(19) Europäisches Patentamt
European Patent Office
Office européen des brevets

(11) EP 1 179 012 B9

(12) **CORRECTED EUROPEAN PATENT SPECIFICATION**
Note: Bibliography reflects the latest situation

(15) Correction information:
Corrected version no 1 (W1 B1)
Corrections, see page(s) 95-131

(48) Corrigendum issued on:
10.09.2003 Bulletin 2003/37

(45) Date of publication and mention of the grant of the patent:
23.10.2002 Bulletin 2002/43

(21) Application number: **00932570.5**

(22) Date of filing: **17.05.2000**

(51) Int Cl.[7]: **C07K 14/115**, C07K 14/135, C07K 14/155, C07K 14/16, A61K 38/16, A61P 31/12

(86) International application number:
PCT/US00/13651

(87) International publication number:
WO 00/069902 (23.11.2000 Gazette 2000/47)

(54) LONG LASTING FUSION PEPTIDE INHIBITORS OF VIRAL INFECTION

LANG WIRKENDE PEPTIDINHIBITOREN DER VIRUSFUSION MIT KÖRPERZELLEN BEI VIRALEN INFEKTIONEN

PEPTIDES HYBRIDES INHIBITEURS A ACTION PROLONGEE DES INFECTIONS VIRALES

(84) Designated Contracting States:
AT BE CH CY DE DK ES FI FR GB GR IE IT LI LU MC NL PT SE
Designated Extension States:
AL LT LV MK RO SI

(30) Priority: **17.05.1999 US 134406 P**
10.09.1999 US 153406 P

(43) Date of publication of application:
13.02.2002 Bulletin 2002/07

(60) Divisional application:
02014617.1 / 1 264 840

(73) Proprietor: **Conjuchem, Inc.**
Montréal, Québec H2X 3Y8 (CA)

(72) Inventors:
- **Bridon, Dominique P.**
Outremont, Quebec H2V 2B2 (CA)
- **Dufresne, Robert S.**
Wellesley, MA 02181 (US)
- **Boudjellab, Nissab**
Dorval, Quebec H9S 3X1 (CA)
- **Robitaille, Martin**
Montreal, Quebec H2X 3Y8 (CA)
- **Milner, Peter G.**
Los Altos Hills, CA 94022 (US)

(74) Representative: **Roques, Sarah Elizabeth et al**
J.A. Kemp & Co.
14 South Square
Gray's Inn
London WC1R 5JJ (GB)

(56) References cited:
EP-A- 0 602 290
WO-A-95/10302
WO-A-99/24074
WO-A-99/24075
WO-A-99/48536
US-A- 5 614 487

EP 1 179 012 B9

Description

FIELD OF THE INVENTION

[0001] This invention relates to modified peptides that are inhibitors of viral activity and/or exhibit antifusogenic properties. In particular, this invention relates to modified peptide inhibitors of human immunodeficiency virus (HIV), respiratory syncytial virus (RSV), human parainfluenza virus (HPV), measles virus (MeV), and simian immunodeficiency virus (SIV) with long duration of action for the treatment of the respective viral infections. The invention also relates to conjugates of the modified peptides and endogenous carriers, particularly conjugates of the modified peptides and various mobile blood components, particularly mobile endogenous proteins.

BACKGROUND OF THE INVENTION

[0002] Membrane fusion events, while commonplace in normal cell biological processes, are also involved in a variety of disease states, including, for example the entry of enveloped viruses into cells. Peptides are known that inhibit or otherwise disrupt membrane fusion-associated events, including, for example, inhibiting retroviral transmission to uninfected cells. As an example, the synthetic peptides DP-107 and DP-178 derived from separate domains within the human immunodeficiency virus type 1 ("HIV-1") transmembrane ("TM") glycoprotein gp41, are potent inhibitors of HIV-1 infection and HIV induced cell-cell fusion.

[0003] Lambert, et al., "Peptides from Conserved Regions of Paramyxovirus Fusion (F) Proteins are Potent Inhibitors of Viral Fusion," Proc. Natl. Acad. Science U.S.A., March 5, 1996, Vol. 93 (5), pp. 2186-91, discloses that the synthetic peptides DP-107 and DP-178 (T-20), derived from separate domains within the human immunodeficiency virus type 1 (HIV-1) transmembrane (TM) protein, gp41, are potent inhibitors of HIV-1 infection and fusion. Using a computer searching strategy (computerized antiviral searching technology, C.A.S.T.) based on the predicted secondary structure of DP-107 and DP-178 (T-20), Lambert, et al. identified conserved heptad repeat domains analogous to the DP-107 and DP-178 regions of HIV-1 gp41 within the glycoproteins of other fusogenic viruses. Antiviral peptides derived from three representative paramyxoviruses, respiratory syncytial virus (RSV), human parainfluenza virus type 3 (HPIV-3), and measles virus (MV) blocked homologous virus-mediated syncytium formation and exhibited EC_{50} values in the range 0.015-0.250 μM. Moreover, these peptides were highly selective for the virus of origin.

[0004] U.S. Patent Nos. 6,013,263, 6,017,536 and 6,020,459 incorporated herein in their entirety, likewise disclose that the 36 amino acid peptide DP178 corresponding to amino acids 638 to 673 of gp41 from the HIV-1 isolate LAI (HIV-1_{LAI}), and the 38 amino acid peptide DP107 corresponding to amino acids 558-595 of gp41 from the HIV-1_{LAI}, both exhibit potent anti-HIV-1 activity.

[0005] While many of the anti-viral or anti-fusogenic peptides described in the art exhibit potent anti-viral and/or anti-fusogenic activity, these peptides suffer from short plasma half-lifes *in vivo,* primarily due to rapid serum clearance and peptidase and protease activity. This in turn greatly reduces the effective anti-viral activity of the peptides. There is therefore a need for a method of prolonging the half-life of existing anti-viral and/or anti-fusogenic peptides and providing for longer duration of action of these peptides *in vivo.*

SUMMARY OF THE INVENTION

[0006] The present invention meets these and other needs and is directed to modified peptides having anti-viral activity and/or anti-fusogenic activity. These modified peptides provide for an increased stability *in vivo* and a reduced susceptibility to peptidase or protease degradation. These modified peptides thereby minimize, e.g., the need for more frequent, or even continual, administration of the peptides. The products of varying embodiments of the present invention can be used, e.g., as a prophylactic against and/or treatment for infection of a number of viruses, including human immunodeficiency virus (HIV), human respiratory syncytial virus (RSV), human parainfluenza virus (HPV), measles virus (MeV) and simian immunodeficiency virus (SIV). Modification of other peptides involved in viral transfection (e. g., Hepatitis, Epstein Barr and other related viruses) is also within the scope of the invention.

[0007] This invention relates to chemically reactive modifications of peptides exhibiting anti-viral and/or anti-fusogenic activity such that the modified peptides can react with available functionalities on blood components to form stable covalent bonds. The reactive group is a maleimide which is reactive with a thiol group on a blood protein, including a mobile blood protein such as albumin. Accordingly, the invention provides a modified anti-viral peptide comprising: a peptide that exhibits anti-viral activity, and a maleimide group which is reactive with a thiol group on blood components to form stable covalent bonds. Typically, the blood component is serum albumin.

[0008] In particular, the invention relates to such chemically reactive modifications wherein the peptide is DP107 or DP178 or analogs thereof, including peptides comprised of amino acid sequences from other (non-HIV) viruses that correspond to the gp41 region of HIV from which DP107 and DP178 are derived and that exhibit anti-viral or anti-

fusogenic activity. More particularly, these peptides preferably exhibit anti-viral activity against, among others, human immunodeficiency virus (HIV), human respiratory syncytial virus (RSV), human parainfluenza virus (HPV), measles virus (MeV) and simian immunodeficiency virus (SIV). The invention also relates to such chemically reactive modifications of the peptides of SEQ ID NO:1 to SEQ ID NO:86. In particular, peptides selected from the group consisting of SEQ ID NO:1 to SEQ ID NO:9, SEQ ID NO:10 to SEQ ID NO:30, SEQ ID NO:31 to SEQ ID NO: 62, SEQ ID NO: 63 to SEQ ID NO:73 or SEQ ID NO: 74 to SEQ ID NO: 86.

[0009] The invention also relates to compositions for use in the prevention and/or treatment of viral infection comprising a peptide that exhibits anti-viral activity modified with a reactive group as described. More particularly, the invention relates to such compositions for use in the prevention and/or treatement of acquired immune deficiency syndrome (AIDS), human respiratory syncytial virus (RSV), human parainfluenza virus (HPV) and measles virus (MeV) comprising a peptide that exhibit anti-viral activity against HIV, RSV, HPV and MeV respectively, modified with a maleimide group which is reactive with a thiol group on blood components, to form stable covalent bonds. Compositions may also be made for the prevention/treatment of simian immunodeficiency virus (SIV).

BRIEF DESCRIPTION OF THE TABLES

[0010] The invention will be better understood by reference to the Tables, in which:

Table 1 lists the commonly occurring amino acids together with their one letter and three letter abbreviations, and common protecting groups.
Table 2 shows DP178 carboxy truncations.
Table 3 shows DP178 amino truncations.
Table 4 shows DP107 carboxy truncations.
Table 5 shows DP107 amino truncations.
Table 6 shows HIV-2_{NIHZ} DP178 analog carboxy truncations.
Table 7 shows HIV-2_{NIHZ} DP178 analog amino truncations.
Table 8 shows RSV F2 region DP107 analog carboxy truncations.
Table 9 shows RSV F2 region DP107 analog amino truncations.
Table 10 shows RSV F1 region DP178 analog carboxy truncations.
Table 11 shows RSV F I region DP178 analog amino truncations.
Table 12 shows HPV3 F1 region DP 178 analog carboxy truncations.
Table 13 shows HPV3 F1 region DP 178 analog amino truncations.
Table 14 shows HPV3 F1 region DP 107 analog carboxy truncations.
Table 15 shows HPV3 F1 region DP107 analog amino truncations.
Table 16 shows representative anti-RSV peptides.
Table 17 shows representative anti-HPV3 peptides.
Table 18 shows representative anti-SIV peptides.
Table 19 shows representative anti-MeV peptides.

BRIEF DESCRIPTION OF SEQUENCE LISTING

[0011] The invention will be better understood by reference to the Sequence Listing, in which:

SEQ ID NO:1 shows the peptide sequence of DP178.
SEQ ID NO:2 shows the peptide sequence of DP107
SEQ ID NO:3-9 show peptide sequences of certain DP178 analogs.
SEQ ID NO:10-30 show the peptide sequences of RSV F1 region and F2 region corresponding to DP178 and DP107, and representtive anti-RSV peptides;
SEQ ID NO:31-62 show the peptide sequences of HPIV3 F1 region corresponding to DP178 and DP107, and representative anti-HPIV3 peptides;
SEQ ID NO:63-73 show peptide sequences of SIV corresponding to DP178 and representative anti-SIV peptides; and
SEQ ID NO:74-78 show peptide sequences of MeV corresponding to DP178 and representative anti-MeV peptides.

DETAILED DESCRIPTION OF THE INVENTION

[0012] To ensure a complete understanding of the invention the following definitions are provided:

Anti-viral peptides: As used herein, anti-viral peptides shall refer to peptides that inhibit viral infection of cells, by, for example, inhibiting cell-cell fusion or free virus infection. The route of infection may involve membrane fusion, as occurs in the case of enveloped viruses, or some other fusion event involving viral and cellular structures. Peptides that inhibit viral infection by a particular virus may be referenced with respect to that particular virus, e. g., anti-HIV peptide, anti-RSV peptide, etc.

Antifusogenic peptides: Antifusogenic peptides are peptides demonstrating an ability to inhibit or reduce the level of membrane fusion events between two or more entities, e.g., virus-cell or cell-cell, relative to the level of membrane fusion that occurs in the absence of the peptide.

HIV and anti-HIV peptides: The human immunodeficiency virus (HIV), which is responsible for acquired immune deficiency syndrome (AIDS), is a member of the lentivirus family of retroviruses. There are two prevalent types of HIV, HIV-1 and HIV-2, with various strain of each having been identified. HIV targets CD-4+ cells, and viral entry depends on binding of the HIV protein gp41 to CD-4+ cell surface receptors. Anti-HIV peptides refer to peptides that exhibit anti-viral activity against HIV, including inhibiting CD-4+ cell infection by free virus and/or inhibiting HIV-induced syncytia formation between infected and uninfected CD-4+ cells.

SIV and anti-SIV peptides: Simian immunodeficiency viruses (SIV) are lentiviruses that cause acquired immunodeficiency syndrome (AIDS)-like illnesses in susceptible monkeys. Anti-SIV peptides are peptides that exhibit anti-viral activity against SIV, including inhibiting of infection of cells by the SIV virus and inhibiting syncytia formation between infected and uninfected cells.

RSV and anti-RSV peptides: Respiratory syncytial virus (RSV) is a respiratory pathogen, especially dangerous in infants and small children where it can cause bronchiolitis (inflammation of the small air passages) and pneumonia. RSVs are negative sense, single stranded RNA viruses and are members of the *Paramyxoviridae* family of viruses. The route of infection of RSV is typically through the mucous membranes by the respiratory tract, i.e., nose, throat, windpipe and bronchi and bronchioles. Anti-RSV peptides are peptides that exhibit anti-viral activity against RSV, including inhibiting mucous membrane cell infection by free RSV virus and syncytia formation between infection and uninfected cells.

HPV and anti-HPV peptides: Human parainfluenza virus (HPIV or HPV), like RSV, is another leading cause of respiratory tract disease, and like RSVs, are negative sense, single stranded RNA viruses that are members of the *Paramyxoviridae* family of viruses. There are four recognized serotypes of HPIV -- HPIV-1, HPIV-2, HPIV-3 and HPIV-4. HPIV-1 is the leading cause of croup in children, and both HPIV-1 and HPIV-2 cause upper and lower respiratory tract illnesses. HPIV-3 is more often associated with bronchiolitis and pneumonia. Anti-HPV peptides are peptides that exhibit anti-viral activity against HPV, including inhibiting infection by free HPV virus and syncytia formation between infected and uninfected cells.

MeV and anti-Mev peptides: Measles virus (VM or MeV) is an enveloped negative, single-stranded RNA virus belonging to the *Paramyxoviridae* family of viruses. Like RSV and HPV, MeV causes respiratory disease, and also produces an immuno-suppression responsible for additional, opportunistic infections. In some cases, MeV can establish infection of the brain leading to severe neurlogical complications. Anti-MeV peptides are peptides that exhibit anti-viral activity against MeV, including inhibiting infection by free MeV virus and syncytia formation between infected and uninfected cells.

DP-178 and DP178 analogs: Unless otherwise indicated explicitly or by context, DP-178 means the 36 amino acid DP-178 peptide corresponding to amino acid residues 638-673 of the gp41 glycoprotein of HIV-1 isolate LAI (HIV_{LAI}) and having the sequence:

YTSLIHSLIEESQNQQEKNEQELLELDKWASLWNWF (SEQ ID NO:1)

as well as truncations, deletions and/or insertions thereof. Truncations of the DP178 peptide may comprise peptides of between 3-36 amino acids. Deletions consist of the removal of one or more amino acid residues from the DP178 peptide, and may involve the removal of a single contiguous portion of the peptide sequence or multiple portions. Insertions may comprise single amino acid residues or stretches of residues and may be made at the carboxy or amino terminal end of the DP178 peptide or at a position internal to the peptide.

DP178 peptide analogs are peptides whose amino acid sequences are comprised of the amino acid sequences of peptide regions of viruses other than HIV-1_{LAI} that correspond to the gp41 region from which DP178 was derived, as well as an truncations, deletions or insertions thereof. Such other viruses may include, but are not limited to, other HIV isolates such as HIV-2_{NIHZ}, respiratory syncytial virus (RSV), human parainfluenza virus (HPV), simian immunodeficiency virus (SIV), and measles virus (MeV). DP178 analogs also refer to those peptide sequences identified or recognized by the ALLMOTI5, 107x178x4 and PLZIP search motifs described in U.S. Patent Nos. 6,013,263, 6,017,536 and 6,020,459 and incorporated herein, having structural and/or amino acid motif similarity to DP178. DP178 analogs further refer to peptides described as "DP178-like" as that term is defined in U.S. Patent Nos. 6,013,263, 6,017,536 and 6,020,459.

DP-107 and DP107 analogs: Unless otherwise indicated explicitly or by context, DP-107 means the 38 amino acid DP-107 peptide corresponding to amino acid residues 558-595 of the gp41 protein of HIV-1 isolate LAI (HIV_{LAI}) and having the sequence:

NNLLRAIEAQQHLLQLTVWQIKQLQARILAVERYLKDQ (SEQ ID NO:2)

as well as truncations, deletions and/or insertions thereof. Truncations of the DP107 peptide may comprise peptides of between 3-38 amino acids. Deletions consist of the removal of one or more amino acid residues from the DP107 peptide, and may involve the removal of a single contiguous portion of the peptide sequence or multiple portions. Insertions may comprise single amino acid residues or stretches of residues and may be made at the carboxy or amino terminal end of the DP107 peptide or at a position internal to the peptide.

DP107 peptide analogs are peptides whose amino acid sequences are comprised of the amino acid sequences of peptide regions of viruses other than HIV-1_{LAI} that correspond to the gp41 region from which DP107 was derived, as well as truncations, deletions and/or insertions thereof. Such other viruses may include, but are not limited to, other HIV isolates such as HIV-2_{NIHZ}, respiratory syncytial virus (RSV), human parainfluenza virus (HPV), simian immunodeficiency virus (SIV), and measles virus (MeV). DP107 analogs also refer to those peptide sequences identified or recognized by the ALLMOTI5, 107x178x4 and PLZIP search motifs described in U.S. Patent Nos. 6,013,263, 6,017,536 and 6,020,459 and incorporated herein, having structural and/or amino acid motif similarity to DP107. DP107 analogs further refer to peptides described as "DP107-like" as that term is defined in U.S. Patent Nos. 6,013,263, 6,017,536 and 6,020,459.

Reactive Groups: Reactive groups are chemical groups capable of forming a covalent bond. Such reactive groups are coupled or bonded to a DP-107 or DP-178 peptide or analogs thereof or other anti-viral or anti-fusogenic peptide of interest. Reactive groups will generally be stable in an aqueous environment and will usually be carboxy, phosphoryl, or convenient acyl group, either as an ester or a mixed anhydride, or an imidate, thereby capable of forming a covalent bond with functionalities such as an amino group, a hydroxy or a thiol at the target site on mobile blood components. For the most part, the esters will involve phenolic compounds, or be thiol esters, alkyl esters, phosphate esters, or the like.

Functionalities: Functionalities are groups on blood components to which reactive groups on modified anti-viral peptides react to form covalent bonds. Functionalities include hydroxyl groups for bonding to ester reactive entities; thiol groups for bonding to maleimides, imidates and thioester groups; amino groups for bonding to carboxy, phosphoryl or acyl groups and carboxyl groups for bonding to amino groups.

Blood Components: Blood components may be either fixed or mobile. Fixed blood components are non-mobile blood components and include tissues, membrane receptors, interstitial proteins, fibrin proteins, collagens, platelets, endothelial cells, epithelial cells and their associated membrane and membraneous receptors, somatic body cells, skeletal and smooth muscle cells, neuronal components, osteocytes and osteoclasts and all body tissues especially those associated with the circulatory and lymphatic systems. Mobile blood components are blood components that do not have a fixed situs for any extended period of time, generally not exceeding 5, more usually one minute. These blood components are not membrane-associated and are present in the blood for extended periods of time and are present in a minimum concentration of at least 0.1 μg/ml. Mobile blood components include serum albumin, transferrin, ferritin and immunoglobulins such as IgM and IgG. The half-life of mobile blood components is at least about 12 hours.

Protective Groups: Protective groups are chemical moieties utilized to protect peptide derivatives from reacting

with themselves. Various protective groups are disclosed herein and in U.S. 5,493,007, which is hereby incorporated by reference. Such protective groups include acetyl, fluorenylmethyloxycarbonyl (Fmoc), t-butyloxycarbonyl (Boc), benzyloxycarbonyl (CBZ), and the like. The specific protected amino acids are depicted in Table 1.

TABLE 1

NATURAL AMINO ACIDS AND THEIR ABBREVIATIONS			
Name	3-Letter Abbreviation	1-Letter Abbreviation	Modified Amino Acids
Alanine	Ala	A	Fmoc-Ala-OH
Arginine	Arg	R	Fmoc-Arg(Pbf)-OH
Asparagine	Asn	N	Fmoc-Asn(Trt)-OH
Aspartic acid	Asp	D	Asp(tBu)-OH
Cysteine	Cys	C	Fmoc-Cys(Trt)
Glutamic acid	Glu	E	Fmoc-Glu(tBu)-OH
Glutamine	Gln	Q	Fmoc-Gln(Trt)-OH
Glycine	Gly	G	Fmoc-Gly-OH
Histidine	His	H	Fmoc-His(Trt)-OH
Isoleucine	Iie	I	Fmoc-Ile-OH
Leucine	Leu	L	Fmoc-Leu-OH
Lysine	Lys	Z	Boc-Lys(Aloc)-OH
Lysine	Lys	X	Fmoc-Lys(Aloc)-OH
Lysine	Lys	K	Fmoc-Lys(Mtt)-OH
Methionine	Met	M	Fmoc-Met-OH
Phenylalanine	Phe	F	Fmoc-Phe-OH
Proline	Pro	P	Fmoc-Pro-OH
Serine	Ser	S	Fmoc-Ser(tBu)-OH
Threonine	Thr	T	Fmoc-Thr(tBu)-OH
Tryptophan	Trp	W	Fmoc-Trp(Boc)-OH
Tyrosine	Tyr	Y	Boc-Tyr(tBu)-OH
Valine	Val	V	Fmoc-Val-OH

Linking Groups: Linking (spacer) groups are chemical moieties that link or connect reactive entities to antiviral or antifusogenic peptides. Linking groups may comprise one or more alkyl moeities, alkoxy moeity, alkenyl moeity, alkynyl moeity or amino moeity substituted by alkyl moeities, cycloalkyl moeity, polycyclic moeity, aryl moeity, polyaryl moeities, substituted aryl moeities, heterocyclic moeities, and substituted heterocyclic moeities. Linking groups may also comprise poly ethoxy amino acids, such as AEA ((2-amino) ethoxy acetic acid) or a preferred linking group AEEA ([2-(2-amino) ethoxy)] ethoxy acetic acid.

Sensitive Functional Groups - A sensitive functional group is a group of atoms that represents a potential reaction site on an antiviral and/or antifusogenic peptide. If present, a sensitive functional group may be chosen as the attachment point for the linker-reactive group modification. Sensitive functional groups include but are not limited to carboxyl, amino, thiol, and hydroxyl groups.

Modified Peptides - A modified peptide is an antiviral and/or antifusogenic peptide that has been modified by attaching a reactive group. The reactive group may be attached to the peptide either via a linking group, or optionally without using a linking group. It is also contemplated that one or more additional amino acids may be added to the peptide to facilitate the attachment of the reactive entity. Modified peptides may be administered *in vivo* such that conjugation with blood components occurs *in vivo*, or they may be first conjugated to blood components *in vitro*

and the resulting conjugated peptide (as defined below) administered *in vivo.*

Conjugated Peptides - A conjugated peptide is a modified peptide that has been conjugated to a blood component via a covalent bond formed between the reactive group of the modified peptide and the functionalities of the blood component, with or without a linking group. As used throughout this application, the term "conjugated peptide" can be made more specific to refer to particular conjugated peptides, for example "conjugated DP178" or "conjugated DP107."

Taking into account these definitions, the present invention takes advantage of the properties of existing anti-viral and antifusogenic peptides. The viruses that may be inhibited by the peptides include, but are not limited to all strains of viruses listed, e.g., in U.S. Patent Nos. 6,013,263, 6,017,536 and 6,020,459 at Tables V-VII and IX-XIV therein. These viruses include, e.g., human retroviruses, including HTV-1, HIV-2, and human T-lympocyte viruses (HTLV-I and HTLV-II), and non-human retroviruses, including bovine leukosis virus, feline sarcoma virus, feline leukemia virus, simian immunodeficiency virus (SIV), simian sarcoma virus, simian leukemia, and sheep progress pneumonia virus. Non-retroviral viruses may also be inhibited by the peptides of the present invention, including human respiratory syncytial virus (RSV), canine distemper virus, Newcastle Disease virus, human parainfluenza virus (HPIV), influenza viruses, measles viruses (MeV), Epstein-Barr viruses, hepatitis B viruses, and simian Mason-Pfizer viruses. Non-enveloped viruses may also be inhibited by the peptides of the present invention, and include, but are not limited to, picomaviruses such as polio viruses, hepatitis A virus, enteroviruses, echoviruses, coxsackie viruses, papovaviruses such as papilloma virus, parvoviruses, adenoviruses, and reoviruses.

[0013] As an example, the mechanism of action of HIV fusion peptides has been described as discussed in the background section of this application and antiviral and antifusogenic properties of the peptides have been well established. A synthetic peptide corresponding to the carboxyl-terminal ectodomain sequence (for instance, amino acid residues 643-678 of HIV-1 class B, of the LAI strain or residues 638-673 from similar strain as well as residues 558-595) has been shown to inhibit virus-mediated cell-cell fusion completely at low concentration. The fusion peptide competes with the leucine zipper region of the native viral gp41 thus resulting in the interference of the fusion/infection of the virus into the cell.

[0014] The focus of the present invention is to modify a selected anti-viral and/or antifusogenic peptide with the DAC (Drug Activity Complex) technology to confer to this peptide improved bio-availability, extended half-life and better distribution through selective conjugation of the peptide onto a protein carrier but without modifying the peptide's anti-viral properties. The carrier of choice (but not limited to) for this invention would be albumin conjugated through its free thiol by an anti-viral and/or antifusogenic peptide modified with a maleimide moiety.

[0015] Several peptide sequences have been described in the literature as highly potent for the prevention of HIV-1 fusion/infection. As examples, peptide DP178 binds to a conformation of gp41 that is relevant for fusion. Thus in one embodiment of the invention, DP178 and DP178-like peptides are modified. Likewise, other embodiments of the invention include modification of DP107 and DP107-like peptide for use against HIV, as well as peptides analagous to DP107 and DP178 that are found in RSV, HPV, MeV and SIV viruses.

1. DP178 and DP107

A. DP178 Peptides

[0016] The DP178 peptide corresponds to amino acid residues 638 to 673 of the transmembrane protein gp41 from the HIV-1_{LAI} isolate, and has the 36 amino acid sequence (reading from amino to carboxy terminus):

NH_2-YTSLIHSLIEESQNQQEKNEQELLELDKWASLWNWF-COOH (SEQ ID NO:1)

[0017] In addition to the full-length DP178 36-mer, the peptides of this invention include truncations of the DP178 peptide comprising peptides of between 3 and 36 amino acid residues (i.e., peptides ranging in size from a tripeptide to a 36-mer polypeptide), These truncated peptides are shown in Tables 2 and 3.

[0018] In addition amino acid substitutions of the DP178 peptide are also within the scope of the invention. HIV-1 and HIV-2 enveloped proteins are structurally distinct, but there exists a striking amino acid conservation within the DP178-corresponding regions of HIV-1 and HIV-2. The amino acid conservation is of a periodic nature, suggesting

some conservation of structure and/or function. Therefore, one possible class of amino acid substitutions would include those amino acid changes which are predicted to stabilize the structure of the DP178 peptides of the invention. Utilizing the DP178 and DP178 analog sequences described herein, the skilled artisan can readily compile DP178 consensus sequences and ascertain from these, conserved amino acid residues which would represent preferred amino acid substitutions.

[0019] The amino acid substitutions may be of a conserved or non-conserved nature. Conserved amino acid substitutions consist of replacing one or more amino acids of the DP178 peptide sequence with amino acids of similar charge, size, and/or hydrophobicity characteristics, such as, for example, a glutamic acid (E) to aspartic acid (D) amino acid substitution. Non-conserved substitutions consist of replacing one or more amino acids of the DP178 peptide sequence with amino acids possessing dissimilar charge, size, and/or hydrophobicity characteristics, such as, for example, a glutamic acid (E) to valine (V) substitution.

[0020] Amino acid insertions of DP178 may consist of single amino acid residues or stretches of residues. The insertions may be made at the carboxy or amino terminal end of the DP178 or DP178 truncated peptides, as well as at a position internal to the peptide.

[0021] Such insertions will generally range from 2 to 15 amino acids in length. It is contemplated that insertions made at either the carboxy or amino terminus of the peptide of interest may be of a broader size range, with about 2 to about 50 amino acids being preferred. One or more such insertions may be introduced into DP178 or DP178 truncations, as long as such insertions result in peptides which may still be recognized by the 107x178x4, ALLMOTI5 or PLZIP search motifs described above.

[0022] Preferred amino or carboxy terminal insertions are peptides ranging from about 2 to about 50 amino acid residues in length, corresponding to gp41 protein regions either amino to or carboxy to the actual DP178 gp41 amino acid sequence, respectively. Thus, a preferred amino terminal or carboxy terminal amino acid insertion would contain gp41 amino acid sequences found immediately amino to or carboxy to the DP178 region of the gp41 protein.

[0023] Deletions of DP178 or DP178 truncations are also within the scope of this invention. Such deletions consist of the removal of one or more amino acids from the DP178 or DP178-like peptide sequence, with the lower limit length of the resulting peptide sequence being 4 to 6 amino acids.

[0024] Such deletions may involve a single contiguous or greater than one discrete portion of the peptide sequences. One or more such deletions may be introduced into DP178 or DP178 truncations, as long as such deletions result in peptides which may still be recognized by the 107x178x4, ALLMOTI5 or PLZIP search motifs described above.

B. DP107 Peptides

[0025] DP107 is a 38 amino acid peptide which exhibits potent antiviral activity, and corresponds to residues 558 to 595 of HIV-1_{LAI} isolate transmembrane (TM) gp41 glycoprotein, as shown here:

NH_2-NNLLRAIEAQQHLLQLTVWQIKQLQARILAVERYLKDQ-COOH

(SEQ ID NO:2)

[0026] In addition to the full-length DP107 38-mer, the DP107 peptides include truncations of the DP107 peptide comprising peptides of between 3 and 38 amino acid residues (i.e., peptides ranging in size from a tripeptide to a 38-mer polypeptide), These peptides are shown in Tables 4 and 5, below.

[0027] In addition, amino acid substitutions of the DP178 peptide are also within the scope of the invention. As for DP178, there also exists a striking amino acid conservation within the DP107-corresponding regions of HIV-1 and HIV-2, again of a periodic nature, suggesting conservation of structure and/or function. Therefore, one possible class of amino acid substitutions includes those amino acid changes predicted to stabilize the structure of the DP107 peptides of the invention. Utilizing the DP107 and DP107 analog sequences described herein, the skilled artisan can readily compile DP107 consensus sequences and ascertain from these, conserved amino acid residues which would represent preferred amino acid substitutions.

[0028] The amino acid substitutions may be of a conserved or non-conserved nature. Conserved amino acid substitutions consist of replacing one or more amino acids of the DP107 peptide sequence with amino acids of similar charge, size, and/or hydrophobicity characteristics, such as, for example, a glutamic acid (E) to aspartic acid (D) amino acid substitution. Non-conserved substitutions consist of replacing one or more amino acids of the DP107 peptide sequence with amino acids possessing dissimilar charge, size, and/or hydrophobicity characteristics, such as, for example, a glutamic acid (E) to valine (V) substitution.

[0029] Amino acid insertions may consist of single amino acid residues or stretches of residues. The insertions may

be made at the carboxy or amino terminal end of the DP107 or DP107 truncated peptides, as well as at a position internal to the peptide.

[0030] Such insertions will generally range from 2 to 15 amino acids in length. It is contemplated that insertions made at either the carboxy or amino terminus of the peptide of interest may be of a broader size range, with about 2 to about 50 amino acids being preferred. One or more such insertions may be introduced into DP107 or DP107 truncations, as long as such insertions result in peptides which may still be recognized by the 107x178x4, ALLMOTI5 or PLZIP search motifs described above.

[0031] Preferred amino or carboxy terminal insertions are peptides ranging from about 2 to about 50 amino acid residues in length, corresponding to gp41 protein regions either amino to or carboxy to the actual DP107 gp41 amino acid sequence, respectively. Thus, a preferred amino terminal or carboxy terminal amino acid insertion would contain gp41 amino acid sequences found immediately amino to or carboxy to the DP107 region of the gp41 protein.

[0032] Deletions of DP107 or DP107 truncations are also within the scope of this invention. Such deletions consist of the removal of one or more amino acids from the DP107 or DP107-like peptide sequence, with the lower limit length of the resulting peptide sequence being 4 to 6 amino acids.

[0033] Such deletions may involve a single contiguous or greater than one discrete portion of the peptide sequences. One or more such deletions may be introduced into DP107 or DP107 truncations, as long as such deletions result in peptides which may still be recognized by the 107x178x4, ALLMOTI5 or PLZIP search motifs.

[0034] DP107 and DP107 truncations are more fully described in U.S. Patent No. 5,656,480, which is incorporated herein by reference in its entirety

2. DP107 and DP178 Analogs

[0035] Peptides corresponding to analogs of the DP178, DP178 truncations, DP107 and DP107 truncation sequences of the invention, described, above, may be found in other viruses, including, for example, non-HIV-1 enveloped viruses, non-enveloped viruses and other non-viral organisms.

[0036] Such DP178 and DP107 analogs may, for example, correspond to peptide sequences present in transmembrane ("TM") proteins of enveloped viruses and may, correspond to peptide sequences present in non enveloped and nonviral organisms. Such peptides may exhibit antifusogenic activity, antiviral activity, most particularly antiviral activity which is specific to the virus in which their native sequences are found, or may exhibit an ability to modulate intracellular processes involving coiled-coil peptide structures.

A. DP178 analogs

[0037] DP178 analogs are peptides whose amino acid sequences are comprised of the amino acid sequences of peptide regions of, for example, other (i.e., other than HIV-1) viruses that correspond to the gp41 peptide region from which DP178 was derived. Such viruses may include, but are not limited to, other HIV-1 isolates and HIV-2 isolates.

[0038] DP178 analogs derived from the corresponding gp41 peptide region of other (i.e., non HIV-1LAI) HIV-1 isolates may include, for example, peptide sequences as shown below.

NH2-YTNTIYTLLEESQNQQEKNEQELLELDKWASLWNWF-COOH (SEQ ID NO:3)

NH2-YTGIIYNLLEESQNQQEKNEQELLELDKWANLWNWF-COOH (SEQ ID NO:4)

NH2-YTSLIYSLLEKSQIQQEKNEQELLELDKWASLWNWF-COOH(SEQ ID NO:5)

[0039] The peptides of SEQ ID NO:3, SEQ ID NO:4 and SEQ ID NO:5 are derived from HIV-1_{SF2}, HIV-1_{RF}, and HIV-1_{MN}, respectively. Other DP178 analogs include those derived from HIV-2, including the peptides of SEQ ID NO:6 and

SEQ ID NO:7, which are derived from HIV-2_{ROD} and HIV-2_{NIHZ}, respectively. Still other useful analogs include the peptides of SEQ ID NO:8 and SEQ ID NO:9, which have been demonstrated to exhibit anti-viral activity.

[0040] In the present invention, it is preferred that the DP178 analogs represent peptides whose amino acid sequences correspond to the DP178 region of the gp41 protein, it is also contemplated that the peptides disclosed herein may, additionally, include amino sequences, ranging from about 2 to about 50 amino acid residues in length, corresponding to gp41 protein regions either amino to or carboxy to the actual DP178 amino acid sequence.

