	Ala 60	Leu	Gly	Lys	Asp
	Pro 65	Ala	Val	Gly	Leu	Pro 70	Val	Ala	Cys	Ala	Ile 75	Glu	Met	Ile	His	Thr 80
25	Tyr	Ser	Leu	Ile	His 85	Asp	Asp	Leu	Pro	Ser 90	Met	Asp	Asn	Asp	Asp 95	Leu
	Arg	Arg	Gly	Lys 100	Pro	Thr	Asn	His	L y s 105	Val	Phe	Gly	Glu	Ala 110	Met	Ala
30	Ile	Leu	Ala 115	Gly	Asp	Gly	Leu	Leu 120	Thr	Tyr	Ala	Phe	Gln 125	Leu	Ile	Thr
	Glu	Ile 130	Asp	Asp	Glu	Arg	Ile 135	Pro	Pro	Ser	Val	Arg 140	Leu	Arg	Leu	Ile
35	Glu 145	Arg	Leu	Ala	Lys	A la 150	Ala	Gly	Pro	Glu	Gly 155	Met	Val	Ala	Gly	Gln 160
40	Ala	Ala	Asp	Met	Glu 165	Gly	Glu	Gly	Lys	Thr 170		Thr	Leu	Ser	Glu 175	Leu
	Glu	Tyr	Ile	His 180	Arg	His	Lys	Thr	Gly 185	Lys	Met	Leu	Gln	Tyr 190	Ser	Val
45	His	Ala	Gly 195	Ala	Leu	Ile	Gly	Gly 200	Ala	Asp	Ala	A rg	Gln 205	Thr	Arg	Glu
	Leu	Asp 210	Glu	Phe	Ala	Ala	His 215	Leu	Gly	Leu	Ala	Phe 220	Gln	Ile	Arg	Asp
50	Asp 225	Ile	Leu	Asp	Ile	Glu 230	Gly	Ala	Glu	Glu	Lys 235	Ile	Gly	Lys	Pro	Val 240
	Gly	Ser	Asp	Gln	Ser 245	Asn	Asn	Lys	Ala	Thr 250	Tyr	Pro	Ala	Leu	Leu 255	Ser