[0041] Table 6 and Table 7 show some possible truncations of the HIV-2_{NIHZ} DP178 analog, which may comprise peptides of between 3 and 36 amino acid residues (i.e., peptides ranging in size from a tripeptide to a 36-mer polypeptide). Peptide sequences in these tables are listed from amino (left) to carboxy (right) terminus.

B. Additional DP178 Analogs and DP107 Analogs

[0042] DP178 and DP107 analogs are recognized or identified, for example, by utilizing one or more of the 107x178x4, ALLMOTI5 or PLZIP computer-assisted search strategies described above. The search strategy identifies additional peptide regions which are predicted to have structural and/or amino acid sequence features similar to those of DP107 and/or DP178.

[0043] The search strategies are described fully in the example presented in Section 9 of US Patent Nos. 6,013,263, 6,017,536 and 6,020,459. While this search strategy is based, in part, on a primary amino acid motif deduced from DP107 and DP178, it is not based solely on searching for primary amino acid sequence homologies, as such protein sequence homologies exist within, but not between major groups of viruses. For example, primary amino acid sequence homology is high within the TM protein of different strains of HIV-1 or within the TM protein of different isolates of simian immunodeficiency virus (SIV).

[0044] The computer search strategy disclosed in US Patent Nos. 6,013,263, 6,017,536 and 6,020,459 successfully identified regions of proteins similar to DP107 or DP178. This search strategy was designed to be used with a commercially-available sequence database package, preferably PC/Gene.

[0045] In US Patent Nos. 6,013,263, 6,017,536 and 6,020,459, a series of search motifs, the 107x178x4, ALLMOTI5 and PLZIP motifs, were designed and engineered to range in stringency from strict to broad, with 107x178x4 being preferred. The sequences identified via such search motifs, such as those listed in Tables V-XIV, of US Patent Nos. 6,013,263, 6,017,536 and 6,020,459 and included in this application by incorporation by reference, potentially exhibit antifusogenic, such as antiviral, activity, may additionally be useful in the identification of antifusogenic, such as antiviral, compounds.

3. Other Anti-Viral Peptides

A. Anti-RSV Peptides

[0046] Anti-RSV peptides include DP178 and/or DP107 analogs identified from corresponding peptide sequences in RSV which have further been identified to inhibit viral infection by RSV. Such peptides of interest include the peptides of Table 16 and peptides of SEQ ID NO: 10 to SEQ ID NO:30. Of particular interest are the following peptides:

YTSVITIELSNIKENKCNGAKVKLIKQELDKYK (SEQ ID NO:14)
TSVITIELSNIKENKCNGAKVKLIKQELDKYKN (SEQ ID NO:15)
VITIELSNIKENKCNGAKVKLIKQELDKYKNAV (SEQ ID NO:16)

IALLSTNKAVVSLSNGVSVLTSKVLDLKNYIDK (SEQ ID NO:29)

The peptide of SEQ ID NO:10 is derived from the F2 region of RSV and was identified in U.S. Patent Nos. 6,103,236 and 6,020,459 using the search motifs described as corresponding to DP107 and DP178 peptides (i.e., "DP107/178 like"). The peptides of SEQ ID NO: 14 to SEQ ID NO: 16 each have amino acid sequences contained within the peptide of SEQ ID NO:10 and each has been shown to exhibit anti-RSV activity, in particular, inhibiting fusion and syncytia formation between RSV-infected and uninfected Hep-2 cells at concentrations of less than 50 μg/ml.

[0047] The peptide of SEQ ID NO:11 is derived from the F1 region of RSV and was identified in U.S. Patent Nos. 6,103,236 and 6,020,459 using the search motifs described as corresponding to DP107 (i.e., "DP107-like"). The peptide of SEQ ID NO:29 contains amino acid sequences contained within the peptide of SEQ ID NO:10 and likewise has been shown to exhibit anti-RSV activity, in particular, inhibiting fusion and syncytia formation between RSV-infected and uninfected Hep-2 cells at concentrations of less than 50 μg/ml.

[0048] Therefore in one embodiment of the invention, the peptide is selected from the group consisting of SEQ ID NO:14 to SEQ ID NO:17 and SEQ ID NO:29.

B. Anti-HPIV Peptides

[0049] Anti-HPIV peptides include DP178 and/or DP107 analogs identified from corresponding peptide sequences in HPIV and which have further been identified to inhibit viral infection by HPIV. Such peptides of interest include the peptides of Table 17 and SEQ ID NO:31 to SEQ ID NO:62. Of particular interest are the following peptides:

VEAKQARSDIEKLKEAIRDTNKAVQSVQSSIGNLI (SEQ ID NO:52)
RSDIEKLKEAIRDTNKAVQSVQSSIGNLIVAIKSV (SEQ ID NO:58)

NSVALDPIDISIELNKAKSDLEESKEWIRRSNQKL (SEQ ID NO:35)
ALDPIDISIELNKAKSDLEESKEWIRRSNQKLDSI (SEQ ID NO:38)
LDPIDISIELNKAKSDLEESKEWIRRSNQKLDSIG (SEQ ID NO:39)
DPIDISIELNKAKSDLEESKEWIRRSNQKLDSIGN (SEQ ID NO:40)
PIDISIELNKAKSDLEESKEWIRRSNQKLDSIGNW (SEQ ID NO:41)
IDISIELNKAKSDLEESKEWIRRSNQKLDSIGNWH (SEQ ID NO:42)

The peptide of SEQ ID NO:31 is derived from the F1 region of HPIV-3 and was identified in U.S. Patent Nos. 6,103,236 and 6,020,459 using the search motifs described as corresponding to DP107 (i.e., "DP107-like"). The peptides of SEQ ID NO:52 and SEQ ID NO:58 each have amino acid sequences contained within the peptide of SEQ ID NO:30 and each has been shown to exhibit anti-HPIV-3 activity, in particular, inhibiting fusion and syncytia formation between HPIV-3-infected Hep2 cells and uninfected CV-1W cells at concentrations of less than 1 μg/ml.

[0050] The peptide of SEQ ID NO:32 is also derived from the F1 region of HPIV-3 and was identified in U.S. Patent Nos. 6,103,236 and 6,020,459 using the search motifs described as corresponding to DP178 (i.e., "DP178-like"). The peptides of SEQ ID NO:35 and SEQ ID NO:38 to SEQ ID NO:42 each have amino acid sequences contained within the peptide of SEQ ID NO:32 and each also has been shown to exhibit anti-HPIV-3 activity, in particular, inhibiting fusion and syncytia formation between HPIV-3-infected Hep2 cells and uninfected CV-1W cells at concentrations of less than 1 μg/ml.

[0051] Therefore in one embodiment of the invention, the peptide is selected from the group consisting of SEQ ID NO: 35, SEQ ID NO:38 to SEQ ID NO:42, SEQ ID NO:52 and SEQ ID NO:58.

C. Anti-MeV Peptides

[0052] Anti-MeV peptides are DP178 and/or DP107 analogs identified from corresponding peptide sequences in measles virus (MeV) which have further been identified to inhibit viral infection by the measles virus. Such peptides of particular interest include the peptides of Table 19 and peptides of SEQ ID NO:74 to SEQ ID NO:86. Of particular interest are the peptides listed below.

HRIDLGPPISLERLDVGTNLGNAIAKLEAKELLE (SEQ ID NO:77)
IDLGPPISLERLDVGTNLGNAIAKLEAKELLESS (SEQ ID NO:79)
LGPPISLERLDVGTNLGNAIAKLEAKELLESSDQ (SEQ ID NO:81)
PISLERLDVGTNLGNAIAKLEAKELLESSDQILR (SEQ ID NO:84)

Sequences derived from measles virus were identified in U.S. Patent Nos. 6,103,236 and 6,020,459 using the search motifs described as corresponding to DP178 (i.e., "DP178-like"). The peptides of SEQ ID NO:77, SEQ ID NO:79, SEQ ID NO:81 and SEQ ID NO:83 each have amino acid sequences so identified, and each has been shown to exhibit anti-MeV activity, in particular, inhibiting fusion and syncytia formation between MeV-infected Hep2 and uninfected Vero cells at concentrations of less than 1 μg/ml.

[0053] Therefore in one embodiment of the invention, the peptide is selected from the group consisting of SEQ ID NO:77, SEQ ID NO:79, SEQ ID NO:81 and SEQ ID NO:84.

D. Anti-SIV Peptides

[0054] Anti-SIV peptides are DP 178 and/or DP107 analogs identified from corresponding peptide sequences in SIV which have further been identified to inhibit viral infection by SIV. Such peptides of interest include the peptides of Table 18 and peptides of SEQ ID NO:63 to SEQ ID NO:73. Of particular interest are the following peptides:

WQEWERKVDFLEENITALLEEAQIQQEKNMYELQK (SEQ ID NO:64)
QEWERKVDFLEENITALLEEAQIQQEKNMYELQKL (SEQ ID NO:65)
EWERKVDFLEENITALLEEAQIQQEKNMYELQKLN (SEQ ID NO:66)
WERKVDFLEENITALLEEAQIQQEKNMYELQKLNS (SEQ ID NO:67)
ERKVDFLEENITALLEEAQIQQEKNMYELQKLNSW (SEQ ID NO:68)
RKVDFLEENITALLEEAQIQQEKNMYELQKLNSWD (SEQ ID NO:69)
KVDFLEENITALLEEAQIQQEKNMYELQKLNSWDV (SEQ ID NO:70)
VDFLEENITALLEEAQIQQEKNMYELQKLNSWDVF (SEQ ID NO:71)
DFLEENITALLEEAQIQQEKNMYELQKLNSWDVFG (SEQ ID NO:72)
FLEENITALLEEAQIQQEKNMYELQKLNSWDVFGN (SEQ ID NO:73)

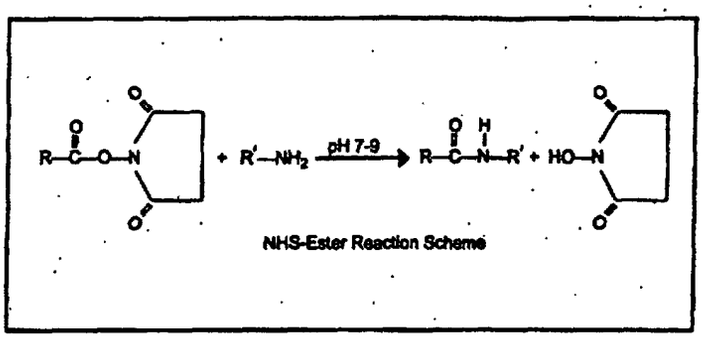
Sequences derived from SIV transmembrane fusion protein were identified in U.S. Patent Nos. 6,103,236 and 6,020,459 using the search motifs described as corresponding to DP178 (i.e., "DP178-like"). The peptides of SEQ ID NO:64 to SEQ ID NO:73 each have amino acid sequences so identified, and each has been shown to exhibit potent anti-SIV activity as crude peptides.

4. Modification of Anti-Viral and Antifusogenic Peptides

[0055] The invention contemplates modifying peptides that exhibit anti-viral and/or antifusogenic activity, including such modifications of DP-107 and DP-178 and analogs thereof. Such modified peptides can react with the available reactive functionalities on blood components via covalent linkages. The invention also relates to such modifications, such combinations with blood components, and methods for their use. These methods include extending the effective therapeutic life of the conjugated anti-viral peptides derivatives as compared to administration of the unconjugated peptides to a patient. The modified peptides are of a type designated as a DAC (Drug Affinity Complex) which comprises the anti-viral peptide molecule and a linking group together with a chemically reactive group capable of reaction with a reactive functionality of a mobile blood protein. By reaction with the blood component or protein the modified peptide, or DAC, may be delivered via the blood to appropriate sites or receptors.

[0056] The functionality on the protein will be a thiol group and the reactive group will be a maleimido-containing group such as gamma-maleimide-butyralamide (GMBA) or maleimidopropionic acid (MPA). The invention therefore provides a DAC wherein the maleimide group is coupled to the peptide via a linking group.

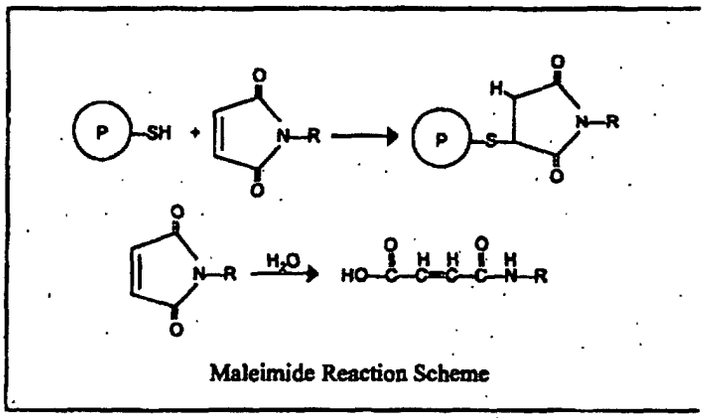
[0057] Primary amines are the principal targets for NHS esters. Accessible α-amine groups present on the N-termini of proteins react with NHS esters. However, α-amino groups on a protein may not be desirable or available for the NHS coupling. While five amino acids have nitrogen in their side chains, only the ε-amine of lysine reacts significantly with NHS esters. An amide bond is formed when the NHS ester conjugation reaction reacts with primary amines releasing N-hydroxysuccinimide as demonstrated in the schematic below.



NHS-Ester Reaction Scheme

[0058] The functional group on this protein will be a thiol group and the chemically reactive group will be a maleimido-

containing group such as MPA or GMBA (gamma-maleimide-butyralamide). The maleimido group is most selective for sulfhydryl groups on peptides when the pH of the reaction mixture is kept between 6.5 and 7.4. At pH 7.0, the rate of reaction of maleimido groups with sulfhydryls is 1000-fold faster than with amines. A stable thioether linkage between the maleimido group and the sulfhydryl is formed which cannot be cleaved under physiological conditions, as demonstrated in the following schematic.



A. Specific Labeling.

[0059] Preferably, the modified peptides of this invention are designed to specifically react with thiol groups on mobile blood proteins. Such reaction is preferably established by covalent bonding of the peptide modified with a maleimide link (e.g. prepared from GMBS, MPA or other maleimides) to a thiol group on a mobile blood protein such as serum albumin or IgG.

[0060] Under certain circumstances, specific labeling with maleimides offers several advantages over non-specific labeling of mobile proteins with groups such as NHS and sulfo-NHS. Thiol groups are less abundant *in vivo* than amino groups. Therefore, the maleimide-modified peptides of this invention, i.e., maleimide peptides, will covalently bond to fewer proteins. For example, in albumin (the most abundant blood protein) there is only a single thiol group. Thus, peptide-maleimide-albumin conjugates will tend to comprise approximately a 1:1 molar ratio of peptide to albumin. In addition to albumin, IgG molecules (class II) also have free thiols. Since IgG molecules and serum albumin make up the majority of the soluble protein in blood they also make up the majority of the free thiol groups in blood that are available to covalently bond to maleimide-modified peptides.

[0061] Further, even among free thiol-containing blood proteins, including IgGs, specific labeling with maleimides leads to the preferential formation of peptide-maleimide-albumin conjugates, due to the unique characteristics of albumin itself. The single free thiol group of albumin, highly conserved among species, is located at amino acid residue 34 (Cys^{34}). It has been demonstrated recently that the Cys^{34} of albumin has increased reactivity relative to free thiols on other free thiol-containing proteins. This is due in part to the very low pK value of 5.5 for the Cys^{34} of albumin. This is much lower than typical pK values for cysteine residues in general, which are typically about 8. Due to this low pK, under normal physiological conditions Cys^{34} of albumin is predominantly in the ionized form, which dramatically increases its reactivity. In addition to the low pK value of Cys^{34}, another factor which enhances the reactivity of Cys^{34} is its location, which is in a crevice close to the surface of one loop of region V of albumin. This location makes Cys^{34} very available to ligands of all kinds, and is an important factor in Cys^{34}'s biological role as free radical trap and free thiol scavenger. These properties make Cys^{34} highly reactive with maleimide-peptides, and the reaction rate acceleration can be as much as 1000-fold relative to rates of reaction of maleimide-peptides with other free-thiol containing proteins.

[0062] Another advantage of peptide-maleimide-albumin conjugates is the reproducibility associated with the 1:1 loading of peptide to albumin specifically at Cys^{34}. Other techniques, such as glutaraldehyde, DCC, EDC and other chemical activations of, e.g, free amines, lack this selectivity. For example, albumin contains 52 lysine residues, 25-30 of which are located on the surface of albumin and therefore accessible for conjugation. Activating these lysine residues, or alternatively modifying peptides to couple through these lysine residues, results in a heterogenous population of conjugates. Even if 1:1 molar ratios of peptide to albumin are employed, the yield will consist of multiple conjugation products, some containing 0, 1, 2 or more peptides per albumin, and each having peptides randomly coupled at any one or more of the 25-30 available lysine sites. Given the numerous possible combinations, characterization of the exact composition and nature of each conjugate batch becomes difficult, and batch-to-batch reproducibility is all but

impossible, making such conjugates less desirable as a therapeutic. Additionally, while it would seem that conjugation through lysine residues of albumin would at least have the advantage of delivering more therapeutic agent per albumin molecule, studies have shown that a 1:1 ratio of therapeutic agent to albumin is preferred. In an article by Stehle, et al., "The Loading Rate Determines Tumor Targeting properties of Methotrexate-Albumin Conjugates in Rats," Anti-Cancer Drugs, Vol. 8, pp. 677-685 (1988), incorporated herein in its entirety, the authors report that a 1:1 ratio of the anti-cancer methotrexate to albumin conjugated via glutaraldehyde gave the most promising results. These conjugates were preferentially taken up by tumor cells, whereas conjugates bearing 5:1 to 20:1 methotrexate molecules had altered HPLC profiles and were quickly taken up by the liver *in vivo.* It is postulated that at these higher ratios, conformational changes to albumin diminish its effectiveness as a therapeutic carrier.

[0063] Through controlled administration of maleimide-peptides *in vivo*, one can control the specific labeling of albumin and IgG *in vivo.* In typical administrations, 80-90% of the administered maleimide-peptides will label albumin and less than 5% will label IgG. Trace labeling of free thiols such as glutathione will also occur. Such specific labeling is preferred for *in vivo* use as it permits an accurate calculation of the estimated half-life of the administered agent.

[0064] In addition to providing controlled specific *in vivo* labeling, maleimide-peptides can provide specific labeling of serum albumin and IgG *ex vivo.* Such *ex vivo* labeling involves the addition of maleimide-peptides to blood, serum or saline solution containing serum albumin and/or IgG. Once conjugation has occurred *ex vivo* with the maleimide-peptides, the blood, serum or saline solution can be readministered to the patient's blood for *in vivo* treatment.

[0065] In contrast to NHS-peptides, maleimide-peptides are generally quite stable in the presence of aqueous solutions and in the presence of free amines. Since maleimide-peptides will only react with free thiols, protective groups are generally not necessary to prevent the maleimide-peptides from reacting with itself. In addition, the increased stability of the modified peptide permits the use of further purification steps such as HPLC to prepare highly purified products suitable for *in vivo* use. Lastly, the increased chemical stability provides a product with a longer shelf life.

5. Synthesis of Modified Anti-Viral and Anti-Fusogenic Peptides

A. Peptide Synthesis

[0066] Anti-viral and/or anti-fusogenic peptides according to the present invention may be synthesized by standard methods of solid phase peptide chemistry known to those of ordinary skill in the art. For example, peptides may be synthesized by solid phase chemistry techniques following the procedures described by Steward and Young (Steward, J. M. and Young, J. D., Solid Phase Peptide Synthesis, 2nd Ed., Pierce Chemical Company, Rockford, III., (1984) using an Applied Biosystem synthesizer. Similarly, multiple peptide fragments may be synthesized then linked together to form larger peptides. These synthetic peptides can also be made with amino acid substitutions at specific locations.

[0067] For solid phase peptide synthesis, a summary of the many techniques may be found in J. M. Stewart and J. D. Young, Solid Phase Peptide Synthesis, W. H. Freeman Co. (San Francisco), 1963 and J. Meienhofer, Hormonal Proteins and Peptides, vol. 2, p. 46, Academic Press (New York), 1973. For classical solution synthesis see G. Schroder and K. Lupke, The Peptides, Vol. 1, Acacemic Press (New York). In general, these methods comprise the sequential addition of one or more amino acids or suitably protected amino acids to a growing peptide chain. Normally, either the amino or carboxyl group of the first amino acid is protected by a suitable protecting group. The protected or derivatized amino acid is then either attached to an inert solid support or utilized in solution by adding the next amino acid in the sequence having the complimentary (amino or carboxyl) group suitably protected and under conditions suitable for forming the amide linkage. The protecting group is then removed from this newly added amino acid residue and the next amino acid (suitably protected) is added, and so forth.

[0068] After all the desired amino acids have been linked in the proper sequence, any remaining protecting groups (and any solid support) are removed sequentially or concurrently to afford the final polypeptide. By simple modification of this general procedure, it is possible to add more than one amino acid at a time to a growing chain, for example, by coupling (under conditions which do not racemize chiral centers) a protected tripeptide with a properly protected dipeptide to form, after deprotection, a pentapeptide.

[0069] A particularly preferred method of preparing compounds of the present invention involves solid phase peptide synthesis wherein the amino acid α-N-terminal is protected by an acid or base sensitive group. Such protecting groups should have the properties of being stable to the conditions of peptide linkage formation while being readily removable without destruction of the growing peptide chain or racemization of any of the chiral centers contained therein. Suitable protecting groups are 9-fluorenylmethyloxycarbonyl (Fmoc), t-butyloxycarbonyl (Boc), benzyloxycarbonyl (Cbz), biphenylisopropyloxycarbonyl ,t-amyloxycarbonyl, isobornyloxycarbonyl, α, α-dimethyl-3,5-dimethoxybenzyloxycarbonyl, o-nitrophenylsulfenyl, 2-cyano-t-butyloxycarbonyl, and the like. The 9-fluorenyl-methyloxycarbonyl (Fmoc) protecting group is particularly preferred for the synthesis of the peptides of the present invention. Other preferred side chain protecting groups are, for side chain amino groups like lysine and arginine, 2,2,5,7,8-pentamethylchroman-6-sulfonyl (pmc), nitro, p-toluenesulfonyl, 4-methoxybenzene-sulfonyl, Cbz, Boc, and adamantyloxycarbonyl; for tyrosine,

benzyl, o-bromobenzyloxycarbonyl, 2,6-dichlorobenzyl, isopropyl, t-butyl (t-Bu), cyclohexyl, cyclopenyl and acetyl (Ac); for serine, t-butyl, benzyl and tetrahydropyranyl; for histidine, trityl, benzyl, Cbz, p-toluenesulfonyl and 2,4-dinitrophenyl; for tryptophan, formyl; for asparticacid and glutamic acid, benzyl and t-butyl and for cysteine, triphenylmethyl (trityl).

[0070] In the solid phase peptide synthesis method, the α-C-terminal amino acid is attached to a suitable solid support or resin. Suitable solid supports useful for the above synthesis are those materials which are inert to the reagents and reaction conditions of the stepwise condensation-deprotection reactions, as well as being insoluble in the media used. The preferred solid support for synthesis of α-C-terminal carboxy peptides is 4-hydroxymethylphenoxymethyl-copoly (styrene-1% divinylbenzene). The preferred solid support for α-C-terminal amide peptides is the 4-(2',4'-dimethoxyphenyl-Fmoc-aminomethyl)phenoxyacetamidoethyl resin available from Applied Biosystems (Foster City, Calif.). The α-C-terminal amino acid is coupled to the resin by means ofN,N'-dicyclohexylcarbodiimide (DCC), N,N'-diisopropylcarbodiimide (DIC) or O-benzotriazol-1-yl-N,N,N',N'-tetramethyluronium-hexafluorophosphate (HBTU), with or without 4-dimethylaminopyridine (DMAP), 1-hydroxybenzotriazole (HOBT), benzotriazol-1-yloxy-tris(dimethylamino)phosphonium-hexafluorophosphate (BOP) or bis(2-oxo-3-oxazolidinyl)phosphine chloride (BOPCl), mediated coupling for from about 1 to about 24 hours at a temperature of between 10° and 50°C in a solvent such as dichloromethane or DMF.

[0071] When the solid support is 4-(2',4'-dimethoxyphenyl-Fmoc-aminomethyl)phenoxy-acetamidoethyl resin, the Fmoc group is cleaved with a secondary amine, preferably piperidine, prior to coupling with the α-C-terminal amino acid as described above. The preferred method for coupling to the deprotected 4-(2',4'-dimethoxyphenyl-Fmoc-aminomethyl)phenoxyacetamidoethyl resin is O-benzotriazol-1-yl-N,N,N',N'-tetramethyluroniumhexafluoro-phosphate (HBTU, 1 equiv.) and 1-hydroxybenzotriazole (HOBT, 1 equiv.) in DMF. The coupling of successive protected amino acids can be carried out in an automatic polypeptide synthesizer as is well known in the art. In a preferred embodiment, the α-N-terminal amino acids of the growing peptide chain are protected with Fmoc. The removal of the Fmoc protecting group from the α-N-terminal side of the growing peptide is accomplished by treatment with a secondary amine, preferably piperidine. Each protected amino acid is then introduced in about 3-fold molar excess, and the coupling is preferably carried out in DMF. The coupling agent is normally O-benzotriazol-1-yl-N,N,N',N'-tetramethyluroniumhexafluorophosphate (HBTU, 1 equiv.) and 1-hydroxybenzotriazole (HOBT, 1 equiv.).

[0072] At the end of the solid phase synthesis, the polypeptide is removed from the resin and deprotected, either in successively or in a single operation. Removal of the polypeptide and deprotection can be accomplished in a single operation by treating the resin-bound polypeptide with a cleavage reagent comprising thioanisole, water, ethanedithiol and trifluoroacetic acid. In cases wherein the α-C-terminal of the polypeptide is an alkylamide, the resin is cleaved by aminolysis with an alkylamine. Alternatively, the peptide may be removed by transesterification, e.g. with methanol, followed by aminolysis or by direct transamidation. The protected peptide may be purified at this point or taken to the next step directly. The removal of the side chain protecting groups is accomplished using the cleavage cocktail described above. The fully deprotected peptide is purified by a sequence of chromatographic steps employing any or all of the following types: ion exchange on a weakly basic resin (acetate form); hydrophobic adsorption chromatography on underivitized polystyrene-divinylbenzene (for example, Amberlite XAD); silica gel adsorption chromatography; ion exchange chromatography on carboxymethylcellulose; partition chromatography, e.g. on Sephadex G-25, LH-20 or countercurrent distribution; high performance liquid chromatography (HPLC), especially reverse-phase HPLC on octyl- or octadecylsilyl-silica bonded phase column packing.

[0073] Molecular weights of these ITPs are determined using Fast Atom Bombardment (FAB) Mass Spectroscopy.

(1) N-Terminal Protective Groups

[0074] As discussed above, the term "N-protecting group" refers to those groups intended to protect the α-N-terminal of an amino acid or peptide or to otherwise protect the amino group of an amino acid or peptide against undesirable reactions during synthetic procedures. Commonly used N-protecting groups are disclosed in Greene, "Protective Groups In Organic Synthesis," (John Wiley & Sons, New York (1981)), which is hereby incorporated by reference. Additionally, protecting groups can be used as pro-drugs which are readily cleaved *in vivo*, for example, by enzymatic hydrolysis, to release the biologically active parent. α-N-protecting groups comprise loweralkanoyl groups such as formyl, acetyl ("Ac"), propionyl, pivaloyl, t-butylacetyl and the like; other acyl groups include 2-chloroacetyl, 2-bromoacetyl, trifluoroacetyl, trichloroacetyl, phthalyl, o-nitrophenoxyacetyl, -chlorobutyryl, benzoyl, 4-chlorobenzoyl, 4-bromobenzoyl, 4-nitrobenzoyl and the like; sulfonyl groups such as benzenesulfonyl, p-toluenesulfonyl and the like; carbamate forming groups such as benzyloxycarbonyl, p-chlorobenzyloxycarbonyl, p-methoxybenzyloxycarbonyl, p-nitrobenzyloxycarbonyl, 2-nitrobenzyloxycarbonyl, p-bromobenzyloxycarbonyl, 3,4-dimethoxybenzyloxycarbonyl, 3,5-dimethoxybenzyloxycarbonyl, 2,4-dimethoxybenzyloxycarbonyl, 4-ethoxybenzyloxycarbonyl, 2-nitro-4,5-dimethoxybenzyloxycarbonyl, 3,4,5-trimethoxybenzyloxycarbonyl, 1-(p-biphenylyl)-1-methylethoxycarbonyl, α,α-dimethyl-3,5-dimethoxybenzyloxycarbonyl, benzhydryloxycarbonyl, t-butyloxycarbonyl (Boc), diisopropylmethoxycarbonyl, isopropyloxycarbonyl, ethoxycarbonyl, methoxycarbonyl, allyloxycarbonyl, 2,2,2,-trichloroethoxycarbonyl, phenoxycarbonyl, 4-nitrophenoxycarbonyl, fluorenyl-9-methoxycarbonyl, cyclopentyloxycarbonyl, adamantyloxycarbonyl, cy-

clohexyloxycarbonyl, phenylthiocarbonyl and the like; arylalkyl groups such as benzyl, triphenylmethyl, benzyloxymethyl, 9-fluorenylmethyloxycarbonyl (Fmoc) and the like and silyl groups such as trimethylsilyl and the like.

(2) Carboxy Protective Groups

[0075] As discussed above, the term "carboxy protecting group" refers to a carboxylic acid protecting ester or amide group employed to block or protect the carboxylic acid functionality while the reactions involving other functional sites of the compound are performed. Carboxy protecting groups are disclosed in Greene, "Protective Groups in Organic Synthesis" pp. 152-186 (1981), which is hereby incorporated by reference. Additionally, a carboxy protecting group can be used as a pro-drug whereby the carboxy protecting group can be readily cleaved *in vivo,* for example by enzymatic hydrolysis, to release the biologically active parent. Such carboxy protecting groups are well known to those skilled in the art, having been extensively used in the protection of carboxyl groups in the penicillin and cephalosporin fields as described in U.S. Pat. Nos. 3,840,556 and 3,719,667, the disclosures of which are hereby incorporated herein by reference. Representative carboxy protecting groups are C_1 -C_8 loweralkyl (e.g., methyl, ethyl or t-butyl and the like); arylalkyl such as phenethyl or benzyl and substituted derivatives thereof such as alkoxybenzyl or nitrobenzyl groups and the like; arylalkenyl such as phenylethenyl and the like; aryl and substituted derivatives thereofsuch as 5-indanyl and the like; dialkylaminoalkyl such as dimethylaminoethyl and the like); alkanoyloxyalkyl groups such as acetoxymethyl, butyryloxymethyl, valeryloxymethyl, isobutyryloxymethyl, isovaleryloxymethyl, 1-(propionyloxy)-1-ethyl, 1-(pivaloyloxyl)-1-ethyl, 1-methyl-1-(propionyloxy)-1-ethyl, pivaloyloxymethyl, propionyloxymethyl and the like; cycloalkanoyloxyalkyl groups such as cyclopropylcarbonyloxymethyl, cyclobutylcarbonyloxymethyl, cyclopentylcarbonyloxymethyl, cyclohexylcarbonyloxymethyl and the like; aroyloxyalkyl such as benzoyloxymethyl, benzoyloxyethyl and the like; arylalkylcarbonyloxyalkyl such as benzylcarbonyloxymethyl, 2-benzylcarbonyloxyethyl and the like; alkoxycarbonylalkyl or cycloalkyloxycarbonylalkyl such as methoxycarbonylmethyl, cyclohexyloxycarbonylmethyl, 1-methoxycarbonyl-1-ethyl and the like; alkoxycarbonyloxyalkyl or cycloalkyloxycarbonyloxyalkyl such as methoxycarbonyloxymethyl, t-butyloxycarbonyloxymethyl, 1-ethoxycarbonyloxy-1-ethyl, 1-cyclohexyloxycarbonyloxy-1-ethyl and the like; aryloxycarbonyloxyalkyl such as 2-(phenoxycarbonyloxy)ethyl, 2-(5-indanyloxycarbonyloxy)ethyl and the like; alkoxyalkylcarbonyloxyalkyl such as 2-(1-methoxy-2-methylpropan-2-oyloxy)ethyl and like; arylalkyloxycarbonyloxyalkyl such as 2-(benzyloxycarbonyloxy)ethyl and the like; arylalkenyloxycarbonyloxyalkyl such as 2-(3-phenylpropen-2-yloxycarbonyloxy)ethyl and the like; alkoxycarbonylaminoalkyl such as t-butyloxycarbonylaminomethyl and the like; alkylaminocarbonylaminoalkyl such as methylaminocarbonylaminomethyl and the like; alkanoylaminoalkyl such as acetylaminomethyl and the like; heterocycliccarbonyloxyalkyl such as 4-methylpiperazinylcarbonyloxymethyl and the like; dialkylaminocarbonylalkyl such as dimethylaminocarbonylmethyl, diethylaminocarbonylmethyl and the like; (5-(loweralkyl)-2-oxo-1,3-dioxolen-4-yl)alkyl such as (5-t-butyl-2-oxo-1,3-dioxolen-4-yl)methyl and the like; and (5-phenyl-2-oxo-1,3-dioxolen-4-yl)alkyl such as (5-phenyl-2-oxo-1,3-dioxolen-4-yl)methyl and the like.

[0076] Representative amide carboxy protecting groups are aminocarbonyl and loweralkylaminocarbonyl groups.

[0077] Preferred carboxy-protected compounds of the invention are compounds wherein the protected carboxy group is a loweralkyl, cycloalkyl or arylalkyl ester, for example, methyl ester, ethyl ester, propyl ester, isopropyl ester, butyl ester, sec-butyl ester, isobutyl ester, amyl ester, isoamyl ester, octyl ester, cyclohexyl ester, phenylethyl ester and the like or an alkanoyloxyalkyl, cycloalkanoyloxyalkyl, aroyloxyalkyl or an arylalkylcarbonyloxyalkyl ester. Preferred amide carboxy protecting groups are loweralkylaminocarbonyl groups. For example, aspartic acid may be protected at the α-C-terminal by an acid labile group (e.g. t-butyl) and protected at the β-C-terminal by a hydrogenation labile group (e.g. benzyl) then deprotected selectively during synthesis.

B. Peptide Modification

[0078] The manner of producing the modified peptides of the present invention will vary widely, depending upon the nature of the various elements comprising the peptide. The synthetic procedures will be selected so as to be simple, provide for high yields, and allow for a highly purified stable product. Normally, the chemically reactive group will be created at the last stage of the synthesis, for example, with a carboxyl group, esterification to form an active ester. Specific methods for the production of modified peptides of the present invention are described below.

[0079] Specifically, the selected peptide is first assayed for anti-viral activity, and then is modified with the linking group only at either the N-terminus, C-terminus or interior of the peptide. The anti-viral activity of this modified peptide-linking group is then assayed. If the anti-viral activity is not reduced dramatically (i.e., reduced less than 10-fold), then the stability of the modified peptide-linking group is measured by its *in vivo* lifetime. If the stability is not improved to a desired level, then the peptide is modified at an alternative site, and the procedure is repeated until a desired level of anti-viral and stability is achieved.

[0080] More specifically, each peptide selected to undergo modification with a linker and a reactive entity group will be modified according to the following criteria: if a terminal carboxylic group is available on the peptide and is not critical

for the retention of anti-viral activity, and no other sensitive functional group is present on the peptide, then the carboxylic acid will be chosen as attachment point for the linker-reactive group modification. If the terminal carboxylic group is involved in anti-viral activity, or if no carboxylic acids are available, then any other sensitive functional group not critical for the retention of anti-viral activity will be selected as the attachment point for the linker-reactive entity modification. If several sensitive functional groups are available on a a peptide, a combination of protecting groups will be used in such a way that after addition of the linker/reactive entity and deprotection of all the protected sensitive functional groups, retention of anti-viral activity is still obtained. If no sensitive functional groups are available on the peptide, or if a simpler modification route is desired, synthetic efforts will allow for a modification of the original peptide in such a way that retention of anti-viral is maintained. In this case the modification will occur at the opposite end of the peptide.

[0081] A maleimide derivative may also be synthesized from a peptide containing a free amino group and a free carboxylic acid. To produce a maleimide derivative from a amino derivatized molecule, one can use N-[γ-maleimidobutyryloxy]succinimide ester (GMBS) and triethylamine in DMF. The succinimide ester group will react with the free amino and the maleimide derivative will be purified from the reaction mixture by crystallization or by chromatography on silica or by HPLC.

[0082] Finally, a maleimide derivative may be synthesized from a peptide containing multiple other sensitive functional groups and no free carboxylic acids. When the selected molecule contains no carboxylic acid, an array of bifunctional crosslinking reagents can be used to convert the molecule into a reactive NHS derivative. For instance maleimidopropionic acid (MPA) can be coupled to the free amine to produce a maleimide derivative through reaction of the free amine with the carboxylic group of MPA using HBTU/HOBt/DIEA activation in DMF.

[0083] Many other commercially available heterobifunctional crosslinking reagents can alternatively be used when needed. A large number of bifunctional compounds are available for linking to entities. Illustrative reagents include: azidobenzoyl hydrazide, N-[4-(p-azidosalicylamino)butyl]-3'-[2'-pyridyldithio)propionamide), bis-sulfosuccinimidyl suberate, dimethyl adipimidate, disuccinimidyl tartrate, N-y-maleimidobutyryloxysuccinimide ester, N-hydroxy sulfosuccinimidyl-4-azidobenzoate, N-succinimidyl [4-azidophenyl]-1,3'-dithiopropionate, N-succinimidyl [4-iodoacetyl]aminobenzoate, glutaraldehyde, and succinimidyl 4-[N-maleimidomethyl]cyclohexane-1-carboxylate.

6. Uses of Modified Anti-Viral Peptides

[0084] Modified anti-viral peptides of the invention may be used as a therapeutic agent in the treatment of patients who are suffering from viral infection, and can be administered to patients according to the methods described below and other methods known in the art. Effective therapeutic dosages of the modified peptides may be determined through procedures well known by those in the art and will take into consideration any concerns over potential toxicity of the peptide.

[0085] The modified peptides can also be administered prophylactically to previously uninfected individuals. This can be advantageous in cases where an individual has been subjected to a high risk of exposure to a virus, as can occur when individual has been in contact with an infected individual where there is a high risk of viral transmission. This can be expecially advantageous where there is known cure for the virus, such as the HIV virus. As a example, prophylactic administration of a modified anti-HIV peptide would be advantageous in a situation where a health care worker has been exposed to blood from an HIV-infected individual, or in other situations where an individual engaged in high-risk activities that potentially expose that individual to the HIV virus.

7. Administration of Modified Anti-Viral and Anti-Fusogenic Peptides

[0086] Generally, the modified peptides will be administered in a physiologically acceptable medium, e.g. deionized water, phosphate buffered saline (PBS), saline, aqueous ethanol or other alcohol, plasma, proteinaceous solutions, mannitol, aqueous glucose, alcohol, vegetable oil, or the like. Other additives which may be included include buffers, where the media are generally buffered at a pH in the range of about 5 to 10, where the buffer will generally range in concentration from about 50 to 250 mM, salt, where the concentration of salt will generally range from about 5 to 500 mM, physiologically acceptable stabilizers, and the like. The compositions may be lyophilized for convenient storage and transport.

[0087] The subject modified peptides will for the most part be administered parenterally, such as intravenously (IV), intraarterially (IA), intramuscularly (IM), subcutaneously (SC), or the like. Administration may in appropriate situations be by transfusion. In some instances, where reaction of the functional group is relatively slow, administration may be oral, nasal, rectal, transdermal or aerosol, where the nature of the conjugate allows for transfer to the vascular system. Usually a single injection will be employed although more than one injection may be used, if desired. The modified peptides may be administered by any convenient means, including syringe, trocar, catheter, or the like.

[0088] The particular manner of administration will vary depending upon the amount to be administered, whether a single bolus or continuous administration, or the like. Preferably, the administration will be intravascularly, where the

site of introduction is not critical to this invention, preferably at a site where there is rapid blood flow, e.g., intravenously, peripheral or central vein. Other routes may find use where the administration is coupled with slow release techniques or a protective matrix. The intent is that the modified peptide be effectively distributed in the blood, so as to be able to react with the blood components. The concentration of the conjugate will vary widely, generally ranging from about 1 pg/ml to 50 mg/ml. The total administered intravascularly will generally be in the range of about 0.1 mg/ml to about 10 mg/ml, more usually about 1 mg/ml to about 5 mg/ml.

[0089] By bonding to long-lived components of the blood, such as immunoglobulin, serum albumin, red blood cells and platelets, a number of advantages ensue. The activity of the peptide is extended for days to weeks. Only one administration need be given during this period of time. Greater specificity can be achieved, since the active compound will be primarily bound to large molecules, where it is less likely to be taken up intracellularly to interfere with other physiological processes.

8. Monitoring the Presence of Modified Peptides

[0090] The blood of the mammalian host may be monitored for the presence of the modified peptide compound one or more times. By taking a portion or sample of the blood of the host, one may determine whether the peptide has become bound to the long-lived blood components in sufficient amount to be therapeutically active and, thereafter, the level of the peptide compound in the blood. If desired, one may also determine to which of the blood components the peptide is bound. This is particularly important when using non-specific modified peptides. For specific maleimide-modified peptides, it is much simpler to calculate the half life of serum albumin and IgG.

A. Immuno Assays

[0091] Another aspect of this invention relates to methods for determining the concentration of the anti-viral peptides and/or analogs, or their derivatives and conjugates in biological samples (such as blood) using antibodies specific for the peptides, peptide analogs or their derivatives and conjugates, and to the use of such antibodies as a treatment for toxicity potentially associated with such peptides, analogs, and/or their derivatives or conjugates. This is advantageous because the increased stability and life of the peptides in vivo in the patient might lead to novel problems during treatment, including increased possibility for toxicity.

[0092] The use of anti-therapeutic agent antibodies, either monoclonal or polyclonal, having specificity for a particular peptide, peptide analog or derivative thereof, can assist in mediating any such problem. The antibody may be generated or derived from a host immunized with the particular peptide, analog or derivative thereof, or with an immunogenic fragment of the agent, or a synthesized immunogen corresponding to an antigenic determinant of the agent. Preferred antibodies will have high specificity and affinity for native, modified and conjugated forms of the peptide, peptide analog or derivative. Such antibodies can also be labeled with enzymes, fluorochromes, or radiolables.

[0093] Antibodies specific for modified peptides may be produced by using purified peptides for the induction of peptide-specific antibodies. By induction of antibodies, it is intended not only the stimulation of an immune response by injection into animals, but analogous steps in the production of synthetic antibodies or other specific binding molecules such as screening of recombinant immunoglobulin libraries. Both monoclonal and polyclonal antibodies can be produced by procedures well known in the art.

[0094] The anti-peptide antibodies may be used to treat toxicity induced by administration of the modified peptide, analog or derivative thereof, and may be used ex vivo or in vivo. Ex vivo methods would include immuno-dialysis treatment for toxicity employing anti-therapeutic agent antibodies fixed to solid supports. In vivo methods include administration of anti-therapeutic agent antibodies in amounts effective to induce clearance of antibody-agent complexes.

[0095] The antibodies may be used to remove the modified peptides, analogs or derivatives thereof, and conjugates thereof, from a patient's blood ex vivo by contacting the blood with the antibodies under sterile conditions. For example, the antibodies can be fixed or otherwise immobilized on a column matrix and the patient's blood can be removed from the patient and passed over the matrix. The modified peptide, peptide analogs, derivatives or conjugates will bind to the antibodies and the blood containing a low concentration of peptide, analog, derivative or conjugate, then may be returned to the patient's circulatory system. The amount of peptide compound removed can be controlled by adjusting the pressure and flow rate.

[0096] Preferential removal of the peptides, analogs, derivatives and conjugates from the plasma component of a patient's blood can be effected, for example, by the use of a semipermeable membrane, or by otherwise first separating the plasma component from the cellular component by ways known in the art prior to passing the plasma component over a matrix containing the anti-therapeutic antibodies. Alternatively the preferential removal of peptide-conjugated blood cells, including red blood cells, can be effected by collecting and concentrating the blood cells in the patient's blood and contacting those cells with fixed anti-therapeutic antibodies to the exclusion of the serum component of the patient's blood.

[0097] The anti-therapeutic antibodies can be administered in vivo, parenterally, to a patient that has received the peptide, analogs, derivatives or conjugates for treatment. The antibodies will bind peptide compounds and conjugates. Once bound the peptide activity will be hindered if not completely blocked thereby reducing the biologically effective concentration of peptide compound in the patient's bloodstream and minimizing harmful side effects. In addition, the bound antibody-peptide complex will facilitate clearance of the peptide compounds and conjugates from the patient's blood stream.

[0098] The invention having been fully described can be further appreciated and understood with reference to the following non-limiting examples.

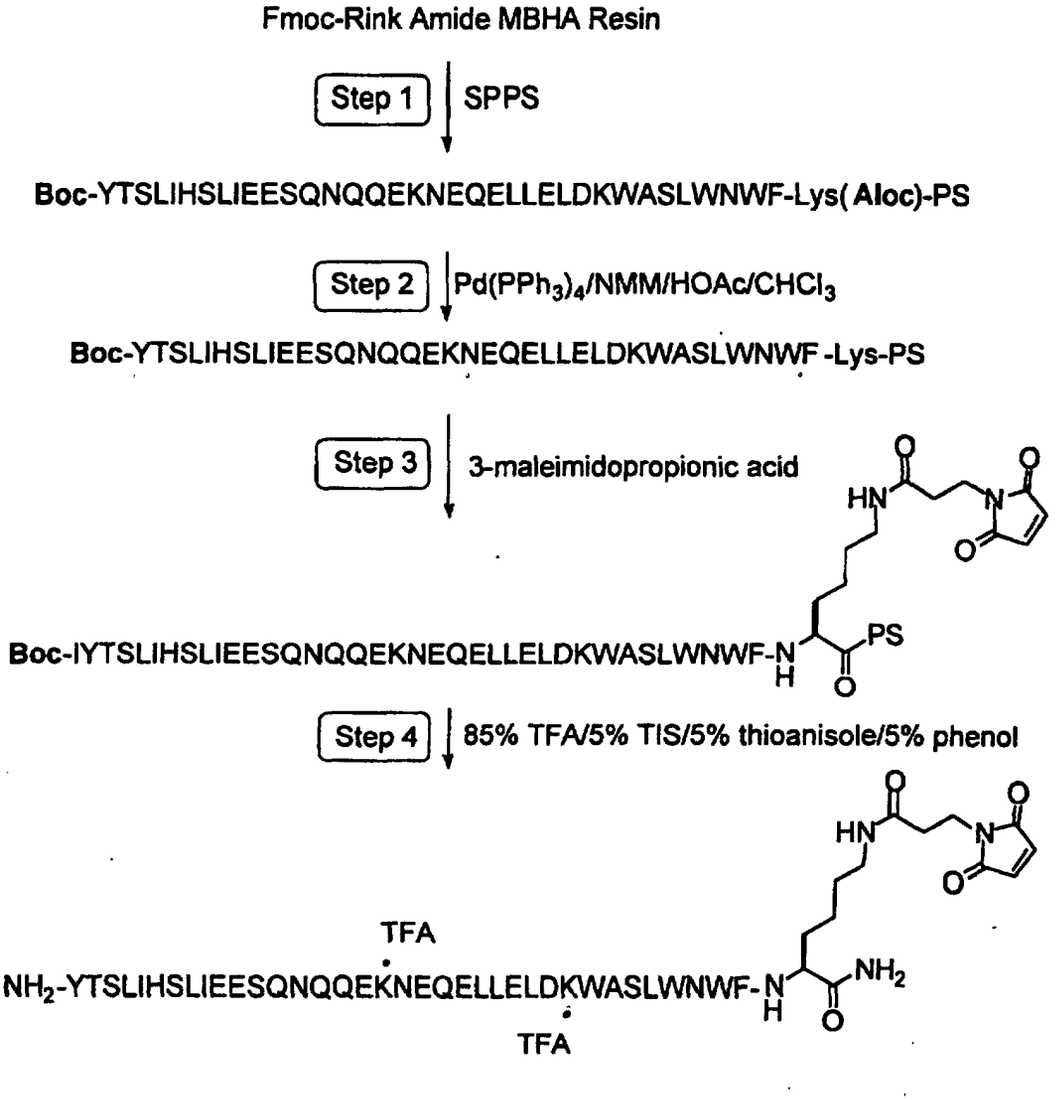
Example 1

Preparation of a Modified DP 178 --Synthesis of YTSLIHSLIEESQNQQEKNEQELLELDKWASLWNWFK(MPA)-NH_2

[0099] In this example, DP178 (SEQ ID NO:1) is synthesized and modified to include a linker and maleimide group according to the following synthesis scheme. As reported in U.S. Pat. Nos. 6,013,236 and 6,020,459, DP178 is a potent inhibitor of HIV-1, and inhibits both cell-induced syncytia formation between HIV-1 infected and uninfected cells and infection of uninfected cells be cell-free HIV-1 virus.

[0100] Solid phase peptide synthesis of the modified peptide on a 100 μmole scale is performed using manual solid-phase synthesis, a Symphony Peptide Synthesizer and Fmoc protected Rink Amide MBHA. The following protected amino acids are sequentially added to resin: Fmoc-Lys(Aloc)-OH, Fmoc-Phe-OH, Fmoc-Trp(Boc)-OH, Fmoc-Asn(Trt)-OH, Fmoc-Trp(Boc)-OH, Fmoc-Leu-OH, Fmoc-Ser(tBu)-OH, Fmoc-Ala-OH, Fmoc-Trp(Boc)-OH, Fmoc-Lys(Boc)-OH, Fmoc-Asp(tBu)-OH, Fmoc-Leu-OH, Fmoc-Glu(tBu)-OH, Fmoc-Leu-OH, Fmoc-Leu-OH, Fmoc-Glu(tBu)-OH, Fmoc-Gln(Trt)-OH, Fmoc-Glu(tBu)-OH, Fmoc-Met-OH, Fmoc-Lys(Boc)-OH, Fmoc-Glu(tBu)-OH, Fmoc-Gln(Trt)-OH, Fmoc-Asn(Trt)-OH, Fmoc-Lys(Boc)-OH, Fmoc-Glu(tBu)-OH; Fmoc-Gln(Trt)-OH, Fmoc-Gln(Trt)-OH, Fmoc-Asn(Trt)-OH, Fmoc-Gln(Trt)-OH, Fmoc-Ser(tBu)-OH, Fmoc-Glu(tBu)-OH, Fmoc-Glu(tBu)-OH, Fmoc-Ile-OH, Fmoc-Leu-OH, Fmoc-Ser(tBu)-OH, Fmoc-His(Boc)-OH, Fmoc-Ile-OH, Fmoc-Leu-OH, Fmoc-Ser(tBu)-OH, Fmoc-Thr(tBu)-OH, Fmoc-Tyr(tBu)-OH.They are dissolved in *N,N*-dimethylformamide (DMF) and, according to the sequence, activated using O-benzotriazol-1-yl-*N,N, N',N'*-tetramethyl-uronium hexafluorophosphate (HBTU) and Diisopropylethylamine (DIEA). Removal of the Fmoc protecting group is achieved using a solution of 20% (V/V) piperidine in *N,N*-dimethylformamide (DMF) for 20 minutes (step 1). At the end of the synthesis.The selective deprotection of the Lys (Aloc) group is performed manually and accomplished by treating the resin with a solution of 3 eq of $Pd(PPh_3)_4$ dissolved in 5 mL of $CHCl_3$:NMM: HOAc (18:1:0.5) for 2 h (Step 2). The resin is then washed with $CHCl_3$ (6 x 5 mL), 20% HOAc in DCM (6 x 5 mL), DCM (6 x 5 mL), and DMF (6 x 5 mL). The synthesis is then re-automated for the addition of the 3-maleimidopropionic acid (Step 3). Between every coupling, the resin is washed 3 times with *N,N*-dimethylformamide (DMF) and 3 times with isopropanol. The peptide is cleaved from the resin using 85% TFA/5% TIS/5% thioanisole and 5% phenol, followed by precipitation by dry-ice cold Et_2O (Step 4). The product is purified by preparative reversed phased HPLC using a Varian (Rainin) preparative binary HPLC system: gradient elution of 30-55% B (0.045% TFA in H_2O (A) and 0.045% TFA in CH_3CN (B)) over 180 min at 9.5 mL/min using a Phenomenex Luna 10 μ phenyl-hexyl, 21 mm x 25 cm column and UV detector (Varian Dynamax UVD II) at λ214 and 254 nm to afford the desired modified peptide (i.e., DAC) in >95% purity, as determined by RP-HPLC.

DP-178 C



Example 2

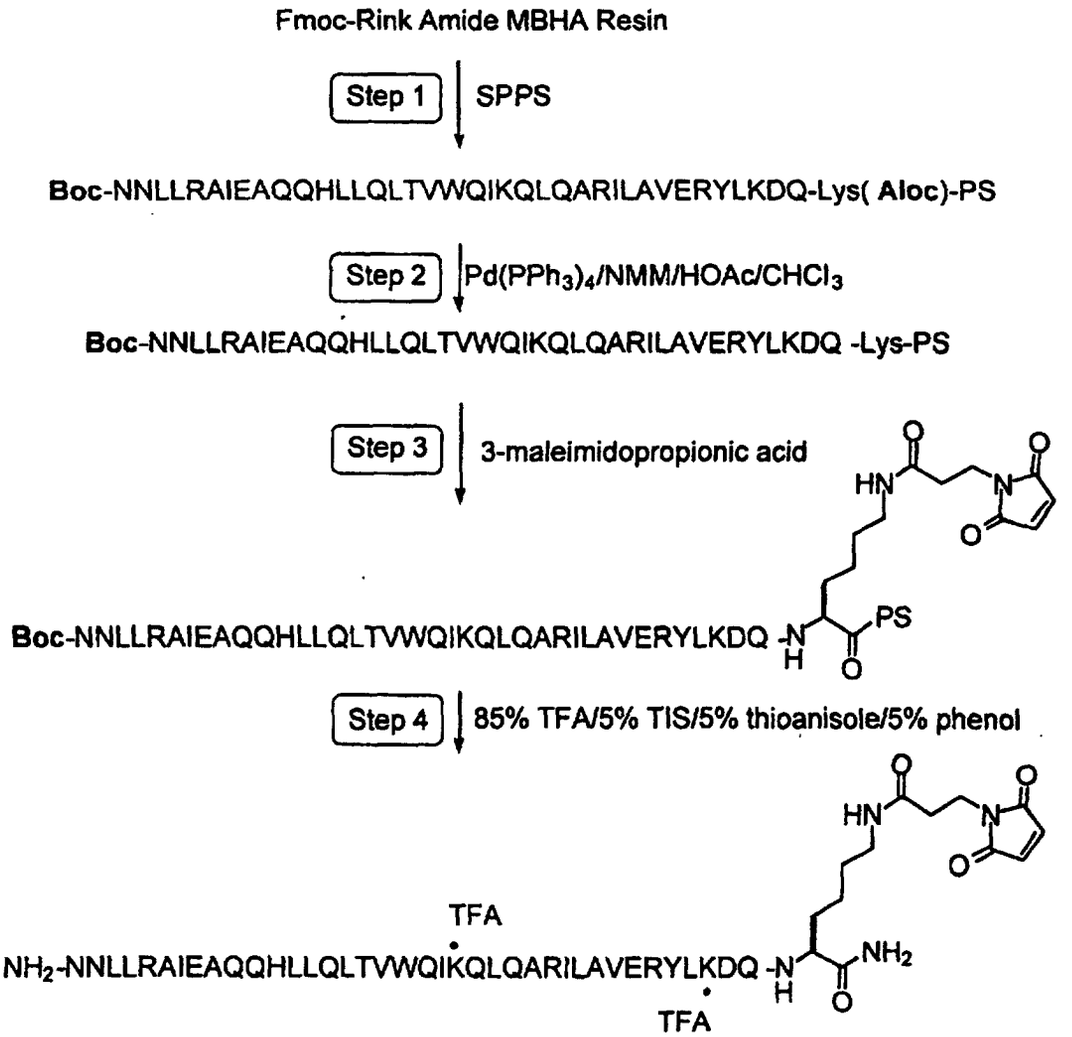
Preparation of a Modified DP107--Synthesis of NNLLRAIEAQQHLLQLTVWQIKQLQARILAVERYLKDQK(MPA) NH_2

[0101] In this example, DP107 (SEQ ID NO:2) is synthesized and modified to include a linker and maleimide group according to the following synthesis scheme. As reported in U.S. Pat. Nos. 6,013,236 and 6,020,459, DP107 exhibits potent antiviral activity against HIV.

[0102] Solid phase peptide synthesis of the modified peptide on a 100 μmole scale is performed using manual solid-phase synthesis, a Symphony Peptide Synthesizer and Fmoc protected Rink Amide MBHA. The following protected amino acids are sequentially added to resin: Fmoc-Lys(Aloc)-OH, Fmoc-Gln(Trt)-OH, Fmoc-Asp(tBu)-OH, Fmoc-Lys (Boc)-OH, Fmoc-Leu-OH, Fmoc-Tyr(tBu)-OH, Fmoc-Arg(Pbf)-OH, Fmoc-Glu(tBu)-OH, Fmoc-Val-OH, Fmoc-Ala-OH, Fmoc-Leu-OH, Fmoc-Ile-OH, Fmoc-Arg(Pbf)-OH, Fmoc-Ala-OH, Fmoc-Gln(Trt)-OH, Fmoc-Leu-OH, Fmoc-Gln(Trt) -OH, Fmoc-Lys(Boc)-OH, Fmoc-Ile-OH, Fmoc-Gln(Trt)-OH, Fmoc-Trp(Boc)-OH, Fmoc-Val-OH, Fmoc-Thr(tBu)-OH, Fmoc-Leu-OH, Fmoc-Gln(Trt)-OH, Fmoc-Leu-OH, Fmoc-Leu-OH, Fmoc-His(Boc)-OH, Fmoc-Gln(Trt)-OH, Fmoc-Gln

(Trt)-OH, Fmoc-Ala-OH, Fmoc-Glu(tBu)-OH, Fmoc-Ile-OH, Fmoc-Ala-OH, Fmoc-Arg(Pbf)-OH, Fmoc-Leu-OH, Fmoc-Leu-OH, Fmoc-Asn(Trt)-OH, Fmoc-Asn(Trt)-OH,They are dissolved in *N,N*-dimethylformamide (DMF) and, according to the sequence, activated using O-benzotriazol-1-yl-*N, N, N', N'*-tetramethyl-uronium hexafluorophosphate (HBTU) and Diisopropylethylamine (DIEA). Removal of the Fmoc protecting group is achieved using a solution of 20% (V/V) piperidine in *N,N*-dimethylformamide (DMF) for 20 minutes (step 1). At the end of the synthesis.The selective deprotection of the Lys (Aloc) group is performed manually and accomplished by treating the resin with a solution of 3 eq of $Pd(PPh_3)_4$ dissolved in 5 mL of $CHCl_3$:NMM:HOAc (18:1:0.5) for 2 h (Step 2). The resin is then washed with $CHCl_3$ (6 x 5 mL), 20% HOAc in DCM (6 x 5 mL), DCM (6 x 5 mL), and DMF (6 x 5 mL). The synthesis is then re-automated for the addition of the 3-maleimidopropionic acid (Step 3). Between every coupling, the resin is washed 3 times with *N,N*-dimethylformamide (DMF) and 3 times with isopropanol. The peptide is cleaved from the resin using 85% TFA/5% TIS/5% thioanisole and 5% phenol, followed by precipitation by dry-ice cold Et_2O (Step 4). The product is purified by preparative reversed phased HPLC using a Varian (Rainin) preparative binary HPLC system: gradient elution of 30-55% B (0.045% TFA in H_2O (A) and 0.045% TFA in CH_3CN (B)) over 180 min at 9.5 mL/min using a Phenomenex Luna 10 μ phenyl-hexyl, 21 mm x 25 cm column and UV detector (Varian Dynamax UVD II) at λ214 and 254 nm to afford the desired modified peptide (i.e., DAC) in >95% purity, as determined by RP-HPLC.

DP-107 C

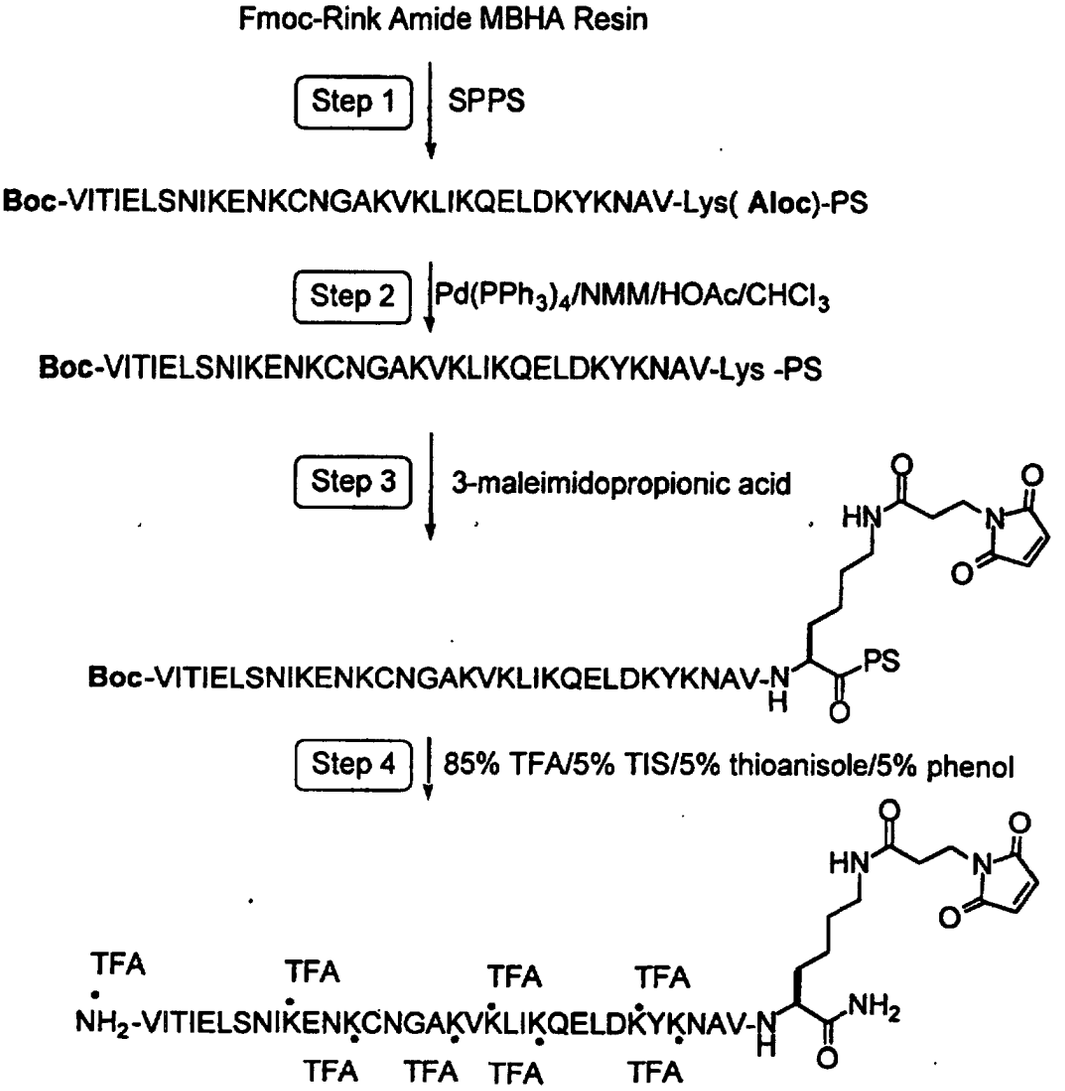


Example 3

Preparation of a Modified anti-RSV peptide (C terminal)

[0103] In this example, the peptide VITIELSNIKENKCNGAKVKLIKQELDKYKNAV (SEQ ID NO:16) is modified to include a linker and maleimide group according to the synthesis scheme set forth below. As reported in U.S. Pat. Nos. 6,013,236 and 6,020,459, the native sequence (SEQ ID NO.) inhibits viral infection of respiratory syncytial virus (RSV), including inhibiting fusion and syncytia formation between RSV-infected and uninfected Hep-2 cells.

[0104] Solid phase peptide synthesis of the modified peptide on a 100 μmole scale is performed using manual solid-phase synthesis, a Symphony Peptide Synthesizer and Fmoc protected Rink Amide MBHA. The following protected amino acids are sequentially added to resin: Fmoc-Lys(Aloc)-OH, Fmoc-Val-OH, Fmoc-Ala-OH, Fmoc-Asn(Trt)-OH, Fmoc-Lys(Boc)-OH, Fmoc-Tyr(tBu)-OH, Fmoc-Lys(Boc)-OH, Fmoc-Asp(tBu)-OH, Fmoc-Leu-OH, Fmoc-Glu(tBu)-OH, Fmoc-Gln(Trt)-OH, Fmoc-Lys(Boc)-OH, Fmoc-Ile-OH, Fmoc-Leu-OH, Fmoc-Lys(Boc)-OH, Fmoc-Val-OH, Fmoc-Lys (Boc)-OH, Fmoc-Ala-OH, Fmoc-Gly-OH, Fmoc-Asn(Trt)-OH, Fmoc-Cys(Trt)-OH, Fmoc-Lys(Boc)-OH, Fmoc-Asn(Trt) -OH, Fmoc-Glu(tBu)-OH, Fmoc-Lys(Boc)-OH, Fmoc-Ile-OH, Fmoc-Asn(Trt)-OH, Fmoc-Ser(tBu)-OH, Fmoc-Leu-OH, Fmoc-Glu(tBu)-OH, Fmoc-Ile-OH, Fmoc-Thr(tBu)-OH, Fmoc-Ile-OH, Fmoc-Val-OH.They are dissolved in *N,N*-dimethylformamide (DMF) and, according to the sequence, activated using O-benzotriazol-1-yl-*N, N, N′, N′*-tetramethyl-uronium hexafluorophosphate (HBTU) and Diisopropylethylamine (DIEA). Removal of the Fmoc protecting group is achieved using a solution of 20% (V/V) piperidine in *N,N*-dimethylformamide (DMF) for 20 minutes (step 1).The selective deprotection of the Lys (Aloe) group is performed manually and accomplished by treating the resin with a solution of 3 eq of $Pd(PPh_3)_4$ dissolved in 5 mL of $CHCl_3$:NMM:HOAc (18:1:0.5) for 2 h (Step 2). The resin is then washed with $CHCl_3$ (6 x 5 mL), 20% HOAc in DCM (6 x 5 mL), DCM (6 x 5 mL), and DMF (6 x 5 mL). The synthesis is then re-automated for the addition of the 3-maleimidopropionic acid (Step 3). Between every coupling, the resin is washed 3 times with *N,N*-dimethylformamide (DMF) and 3 times with isopropanol. The peptide is cleaved from the resin using 85% TFA/5% TIS/5% thioanisole and 5% phenol, followed by precipitation by dry-ice cold Et_2O (Step 4). The product is purified by preparative reversed phased HPLC using a Varian (Rainin) preparative binary HPLC system: gradient elution of 30-55% B (0.045% TFA in H_2O (A) and 0.045% TFA in CH_3CN (B)) over 180 min at 9.5 mL/min using a Phenomenex Luna 10 μ phenyl-hexyl, 21 mm x 25 cm column and UV detector (Varian Dynamax UVD II) at λ214 and 254 nm to afford the desired modified peptide (i.e., DAC) in >95% purity, as determined by RP-HPLC.



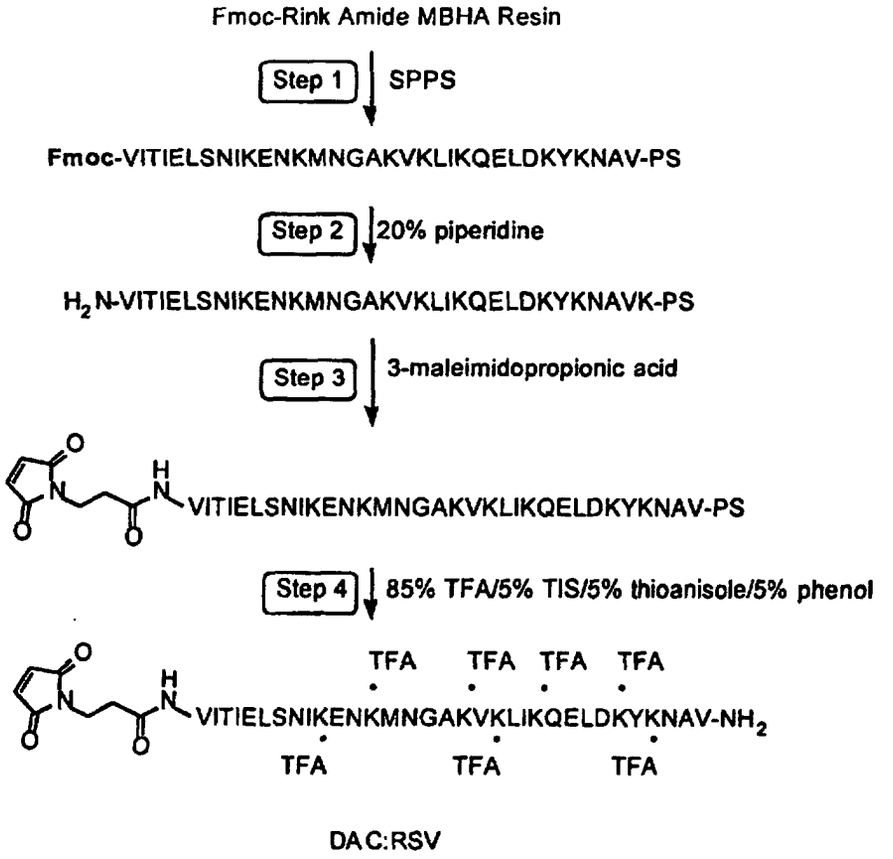
Example 4

Preparation of a Modified anti-RSV peptide (T-N terminal)

[0105] In this example, the peptide VITIELSNIKENKCNGAKVKLIKQELDKYKNAV (SEQ ID NO:17), which corresponds to the peptide of SEQ ID NO:16 but where a Cysteine (C) has been substitututed for the Methionine (M), residue is synthesized and modified to include a linker and maleimide group according to the synthesis scheme set forth below. As reported in U.S. Pat. Nos. 6,013,236 and 6,020,459, the native sequence (SEQ ID NO:16) inhibits viral infection of respiratory syncytial virus (RSV), including inhibiting fusion and syncytia formation between RSV-infected and uninfected Hep-2 cells.