15 Claims

20

30

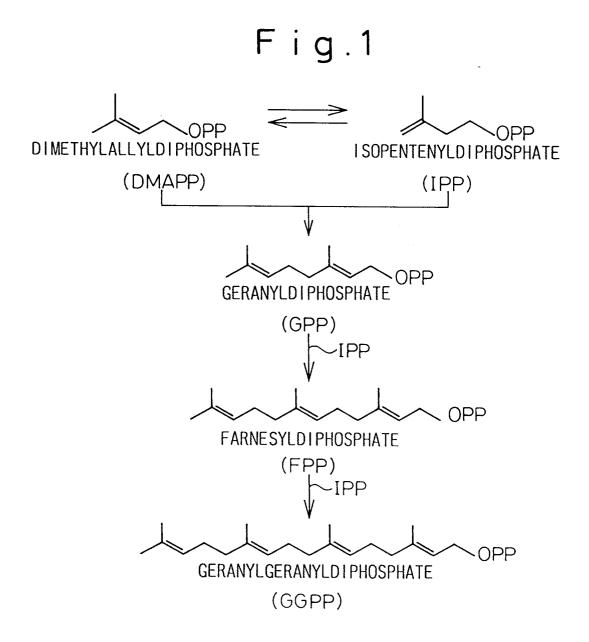
40

45

50

- 1. A process for production of a gene coding for a mutated farnesyldiphosphate synthase capable of synthesizing geranyldiphosphate synthase comprising the steps of:
 - (1) subjecting a gene coding for a farnesyldiphosphate synthase to mutagenesis;
 - (2) expressing the genes subjected to the mutagenesis; and
 - (3) selecting a gene coding for a mutated farnesyldiphosphate synthase capable of synthesizing geranylgeranyldiphosphate.
- 25 **2.** A process according to claim 1, wherein the gene coding for mutated farnesyldiphosphate synthase capable of synthesizing geranylgeranyldiphosphate is derived from <u>Bacillus</u> stearothermophilus.
 - **3.** A gene coding for a mutated farnesyldiphosphate synthase capable of synthesizing geranyldiphosphate synthase, obtainable according to a process of claim 1.
 - **4.** A process for production of a mutated farnesyldiphosphate synthase capable of synthesizing geranylgeranyldiphosphate, comprising the step of expressing a gene of claim 3.
- **5.** A mutated farnesyldiphosphate synthase capable of synthesizing geranylgeranyldiphosphate, obtainable according to a process of claim 4.
 - **6.** A process for production of geranylgeranyldiphosphate or geranylgeranyol, comprising the step of acting a mutated farnesyldiphosphate synthase capable of synthesizing geranylgeranyldiphosphate on a substrate selected from the group consisting of isopentenyldiphosphate, dimethylallyldiphosphate, geranyldiphosphate and farnesyldiphosphate.
 - 7. A geranylgeranyldiphosphate synthase having an amino acid sequence modified from a native amino acid sequence of a farnesyldiphosphate synthase wherein the modification comprises deletion of one to a few amino acid residues, addition of one to a few amino acid residues or replacement of one to a few amino acid residues with other amino acid residues, or a combination of said modification.
 - 8. A geranylgeranyldiphosphate synthase according to claim 7, wherein the modification is present on at least one of the positions 34, 59, 81, 157, 182, 239, 265 and 275 of farnesyldiphosphate synthase of Bacillus stearother-mophilus origin, or one to a few corresponding positions in an amino acid sequence of a farnesyldiphosphate synthase of other origin.
 - 9. A geranylgeranyldiphosphate synthase having an amino acid sequence shown in SEQ ID NO: 1.
 - 10. A geranylgeranyldiphosphate synthase having an amino acid sequence shown in SEQ ID NO: 2.
 - 11. A geranylgeranyldiphosphate synthase having an amino acid sequence shown in SEQ ID NO: 3.
 - 12. A geranylgeranyldiphosphate synthase having an amino acid sequence shown in SEQ ID NO: 4.

13. A gene coding for a geranylgeranyldiphosphate synthase according to claim 7. 14. A gene coding for a geranylgeranyldiphosphate synthase according to claim 8. **15.** A gene coding for a geranylgeranyldiphosphate synthase according to claim 9. **16.** A gene coding for a geranylgeranyldiphosphate synthase according to claim 10. 17. A gene coding for a geranylgeranyldiphosphate synthase according to claim 11. 10 18. A gene coding for a geranylgeranyldiphosphate synthase according to claim 12. 19. An expression vector comprising a gene according to claim 13. 20. An expression vector comprising a gene according to claim 14. 21. An expression vector comprising a gene according to claim 15. 22. An expression vector comprising a gene according to claim 16. 20 23. An expression vector comprising a gene according to claim 17. 24. An expression vector comprising a gene according to claim 18. 25. A recombinant host transformed with an expression vector according to claim 19. 26. A recombinant host transformed with an expression vector according to claim 20. 27. A recombinant host transformed with an expression vector according to claim 21. 30 28. A recombinant host transformed with an expression vector according to claim 22. 29. A recombinant host transformed with an expression vector according to claim 23. 30. A recombinant host transformed with an expression vector according to claim 24. 35 40 45 50 55