[0106] Solid phase peptide synthesis of the modified peptide on a 100 μmole scale is performed using manual solid-phase synthesis, a Symphony Peptide Synthesizer and Fmoc protected Rink Amide MBHA. The following protected amino acids are sequentially added to resin: Fmoc-Val-OH, Fmoc-Ala-OH, Fmoc-Asn(Trt)-OH, Fmoc-Lys(Boc)-OH, Fmoc-Tyr(tBu)-OH, Fmoc-Lys(Boc)-OH, Fmoc-Asp(tBu)-OH, Fmoc-Leu-OH, Fmoc-Glu(tBu)-OH, Fmoc-Gln(Trt)-OH, Fmoc-Lys(Boc)-OH, Fmoc-Ile-OH, Fmoc-Leu-OH, Fmoc-Lys(Boc)-OH, Fmoc-Val-OH, Fmoc-Lys(Boc)-OH, Fmoc-Ala-OH, Fmoc-Gly-OH, Fmoc-Asn(Trt)-OH, **Fmoc-Met-OH,** Fmoc-Lys(Boc)-OH, Fmoc-Asn(Trt)-OH, Fmoc-Glu(tBu)-OH, Fmoc-Lys(Boc)-OH, Fmoc-Ile-OH, Fmoc-Asn(Trt)-OH, Fmoc-Ser(tBu)-OH, Fmoc-Leu-OH, Fmoc-Glu(tBu)-OH, Fmoc-Ile-OH, Fmoc-Thr(tBu)-OH, Fmoc-Ile-OH, Fmoc-Val-OH.They are dissolved in *N,N*-dimethylformamide (DMF) and, according to the sequence, activated using O-benzotriazol-1-yl-*N, N, N',N'*-tetramethyl-uronium hexafluorophos-

phate (HBTU) and Diisopmpylethylamine (DIEA). Removal of the Fmoc protecting group is achieved using a solution of 20% (V/V) piperidine in *N,N*-dimethylformamide (DMF) for 20 minutes (step 1). The synthesis is then re-automated for the addition of the 3-maleimidopropionic acid (Step 3). Between every coupling, the resin is washed 3 times with *N,N*-dimethylformamide (DMF) and 3 times with isopropanol. The peptide is cleaved from the resin using 85% TFA/ 5% TIS/5% thioanisole and 5% phenol, followed by precipitation by dry-ice cold Et_2O (Step 4). The product is purified by preparative reversed phased HPLC using a Varian (Rainin) preparative binary HPLC system: gradient elution of 30-55% B (0.045% TFA in H_2O (A) and 0.045% TFA in CH_3CN (B)) over 180 min at 9.5 mL/min using a Phenomenex Luna 10 μ phenyl-hexyl, 21 mm x 25 cm column and UV detector (Varian Dynamax UVD II) at λ214 and 254 nm to afford the desired modified peptide (i.e., DAC) in >95% purity, as determined by RP-HPLC.



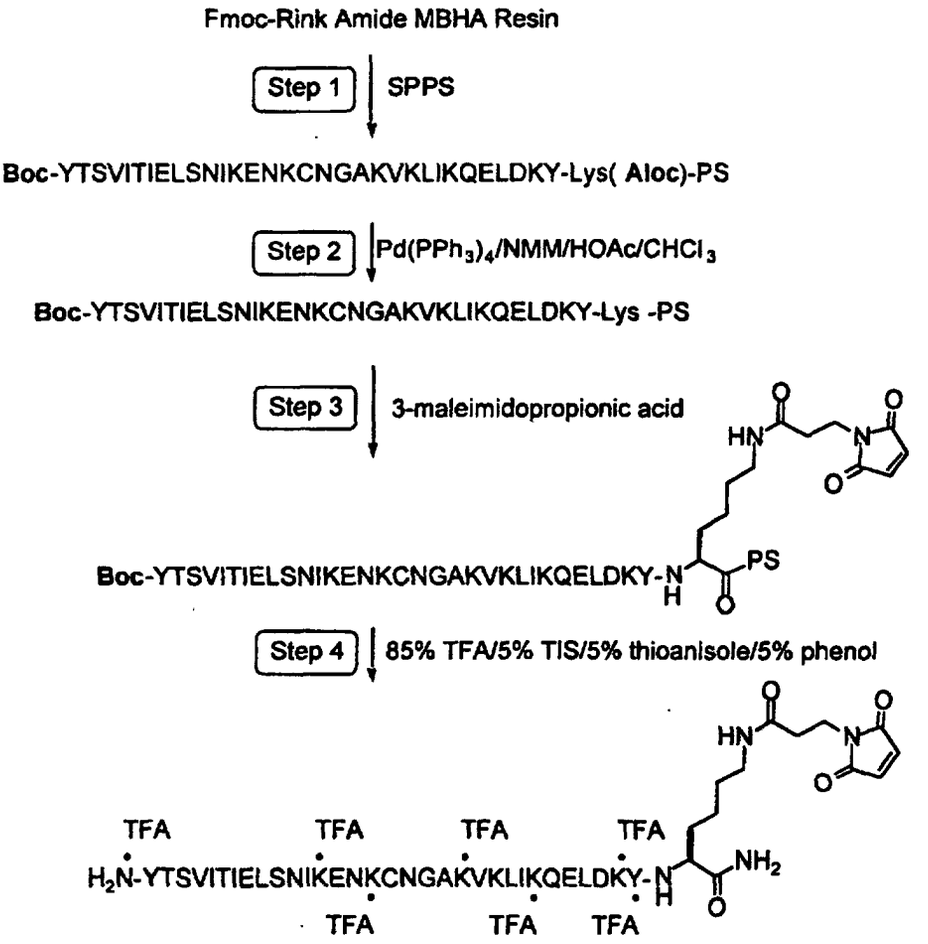
Example 5

Preparation of a Modified anti-RSV peptide

[0107] In this example, the peptide SEQ ID NO:14 is synthesized and modified to include a linker and maleimide group according to the synthesis scheme set forth below. As reported in U.S. Pat. Nos. 6,013,236 and 6,020,459, SEQ ID NO:14 inhibits viral infection of respiratory syncytial virus (RSV), including inhibiting fusion and syncytia formation between RSV-infected and uninfected Hep-2 cells.

[0108] Solid phase peptide synthesis of the modified peptide on a 100 μmole scale is performed using manual solid-phase synthesis, a Symphony Peptide Synthesizer and Fmoc protected Rink Amide MBHA. The following protected amino acids are sequentially added to resin: Fmoc-Lys(Aloc)-OH, Fmoc-Lys(Boc)-OH, Fmoc-Tyr(tBu)-OH, Fmoc-Lys (Boc)-OH, Fmoc-Asp(tBu)-OH, Fmoc-Leu-OH, Fmoc-Glu(tBu)-OH, Fmoc-Gln(Trt)-OH, Fmoc-Lys(Boc)-OH, Fmoc-Ile-OH, Fmoc-Leu-OH, Fmoc-Lys(Boc)-OH, Fmoc-Val-OH, Fmoc-Lys(Boc)-OH, Fmoc-Ala-OH, Fmoc-Gly-OH, Fmoc-Asn(Trt)-OH, Fmoc-Cys(Trt)-OH, Fmoc-Lys(Boc)-OH, Fmoc-Asn(Trt)-OH, Fmoc-Glu(tBu)-OH, Fmoc-Lys(Boc)-OH, Fmoc-Ile-OH, Fmoc-Asn(Trt)-OH, Fmoc-Ser(tBu)-OH, Fmoc-Leu-OH, Fmoc-Glu(tBu)-OH, Fmoc-Ile-OH, Fmoc-Thr (tBu)-OH, Fmoc-Ile-OH, Fmoc-Val-OH, Fmoc-Ser(tBu)-OH, Fmoc-Thr(tBu)-OH, Fmoc-Tyr(tBu)-OH.They are dissolved in *N,N*-dimethylformamide (DMF) and, according to the sequence, activated using O-benzotriazol-1-yl-*N, N, N',N'*-

tetramethyl-uronium hexafluorophosphate (HBTU) and Diisopropylethylamine (DIEA). Removal of the Fmoc protecting group is achieved using a solution of 20% (V/V) piperidine in *N,N*-dimethylformamide (DMF) for 20 minutes (step 1). The selective deprotection of the Lys (Aloc) group is performed manually and accomplished by treating the resin with a solution of 3 eq of $Pd(PPh_3)_4$ dissolved in 5 mL of $CHCl_3$:NMM:HOAc (18:1:0.5) for 2 h (Step 2). The resin is then washed with $CHCl_3$ (6 x 5 mL), 20% HOAc in DCM (6 x 5 mL), DCM (6 x 5 mL), and DMF (6 x 5 mL). The synthesis is then re-automated for the addition of the 3-maleimidopropionic acid (Step 3). Between every coupling, the resin is washed 3 times with *N,N*-dimethylformamide (DMF) and 3 times with isopropanol. The peptide is cleaved from the resin using 85% TFA/5% TIS/5% thioanisole and 5% phenol, followed by precipitation by dry-ice cold Et_2O (Step 4). The product is purified by preparative reversed phased HPLC using a Varian (Rainin) preparative binary HPLC system: gradient elution of 30-55% B (0.045% TFA in H_2O (A) and 0.045% TFA in CH_3CN (B)) over 180 min at 9.5 mL/min using a Phenomenex Luna 10 μ phenyl-hexyl, 21 mm x 25 cm column and UV detector (Varian Dynamax UVD II) at 214 and 254 nm to afford the desired modified peptide (i.e., DAC) in >95% purity, as determined by RP-HPLC.



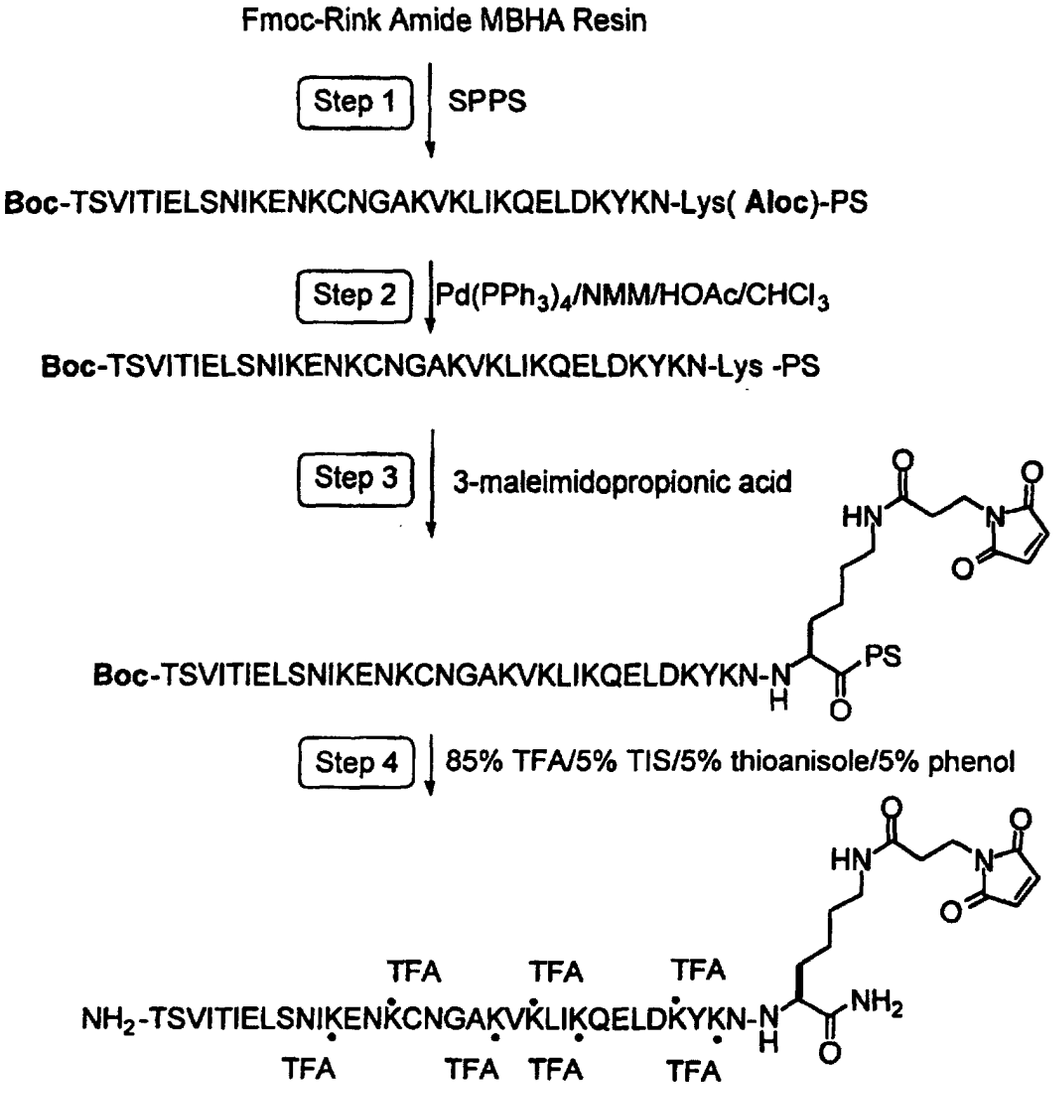
Example 6 (T-143)

Preparation of a Modified anti-RSV peptide

[0109] In this example, the peptide SEQ ID NO:15 is synthesized and modified to include a linker and maleimide group according to the synthesis scheme set forth below. As reported in U.S. Pat. Nos. 6,013,236 and 6,020,459, SEQ ID NO:15 inhibits viral infection of respiratory syncytial virus (RSV), including inhibiting fusion and syncytia formation between RSV-infected and uninfected Hep-2 cells.

[0110] Solid phase peptide synthesis of the modified peptide analog on a 100 μmole scale is performed using manual solid-phase synthesis, a Symphony Peptide Synthesizer and Fmoc protected Rink Amide MBHA. The following protected amino acids are sequentially added to resin: Fmoc-Lys(Aloc)-OH, Fmoc-Asn(Trt)-OH, Fmoc-Lys(Boc)-OH, Fmoc-Tyr(tBu)-OH, Fmoc-Lys(Boc)-OH, Fmoc-Asp(tBu)-OH, Fmoc-Leu-OH, Fmoc-Glu(tBu)-OH, Fmoc-Gln(Trt)-OH, Fmoc-Lys(Boc)-OH, Fmoc-Ile-OH, Fmoc-Leu-OH, Fmoc-Lys(Boc)-OH, Fmoc-Val-OH, Fmoc-Lys(Boc)-OH, Fmoc-

Ala-OH, Fmoc-Gly-OH, Fmoc-Asn(Trt)-OH, Fmoc-Cys(Trt)-OH, Fmoc-Lys(Boc)-OH, Fmoc-Asn(Trt)-OH, Fmoc-Glu (tBu)-OH, Fmoc-Lys(Boc)-OH, Fmoc-Ile-OH, Fmoc-Asn(Trt)-OH, Fmoc-Ser(tBu)-OH, Fmoc-Leu-OH, Fmoc-Glu(tBu) -OH, Fmoc-Ile-OH, Fmoc-Thr(tBu)-OH, Fmoc-Ile-OH, Fmoc-Val-OH, Fmoc-Ser(tBu)-OH, Fmoc-Thr(tBu)-OH.They are dissolved in *N,N*-dimethylformamide (DMF) and, according to the sequence, activated using O-benzotriazol-1-yl-*N, N, N', N'*-tetramethyl-uronium hexafluorophosphate (HBTU) and Diisopropylethylamine (DIEA). Removal of the Fmoc protecting group is achieved using a solution of 20% (V/V) piperidine in *N,N*-dimethylformamide (DMF) for 20 minutes (step 1).The selective deprotection of the Lys (Aloc) group is performed manually and accomplished by treating the resin with a solution of 3 eq of $Pd(PPh_3)_4$ dissolved in 5 mL of $CHCl_3$:NMM:HOAc (18:1:0.5) for 2 h (Step 2). The resin is then washed with $CHCl_3$ (6 x 5 mL), 20% HOAc in DCM (6 x 5 mL), DCM (6 x 5 mL), and DMF (6 x 5 mL). The synthesis is then re-automated for the addition of the 3-maleimidopropionic acid (Step 3). Between every coupling, the resin is washed 3 times with *N,N*-dimethylformamide (DMF) and 3 times with isopropanol. The peptide is cleaved from the resin using 85% TFA/5% TIS/5% thioanisole and 5% phenol, followed by precipitation by dry-ice cold Et_2O (Step 4). The product is purified by preparative reversed phased HPLC using a Varian (Rainin) preparative binary HPLC system: gradient elution of 30-55% B (0.045% TFA in H_2O (A) and 0.045% TFA in CH_3CN (B)) over 180 min at 9.5 mL/min using a Phenomenex Luna 10 μ phenyl-hexyl, 21 mm x 25 cm column and UV detector (Varian Dynamax UVD II) at λ214 and 254 nm to afford the desired modified peptide (i.e:, DAC) in >95% purity, as determined by RP-HPLC.

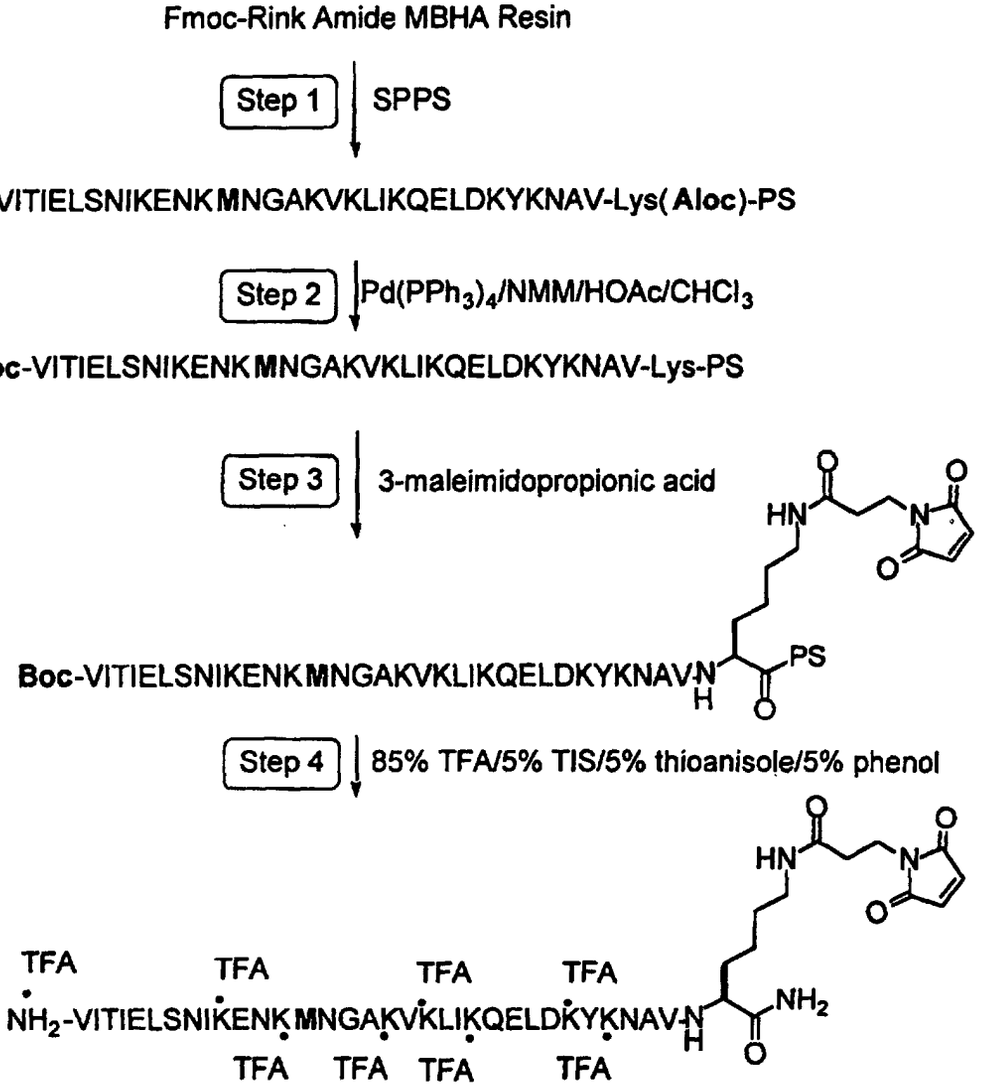


Example 7

Preparation of a Modified anti-RSV peptide (C Terminal)

[0111] In this example, the peptide SEQ ID NO:17), which corresponds to SEQ ID NO:16 with a cysteine (C) substituted for the Methionine (M), is synthesized and modified to include a linker and maleimide group according to the synthesis scheme set forth below. As reported in U.S. Pat. Nos. 6,013,236 and 6,020,459, the native sequence SEQ ID NO:16. inhibits viral infection of respiratory syncytial virus (RSV), including inhibiting fusion and syncytia formation between RSV-infected and uninfected Hep-2 cells.

[0112] Solid phase peptide synthesis of the modified peptide on a 100 μmole scale is performed using manual solid-phase synthesis, a Symphony Peptide Synthesizer and Fmoc protected Rink Amide MBHA. The following protected amino acids are sequentially added to resin: Fmoc-Lys(Aloc)-OH, Fmoc-Val-OH, Fmoc-Ala-OH, Fmoc-Asn(Trt)-OH, Fmoc-Lys(Boc)-OH, Fmoc-Tyr(tBu)-OH, Fmoc-Lys(Boc)-OH, Fmoc-Asp(tBu)-OH, Fmoc-Leu-OH, Fmoc-Glu(tBu)-OH, Fmoc-Gln(Trt)-OH, Fmoc-Lys(Boc)-OH, Fmoc-Ile-OH, Fmoc-Leu-OH, Fmoc-Lys(Boc)-OH, Fmoc-Val-OH, Fmoc-Lys (Boc)-OH, Fmoc-Ala-OH, Fmoc-Gly-OH, Fmoc-Asn(Trt)-OH, **Fmoc-Met-OH,** Fmoc-Lys(Boc)-OH, Fmoc-Asn(Trt)-OH, Fmoc-Glu(tBu)-OH, Fmoc-Lys(Boc)-OH, Fmoc-Ile-OH, Fmoc-Asn(Trt)-OH, Fmoc-Ser(tBu)-OH, Fmoc-Leu-OH, Fmoc-Glu(tBu)-OH, Fmoc-Ile-OH, Fmoc-Thr(tBu)-OH, Fmoc-Ile-OH, Fmoc-Val-OH.They are dissolved in *N,N*-dimethylformamide (DMF) and, according to the sequence, activated using O-benzotriazol-1-yl-*N, N, N′, N′*-tetramethyl-uronium hexafluorophosphate (HBTU) and Diisopropylethylamine (DIEA). Removal of the Fmoc protecting group is achieved using a solution of 20% (V/V) piperidine in *N,N*-dimethylformamide (DMF) for 20 minutes (step 1).The selective deprotection of the Lys (Aloc) group is performed manually and accomplished by treating the resin with a solution of 3 eq of $Pd(PPh_3)_4$ dissolved in 5 mL of $CHCl_3$:NMM:HOAc (18:1:0.5) for 2 h (Step 2). The resin is then washed with $CHCl_3$ (6 x 5 mL), 20% HOAc in DCM (6 x 5 mL), DCM (6 x 5 mL), and DMF (6 x 5 mL). The synthesis is then re-automated for the addition of the 3-maleimidopropionic acid (Step 3). Between every coupling, the resin is washed 3 times with *N,N*-dimethylformamide (DMF) and 3 times with isopropanol. The peptide is cleaved from the resin using 85% TFA/5% TIS/5% thioanisole and 5% phenol, followed by precipitation by dry-ice cold Et_2O (Step 4). The product is purified by preparative reversed phased HPLC using a Varian (Rainin) preparative binary HPLC system: gradient elution of 30-55% B (0.045% TFA in H_2O (A) and 0.045% TFA in CH_3CN (B)) over 180 min at 9.5 mL/min using a Phenomenex Luna 10 μ phenyl-hexyl, 21 mm x 25 cm column and UV detector (Varian Dynamax UVD II) at λ 214 and 254 nm to afford the desired modified peptide (i.e., DAC) in >95% purity, as determined by RP-HPLC.



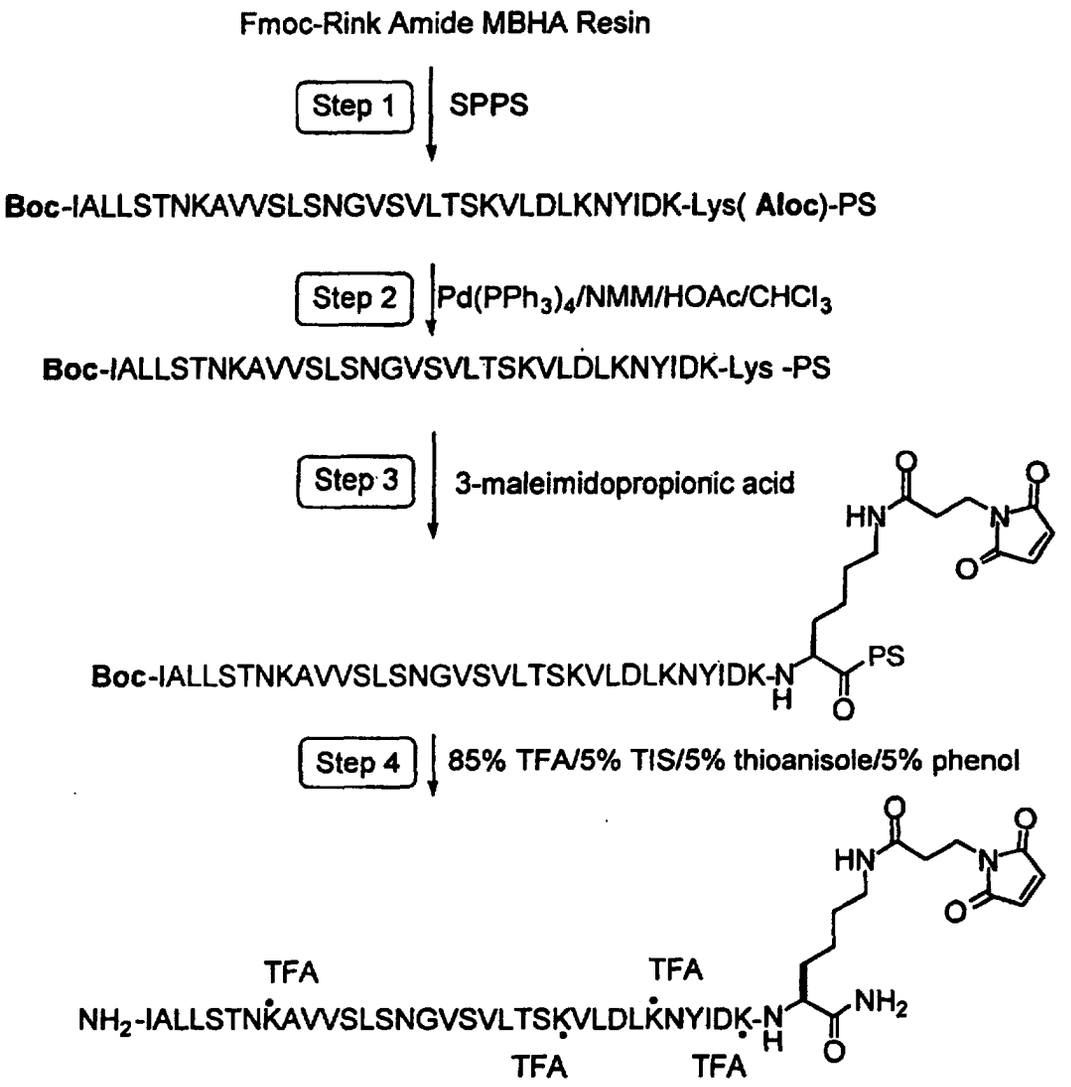
Example 8

Preparation of a Modified anti-RSV peptide

[0113] In this example, the peptide SEQ ID NO:29. is synthesized and modified to include a linker and maleimide group according to the synthesis scheme set forth below. As reported in U.S. Pat. Nos. 6,013,236 and 6,020,459, SEQ ID NO:29 inhibits viral infection of respiratory syncytial virus (RSV), including inhibiting fusion and syncytia formation between RSV-infected and uninfected Hep-2 cells.

[0114] Solid phase peptide synthesis of the modified peptide on a 100 μmole scale is performed using manual solid-phase synthesis, a Symphony Peptide Synthesizer and Fmoc protected Rink Amide MBHA. The following protected amino acids are sequentially added to resin: Fmoc-Lys(Aloc)-OH, Fmoc-Lys(Boc)-OH, Fmoc-Asp(tBu)-OH, Fmoc-Ile-OH, Fmoc-Tyr(tBu)-OH, Fmoc-Asn(Trt)-OH, Fmoc-Lys(Boc)-OH, Fmoc-Leu-OH, Fmoc-Asp(tBu)-OH, Fmoc-Leu-OH, Fmoc-Val-OH, Fmoc-Lys(Boc)-OH, Fmoc-Ser(tBu)-OH, Fmoc-Thr(tBu)-OH, Fmoc-Leu-OH, Fmoc-Val-OH, Fmoc-Ser(tBu)-OH, Fmoc-Val-OH, Fmoc-Gly-OH, Fmoc-Asn(Trt)-OH, Fmoc-Ser(tBu)-OH, Fmoc-Leu-OH, Fmoc-Ser (tBu)-OH, Fmoc-Val-OH, Fmoc-Val-OH, Fmoc-Ala-OH, Fmoc-Lys(Boc)-OH, Fmoc-Asn(Trt)-OH, Fmoc-Thr(tBu)-OH, Fmoc-Ser(tBu)-OH, Fmoc-Leu-OH, Fmoc-Leu-OH, Fmoc-Ala-OH, Fmoc-Ile-OH.They are dissolved in *N,N*-dimethyl-formamide (DMF) and, according to the sequence, activated using O-benzotriazol-I-yl-*N,N, N',N'*-tetramethyl-uronium hexafluorophosphate (HBTU) and Diisopropylethylamine (DIEA). Removal of the Fmoc protecting group is achieved using a solution of 20% (V/V) piperidine in *N,N*-dimethylformamide (DMF) for 20 minutes (step 1).The selective deprotection of the Lys (Aloc) group is performed manually and accomplished by treating the resin with a solution of 3 eq of

$Pd(PPh_3)_4$ dissolved in 5 mL of $CHCl_3$:NMM:HOAc (18:1:0.5) for 2 h (Step 2). The resin is then washed with $CHCl_3$ (6 x 5 mL), 20% HOAc in DCM (6 x 5 mL), DCM (6 x 5 mL), and DMF (6 x 5 mL). The synthesis is then re-automated for the addition of the 3-maleinudopropionic acid (Step 3). Between every coupling, the resin is washed 3 times with *N, N*-dimethylformamide (DMF) and 3 times with isopropanol. The peptide is cleaved from the resin using 85% TFA/5% TIS/5% thioanisole and 5% phenol, followed by precipitation by dry-ice cold Et_2O (Step 4). The product is purified by preparative reversed phased HPLC using a Varian (Rainin) preparative binary HPLC system: gradient elution of 30-55% B (0.045% TFA in H_2O (A) and 0.045% TFA in CH_3CN (B)) over 180 min at 9.5 mL/min using a Phenomenex Luna 10 μ phenyl-hexyl, 21 mm x 25 cm column and UV detector (Varian Dynamax UVD II) at λ 214 and 254 nm to afford the desired modified peptide (i.e., DAC) in >95% purity, as determined by RP-HPLC.



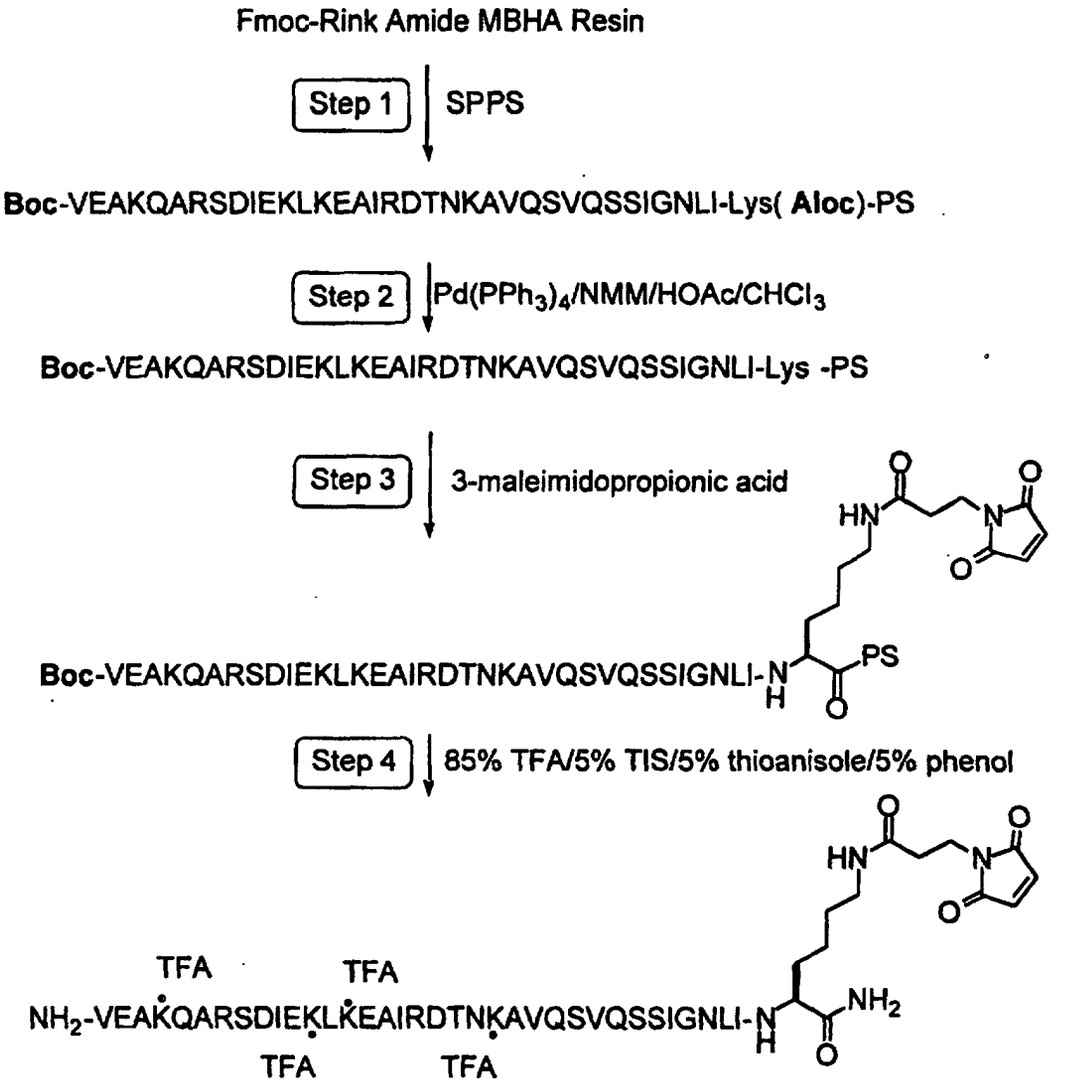
Example 9 (T-173)

Preparation of a Modified anti-HPIV peptide

[0115] In this example, the peptide SEQ ID NO:52. is synthesized and modified to include a linker and maleimide group according to the synthesis scheme set forth below. As reported in U.S. Pat. Nos. 6,013,236 and 6,020,459, SEQ ID NO:52 inhibits viral infection of human parainfluenza virus 3 (HPIV3), including inhibiting fusion and syncytia formation between HPIV3-infected Hep2 cells and uninfected CV-1W cells.

[0116] Solid phase peptide synthesis of the modified peptide on a 100 μmole scale is performed using manual solid-phase synthesis, a Symphony Peptide Synthesizer and Fmoc protected Rink Amide MBHA. The following protected amino acids are sequentially added to resin: Fmoc-Lys(Aloc)-OH, Fmoc-Ile-OH, Fmoc-Leu-OH, Fmoc-Asn(Trt)-OH,

Fmoc-Gly-OH, Fmoc-Ile-OH, Fmoc-Ser(tBu)-OH, Fmoc-Ser(tBu)-OH, Fmoc-Gln(Trt)-OH, Fmoc-Val-OH, Fmoc-Ser (tBu)-OH, Fmoc-Gln(Trt)-OH, Fmoc-Val-OH, Fmoc-Ala-OH, Fmoc-Lys(Boc)-OH, Fmoc-Asn(Trt)-OH, Fmoc-Thr(tBu) -OH, Fmoc-Asp(tBu)-OH, Fmoc-Arg(Pbf)-OH, Fmoc-Ile-OH, Fmoc-Ala-OH, Fmoc-Glu(tBu)-OH, Fmoc-Lys(Boc)-OH, Fmoc-Leu-OH, Fmoc-Lys(Boc)-OH, Fmoc-Glu(tBu)-OH, Fmoc-Ile-OH, Fmoc-Asp(tBu)-OH, Fmoc-Ser(tBu)-OH, Fmoc-Arg(Pbf)-OH, Fmoc-Ala-OH, Fmoc-Gln(Trt)-OH, Fmoc-Lys(Boc)-OH, Fmoc-Ala-OH, Fmoc-Glu(tBu)-OH, Fmoc-Val-OH.They are dissolved in *N,N*-dimethylformamide (DMF) and, according to the sequence, activated using O-benzotriazol-1-yl-*N, N, N', N'*-tetramethyl-uronium hexafluorophosphate (HBTU) and Diisopropylethylamine (DIEA). Removal of the Fmoc protecting group is achieved using a solution of 20% (V/V) piperidine in *N,N*-dimethylformamide (DMF) for 20 minutes (step 1).The selective deprotection of the Lys (Aloc) group is performed manually and accomplished by treating the resin with a solution of 3 eq of $Pd(PPh_3)_4$ dissolved in 5 mL of $CHCl_3$:NMM:HOAc (18:1:0.5) for 2 h (Step 2). The resin is then washed with $CHCl_3$ (6 x 5 mL), 20% HOAc in DCM (6 x 5 mL), DCM (6 x 5 mL), and DMF (6 x 5 mL). The synthesis is then re-automated for the addition of the 3-maleimidopropionic acid (Step 3). Between every coupling, the resin is washed 3 times with *N,N*-dimethylformamide (DMF) and 3 times with isopropanol. The peptide is cleaved from the resin using 85% TFA/5% TIS/5% thioanisole and 5% phenol, followed by precipitation by dry-ice cold Et_2O (Step 4). The product is purified by preparative reversed phased HPLC using a Varian (Rainin) preparative binary HPLC system: gradient elution of 30-55% B (0.045% TFA in H_2O (A) and 0.045% TFA in CH_3CN (B)) over 180 min at 9.5 mL/min using a Phenomenex Luna 10 μ phenyl-hexyl, 21 mm x 25 cm column and UV detector (Varian Dynamax UVD II) at λ 214 and 254 nm to afford the desired modified peptide (i.e., DAC) in >95% purity, as determined by RP-HPLC.

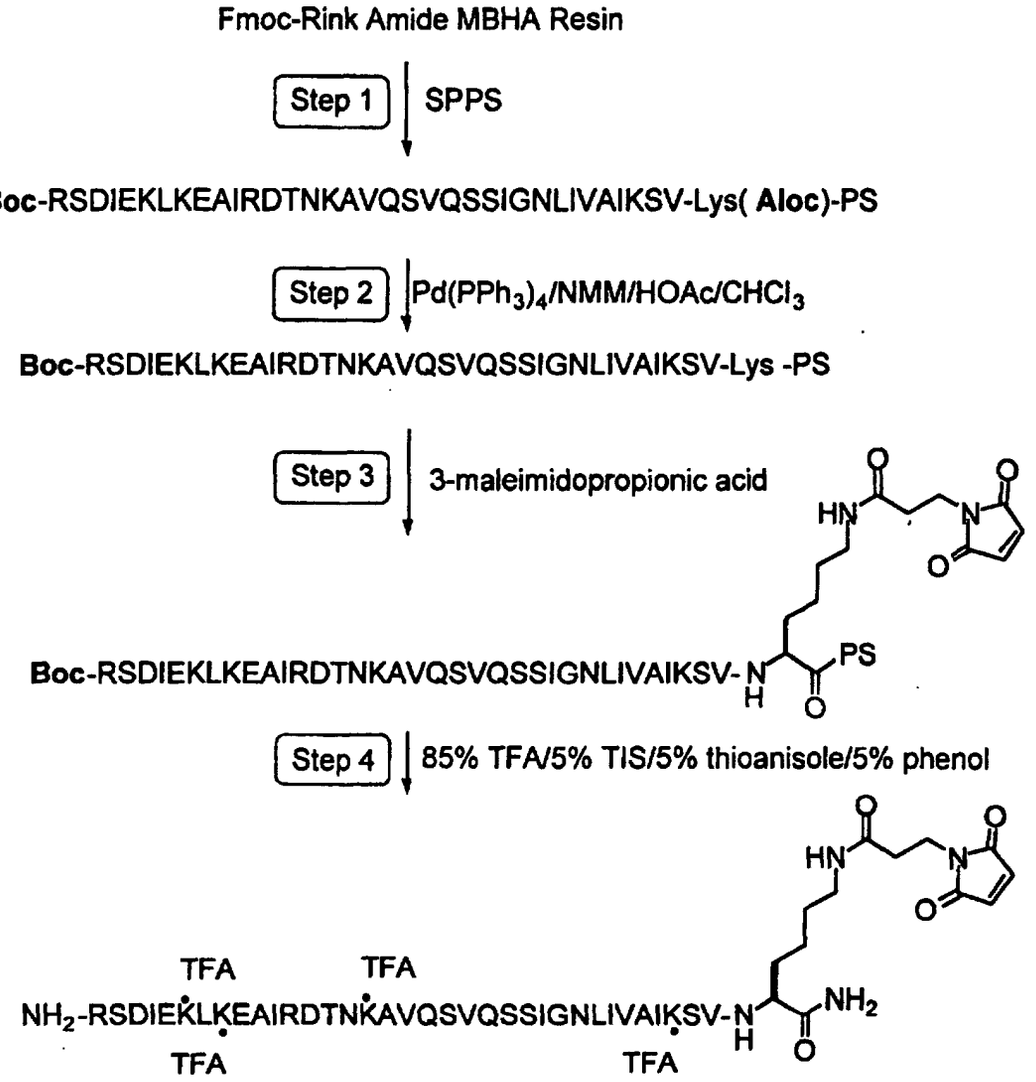


Example 10

Preparation of a Modified anti-HPIV peptide

[0117] In this example, the peptide SEQ ID NO:58 is synthesized and modified to include a linker and maleimide group according to the synthesis scheme set forth below. As reported in U.S. Pat. Nos. 6,013,236 and 6,020,459, SEQ ID NO:58 inhibits viral infection of human parainfluenza virus 3 (HPIV3), including inhibiting fusion and syncytia formation between HPIV3-infected Hep2 cells and uninfected CV-1W cells.

[0118] Solid phase peptide synthesis of the modified peptide on a 100 μmole scale is performed using manual solid-phase synthesis, a Symphony Peptide Synthesizer and Fmoc protected Rink Amide MBHA. The following protected amino acids are sequentially added to resin: Fmoc-Lys(Aloc)-OH, Fmoc-Val-OH, Fmoc-Ser(tBu)-OH, Fmoc-Lys(Boc)-OH, Fmoc-Ile-OH, Fmoc-Ala-OH, Fmoc-Val-OH, Fmoc-Ile-OH, Fmoc-Leu-OH, Fmoc-Asn(Trt)-OH, Fmoc-Gly-OH, Fmoc-Ile-OH, Fmoc-Ser(tBu)-OH, Fmoc-Ser(tBu)-OH, Fmoc-Gln(Trt)-OH, Fmoc-Val-OH, Fmoc-Ser(tBu)-OH, Fmoc-Gln(Trt)-OH, Fmoc-Val-OH, Fmoc-Ala-OH, Fmoc-Lys(Boc)-OH, Fmoc-Asn(Trt)-OH, Fmoc-Thr(tBu)-OH, Fmoc-Asp(tBu)-OH, Fmoc-Arg(Pbf)-OH, Fmoc-Ile-OH, Fmoc-Ala-OH, Fmoc-Glu(tBu)-OH, Fmoc-Lys(Boc)-OH, Fmoc-Leu-OH, Fmoc-Lys(Boc)-OH, Fmoc-Glu(tBu)-OH, Fmoc-Ile-OH, Fmoc-Asp(tBu)-OH, Fmoc-Ser(tBu)-OH, Fmoc-Arg(Pbf)-OH. They are dissolved in *N,N*-dimethylformamide (DMF) and, according to the sequence, activated using O-benzotriazol-1-yl-*N, N, N′, N′*-tetramethyl-uronium hexafluorophosphate (HBTU) and Diisopropylethylamine (DIEA). Removal of the Fmoc protecting group is achieved using a solution of 20% (V/V) piperidine in *N,N*-dimethylformamide (DMF) for 20 minutes (step 1).The selective deprotection of the Lys (Aloc) group is performed manually and accomplished by treating the resin with a solution of 3 eq of $Pd(PPh_3)_4$ dissolved in 5 mL of $CHCl_3$:NMM:HOAc (18:1:0.5) for 2 h (Step 2). The resin is then washed with $CHCl_3$ (6 x 5 mL), 20% HOAc in DCM (6 x 5 mL), DCM (6 x 5 mL), and DMF (6 x 5 mL). The synthesis is then re-automated for the addition of the 3-maleimidopropionic acid (Step 3). Between every coupling, the resin is washed 3 times with *N,N*-dimethylformamide (DMF) and 3 times with isopropanol. The peptide is cleaved from the resin using 85% TFA/5% TIS/5% thioanisole and 5% phenol, followed by precipitation by dry-ice cold Et_2O (Step 4). The product is purified by preparative reversed phased HPLC using a Varian (Rainin) preparative binary HPLC system: gradient elution of 30-55% B (0.045% TFA in H_2O (A) and 0.045% TFA in CH_3CN (B)) over 180 min at 9.5 mL/min using a Phenomenex Luna 10 μ phenyl-hexyl, 21 mm x 25 cm column and UV detector (Varian Dynamax UVD II) at λ 214 and 254 nm to afford the desired modified peptide (i.e., DAC) in >95% purity, as determined by RP-HPLC.



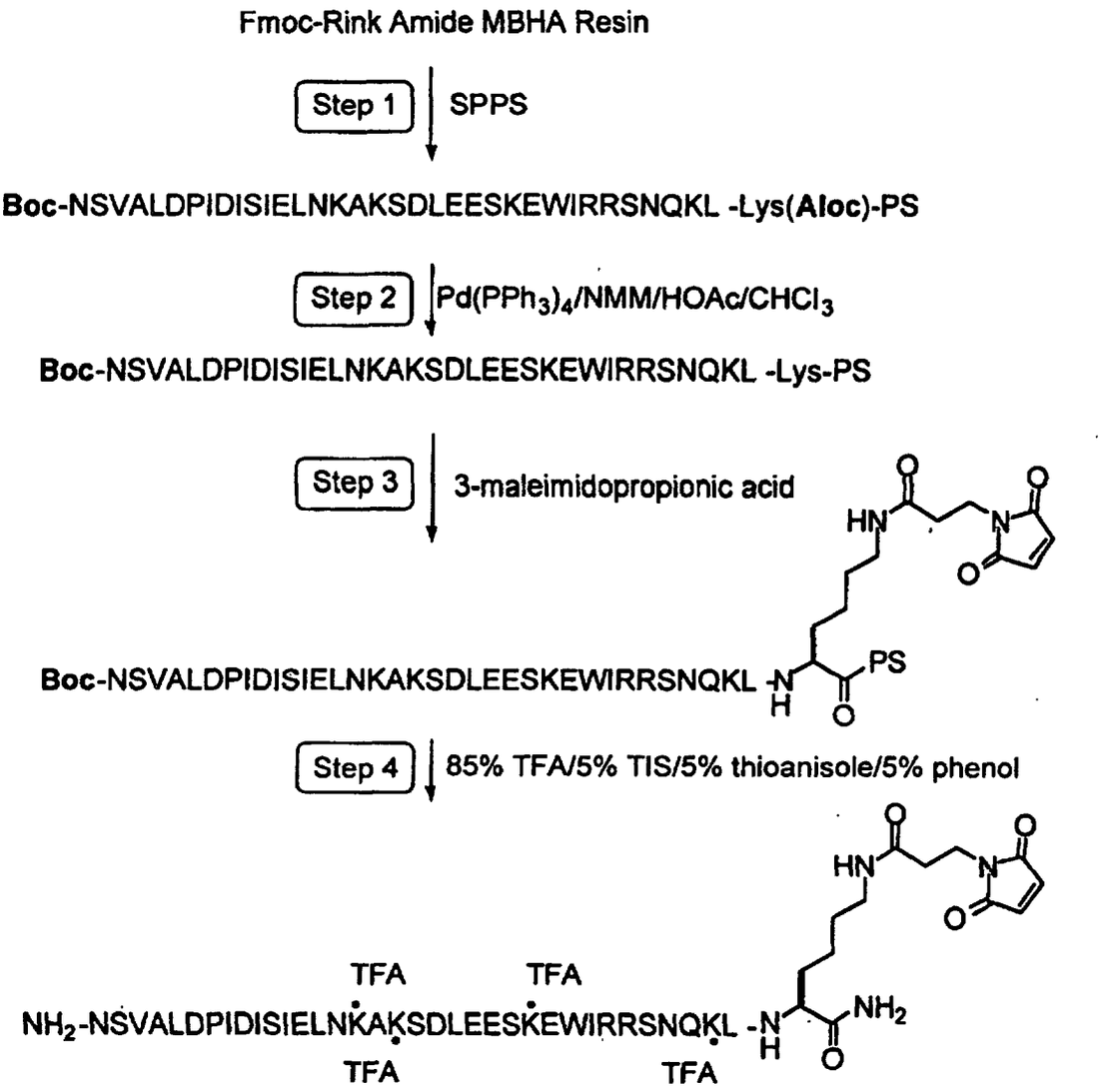
Example 11

Preparation of a Modified anti-HPIV peptide

[0119] In this example, the peptide SEQ ID NO:35 is synthesized and modified to include a linker and maleimide group according to the synthesis scheme set forth below. As reported in U.S. Pat. Nos. 6,013,236 and 6,020,459, SEQ ID NO:35 inhibits viral infection of human parainfluenza virus 3 (HPIV3), including inhibiting fusion and syncytia formation between HPIV3-infected Hep2 cells and uninfected CV-1W cells.

[0120] Solid phase peptide synthesis of the modified peptide on a 100 μmole scale is performed using manual solid-phase synthesis, a Symphony Peptide Synthesizer and Fmoc protected Rink Amide MBHA. The following protected amino acids are sequentially added to resin: Fmoc-Lys(Aloc)-OH, Fmoc-Leu-OH, Fmoc-Lys(Boc)-OH, Fmoc-Gln(Trt)-OH, Fmoc-Asn(Trt)-OH, Fmoc-Ser(tBu)-OH, Fmoc-Arg(Pbf)-OH, Fmoc-Arg(Pbf)-OH, Fmoc-Ile-OH, Fmoc-Trp(Boc)-OH, Fmoc-Glu(tBu)-OH, Fmoc-Lys(Boc)-OH, Fmoc-Ser(tBu)-OH, Fmoc-Glu(tBu)-OH, Fmoc-Glu(tBu)-OH, Fmoc-Leu-OH, Fmoc-Asp(tBu)-OH, Fmoc-Ser(tBu)-OH, Fmoc-Lys(Boc)-OH, Fmoc-Ala-OH, Fmoc-Lys(Boc)-OH, Fmoc-Asn(Trt)-OH, Fmoc-Leu-OH, Fmoc-Ile-OH, Fmoc-Glu(tBu)-OH, Fmoc-Ser(tBu)-OH, Fmoc-Ile-OH, Fmoc-Asp(tBu)-OH, Fmoc-Ile-OH, Fmoc-Pro-OH, Fmoc-Asp(tBu)-OH, Fmoc-Leu-OH, Fmoc-Gln(Trt)-OH Fmoc-Ala-OH, Fmoc-Val-OH, Fmoc-Ser(tBu)-OH, Fmoc-Asn(Trt)-OH.They are dissolved in *N,N*-dimethylformamide (DMF) and, according to the sequence, activated using O-benzotriazol-1-yl-*N, N, N', N'*-tetramethyl-uronium hexafluorophosphate (HBTU) and Di-isopropylethylamine (DIEA). Removal of the Fmoc protecting group is achieved using a solution of 20% (V/V) piperidine

in *N,N*-dimethylformamide (DMF) for 20 minutes (step 1).The selective deprotection of the Lys (Aloc) group is performed manually and accomplished by treating the resin with a solution of 3 eq of $Pd(PPh_3)_4$ dissolved in 5 mL of $CHCl_3$:NMM: HOAc (18:1:0.5) for 2 h (Step 2). The resin is then washed with $CHCl_3$ (6 x 5 mL), 20% HOAc in DCM (6 x 5 mL), DCM (6 x 5 mL), and DMF (6 x 5 mL). The synthesis is then re-automated for the addition of the 3-maleimidopropionic acid (Step 3). Between every coupling, the resin is washed 3 times with *N,N*-dimethylformamide (DMF) and 3 times with isopropanol. The peptide is cleaved from the resin using 85% TFA/5% TIS/5% thioanisole and 5% phenol, followed by precipitation by dry-ice cold Et_2O (Step 4). The product is purified by preparative reversed phased HPLC using a Varian (Rainin) preparative binary HPLC system: gradient elution of 30-55% B (0.045% TFA in H_2O (A) and 0.045% TFA in CH_3CN (B)) over 180 min at 9.5 mL/min using a Phenomenex Luna 10 μ phenyl-hexyl, 21 mm x 25 cm column and UV detector (Varian Dynamax UVD II) at λ 214 and 254 nm to afford the desired modified peptide (i.e., DAC) in >95% purity, as determined by RP-HPLC.



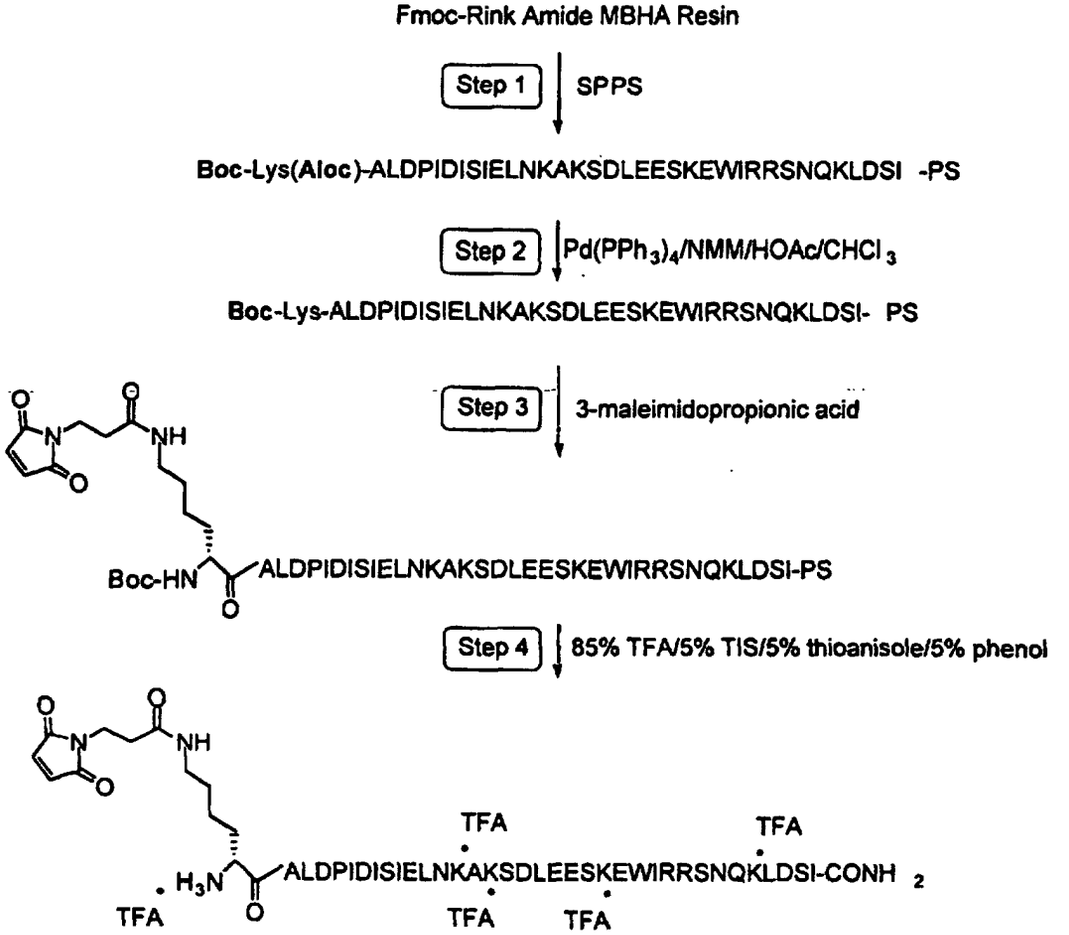
Example 12

Preparation of a Modified anti-HPIV peptide

[0121] In this example, the peptide SEQ ID NO:38 is synthesized and modified to include a linker and maleimide

group according to the synthesis scheme set forth below. As reported in U.S. Pat. Nos. 6,013,236 and 6,020,459, SEQ ID NO: 38 inhibits viral infection of human parainfluenza virus 3 (HPIV3), including inhibiting fusion and syncytia formation between HPIV3-infected Hep2 cells and uninfected CV-1W cells.

[0122] Solid phase peptide synthesis of the modified peptide analog on a 100 μmole scale is performed using manual solid-phase synthesis, a Symphony Peptide Synthesizer and Fmoc protected Rink Amide MBHA. The following protected amino acids are sequentially added to resin: Fmoc-Ile-OH, Fmoc-Ser(tBu)-OH, Fmoc-Asp(tBu)-OH, Fmoc-Leu-OH, Fmoc-Lys(Boc)-OH, Fmoc-Gln(Trt)-OH, Fmoc-Asn(Trt)-OH, Fmoc-Ser(tBu)-OH, Fmoc-Arg(Pbf)-OH, Fmoc-Arg(Pbf)-OH, Fmoc-Ile-OH, Fmoc-Trp(Boc)-OH, Fmoc-Glu(tBu)-OH, Fmoc-Lys(Boc)-OH, Fmoc-Ser(tBu)-OH, Fmoc-Glu(tBu)-OH, Fmoc-Glu(tBu)-OH, Fmoc-Leu-OH, Fmoc-Asp(tBu)-OH, Fmoc-Ser(tBu)-OH, Fmoc-Lys(Boc)-OH, Fmoc-Ala-OH, Fmoc-Lys(Boc)-OH, Fmoc-Asn(Trt)-OH, Fmoc-Leu-OH, Fmoc-Glu(tBu)-OH, Fmoc-Ile-OH, Fmoc-Ser(tBu)-OH, Fmoc-Ile-OH, Fmoc-Asp(tBu)-OH, Fmoc-Ile-OH, Fmoc-Pro-OH, Fmoc-Asp(tBu)-OH, Fmoc-Leu-OH, Fmoc-Ala-OH BOC-Lys(Aloc)-OH.They are dissolved in *N,N*-dimethylformamide (DMF) and, according to the sequence, activated using O-benzotriazol-1-yl-*N, N, N′, N′*-tetramethyl-uronium hexafluorophosphate (HBTU) and Diisopropylethylamine (DIEA). Removal of the Fmoc protecting group is achieved using a solution of 20% (V/V) piperidine in *N,N*-dimethylformamide (DMF) for 20 minutes (step 1).The selective deprotection of the Lys (Aloc) group is performed manually and accomplished by treating the resin with a solution of 3 eq of $Pd(PPh_3)_4$ dissolved in 5 mL of $CHCl_3$:NMM: HOAc (18:1:0.5) for 2 h (Step 2). The resin is then washed with $CHCl_3$ (6 x 5 mL), 20% HOAc in DCM (6 x 5 mL), DCM (6 x 5 mL), and DMF (6 x 5 mL). The synthesis is then re-automated for the addition of the 3-maleimidopropionic acid (Step 3). Between every coupling, the resin is washed 3 times with *N,N*-dimethylformamide (DMF) and 3 times with isopropanol. The peptide is cleaved from the resin using 85% TFA/5% TIS/5% thioanisole and 5% phenol, followed by precipitation by dry-ice cold Et_2O (Step 4). The product is purified by preparative reversed phased HPLC using a Varian (Rainin) preparative binary HPLC system: gradient elution of 30-55% B (0.045% TFA in H_2O (A) and 0.045% TFA in CH_3CN (B)) over 180 min at 9.5 mL/min using a Phenomenex Luna 10 μ phenyl-hexyl, 21 mm x 25 cm column and UV detector (Varian Dynamax UVD II) at λ 214 and 254 nm to afford the desired modified peptide (i.e., DAC) in >95% purity, as determined by RP-HPLC.



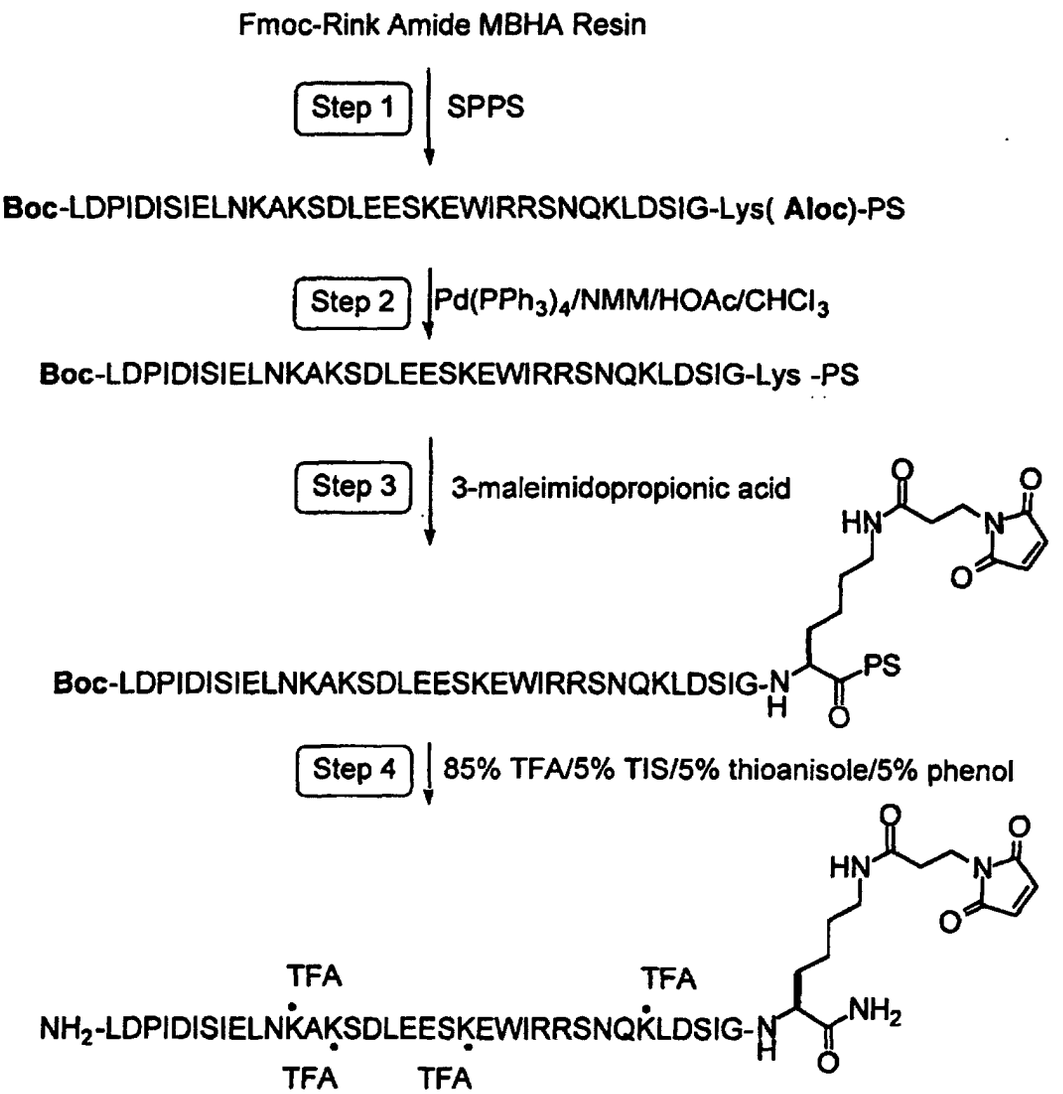
Example 13

Preparation of a Modified anti-HPIV peptide

[0123] In this example, the peptide SEQ ID NO:39 is synthesized and modified to include a linker and maleimide group according to the synthesis scheme set forth below. As reported in U.S. Pat. Nos. 6,013,236 and 6,020,459, SEQ ID NO:39 inhibits viral infection of human parainfluenza virus 3 (HPIV3), including inhibiting fusion and syncytia formation between HPIV3-infected Hep2 cells and uninfected CV-1W cells.

[0124] Solid phase peptide synthesis of the modified peptide on a 100 μmole scale is performed using manual solid-phase synthesis, a Symphony Peptide Synthesizer and Fmoc protected Rink Amide MBHA. The following protected amino acids are sequentially added to resin: Fmoc-Lys(Aloc)-OH, Fmoc-Gly-OH, Fmoc-Ile-OH, Fmoc-Ser(tBu)-OH, Fmoc-Asp(tBu)-OH, Fmoc-Leu-OH, Fmoc-Lys(Boc)-OH, Fmoc-Gly-OH, Fmoc-Asn(Trt)-OH, Fmoc-Ser(tBu)-OH, Fmoc-Arg(Pbf)-OH, Fmoc-Arg(Pbf)-OH, Fmoc-Ile-OH, Fmoc-Trp(Boc)-OH, Fmoc-Glu(tBu)-OH, Fmoc-Lys(Boc)-OH, Fmoc-Ser(tBu)-OH, Fmoc-Glu(tBu)-OH, Fmoc-Glu(tBu)-OH, Fmoc-Leu-OH, Fmoc-Asp(tBu)-OH, Fmoc-Ser(tBu)-OH, Fmoc-Lys(Boc)-OH, Fmoc-Ala-OH, Fmoc-Lys(Boc)-OH, Fmoc-Asn(Trt)-OH, Fmoc-Leu-OH, Fmoc-Glu(tBu)-OH, Fmoc-Ile-OH, Fmoc-Ser(tBu)-OH, Fmoc-Ile-OH, Fmoc-Asp(tBu)-OH, Fmoc-Ile-OH, Fmoc-Pro-OH, Fmoc-Asp(tBu)-OH, Fmoc-Leu-OH.They are dissolved in *N,N*-dimethylformamide (DMF) and, according to the sequence, activated using O-benzotriazol-1-yl-*N, N, N, N'*-tetramethyl-uronium hexafluorophosphate (HBTU) and Diisopropylethylamine (DIEA). Removal of the Fmoc protecting group is achieved using a solution of 20% (V/V) piperidine in *N,N*-dimethylformamide (DMF) for 20 minutes (step 1).The selective deprotection of the Lys (Aloc) group is performed manually and accomplished by treating the resin with a solution of 3 eq of $Pd(PPh_3)_4$ dissolved in 5 mL of $CHCl_3$:NMM:HOAc (18:1:0.5) for 2 h (Step 2). The resin is then washed with $CHCl_3$ (6 x 5 mL), 20% HOAc in DCM (6 x 5 mL), DCM (6 x

5 mL), and DMF (6 x 5 mL). The synthesis is then re-automated for the addition of the 3-maleimidopropionic acid (Step 3). Between every coupling, the resin is washed 3 times with *N,N*-dimethylformamide (DMF) and 3 times with isopropanol. The peptide is cleaved from the resin using 85% TFA/5% TIS/5% thioanisole and 5% phenol, followed by precipitation by dry-ice cold Et_2O (Step 4). The product is purified by preparative reversed phased HPLC using a Varian (Rainin) preparative binary HPLC system: gradient elution of 30-55% B (0.045% TFA in H_2O (A) and 0.045% TFA in CH_3CN (B)) over 180 min at 9.5 mL/min using a Phenomenex· Luna 10 μ phenyl-hexyl, 21 mm x 25 cm column and UV detector (Varian Dynamax UVD II) at λ 214 and 254 nm to afford the desired modified peptide (i.e., DAC) in >95% purity, as determined by RP-HPLC.

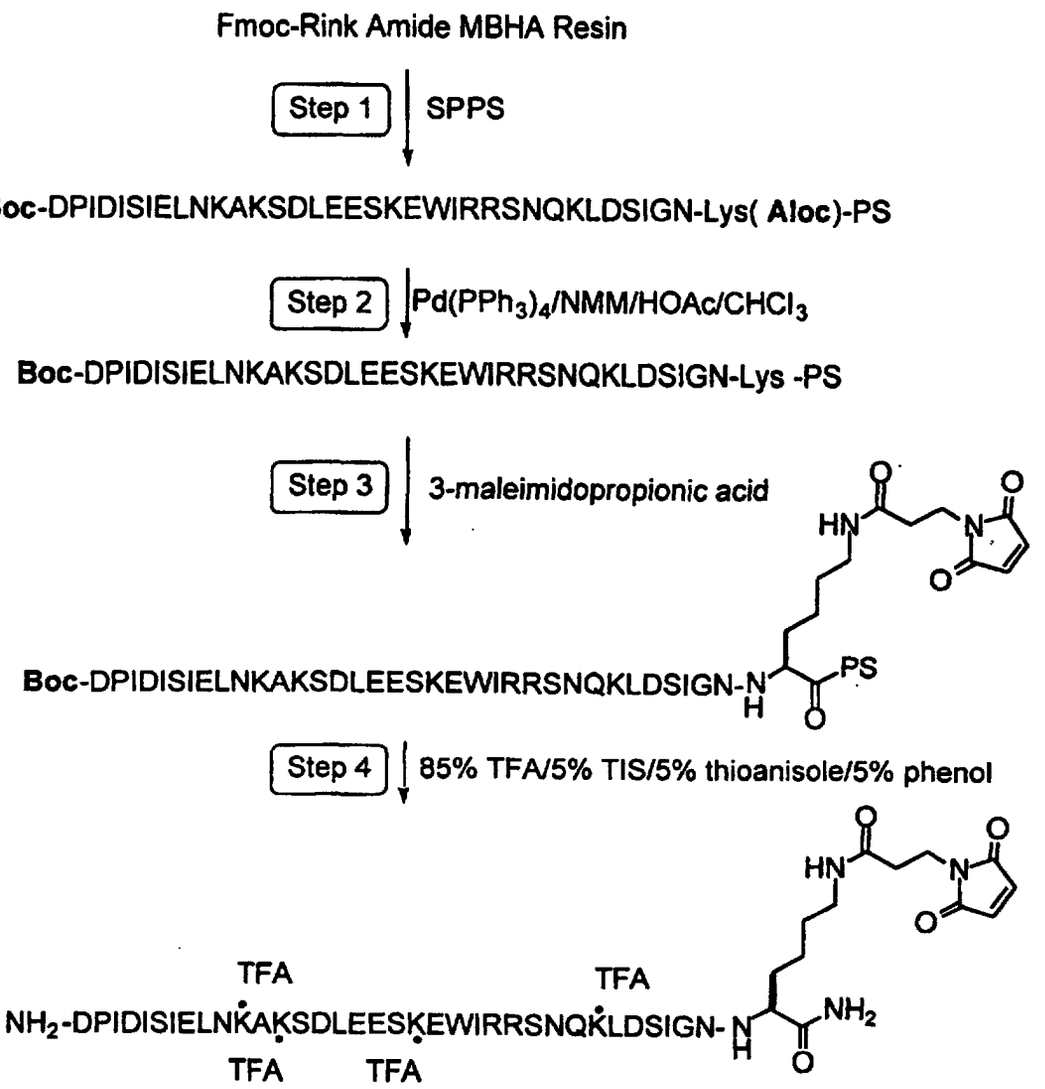


Example 14

Preparation of a Modified anti-HPIV peptide

[0125] In this example, the peptide SEQ ID NO:40 is synthesized and modified to include a linker and maleimide group according to the synthesis scheme set forth below. As reported in U.S. Pat. Nos. 6,013,236 and 6,020,459, SEQ ID NO. inhibits viral infection of human parainfluenza virus 3 (HPIV3), including inhibiting fusion and syncytia formation between HPIV3-infected Hep2 cells and uninfected CV-1W cells.