F i q.2

A YSLEAGGKRIRPL LLLST Q YGALLGGKRLRPF LVYAT LNY NTPGGKLNRGL SVVDT LEY NAIGGKYNRGL TVVVA LEY NTVGGKYNRGL TVVQT	B MIHT YSLIHDDLPSMDNDDLRRGKPIN H AYSLIHDDLPAMDDDDLRRGLPIC HLQ AYFLVADD MMDKSITRRGQP C WYKVPEVGEI LLQ AFFLVADD IMDSSLTRRGQ IC WYQKPGVGLD LLQ AFFLVLDD IMDSSHTRRGQI C WYQKPGIGLD	GQAADM EGEGKTLTLSE GQALDL DAEGKHVPLDA GQLMDL ITAPEDKVDLS GQTLDL LTAPQGNVDLV GQTLDL ITAPQGVDLG
MAQL SVEQFLNEQKQAVETAL SRYIERLEGPAKLKKAM MOFPQQLEACVKQANQAL SRFIAPL PFQNTPVVETM MASEKEIRRERFLNVFPKLVEELNASLLAYGMPKEACOWYAHS LNY NTPGGKLNRGL MNGDQNSDVYAQEKQDFVQHFSQIVRVLTEDEMGHPEIGDAIARLKEV MNGDQNSDVYAQEKQDFVQHFSQIVRVLTEDELGHPEKGDAITRIKEV LEY NTVGGKYNRGL	S C C A E C C VE	LLTYA FQLITEIDDERIPPSVRLRLIERLAKAAGPEGMVA GQAADM EGEGKTLTLSE TL A FSILSDADMPEVSDRDRISMISELASASGIAGMCG GQALDL ML EA AIYKLLKSHFRNEKYYIDITELFHEVTFQTEL CIYRLLKLYCREQPYYLNLIELFLQSSYQTEI CIYRLLKFYCREQPYYLNLELFLQSSYQTEI CATLOL ITAPQGQVDLG
MAQL SVEQFLNEQKÇ MDF PQQLEACVKQAN MASEKE I RRERFLNV MNGDQNSDVYAQEKÇ MNGDQNSDVYAQEKÇ	VRALGKDPAVGLPVA GHMFGVSTNTLDAPAAAVE C YAILSNKTVEQLGQEEYEKVAILGW C FRELVEPRKQDADSLQRAWTVGW C FQELVEPRKQDAESLQRALTVGW C	C A GDG LLTYA A GDALQTL A AINDAF ML EA AINDAN LL EA AINDAL LL EA
25,000	2000	2000

		ESI LEM LEM	YPALLSLAGAKEKLAFHIEAAQRHLRNADVDGAA TYPALLGLEQARKKARDLIODARQSLKQLAEQSLDTS WVINKALELASAEQRKTLDENYGKKDSVAEAKCKKIF WLVVQCLQRATPEQYQILKENYGQKAEKVARVKALYE WLVVQCLLRATPQQRQILEENYGQKDPEKVARVKALY	,	(1) B. STEAROTHERMOPHILAS (2) E. COL! (3) YEAST (4) HUMAN (5) RAT
F i g . 3	ш	(1) LEYIHRH KTGKMLQYSVHAG ALIG G ADAR QTRELDEFAAHL (2) LERIHRH KTGA LIRAAVRLGALS AG DKG RRALPVLDKYAESI (3) KFSLKKHSFIVTF KTAYYSFYLPVAL AMYVAGITDEK DLKQARDVLIPL (4) RFTEKRYKSIVKY KTAFYSFYLPIAA AMYMAGI D G EKEHANAKKILLEM (5) RYTEKRYKSIVKY KTAFYSFYLPIAA AMYMAGI D G EKEHANALKILLEM	(1) GLAFQIRDDILDIEGAEEKI GKPVGSD QSNNKAT YPALLSLAGAKEKLAFHIEAAQRHLRNADVDGAA (2) GLAFQVQDDILDVVGDTA TLGKRQGAD QQLGK S TYPALLGLEQARKKARDLIDDARQSLKQLAEQSLDTS (3) GEYFQIQDDYLDCFGTPEQI GKI GTDIQDN KCS WVINKALELASAEQRKTLDENYGKKDSVAEAKCKKIF (4) GEFFQIQDDYLDLFGDPSVT GKI GTDIQDN KCS WLVVQCLQRATPEQYQILKENYGQKAEKVARVKALYE (5) GEFFQIQDDYLDLFGDPSVT GKV GTDIQDN KCS WLVVQCLLRATPQQRQILEENYGQKDPEKVARVKALY	O	1) 2) A 2) A 3) NDLKIEQLYHEYEESIAKDLKAKISQVDESRGFKADV LTAFLN KVYKRSK 4) ELDLPAVFLQYEEDSYSHIMALIEQYAAPLPPAVF 5) EELDLRSVFFKYEEDSYNRLKSLIEQCSAPLPPSIF LE LANKIYKRRK