[0126] Solid phase peptide synthesis of the modofied peptide on a 100 μmole scale is performed using manual solid-phase synthesis, a Symphony Peptide Synthesizer and Fmoc protected Rink Amide MBHA. The following protected amino acids are sequentially added to resin: Fmoc-Lys(Aloc)-OH, Fmoc-Asn(Trt)-OH, Fmoc-Gly-OH, Fmoc-Ile-OH, Fmoc-Ser(tBu)-OH, Fmoc-Asp(tBu)-OH, Fmoc-Leu-OH, Fmoc-Lys(Boc)-OH, Fmoc-Gly-OH, Fmoc-Asn(Trt)-OH, Fmoc-Ser(tBu)-OH, Fmoc-Arg(Pbf)-OH, Fmoc-Arg(Pbf)-OH, Fmoc-Ile-OH, Fmoc-Trp(Boc)-OH, Fmoc-Glu(tBu)-OH, Fmoc-Lys(Boc)-OH, Fmoc-Ser(tBu)-OH, Fmoc-Glu(tBu)-OH, Fmoc-Glu(tBu)-OH, Fmoc-Leu-OH, Fmoc-Asp(tBu)-OH, Fmoc-Ser(tBu)-OH, Fmoc-Lys(Boc)-OH, Fmoc-Ala-OH, Fmoc-Lys(Boc)-OH, Fmoc-Asn(Trt)-OH, Fmoc-Leu-OH, Fmoc-Glu(tBu)-OH, Fmoc-Ile-OH, Fmoc-Ser(tBu)-OH, Fmoc-Ile-OH, Fmoc-Asp(tBu)-OH, Fmoc-Ile-OH, Fmoc-Pro-OH, Fmoc-Asp(tBu)-OH.They are dissolved in *N,N*-dimethylformamide (DMF) and, according to the sequence, activated using O-benzotriazol-1-yl-*N,N, N',N'*-tetramethyl-uronium hexafluorophosphate (HBTU) and Diisopropyl-ethylamine (DIEA). Removal of the Fmoc protecting group is achieved using a solution of 20% (V/V) piperidine in *N, N*-dimethylformamide (DMF) for 20 minutes (step 1).The selective deprotection of the Lys (Aloc) group is performed manually and accomplished by treating the resin with a solution of 3 eq of $Pd(PPh_3)_4$ dissolved in 5 mL of $CHCl_3$:NMM: HOAc (18:1:0.5) for 2 h (Step 2). The resin is then washed with $CHCl_3$ (6 x 5 mL), 20% HOAc in DCM (6 x 5 mL), DCM (6 x 5 mL), and DMF (6 x 5 mL). The synthesis is then re-automated for the addition of the 3-maleimidopropionic acid (Step 3). Between every coupling, the resin is washed 3 times with *N,N*-dimethylformamide (DMF) and 3 times with isopropanol. The peptide is cleaved from the resin using 85% TFA/5% TIS/5% thioanisole and 5% phenol, followed by precipitation by dry-ice cold Et_2O (Step 4). The product is purified by preparative reversed phased HPLC using a Varian (Rainin) preparative binary HPLC system: gradient elution of 30-55% B (0.045% TFA in H_2O (A) and 0.045% TFA in CH_3CN (B)) over 180 min at 9.5 mL/min using a Phenomenex Luna 10 μ phenyl-hexyl, 21 mm x 25 cm column and UV detector (Varian Dynamax UVD II) at λ 214 and 254 nm to afford the desired modified peptide (i.e., DAC) in >95% purity, as determined by RP-HPLC.



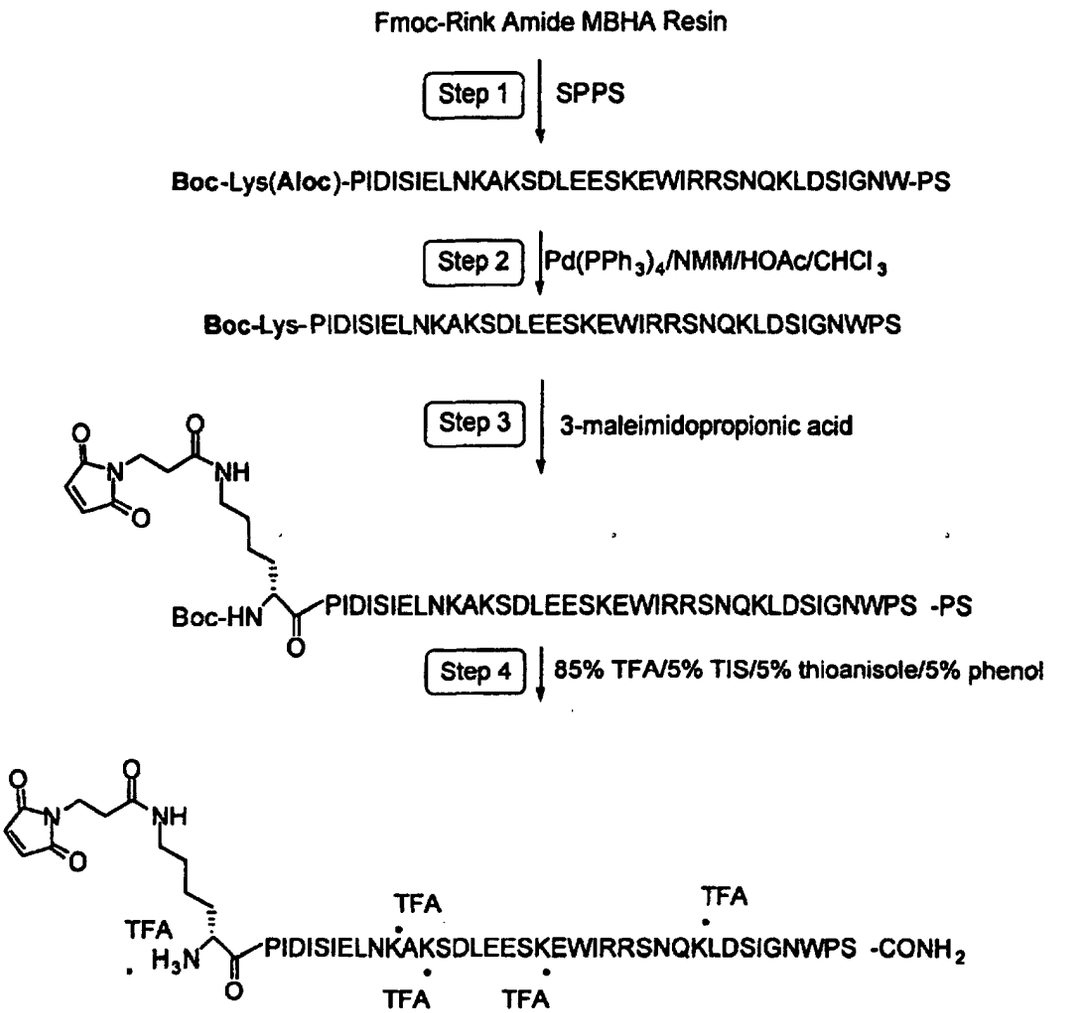
Example 15

Preparation of a Modified anti-HPIV peptide

[0127] In this example, the peptide SEQ ID NO:41 is synthesized and modified to include a linker and maleimide group according to the synthesis scheme set forth below. As reported in U.S. Pat. Nos. 6,013,236 and 6,020,459, SEQ ID NO:41 inhibits viral infection of human parainfluenza virus 3 (HPIV3), including inhibiting fusion and syncytia formation between HPIV3-infected Hep2 cells and uninfected CV-1W cells.

[0128] Solid phase peptide synthesis of the modified peptide on a 100 μmole scale is performed using manual solid-phase synthesis, a Symphony Peptide Synthesizer and Fmoc protected Rink Amide MBHA. The following protected amino acids are sequentially added to resin: Fmoc-Trp(Boc)-OH, Fmoc-Asn(Trt)-OH, Fmoc-Gly-OH, Fmoc-Ile-OH, Fmoc-Ser(tBu)-OH, Fmoc-Asp(tBu)-OH, Fmoc-Leu-OH, Fmoc-Lys(Boc)-OH, Fmoc-Gly-OH, Fmoc-Asn(Trt)-OH, Fmoc-Ser(tBu)-OH, Fmoc-Arg(Pbf)-OH, Fmoc-Arg(Pbf)-OH, Fmoc-Ile-OH, Fmoc-Trp(Boc)-OH, Fmoc-Glu(tBu)-OH, Fmoc-Lys(Boc)-OH, Fmoc-Ser(tBu)-OH, Fmoc-Glu(tBu)-OH, Fmoc-Glu(tBu)-OH, Fmoc-Leu-OH, Fmoc-Asp(tBu)-OH, Fmoc-Ser(tBu)-OH, Fmoc-Lys(Boc)-OH, Fmoc-Ala-OH, Fmoc-Lys(Boc)-OH, Fmoc-Asn(Trt)-OH, Fmoc-Leu-OH, Fmoc-Glu(tBu)-OH, Fmoc-Ile-OH, Fmoc-Ser(tBu)-OH, Fmoc-Ile-OH, Fmoc-Asp(tBu)-OH, Fmoc-Ile-OH, Fmoc-Pro-OH Boc-Lys(Aloc)-OH,. They are dissolved in *N,N*-dimethylformamide (DMF) and, according to the sequence, activated using O-benzotriazol-1-yl-*N,N, N',N'*-tetramethyl-uronium hexafluorophosphate (HBTU) and Diisopropylethylamine

(DIEA). Removal of the Fmoc protecting group is achieved using a solution of 20% (V/V) piperidine in *N,N*-dimethylformamide (DMF) for 20 minutes (step 1).The selective deprotection of the Lys (Aloc) group is performed manually and accomplished by treating the resin with a solution of 3 eq of $Pd(PPh_3)_4$ dissolved in 5 mL of $CHCl_3$:NMM:HOAc (18:1:0.5) for 2 h (Step 2). The resin is then washed with $CHCl_3$ (6 x 5 mL), 20% HOAc in DCM (6 x 5 mL), DCM (6 x 5 mL), and DMF (6 x 5 mL). The synthesis is then re-automated for the addition of the 3-maleimidopropionic acid (Step 3). Between every coupling, the resin is washed 3 times with *N,N*-dimethylformamide (DMF) and 3 times with isopropanol. The peptide is cleaved from the resin using 85% TFA/5% TIS/5% thioanisole and 5% phenol, followed by precipitation by dry-ice cold Et_2O (Step 4). The product is purified by preparative reversed phased HPLC using a Varian (Rainin) preparative binary HPLC system: gradient elution of 30-55% B (0.045% TFA in H_2O (A) and 0.045% TFA in CH_3CN (B)) over 180 min at 9.5 mL/min using a Phenomenex Luna 10 μ phenyl-hexyl, 21 mm x 25 cm column and UV detector (Varian Dynamax UVD II) at λ 214 and 254 nm to afford the desired modified peptide (i.e., DAC) in >95% purity, as determined by RP-HPLC.

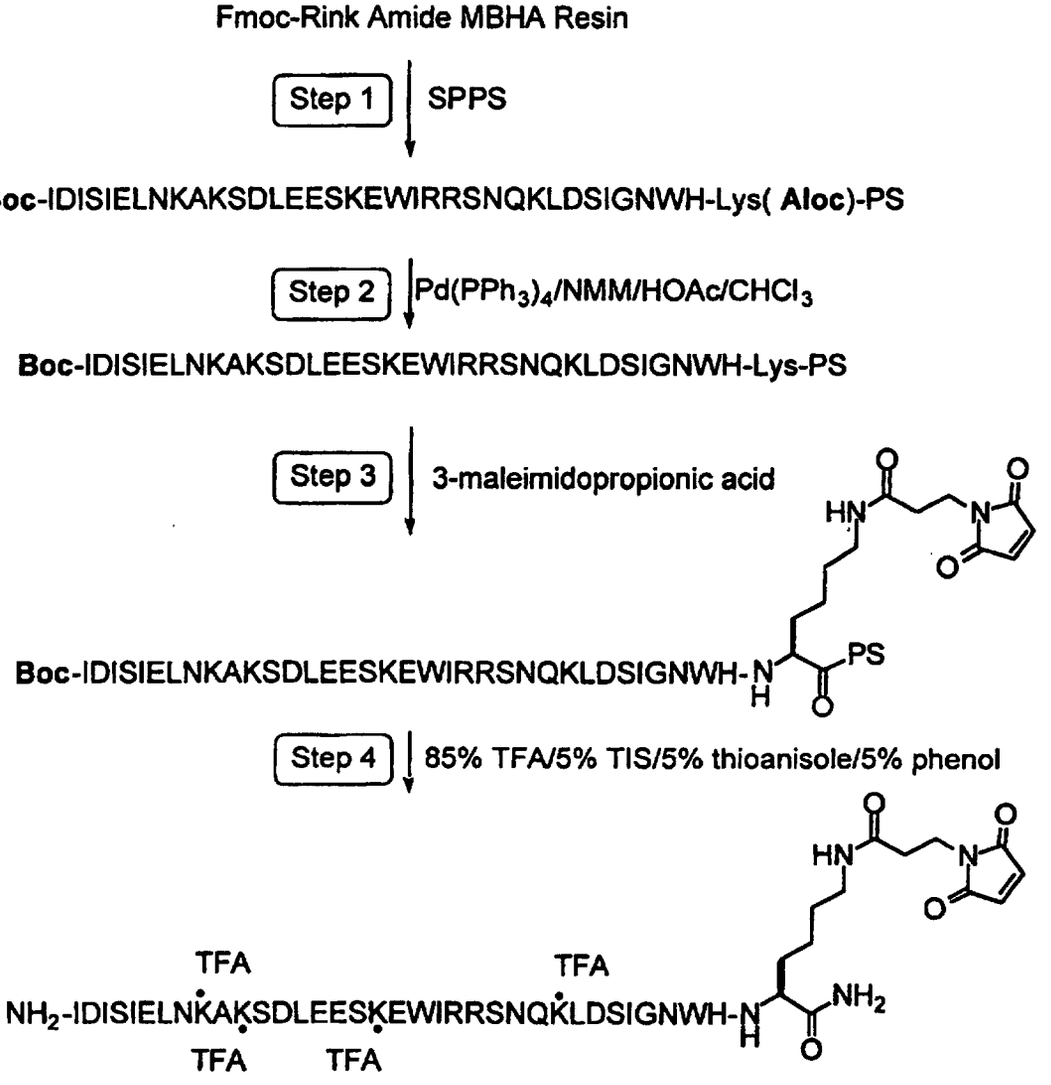


Example 16

Preparation of a Modified anti-HPIV peptide

[0129] In this example, the peptide SEQ ID NO:42 is synthesized and modified to include a linker and maleimide group according to the synthesis scheme set forth below. As reported in U.S. Pat. Nos. 6,013,236 and 6,020,459, SEQ ID NO:42 inhibits viral infection of human parainfluenza virus 3 (HPIV3), including inhibiting fusion and syncytia formation between HPIV3-infected Hep2 cells and uninfected CV-1W cells.

[0130] Solid phase peptide synthesis of the modified peptide on a 100 μmole scale is performed using manual solid-phase synthesis, a Symphony Peptide Synthesizer and Fmoc protected Rink Amide MBHA. The following protected amino acids are sequentially added to resin: Fmoc-Lys(Aloc)-OH, Fmoc-His(Boc)-OH, Fmoc-Trp(Boc)-OH, Fmoc-Asn (Trt)-OH, Fmoc-Gly-OH Fmoc-Ile-OH, Fmoc-Ser(tBu)-OH, Fmoc-Asp(tBu)-OH, Fmoc-Leu-OH, Fmoc-Lys(Boc)-OH, Fmoc-Gly-OH, Fmoc-Asn(Trt)-OH, Fmoc-Ser(tBu)-OH, Fmoc-Arg(Pbf)-OH, Fmoc-Arg(Pbf)-OH, Fmoc-Ile-OH, Fmoc-Trp(Boc)-OH, Fmoc-Glu(tBu)-OH, Fmoc-Lys(Boc)-OH, Fmoc-Ser(tBu)-OH, Fmoc-Glu(tBu)-OH, Fmoc-Glu(tBu)-OH, Fmoc-Leu-OH, Fmoc-Asp(tBu)-OH, Fmoc-Ser(tBu)-OH, Fmoc-Lys(Boc)-OH, Fmoc-Ala-OH, Fmoc-Lys(Boc)-OH, Fmoc-Asn(Trt)-OH, Fmoc-Leu-OH, Fmoc-Glu(tBu)-OH, Fmoc-Ile-OH, Fmoc-Ser(tBu)-OH, Fmoc-Ile-OH, Fmoc-Asp (tBu)-OH, Fmoc-Ile-OH. They are dissolved in *N,N*-dimethylformamide (DMF) and, according to the sequence, activated using O-benzotriazol-1-yl-*N, N, N', N*-tetramethyl-uronium hexafluorophosphate (HBTU) and Diisopropylethylamine (DIEA). Removal of the Fmoc protecting group is achieved using a solution of 20% (V/V) piperidine in *N,N*-dimethylformamide (DMF) for 20 minutes (step 1).The selective deprotection of the Lys (Aloc) group is performed manually and accomplished by treating the resin with a solution of 3 eq of $Pd(PPh_3)_4$ dissolved in 5 mL of $CHCl_3$:NMM: HOAc (18:1:0.5) for 2 h (Step 2). The resin is then washed with $CHCl_3$ (6 x 5 mL), 20% HOAc in DCM (6 x 5 mL), DCM (6 x 5 mL), and DMF (6 x 5 mL). The synthesis is then re-automated for the addition of the 3-maleimidopropionic acid (Step 3). Between every coupling, the resin is washed 3 times with *N,N*-dimethylformamide (DMF) and 3 times with isopropanol. The peptide is cleaved from the resin using 85% TFA/5% TIS/5% thioanisole and 5% phenol, followed by precipitation by dry-ice cold Et_2O (Step 4). The product is purified by preparative reversed phased HPLC using a Varian (Rainin) preparative binary HPLC system: gradient elution of 30-55% B (0.045% TFA in H_2O (A) and 0.045% TFA in CH_3CN (B)) over 180 min at 9.5 mL/min using a Phenomenex Luna 10 μ phenyl-hexyl, 21 mm x 25 cm column and UV detector (Varian Dynamax UVD II) at λ 214 and 254 nm to afford the desired modified peptide (i.e., DAC) in >95% purity, as determined by RP-HPLC



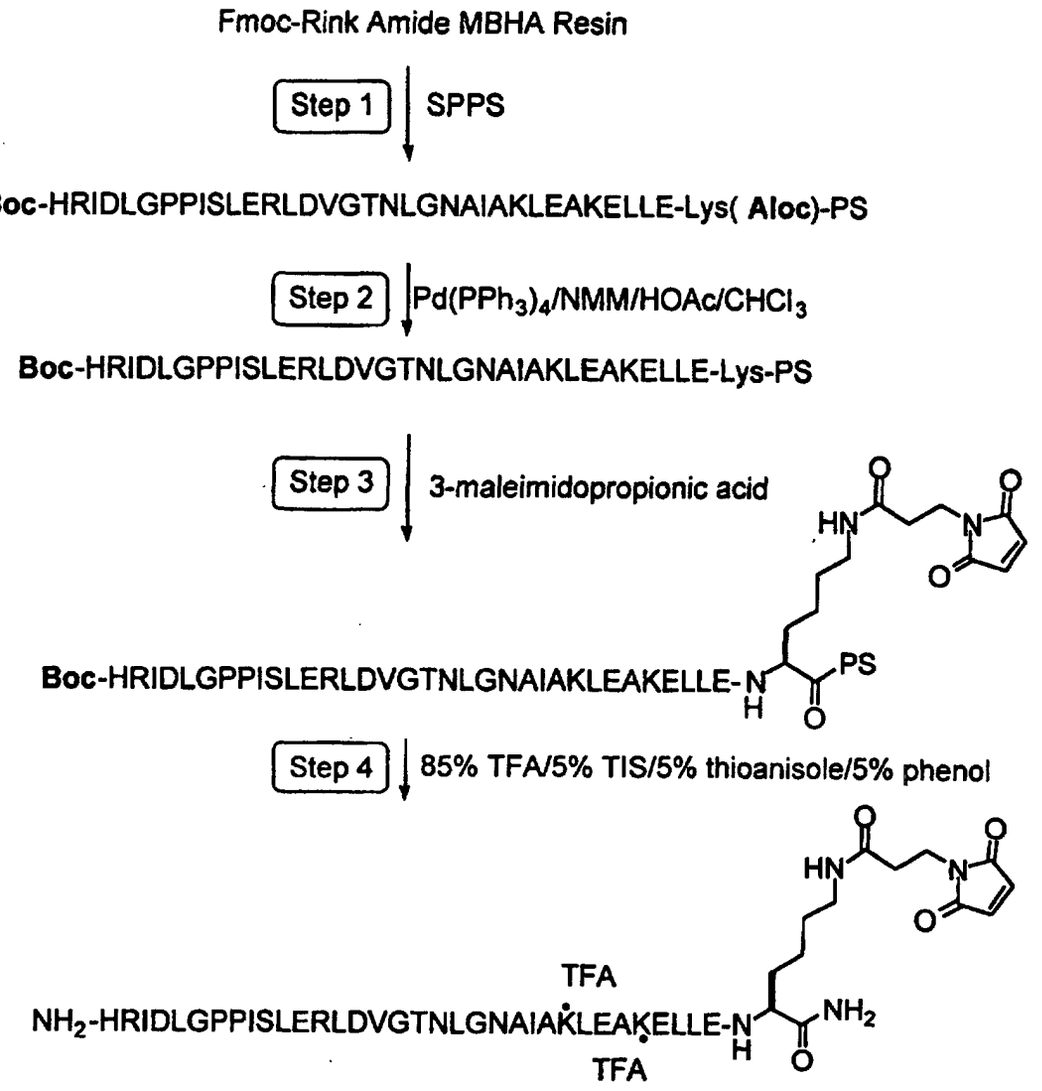
Example 17

Preparation of a Modified anti-MeV peptide

[0131] In this example, the peptide SEQ ID NO:77. is synthesized and modified to include a linker and maleimide group according to the synthesis scheme set forth below. As reported in U.S. Pat. Nos. 6,013,236 and 6,020,459, SEQ ID NO:77 inhibits viral infection of measles virus (MeV), including inhibiting fusion and syncytia formation between MeV-infected and uninfected Vero cells.

[0132] Solid phase peptide synthesis of the modified peptide on a 100 μmole scale is performed using manual solid-phase synthesis, a Symphony Peptide Synthesizer and Fmoc protected Rink Amide MBHA. The following protected amino acids are sequentially added to resin: Fmoc-Lys(Aloc)-OH, Fmoc-Glu(tBu)-OH, Fmoc-Leu-OH, Fmoc-Leu-OH, Fmoc-Glu(tBu)-OH, Fmoc-Lys(Boc)-OH, Fmoc-Ala-OH, Fmoc-Glu(tBu)-OH, Fmoc-Leu-OH, Fmoc-Lys(Boc)-OH, Fmoc-Ala-OH, Fmoc-Ile-OH, Fmoc-Ala-OH, Fmoc-Asn(Trt)-OH, Fmoc-Gly-OH, Fmoc-Leu-OH, Fmoc-Asn(Trt)-OH, Fmoc-Thr(tBu)-OH, Fmoc-Gly-OH, Fmoc-Val-OH, Fmoc-Asp(tBu)-OH, Fmoc-Leu-OH, Fmoc-Arg(Pbf)-OH, Fmoc-Glu (tBu)-OH, Fmoc-Leu-OH, Fmoc-Ser(tBu)-OH, Fmoc-Ile-OH, Fmoc-Pro-OH, Fmoc-Pro-OH, Fmoc-Gly-OH, Fmoc-Leu-OH, Fmoc-Asp(tBu)-OH, Fmoc-Ile-OH, Fmoc-Arg(Pbf)-OH, Fmoc-His(Boc)-OH. They are dissolved in *N,N*-dimethylformamide (DMF) and, according to the sequence, activated using O-benzotriazol-1-yl-*N, N, N′, N′*-tetramethyl-uronium hexafluorophosphate (HBTU) and Diisopropylethylamine (DIEA). Removal of the Fmoc protecting group is achieved using a solution of 20% (V/V) piperidine in *N,N*-dimethylformamide (DMF) for 20 minutes (step 1).The selec-

tive deprotection of the Lys (Aloc) group is performed manually and accomplished by treating the resin with a solution of 3 eq of $Pd(PPh_3)_4$ dissolved in 5 mL of $CHCl_3$:NMM:HOAc (18:1:0.5) for 2 h (Step 2). The resin is then washed with $CHCl_3$ (6 x 5 mL), 20% HOAc in DCM (6 x 5 mL), DCM (6 x 5 mL), and DMF (6 x 5 mL). The synthesis is then re-automated for the addition of the 3-maleimidopropionic acid (Step 3). Between every coupling, the resin is washed 3 times with *N,N*-dimethylformamide (DMF) and 3 times with isopropanol. The peptide is cleaved from the resin using 85% TFA/5% TIS/5% thioanisole and 5% phenol, followed by precipitation by dry-ice cold Et_2O (Step 4). The product is purified by preparative reversed phased HPLC using a Varian (Rainin) preparative binary HPLC system: gradient elution of 30-55% B (0.045% TFA in H_2O (A) and 0.045% TFA in CH_3CN (B)) over 180 min at 9.5 mL/min using a Phenomenex Luna 10 μ phenyl-hexyl, 21 mm x 25 cm column and UV detector (Varian Dynamax UVD II) at λ 214 and 254 nm to afford the desired modified peptide (i.e., DAC) in >95% purity, as determined by RP-HPLC.



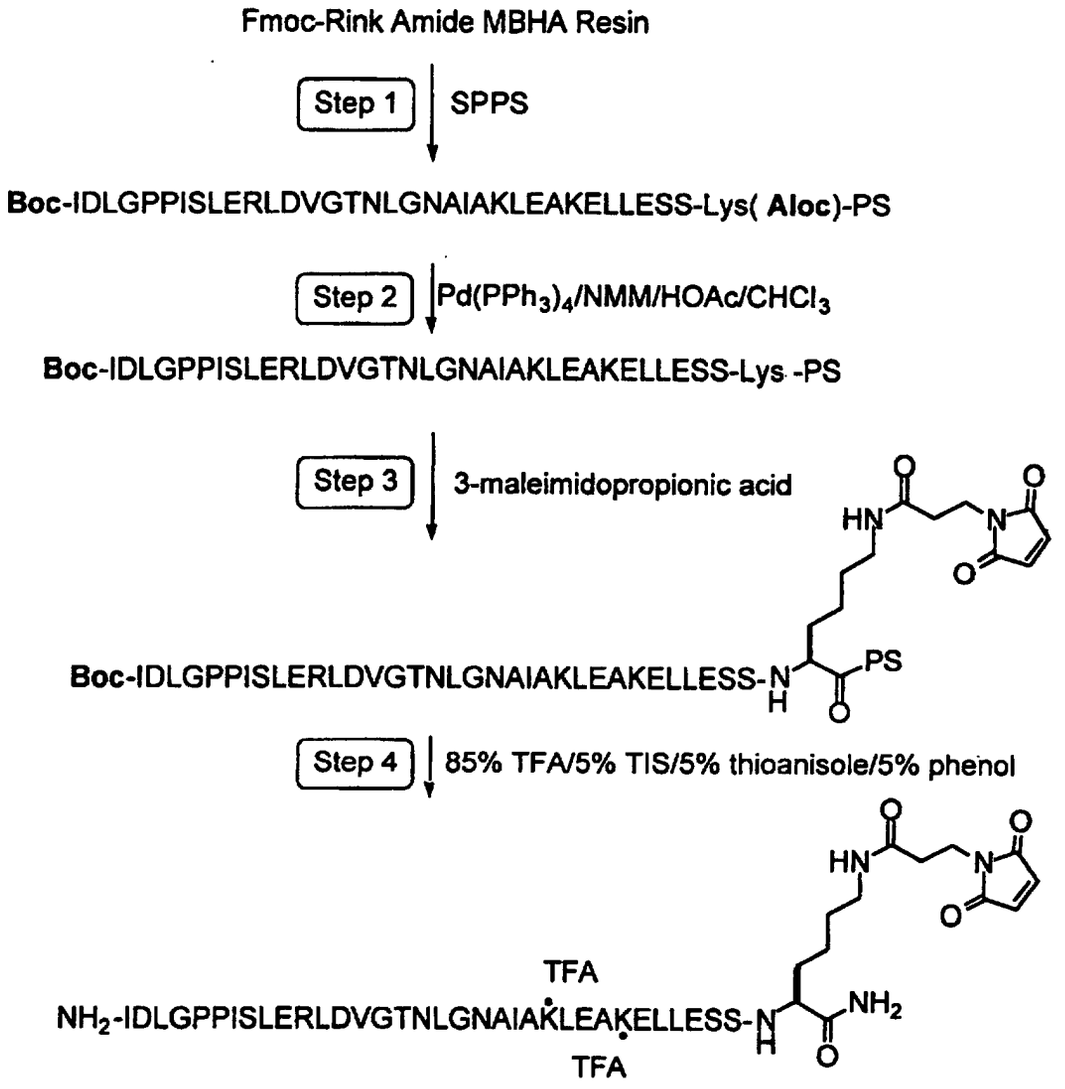
Example 18

Preparation of a Modified anti-MeV peptide

[0133] In this example, the peptide SEQ ID NO:79 is synthesized and modified to include a linker and maleimide group according to the synthesis scheme set forth below. As reported in U.S. Pat. Nos. 6,013,236 and 6,020,459, SEQ ID NO:79 inhibits viral infection of measles virus (MeV), including inhibiting fusion and syncytia formation between

MeV-infected and uninfected Vero cells.

[0134] Solid phase peptide synthesis of the modified peptide on a 100 μmole scale is performed using manual solid-phase synthesis, a Symphony Peptide Synthesizer and Fmoc protected Rink Amide MBHA. The following protected amino acids are sequentially added to resin: Fmoc-Lys(Aloc)-OH, Fmoc-Ser(tBu)-OH, Fmoc-Ser(tBu)-OH, Fmoc-Glu (tBu)-OH, Fmoc-Leu-OH, Fmoc-Leu-OH, Fmoc-Glu(tBu)-OH, Fmoc-Lys(Boc)-OH, Fmoc-Ala-OH, Fmoc-Glu(tBu)-OH, Fmoc-Leu-OH, Fmoc-Lys(Boc)-OH, Fmoc-Ala-OH, Fmoc-Ile-OH, Fmoc-Ala-OH, Fmoc-Asn(Trt)-OH, Fmoc-Gly-OH, Fmoc-Leu-OH, Fmoc-Asn(Trt)-OH, Fmoc-Thr(tBu)-OH, Fmoc-Gly-OH, Fmoc-Val-OH, Fmoc-Asp(tBu)-OH, Fmoc-Leu-OH, Fmoc-Arg(Pbf)-OH, Fmoc-Glu(tBu)-OH, Fmoc-Leu-OH, Fmoc-Ser(tBu)-OH, Fmoc-Ile-OH, Fmoc-Pro-OH, Fmoc-Pro-OH, Fmoc-Gly-OH, Fmoc-Leu-OH, Fmoc-Asp(tBu)-OH, Fmoc-Ile-OH, They are dissolved in *N,N*-dimethyl-formamide (DMF) and, according to the sequence, activated using O-benzotriazol-1-yl-*N, N, N',N'*-tetramethyl-uronium hexafluorophosphate (HBTU) and Diisopropylethylamine (DIEA). Removal of the Fmoc protecting group is achieved using a solution of 20% (V/V) piperidine in *N,N*-dimethylformamide (DMF) for 20 minutes (step 1).The selective deprotection of the Lys (Aloc) group is performed manually and accomplished by treating the resin with a solution of 3 eq of $Pd(PPh_3)_4$ dissolved in 5 mL of $CHCl_3$:NMM:HOAc (18:1:0.5) for 2 h (Step 2). The resin is then washed with $CHCl_3$ (6 x 5 mL), 20% HOAc in DCM (6 x 5 mL), DCM (6 x 5 mL), and DMF (6 x 5 mL). The synthesis is then re-automated for the addition of the 3-maleimidopropionic acid (Step 3). Between every coupling, the resin is washed 3 times with *N,N*-dimethylformamide (DMF) and 3 times with isopropanol. The peptide is cleaved from the resin using 85% TFA/5% TIS/5% thioanisole and 5% phenol, followed by precipitation by dry-ice cold Et_2O (Step 4). The product is purified by preparative reversed phased HPLC using a Varian (Rainin) preparative binary HPLC system: gradient elution of 30-55% B (0.045% TFA in H_2O (A) and 0.045% TFA in CH_3CN (B)) over 180 min at 9.5 mL/min using a Phenomenex Luna 10 μ phenyl-hexyl, 21 mm x 25 cm column and UV detector (Varian Dynamax UVD II) at λ 214 and 254 nm to afford the desired modified peptide (i.e., DAC) in >95% purity, as determined by RP-HPLC



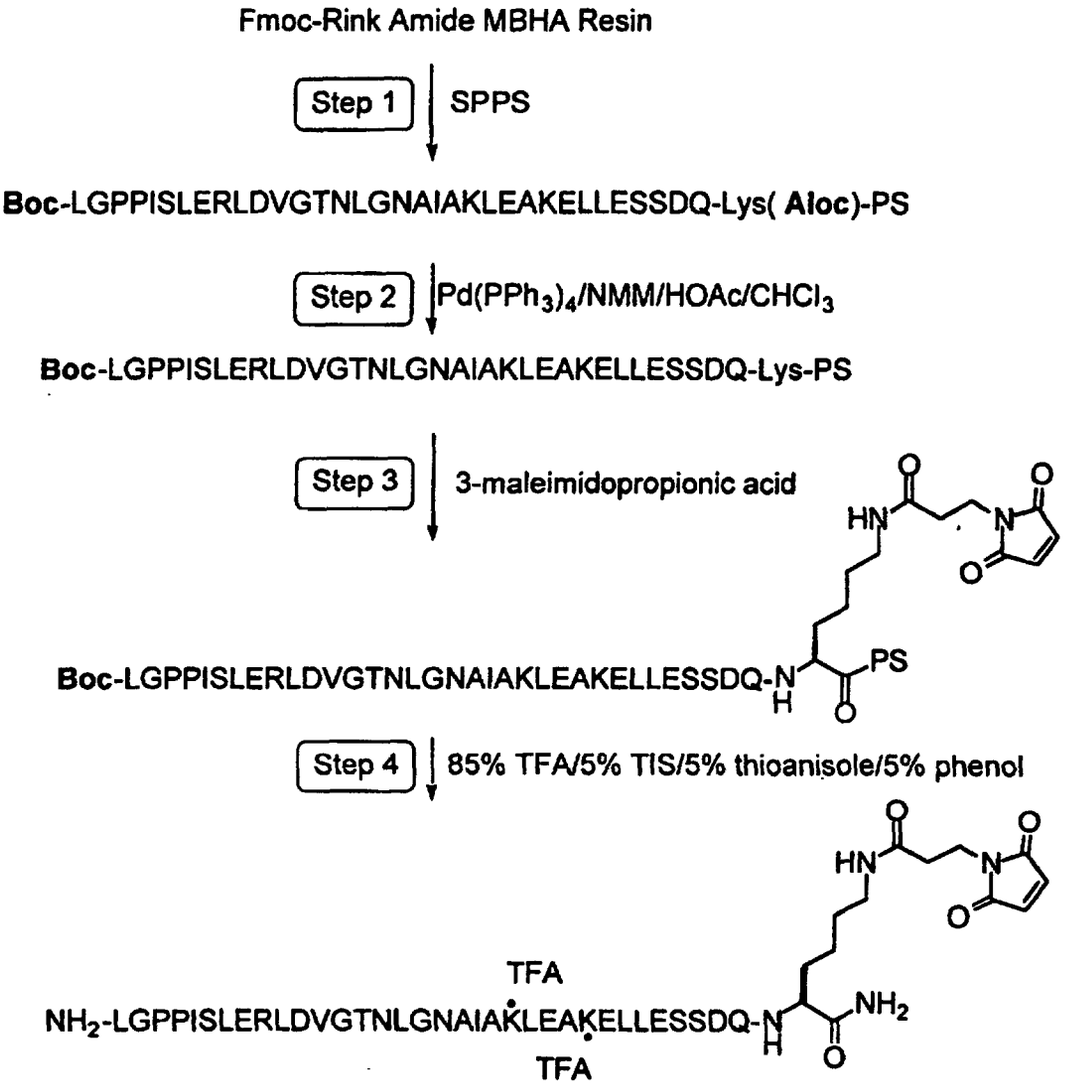
Example 19

Preparation of a Modified anti-MeV peptide

[0135] In this example, the peptide SEQ ID NO:81 is synthesized and modified to include a linker and maleimide group according to the synthesis scheme set forth below. As reported in U.S. Pat. Nos. 6,013,236 and 6,020,459, SEQ ID NO: 79 inhibits viral infection of measles virus (MeV), including inhibiting fusion and syncytia formation between MeV-infected and uninfected Vero cells.

[0136] Solid phase peptide synthesis of the modified peptide on a 100 μmole scale is performed using manual solid-phase synthesis, a Symphony Peptide Synthesizer and Fmoc protected Rink Amide MBHA. The following protected amino acids are sequentially added to resin: Fmoc-Lys(Aloc)-OH, Fmoc-Gln(Trt)-OH, Fmoc-Asp(tBu)-OH,Fmoc-Ser (tBu)-OH, Fmoc-Ser(tBu)-OH, Fmoc-Glu(tBu)-OH, Fmoc-Leu-OH, Fmoc-Leu-OH, Fmoc-Glu(tBu)-OH, Fmoc-Lys(Boc) -OH, Fmoc-Ala-OH, Fmoc-Glu(tBu)-OH, Fmoc-Leu-OH, Fmoc-Lys(Boc)-OH, Fmoc-Ala-OH, Fmoc-Ile-OH, Fmoc-Ala-OH, Fmoc-Asn(Trt)-OH, Fmoc-Gly-OH, Fmoc-Leu-OH, Fmoc-Asn(Trt)-OH, Fmoc-Thr(tBu)-OH, Fmoc-Gly-OH, Fmoc-Val-OH, Fmoc-Asp(tBu)-OH, Fmoc-Leu-OH, Fmoc-Arg(Pbf)-OH, Fmoc-Glu(tBu)-OH, Fmoc-Leu-OH, Fmoc-Ser (tBu)-OH, Fmoc-Ile-OH, Fmoc-Pro-OH, Fmoc-Pro-OH, Fmoc-Gly-OH, Fmoc-Leu-OH. They are dissolved in *N,N*-dimethylformamide (DMF) and, according to the sequence, activated using O-benzotriazol-1-yl-*N, N, N',N'*-tetramethyl-uronium hexafluorophosphate (HBTU) and Diisopropylethylamine (DIEA). Removal of the Fmoc protecting group is achieved using a solution of 20% (V/V) piperidine in *N,N*-dimethylformamide (DMF) for 20 minutes (step 1).The selec-

tive deprotection of the Lys (Aloc) group is performed manually and accomplished by treating the resin with a solution of 3 eq of $Pd(PPh_3)_4$ dissolved in 5 mL of $CHCl_3$:NMM:HOAc (18:1:0.5) for 2 h (Step 2). The resin is then washed with $CHCl_3$ (6 x 5 mL), 20% HOAc in DCM (6 x 5 mL), DCM (6 x 5 mL), and DMF (6 x 5 mL). The synthesis is then re-automated for the addition of the 3-maleimidopropionic acid (Step 3). Between every coupling, the resin is washed 3 times with *N,N*-dimethylformamide (DMF) and 3 times with isopropanol. The peptide is cleaved from the resin using 85% TFA/5% TIS/5% thioanisole and 5% phenol, followed by precipitation by dry-ice cold Et_2O (Step 4). The product is purified by preparative reversed phased HPLC using a Varian (Rainin) preparative binary HPLC system: gradient elution of 30-55% B (0.045% TFA in H_2O (A) and 0.045% TFA in CH_3CN (B)) over 180 min at 9.5 mL/min using a Phenomenex Luna 10 μ phenyl-hexyl, 21 mm x 25 cm column and UV detector (Varian Dynamax UVD II) at λ 214 and 254 nm to afford the desired modified peptide (i.e., DAC) in >95% purity, as determined by RP-HPLC.

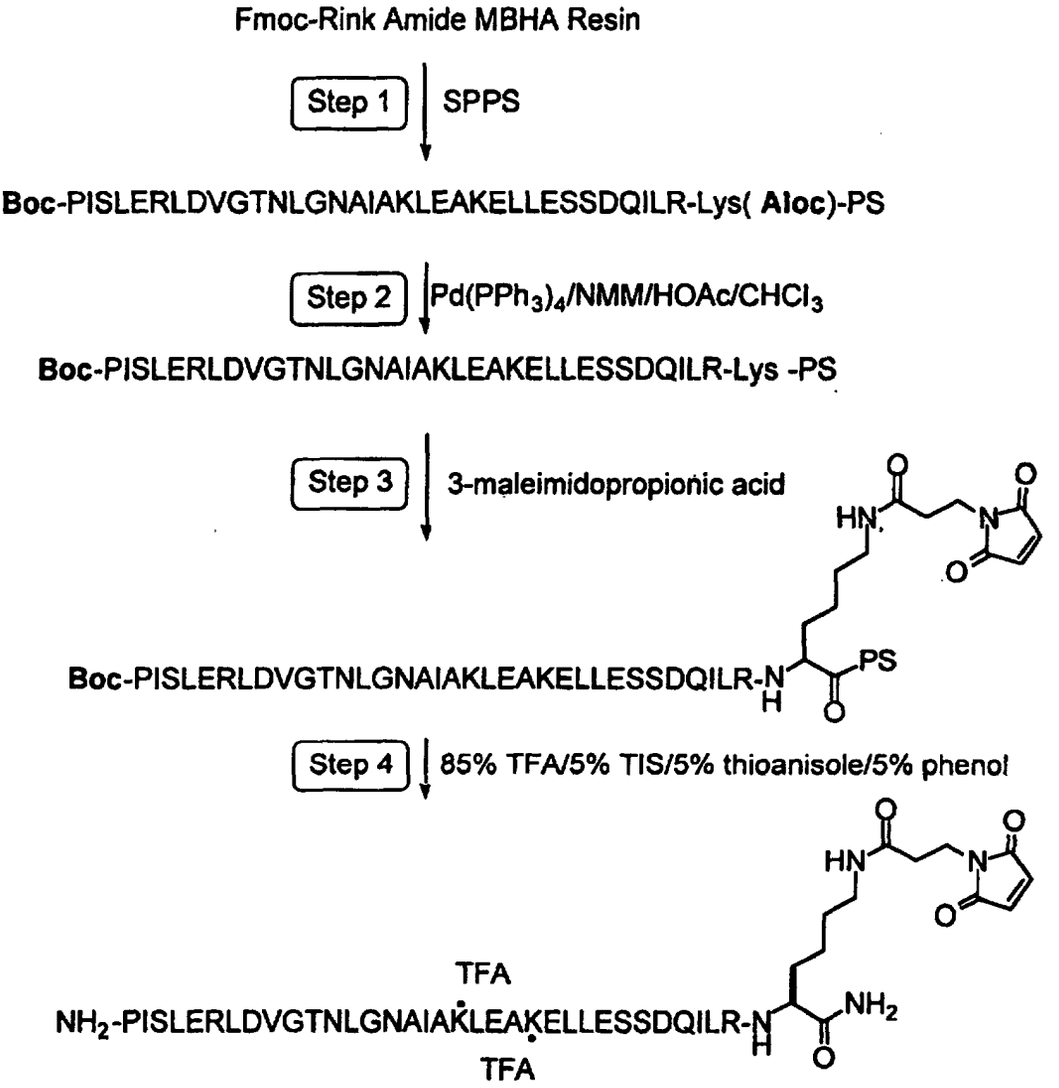


Example 20

Preparation of a Modified anti-MeV peptide

[0137] In this example, the peptide SEQ ID NO:84 is synthesized and modified to include a linker and maleimide group according to the synthesis scheme set forth below. As reported in U.S. Pat. Nos. 6,013,236 and 6,020,459, SEQ ID NO:84 inhibits viral infection of measles virus (MeV), including inhibiting fusion and syncytia formation between MeV-infected and uninfected Vero cells.

[0138] Solid phase peptide synthesis of the modified peptide on a 100 μmole scale is performed using manual solid-phase synthesis, a Symphony Peptide Synthesizer and Fmoc protected Rink Amide MBHA. The following protected amino acids are sequentially added to resin: Fmoc-Lys(Aloc)-OH, Fmoc-Arg(Pbf)-OH, Fmoc-Leu-OH, Fmoc-Ile-OH, Fmoc-Gln(Trt)-OH, Fmoc-Asp(tBu)-OH,Fmoc-Ser(tBu)-OH, Fmoc-Ser(tBu)-OH, Fmoc-Glu(tBu)-OH, Fmoc-Leu-OH, Fmoc-Leu-OH, Fmoc-Glu(tBu)-OH, Fmoc-Lys(Boc)-OH, Fmoc-Ala-OH, Fmoc-Glu(tBu)-OH, Fmoc-Leu-OH, Fmoc-Lys (Boc)-OH, Fmoc-Ala-OH, Fmoc-Ile-OH, Fmoc-Ala-OH, Fmoc-Asn(Trt)-OH, Fmoc-Gly-OH, Fmoc-Leu-OH, Fmoc-Asn (Trt)-OH, Fmoc-Thr(tBu)-OH, Fmoc-Gly-OH, Fmoc-Val-OH, Fmoc-Asp(tBu)-OH, Fmoc-Leu-OH, Fmoc-Arg(Pbf)-OH, Fmoc-Glu(tBu)-OH, Fmoc-Leu-OH, Fmoc-Ser(tBu)-OH, Fmoc-Ile-OH, Fmoc-Pro-OH. They are dissolved in *N,N*-dimethylformamide (DMF) and, according to the sequence, activated using O-benzotriazol-1-yl-*N, N, N′, N′*-tetramethyl-uronium hexafluorophosphate (HBTU) and Diisopropylethylamine (DIEA). Removal of the Fmoc protecting group is achieved using a solution of 20% (V/V) piperidine in *N,N*-dimethylformamide (DMF) for 20 minutes (step 1).The selective deprotection of the Lys (Aloc) group is performed manually and accomplished by treating the resin with a solution of 3 eq of $Pd(PPh_3)_4$ dissolved in 5 mL of $CHCl_3$:NMM:HOAc (18:1:0.5) for 2 h (Step 2). The resin is then washed with $CHCl_3$ (6 x 5 mL), 20% HOAc in DCM (6 x 5 mL), DCM (6 x 5 mL), and DMF (6 x 5 mL). The synthesis is then re-automated for the addition of the 3-maleimidopropionic acid (Step 3). Between every coupling, the resin is washed 3 times with *N,N*-dimethylformamide (DMF) and 3 times with isopropanol. The peptide is cleaved from the resin using 85% TFA/5% TIS/5% thioanisole and 5% phenol, followed by precipitation by dry-ice cold Et_2O (Step 4). The product is purified by preparative reversed phased HPLC using a Varian (Rainin) preparative binary HPLC system: gradient elution of 30-55% B (0.045% TFA in H_2O (A) and 0.045% TFA in CH_3CN (B)) over 180 min at 9.5 mL/min using a Phenomenex Luna 10 μ phenyl-hexyl, 21 mm x 25 cm column and UV detector (Varian Dynamax UVD II) at λ 214 and 254 nm to afford the desired modified peptide (i.e., DAC) in >95% purity, as determined by RP-HPLC.



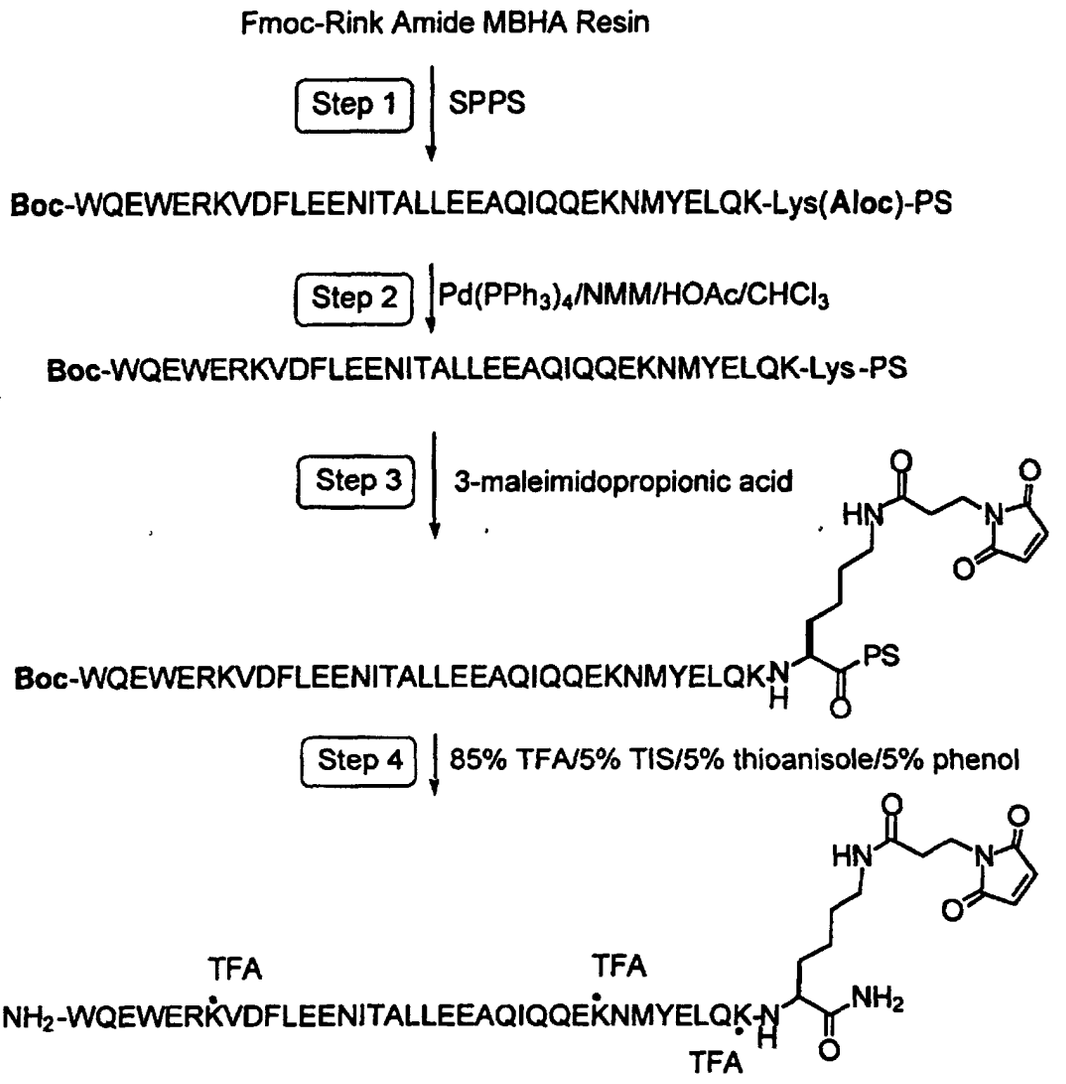
Example 21

Preparation of a Modified anti-SIV peptide

[0139] In this example, the peptide SEQ ID NO:64 is synthesized and modified to include a linker and maleimide group according to the synthesis scheme set forth below. As reported in U.S. Pat. Nos. 6,013,236 and 6,020,459, SEQ ID NO:64. exhibits potent antiviral activity as a crude peptide against simian immunodeficiency virus (SIV).

[0140] Solid phase peptide synthesis of the modified peptide on a 100 μmole scale is performed using manual solid-phase synthesis, a Symphony Peptide Synthesizer and Fmoc protected Rink Amide MBHA. The following protected amino acids are sequentially added to resin: Fmoc-Lys(Aloc)-OH, Fmoc-Lys(Boc)-OH, Fmoc-Gln(Trt)-OH, Fmoc-Leu-OH, Fmoc-Glu(tBu)-OH, Fmoc-Tyr(tBu)-OH, Fmoc-Met-OH, Fmoc-Asn(Trt)-OH, Fmoc-Lys(Boc)-OH, Fmoc-Glu (tBu)-OH, Fmoc-Gln(Trt)-OH, Fmoc-Gln(Trt)-OH, Fmoc-Ile-OH, Fmoc-Gln(Trt)-OH, Fmoc-Ala-OH, Fmoc-Glu(tBu) -OH, Fmoc-Glu(tBu)-OH, Fmoc-Leu-OH, Fmoc-Leu-OH, Fmoc-Ala-OH, Fmoc-Thr(tBu)-OH, Fmoc-Ile-OH, Fmoc-Asn (Trt)-OH, Fmoc-Glu(tBu)-OH, Fmoc-Glu(tBu)-OH, Fmoc-Leu-OH, Fmoc-Phe-OH, Fmoc-Asp(tBu)-OH, Fmoc-Val-OH, Fmoc-Lys(Boc)-OH, Fmoc-Arg(Pbf)-OH, Fmoc-Glu(tBu)-OH, Fmoc-Trp(Boc)-OH, Fmoc-Glu(tBu)-OH, Fmoc-Gln(Trt) -OH, Fmoc-Trp(Boc)-OH. They are dissolved in *N,N*-dimethylformamide (DMF) and, according to the sequence, activated using O-benzotriazol-1-yl-*N, N, N', N'*-tetramethyl-uronium hexafluorophosphate (HBTU) and Diisopropylethylamine (DIEA). Removal of the Fmoc protecting group is achieved using a solution of 20% (V/V) piperidine in *N,N*-

dimethylformamide (DMF) for 20 minutes (step 1).The selective deprotection of the Lys (Aloc) group is performed manually and accomplished by treating the resin with a solution of 3 eq of $Pd(PPh_3)_4$ dissolved in 5 mL of $CHCl_3$:NMM: HOAc (18:1:0.5) for 2 h (Step 2). The resin is then washed with $CHCl_3$ (6 x 5 mL), 20% HOAc in DCM (6 x 5 mL), DCM (6 x 5 mL), and DMF (6 x 5 mL). The synthesis is then re-automated for the addition of the 3-maleimidopropionic acid (Step 3). Between every coupling, the resin is washed 3 times with *N,N*-dimethylformamide (DMF) and 3 times with isopropanol. The peptide is cleaved from the resin using 85% TFA/5% TIS/5% thioanisole and 5% phenol, followed by precipitation by dry-ice cold Et_2O (Step 4). The product is purified by preparative reversed phased HPLC using a Varian (Rainin) preparative binary HPLC system: gradient elution of 30-55% B (0.045% TFA in H_2O (A) and 0.045% TFA in CH_3CN (B)) over 180 min at 9.5 mL/min using a Phenomenex Luna 10 μ phenyl-hexyl, 21 mm x 25 cm column and UV detector (Varian Dynamax UVD II) at λ 214 and 254 nm to afford the desired modified peptide (i.e., DAC) in >95% purity, as determined by RP-HPLC.



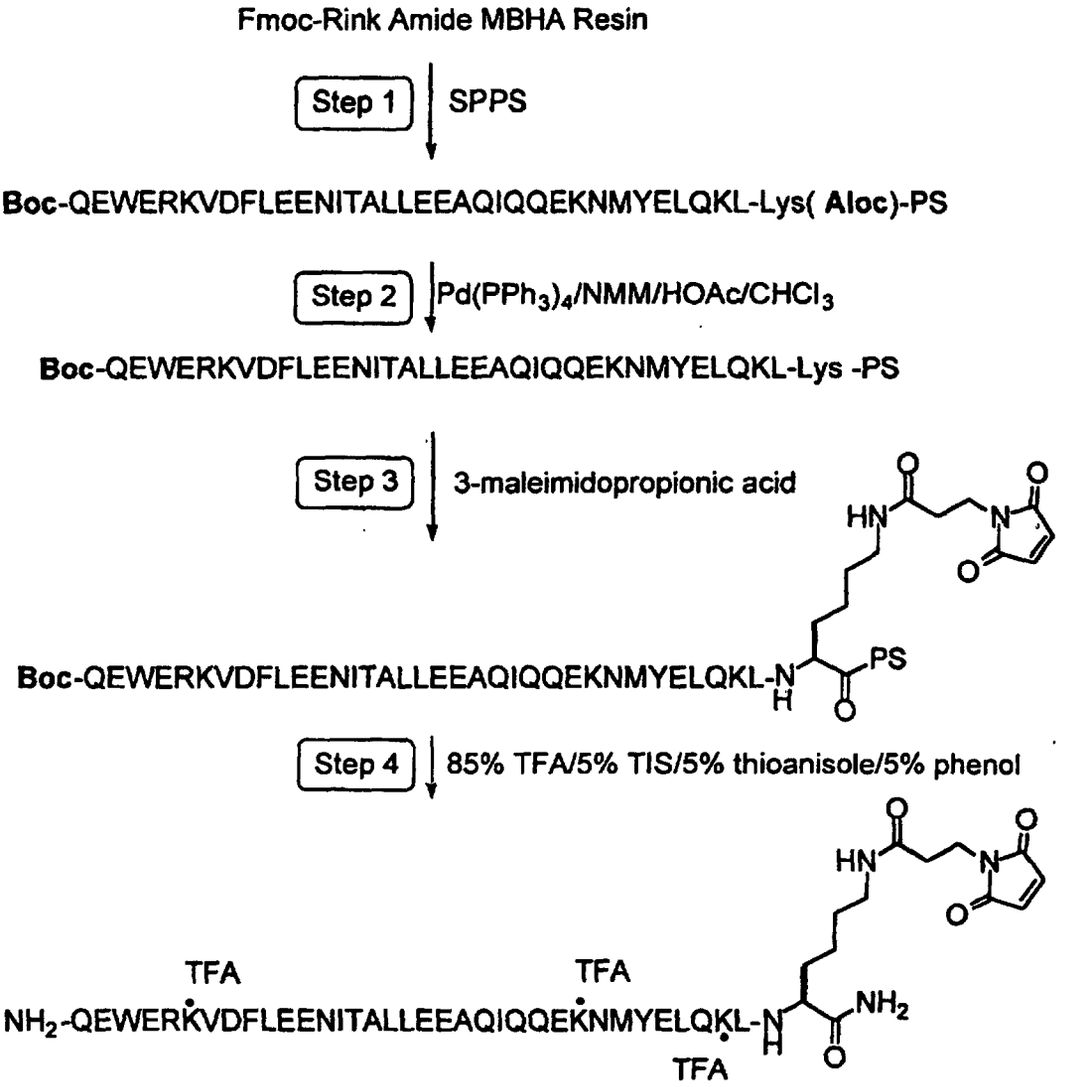
Example 22

Preparation of a Modified anti-SIV peptide

[0141] In this example, the peptide SEQ ID NO:65 is synthesized and modified to include a linker and maleimide

group according to the synthesis scheme set forth below. As reported in U.S. Pat. Nos. 6,013,236 and 6,020,459, SEQ ID NO:65 exhibits potent antiviral activity as a crude peptide against simian immunodeficiency virus (SIV).

[0142] Solid phase peptide synthesis of the modified peptide on a 100 μmole scale is performed using manual solid-phase synthesis, a Symphony Peptide Synthesizer and Fmoc protected Rink Amide MBHA. The following protected amino acids are sequentially added to resin: Fmoc-Lys(Aloc)-OH, Fmoc-Leu-OH, Fmoc-Lys(Boc)-OH, Fmoc-Gln(Trt)-OH, Fmoc-Leu-OH, Fmoc-Glu(tBu)-OH, Fmoc-Tyr(tBu)-OH, Fmoc-Met-OH, Fmoc-Asn(Trt)-OH, Fmoc-Lys(Boc)-OH, Fmoc-Glu(tBu)-OH, Fmoc-Gln(Trt)-OH, Fmoc-Gln(Trt)-OH, Fmoc-Ile-OH, Fmoc-Gln(Trt)-OH, Fmoc-Ala-OH, Fmoc-Glu(tBu)-OH, Fmoc-Glu(tBu)-OH, Fmoc-Leu-OH, Fmoc-Leu-OH, Fmoc-Ala-OH, Fmoc-Thr(tBu)-OH, Fmoc-Ile-OH, Fmoc-Asn(Trt)-OH, Fmoc-Glu(tBu)-OH, Fmoc-Glu(tBu)-OH, Fmoc-Leu-OH, Fmoc-Phe-OH, Fmoc-Asp(tBu)-OH, Fmoc-Val-OH, Fmoc-Lys(Boc)-OH, Fmoc-Arg(Pbf)-OH, Fmoc-Glu(tBu)-OH, Fmoc-Trp(Boc)-OH, Fmoc-Glu(tBu)-OH, Fmoc-Gln(Trt)-OH. They are dissolved in *N,N*-dimethylformamide (DMF) and, according to the sequence, activated using O-benzotriazol-1-yl-*N*,*N*, *N*′,*N*′-tetramethyl-uronium hexafluorophosphate (HBTU) and Diisopropylethylamine (DIEA). Removal of the Fmoc protecting group is achieved using a solution of 20% (V/V) piperidine in *N,N*-dimethylformamide (DMF) for 20 minutes (step 1).The selective deprotection of the Lys (Aloc) group is performed manually and accomplished by treating the resin with a solution of 3 eq of $Pd(PPh_3)_4$ dissolved in 5 mL of $CHCl_3$:NMM:HOAc (18:1:0.5) for 2 h (Step 2). The resin is then washed with $CHCl_3$ (6 x 5 mL), 20% HOAc in DCM (6 x 5 mL), DCM (6 x 5 mL), and DMF (6 x 5 mL). The synthesis is then re-automated for the addition of the 3-maleimidopropionic acid (Step 3). Between every coupling, the resin is washed 3 times with *N,N*-dimethylformamide (DMF) and 3 times with isopropanol. The peptide is cleaved from the resin using 85% TFA/5% TIS/5% thioanisole and 5% phenol, followed by precipitation by dry-ice cold Et_2O (Step 4). The product is purified by preparative reversed phased HPLC using a Varian (Rainin) preparative binary HPLC system: gradient elution of 30-55% B (0.045% TFA in H_2O (A) and 0.045% TFA in CH_3CN (B)) over 180 min at 9.5 mL/min using a Phenomenex Luna 10 μ phenyl-hexyl, 21 mm x 25 cm column and UV detector (Varian Dynamax UVD II) at λ 214 and 254 nm to afford the desired modified peptide (i.e., DAC) in >95% purity, as determined by RP-HPLC.



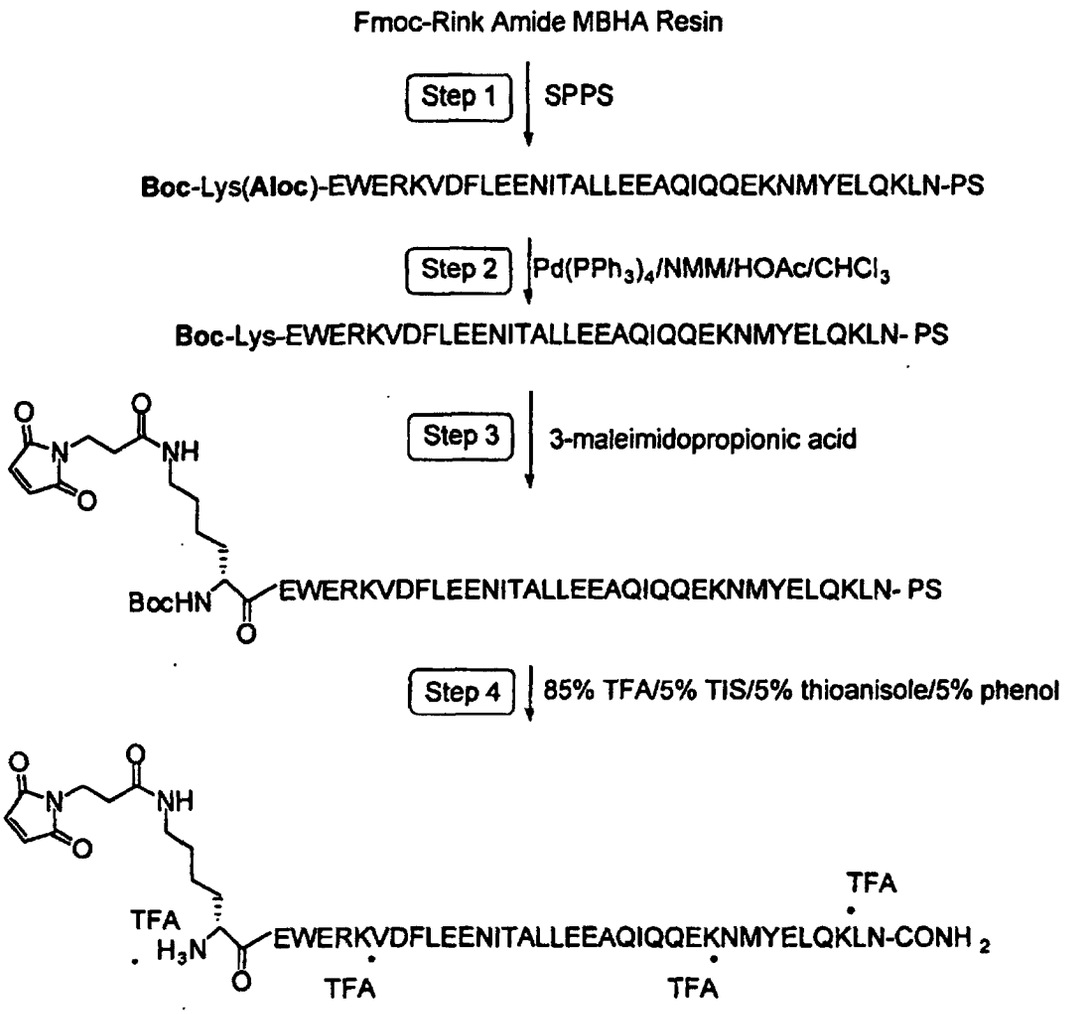
Example 23

Preparation of a Modified anti-SIV peptide

[0143] In this example, the peptide SEQ ID NO:66 is synthesized and modified to include a linker and maleimide group according to the synthesis scheme set forth below. As reported in U.S. Pat. Nos. 6,013,236 and 6,020,459, SEQ ID NO:66 exhibits potent antiviral activity as a crude peptide against simian immunodeficiency virus (SIV).

[0144] Solid phase peptide synthesis of the modified peptide on a 100 μmole scale is performed using manual solid-phase synthesis, a Symphony Peptide Synthesizer and Fmoc protected Rink Amide MBHA. The following protected amino acids are sequentially added to resin: Fmoc-Asn(Trt)-OH, Fmoc-Leu-OH, Fmoc-Lys(Boc)-OH, Fmoc-Gln(Trt)-OH, Fmoc-Leu-OH, Fmoc-Glu(tBu)-OH, Fmoc-Tyr(tBu)-OH, Fmoc-Met-OH, Fmoc-Asn(Trt)-OH, Fmoc-Lys(Boc)-OH, Fmoc-Glu(tBu)-OH, Fmoc-Gln(Trt)-OH, Fmoc-Gln(Trt)-OH, Fmoc-Ile-OH, Fmoc-Gln(Trt)-OH, Fmoc-Ala-OH, Fmoc-Glu(tBu)-OH, Fmoc-Glu(tBu)-OH, Fmoc-Leu-OH, Fmoc-Leu-OH, Fmoc-Ala-OH, Fmoc-Thr(tBu)-OH, Fmoc-Ile-OH, Fmoc-Asn(Trt)-OH, Fmoc-Glu(tBu)-OH, Fmoc-Glu(tBu)-OH, Fmoc-Leu-OH, Fmoc-Phe-OH, Fmoc-Asp(tBu)-OH,

Fmoc-Val-OH, Fmoc-Lys(Boc)-OH, Fmoc-Arg(Pbf)-OH, Fmoc-Glu(tBu)-OH, Fmoc-Trp(Boc)-OH, Fmoc-Glu(tBu)-OH Boc-Lys(Aloc)-OH,. They are dissolved in *N,N*-dimethylformamide (DMF) and, according to the sequence, activated using O-benzotriazol-1-yl-*N, N, N, N'*-tetramethyl-uronium hexafluorophosphate (HBTU) and Diisopropylethylamine (DIEA). Removal of the Fmoc protecting group is achieved using a solution of 20% (V/V) piperidine in *N,N*-dimethylformamide (DMF) for 20 minutes (step 1).The selective deprotection of the Lys (Aloc) group is performed manually and accomplished by treating the resin with a solution of 3 eq of $Pd(PPh_3)_4$ dissolved in 5 mL of $CHCl_3$:NMM:HOAc (18:1:0.5) for 2 h (Step 2). The resin is then washed with $CHCl_3$ (6 x 5 mL), 20% HOAc in DCM (6 x 5 mL), DCM (6 x 5 mL), and DMF (6 x 5 mL). The synthesis is then re-automated for the addition of the 3-maleimidopropionic acid (Step 3). Between every coupling, the resin is washed 3 times with *N,N*-dimethylformamide (DMF) and 3 times with isopropanol. The peptide is cleaved from the resin using 85% TFA/5% TIS/5% thioanisole and 5% phenol, followed by precipitation by dry-ice cold Et_2O (Step 4). The product is purified by preparative reversed phased HPLC using a Varian (Rainin) preparative binary HPLC system: gradient elution of 30-55% B (0.045% TFA in H_2O (A) and 0.045% TFA in CH_3CN (B)) over 180 min at 9.5 mL/min using a Phenomenex Luna 10 μ phenyl-hexyl, 21 mm x 25 cm column and UV detector (Varian Dynamax UVD II) at λ 214 and 254 nm to afford the desired modified peptide (i.e., DAC) in >95% purity, as determined by RP-HPLC.