Fig.4

		2 34
W.T	1:	MAQLSVEQFLNEQKQAVETALSRYIERLEGPAKLKKAMAYSLEAGGKRIR
No.1	1:	
No.2	1:	V
No.3	1:	
No.4	1:	
		59 81
W.T	51:	PLLLLSTVRALGKDPAVGLPVACAIEMIHTYSLIHDDLPSMDNDDLRRGK
No.1	51:	H
No.2	51:	Q
No.3	51:	
No.4	51:	Н
		141
W.T	101:	PTNHKVFGEAMAILAGDGLLTYAFQLITEIDDERIPPSVRLRLIERLAKA
No.1	101:	
No. 2	101:	
No.3	101:	
No. 4	101:	
		157 182
₩.T		AGPEGMVAGQAADMEGEGKTLTLSELEYIHRHKTGKMLQYSVHAGALIGG
No.1	151:	
No.2	151:	
No. 2 No. 3	151 : 151 :	
No.2	151:	
No. 2 No. 3	151 : 151 :	
No. 2 No. 3 No. 4	151 : 151 : 151 :	239
No. 2 No. 3 No. 4	151 : 151 : 151 : 201 :	239 ADARQTRELDEFAAHLGLAFQIRDD1LDIEGAEEKIGKPVGSDQSNNKAT
No. 2 No. 3 No. 4 W. T No. 1	151 : 151 : 151 : 201 : 201 :	239 ADARQTRELDEFAAHLGLAFQIRDD1LDIEGAEEKIGKPVGSDQSNNKAT
No. 2 No. 3 No. 4 W. T No. 1	151 : 151 : 151 : 201 : 201 : 201 :	239 ADARQTRELDEFAAHLGLAFQIRDD1LDIEGAEEKIGKPVGSDQSNNKAT
No. 2 No. 3 No. 4 W.T No. 1 No. 2 No. 3	151 : 151 : 151 : 201 : 201 : 201 :	239 ADARQTRELDEFAAHLGLAFQIRDD1LDIEGAEEKIGKPVGSDQSNNKAT
No. 2 No. 3 No. 4 W. T No. 1	151 : 151 : 151 : 201 : 201 : 201 :	239 ADARQTRELDEFAAHLGLAFQIRDD1LDIEGAEEKIGKPVGSDQSNNKAT
No. 2 No. 3 No. 4 W.T No. 1 No. 2 No. 3	151 : 151 : 151 : 201 : 201 : 201 :	239 ADARQTRELDEFAAHLGLAFQIRDD1LDIEGAEEKIGKPVGSDQSNNKAT R
No. 2 No. 3 No. 4 W. T No. 1 No. 2 No. 3 No. 4	151 : 151 : 151 : 201 : 201 : 201 : 201 :	239 ADARQTRELDEFAAHLGLAFQIRDD1LDIEGAEEKIGKPVGSDQSNNKAT R 265 275
No. 2 No. 3 No. 4 W. T No. 1 No. 2 No. 3 No. 4	151 : 151 : 151 : 201 : 201 : 201 : 201 :	239 ADARQTRELDEFAAHLGLAFQIRDD1LDIEGAEEKIGKPVGSDQSNNKAT R 265 275 YPALLSLAGAKEKLAFHIEAAQRHLRNADVDGAALAYICELVAARDHX
No. 2 No. 3 No. 4 W. T No. 1 No. 2 No. 3 No. 4	151 : 151 : 151 : 201 : 201 : 201 : 201 : 251 :	239 ADARQTRELDEFAAHLGLAFQIRDDILDIEGAEEKIGKPVGSDQSNNKAT R 265 275 YPALLSLAGAKEKLAFHIEAAQRHLRNADVDGAALAYICELVAARDHX S
No. 2 No. 3 No. 4 W. T No. 1 No. 2 No. 3 No. 4	151 : 151 : 151 : 201 : 201 : 201 : 201 : 251 : 251 :	ADARQTRELDEFAAHLGLAFQIRDD1LDIEGAEEKIGKPVGSDQSNNKAT R 265 275 YPALLSLAGAKEKLAFHIEAAQRHLRNADVDGAALAYICELVAARDHX S
No. 2 No. 3 No. 4 W. T No. 1 No. 2 No. 3 No. 4	151 : 151 : 151 : 201 : 201 : 201 : 201 : 251 :	ADARQTRELDEFAAHLGLAFQIRDDILDIEGAEEKIGKPVGSDQSNNKAT R 265 275 YPALLSLAGAKEKLAFHIEAAQRHLRNADVDGAALAYICELVAARDHX S

Fig.5