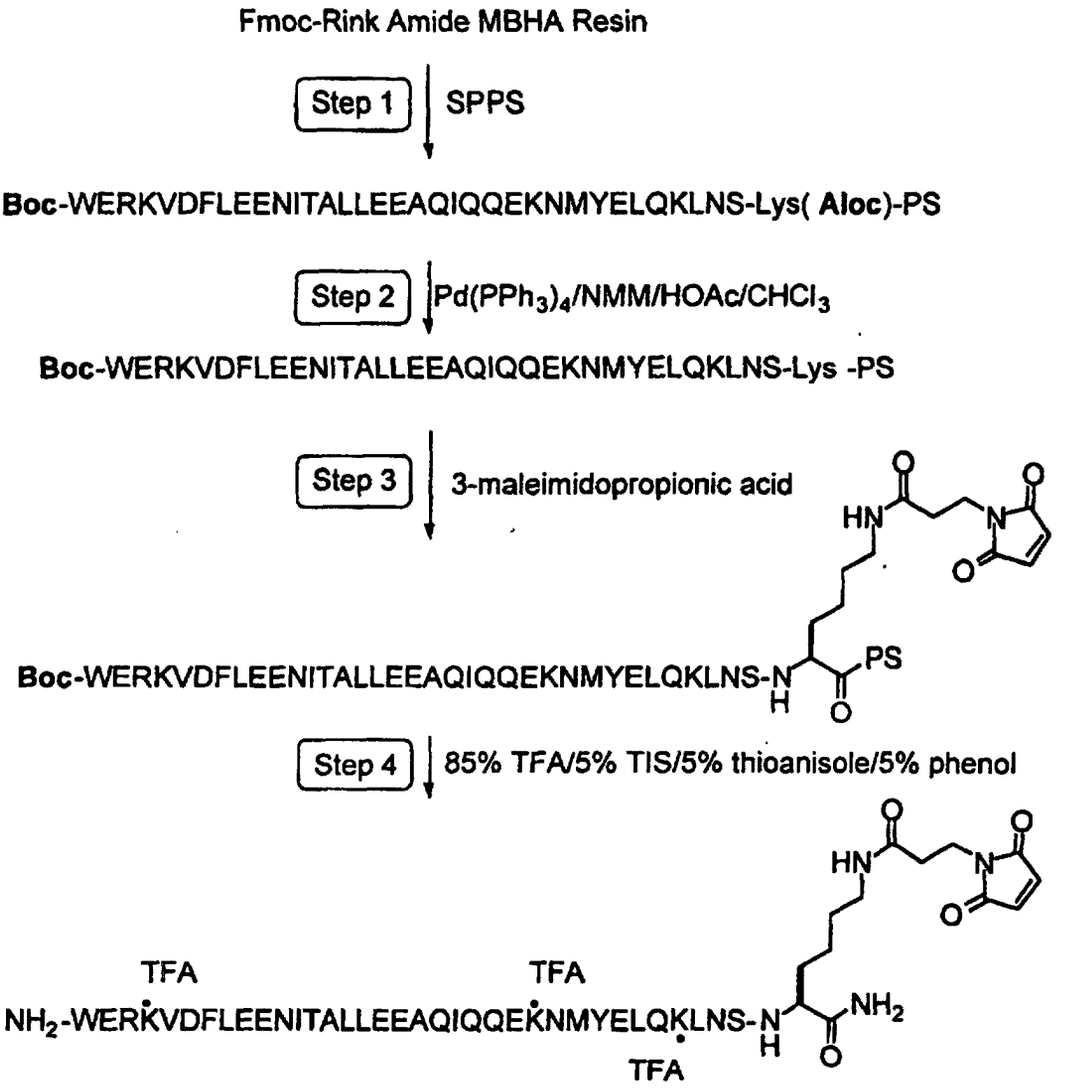
Example 24

Preparation of a Modified anti-SIV peptide

[0145] In this example, the peptide SEQ ID NO:67 is synthesized and modified to include a linker and maleimide

group according to the synthesis scheme set forth below. As reported in U.S. Pat. Nos. 6,013,236 and 6,020,459, SEQ ID NO:67 exhibits potent antiviral activity as a crude peptide against simian immunodeficiency virus (SIV).

[0146] Solid phase peptide synthesis of the modified peptide on a 100 μmole scale is performed using manual solid-phase synthesis, a Symphony Peptide Synthesizer and Fmoc protected Rink Amide MBHA. The following protected amino acids are sequentially added to resin: Fmoc-Lys(Aloc)-OH, Fmoc-Ser(tBu)-OH, Fmoc-Asn(Trt)-OH, Fmoc-Leu-OH, Fmoc-Lys(Boc)-OH, Fmoc-Gln(Trt)-OH, Fmoc-Leu-OH, Fmoc-Glu(tBu)-OH, Fmoc-Tyr(tBu)-OH, Fmoc-Met-OH, Fmoc-Asn(Trt)-OH, Fmoc-Lys(Boc)-OH, Fmoc-Glu(tBu)-OH, Fmoc-Gln(Trt)-OH, Fmoc-Gln(Trt)-OH, Fmoc-Ile-OH, Fmoc-Gln(Trt)-OH, Fmoc-Ala-OH, Fmoc-Glu(tBu)-OH, Fmoc-Glu(tBu)-OH, Fmoc-Leu-OH, Fmoc-Leu-OH, Fmoc-Ala-OH, Fmoc-Thr(tBu)-OH, Fmoc-Ile-OH, Fmoc-Asn(Trt)-OH, Fmoc-Glu(tBu)-OH, Fmoc-Glu(tBu)-OH, Fmoc-Leu-OH, Fmoc-Phe-OH, Fmoc-Asp(tBu)-OH, Fmoc-Val-OH, Fmoc-Lys(Boc)-OH, Fmoc-Arg(Pbf)-OH, Fmoc-Glu(tBu)-OH, Fmoc-Trp(Boc)-OH. They are dissolved in *N,N*-dimethylformamide (DMF) and, according to the sequence, activated using O-benzotriazol-1-yl-*N, N, N, N'*-tetramethyl-uronium hexafluorophosphate (HBTU) and Diisopropylethylamine (DIEA). Removal of the Fmoc protecting group is achieved using a solution of 20% (V/V) piperidine in *N,N*-dimethylformamide (DMF) for 20 minutes (step 1).The selective deprotection of the Lys (Aloc) group is performed manually and accomplished by treating the resin with a solution of 3 eq of $Pd(PPh_3)_4$ dissolved in 5 mL of $CHCl_3$:NMM: HOAc (18:1:0.5) for 2 h (Step 2). The resin is then washed with $CHCl_3$ (6 x 5 mL), 20% HOAc in DCM (6 x 5 mL), DCM (6 x 5 mL), and DMF (6 x 5 mL). The synthesis is then re-automated for the addition of the 3-maleimidopropionic acid (Step 3). Between every coupling, the resin is washed 3 times with *N,N*-dimethylformamide (DMF) and 3 times with isopropanol. The peptide is cleaved from the resin using 85% TFA/5% TIS/5% thioanisole and 5% phenol, followed by precipitation by dry-ice cold Et_2O (Step 4). The product is purified by preparative reversed phased HPLC using a Varian (Rainin) preparative binary HPLC system: gradient elution of 30-55% B (0.045% TFA in H_2O (A) and 0.045% TFA in CH_3CN (B)) over 180 min at 9.5 mL/min using a Phenomenex Luna 10 μ phenyl-hexyl, 21 mm x 25 cm column and UV detector (Varian Dynamax UVD II) at λ 214 and 254 nm to afford the desired modified peptide (i.e., DAC) in >95% purity, as determined by RP-HPLC.



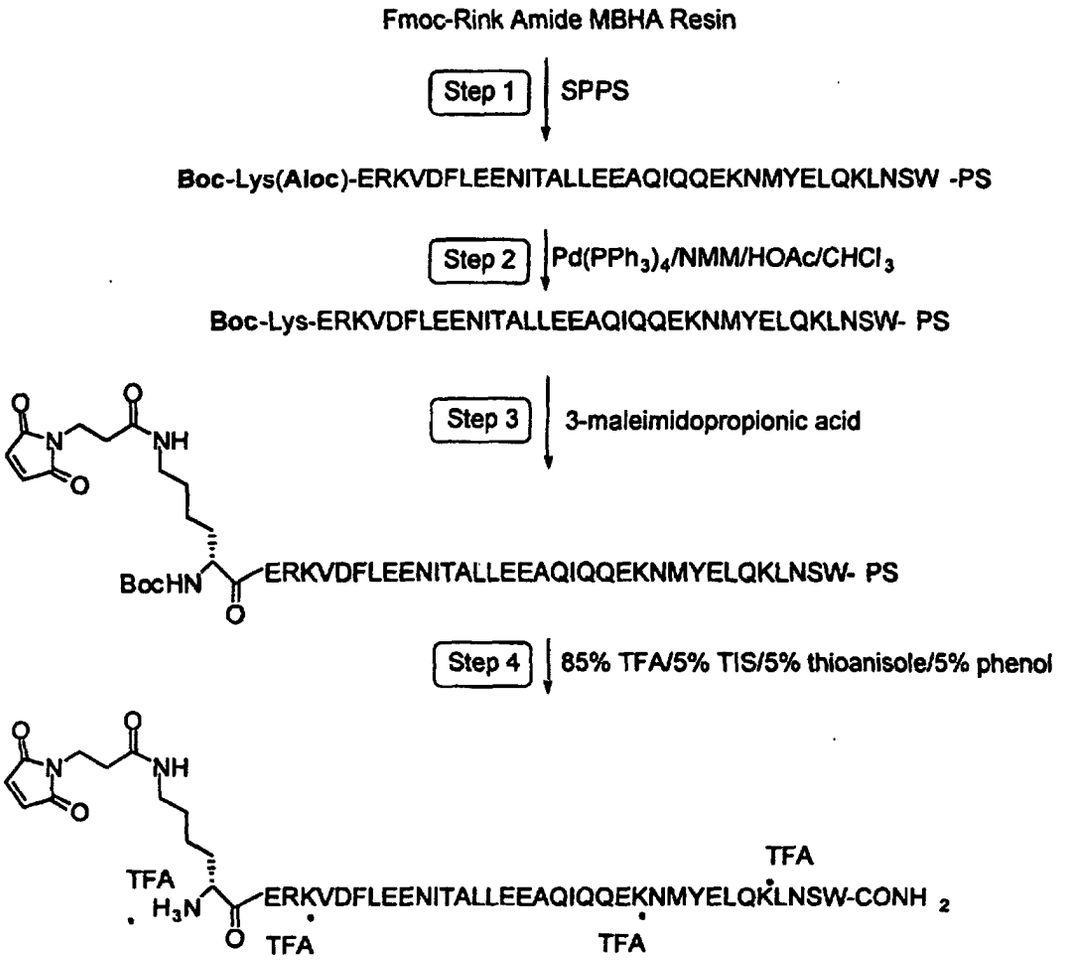
Example 25

Preparation of a Modified anti-SIV peptide

[0147] In this example, the peptide SEQ ID NO:68 is synthesized and modified to include a linker and maleimide group according to the synthesis scheme set forth below. As reported in U.S. Pat. Nos. 6,013,236 and 6,020,459, SEQ ID NO:68 exhibits potent antiviral activity as a crude peptide against simian immunodeficiency virus (SIV).

[0148] Solid phase peptide synthesis of the modified peptide on a 100 μmole scale is performed using manual solid-phase synthesis, a Symphony Peptide Synthesizer and Fmoc protected Rink Amide MBHA. The following protected amino acids are sequentially added to resin: Fmoc-Trp(Boc)-OH, Fmoc-Ser(tBu)-OH, Fmoc-Asn(Trt)-OH, Fmoc-Leu-OH, Fmoc-Lys(Boc)-OH, Fmoc-Gln(Trt)-OH, Fmoc-Leu-OH, Fmoc-Glu(tBu)-OH, Fmoc-Tyr(tBu)-OH, Fmoc-Met-OH, Fmoc-Asn(Trt)-OH, Fmoc-Lys(Boc)-OH, Fmoc-Glu(tBu)-OH, Fmoc-Gln(Trt)-OH, Fmoc-Gln(Trt)-OH, Fmoc-Ile-OH, Fmoc-Gln(Trt)-OH, Fmoc-Ala-OH, Fmoc-Glu(tBu)-OH, Fmoc-Glu(tBu)-OH, Fmoc-Leu-OH, Fmoc-Leu-OH, Fmoc-Ala-OH, Fmoc-Thr(tBu)-OH, Fmoc-Ile-OH, Fmoc-Asn(Trt)-OH, Fmoc-Glu(tBu)-OH, Fmoc-Glu(tBu)-OH, Fmoc-Leu-OH, Fmoc-Phe-OH, Fmoc-Asp(tBu)-OH, Fmoc-Val-OH, Fmoc-Lys(Boc)-OH, Fmoc-Arg(Pbf)-OH, Fmoc-Glu(tBu) -OH Boc-Lys(Aloc)-OH,. They are dissolved in *N,N*-dimethylformamide (DMF) and, according to the sequence, activated using O-benzotriazol-1-yl-*N, N, N′, N′*-tetramethyl-uronium hexafluorophosphate (HBTU) and Diisopropylethylamine (DIEA). Removal of the Fmoc protecting group is achieved using a solution of 20% (V/V) piperidine in *N,N*-dimethylformamide (DMF) for 20 minutes (step 1).The selective deprotection of the Lys (Aloc) group is performed

manually and accomplished by treating the resin with a solution of 3 eq of $Pd(PPh_3)_4$ dissolved in 5 mL of $CHCl_3$:NMM: HOAc (18:1:0.5) for 2 h (Step 2). The resin is then washed with $CHCl_3$ (6 x 5 mL), 20% HOAc in DCM (6 x 5 mL), DCM (6 x 5 mL), and DMF (6 x 5 mL). The synthesis is then re-automated for the addition of the 3-maleimidopropionic acid (Step 3). Between every coupling, the resin is washed 3 times with *N,N*-dimethylformamide (DMF) and 3 times with isopropanol. The peptide is cleaved from the resin using 85% TFA/5% TIS/5% thioanisole and 5% phenol, followed by precipitation by dry-ice cold Et_2O (Step 4). The product is purified by preparative reversed phased HPLC using a Varian (Rainin) preparative binary HPLC system: gradient elution of 30-55% B (0.045% TFA in H_2O (A) and 0.045% TFA in CH_3CN (B)) over 180 min at 9.5 mL/min using a Phenomenex Luna 10 μ phenyl-hexyl, 21 mm x 25 cm column and UV detector (Varian Dynamax UVD II) at λ 214 and 254 nm to afford the desired modified peptide (i.e., DAC) in >95% purity, as determined by RP-HPLC.



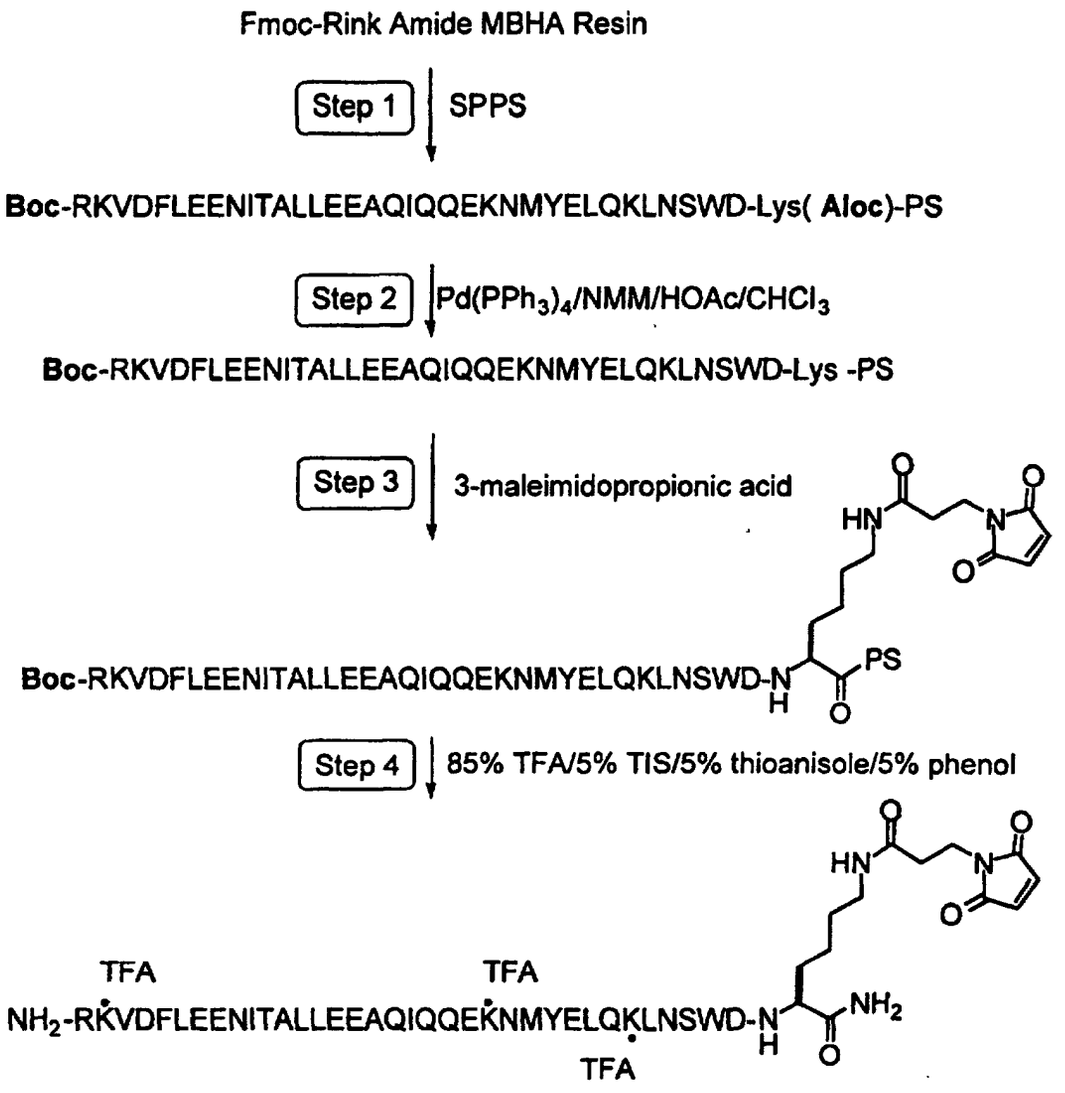
Example 26

Preparation of a Modified anti-SIV peptide

[0149] In this example, the peptide SEQ ID NO:69 is synthesized and modified to include a linker and maleimide group according to the synthesis scheme set forth below. As reported in U.S. Pat. Nos. 6,013,236 and 6,020,459, SEQ ID NO:69 exhibits potent antiviral activity as a crude peptide against simian immunodeficiency virus (SIV).

[0150] Solid phase peptide synthesis of the modified peptide on a 100 μmole scale is performed using manual solid-phase synthesis, a Symphony Peptide Synthesizer and Fmoc protected Rink Amide MBHA. The following protected amino acids are sequentially added to resin: Fmoc-Lys(Aloc)-OH, Fmoc-Asp(tBu)-OH, Fmoc-Trp(Boc)-OH, Fmoc-Ser (tBu)-OH, Fmoc-Asn(Trt)-OH, Fmoc-Leu-OH, Fmoc-Lys(Boc)-OH, Fmoc-Gln(Trt)-OH, Fmoc-Leu-OH, Fmoc-Glu(tBu) -OH, Fmoc-Tyr(tBu)-OH, Fmoc-Met-OH, Fmoc-Asn(Trt)-OH, Fmoc-Lys(Boc)-OH, Fmoc-Glu(tBu)-OH, Fmoc-Gln(Trt)

-OH, Fmoc-Gln(Trt)-OH, Fmoc-Ile-OH, Fmoc-Gln(Trt)-OH, Fmoc-Ala-OH, Fmoc-Glu(tBu)-OH, Fmoc-Glu(tBu)-OH, Fmoc-Leu-OH, Fmoc-Leu-OH, Fmoc-Ala-OH, Fmoc-Thr(tBu)-OH, Fmoc-Ile-OH, Fmoc-Asn(Trt)-OH, Fmoc-Glu(tBu)-OH, Fmoc-Glu(tBu)-OH, Fmoc-Leu-OH, Fmoc-Phe-OH, Fmoc-Asp(tBu)-OH, Fmoc-Val-OH, Fmoc-Lys(Boc)-OH, Fmoc-Arg(Pbf)-OH. They are dissolved in *N,N*-dimethylformamide (DMF) and, according to the sequence, activated using O-benzotriazol-1-yl-*N, N, N,N*'-tetramethyl-uronium hexafluorophosphate (HBTU) and Diisopropylethylamine (DIEA). Removal of the Fmoc protecting group is achieved using a solution of 20% (V/V) piperidine in *N,N*-dimethylformamide (DMF) for 20 minutes (step 1).The selective deprotection of the Lys (Aloc) group is performed manually and accomplished by treating the resin with a solution of 3 eq of $Pd(PPh_3)_4$ dissolved in 5 mL of $CHCl_3$:NMM:HOAc (18:1:0.5) for 2 h (Step 2). The resin is then washed with $CHCl_3$ (6 x 5 mL), 20% HOAc in DCM (6 x 5 mL), DCM (6 x 5 mL), and DMF (6 x 5 mL). The synthesis is then re-automated for the addition of the 3-maleimidopropionic acid (Step 3). Between every coupling, the resin is washed 3 times with *N,N*-dimethylformamide (DMF) and 3 times with isopropanol. The peptide is cleaved from the resin using 85% TFA/5% TIS/5% thioanisole and 5% phenol, followed by precipitation by dry-ice cold Et_2O (Step 4). The product is purified by preparative reversed phased HPLC using a Varian (Rainin) preparative binary HPLC system: gradient elution of 30-55% B (0.045% TFA in H_2O(A) and 0.045% TFA in CH_3CN (B)) over 180 min at 9.5 mL/min using a Phenomenex Luna 10 μ phenyl-hexyl, 21 mm x 25 cm column and UV detector (Varian Dynamax UVD II) at λ 214 and 254 nm to afford the desired modified peptide (i.e., DAC) in >95% purity, as determined by RP-HPLC.

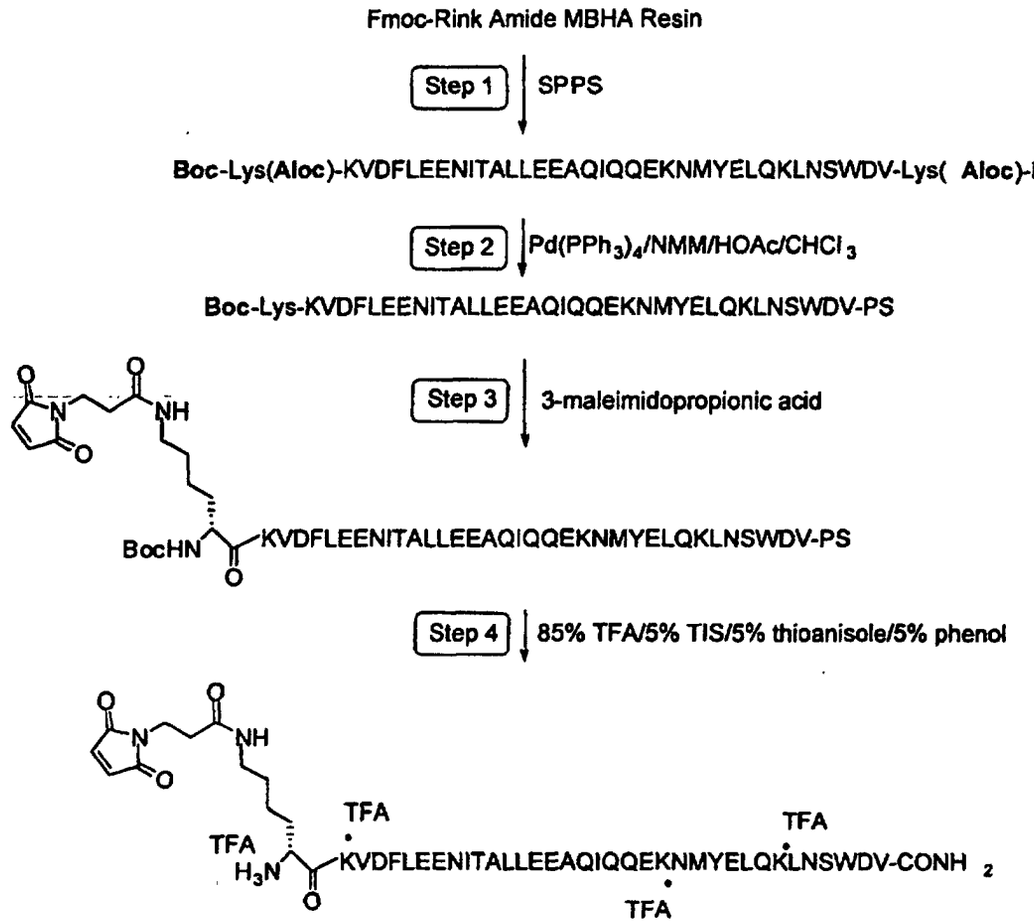


Example 27

Preparation of a Modified anti-SIV peptide

[0151] In this example, the peptide SEQ ID NO:70. is synthesized and modified to include a linker and maleimide group according to the synthesis scheme set forth below. As reported in U.S. Pat. Nos. 6,013,236 and 6,020,459, SEQ ID NO:70 exhibits potent antiviral activity as a crude peptide against simian immunodeficiency virus (SIV).

[0152] Solid phase peptide synthesis of the modified peptide on a 100 μmole scale is performed using manual solid-phase synthesis, a Symphony Peptide Synthesizer and Fmoc protected Rink Amide MBHA. The following protected amino acids are sequentially added to resin: Fmoc-Val-OH, Fmoc-Asp(tBu)-OH, Fmoc-Trp(Boc)-OH, Fmoc-Ser(tBu)-OH, Fmoc-Asn(Trt)-OH, Fmoc-Leu-OH, Fmoc-Lys(Boc)-OH, Fmoc-Gln(Trt)-OH, Fmoc-Leu-OH, Fmoc-Glu(tBu)-OH, Fmoc-Tyr(tBu)-OH, Fmoc-Met-OH, Fmoc-Asn(Trt)-OH, Fmoc-Lys(Boc)-OH, Fmoc-Glu(tBu)-OH, Fmoc-Gln(Trt)-OH, Fmoc-Gln(Trt)-OH, Fmoc-Ile-OH, Fmoc-Gln(Trt)-OH, Fmoc-Ala-OH, Fmoc-Glu(tBu)-OH, Fmoc-Glu(tBu)-OH, Fmoc-Leu-OH, Fmoc-Leu-OH, Fmoc-Ala-OH, Fmoc-Thr(tBu)-OH, Fmoc-Ile-OH, Fmoc-Asn(Trt)-OH, Fmoc-Glu(tBu)-OH, Fmoc-Glu(tBu)-OH, Fmoc-Leu-OH, Fmoc-Phe-OH, Fmoc-Asp(tBu)-OH, Fmoc-Val-OH, Fmoc-Lys(Boc)-OH, Boc-Lys(Aloc)-OH,. They are dissolved in *N,N*-dimethylformamide (DMF) and, according to the sequence, activated using O-benzotriazol-1-yl-*N, N, N′, N′*-tetramethyl-uronium hexafluorophosphate (HBTU) and Diisopropylethylamine (DIEA). Removal of the Fmoc protecting group is achieved using a solution of 20% (V/V) piperidine in *N,N*-dimethylformamide (DMF) for 20 minutes (step 1).The selective deprotection of the Lys (Aloc) group is performed manually and accomplished by treating the resin with a solution of 3 eq of $Pd(PPh_3)_4$ dissolved in 5 mL of $CHCl_3$:NMM:HOAc (18:1:0.5) for 2 h (Step 2). The resin is then washed with $CHCl_3$ (6 x 5 mL), 20% HOAc in DCM (6 x 5 mL), DCM (6 x 5 mL), and DMF (6 x 5 mL). The synthesis is then re-automated for the addition of the 3-maleimidopropionic acid (Step 3). Between every coupling, the resin is washed 3 times with *N,N*-dimethylformamide (DMF) and 3 times with isopropanol. The peptide is cleaved from the resin using 85% TFA/5% TIS/5% thioanisole and 5% phenol, followed by precipitation by dry-ice cold Et_2O (Step 4). The product is purified by preparative reversed phased HPLC using a Varian (Rainin) preparative binary HPLC system: gradient elution of 30-55% B (0.045% TFA in H_2O (A) and 0.045% TFA in CH_3CN (B)) over 180 min at 9.5 mL/min using a Phenomenex Luna 10 μ phenyl-hexyl, 21 mm x 25 cm column and UV detector (Varian Dynamax UVD II) at λ 214 and 254 nm to afford the desired modified peptide (i.e., DAC) in >95% purity, as determined by RP-HPLC.



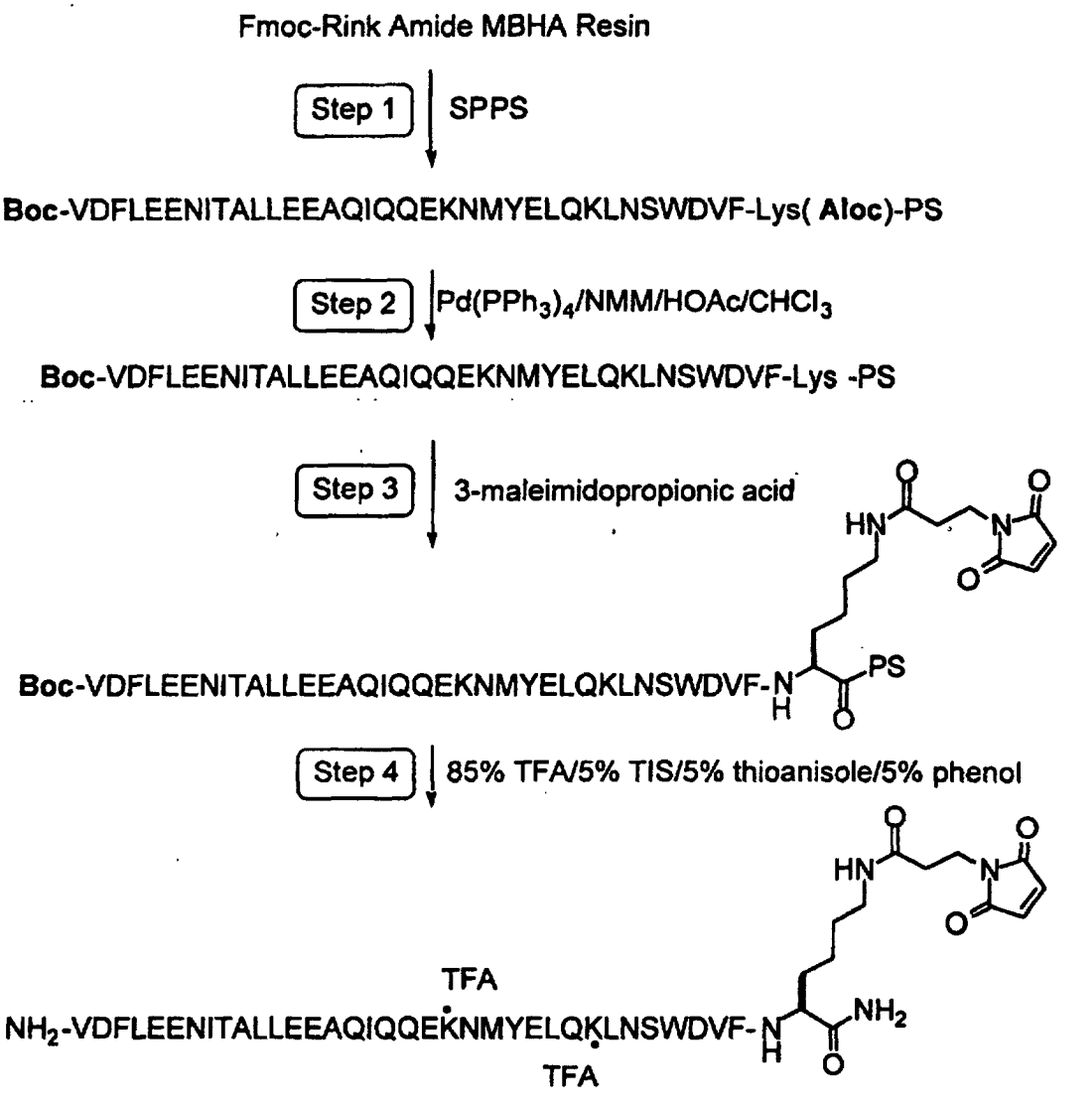
Example 28

Preparation of a Modified anti-SIV peptide

[0153] In this example, the peptide SEQ ID NO:71 is synthesized and modified to include a linker and maleimide group according to the synthesis scheme set forth below. As reported in U.S. Pat. Nos. 6,013,236 and 6,020,459, SEQ ID NO:71 exhibits potent antiviral activity as a crude peptide against simian immunodeficiency virus (SIV).

[0154] Solid phase peptide synthesis of the modified peptide on a 100 μmole scale is performed using manual solid-phase synthesis, a Symphony Peptide Synthesizer and Fmoc protected Rink Amide MBHA. The following protected amino acids are sequentially added to resin: Fmoc-Lys(Aloc)-OH, Fmoc-Phe-OH, Fmoc-Val-OH, Fmoc-Asp(tBu)-OH, Fmoc-Trp(Boc)-OH, Fmoc-Ser(tBu)-OH, Fmoc-Asn(Trt)-OH, Fmoc-Leu-OH, Fmoc-Lys(Boc)-OH, Fmoc-Gln(Trt)-OH, Fmoc-Leu-OH, Fmoc-Glu(tBu)-OH, Fmoc-Tyr(tBu)-OH, Fmoc-Met-OH, Fmoc-Asn(Trt)-OH, Fmoc-Lys(Boc)-OH, Fmoc-Glu(tBu)-OH, Fmoc-Gln(Trt)-OH, Fmoc-Gln(Trt)-OH, Fmoc-Ile-OH, Fmoc-Gln(Trt)-OH, Fmoc-Ala-OH, Fmoc-Glu(tBu)-OH, Fmoc-Glu(tBu)-OH, Fmoc-Leu-OH, Fmoc-Leu-OH, Fmoc-Ala-OH, Fmoc-Thr(tBu)-OH, Fmoc-Ile-OH, Fmoc-Asn(Trt)-OH, Fmoc-Glu(tBu)-OH, Fmoc-Glu(tBu)-OH, Fmoc-Leu-OH, Fmoc-Phe-OH, Fmoc-Asp(tBu)-OH, Fmoc-Val-OH. They are dissolved in *N,N*-dimethylformamide (DMF) and, according to the sequence, activated using O-benzotriazol-1-yl-*N, N, N′, N′*-tetramethyl-uronium hexafluorophosphate (HBTU) and Diisopropylethylamine (DIEA). Removal of the Fmoc protecting group is achieved using a solution of 20% (V/V) piperidine in *N,N*-dimethylformamide (DMF) for 20 minutes (step 1).The selective deprotection of the Lys (Aloc) group is performed manually and accomplished by treating the resin with a solution of 3 eq of $Pd(PPh_3)_4$ dissolved in 5 mL of $CHCl_3$:NMM:HOAc (18:1:0.5) for 2 h (Step 2). The resin is then washed with $CHCl_3$ (6 x 5 mL), 20% HOAc in DCM (6 x 5 mL), DCM (6 x 5 mL), and DMF (6 x 5 mL). The synthesis is then re-automated for the addition of the 3-maleimidopropionic acid (Step 3). Between every coupling, the resin is washed 3 times with *N,N*-dimethylformamide (DMF) and 3 times with isopropanol. The

peptide is cleaved from the resin using 85% TFA/5% TIS/5% thioanisole and 5% phenol, followed by precipitation by dry-ice cold Et_2O (Step 4). The product is purified by preparative reversed phased HPLC using a Varian (Rainin) preparative binary HPLC system: gradient elution of 30-55% B (0.045% TFA in H_2O (A) and 0.045% TFA in CH_3CN (B)) over 180 min at 9.5 mL/min using a Phenomenex Luna 10 μ phenyl-hexyl, 21 mm x 25 cm column and UV detector (Varian Dynamax UVD II) at λ 214 and 254 nm to afford the desired modified peptide (i.e., DAC) in >95% purity, as determined by RP-HPLC.



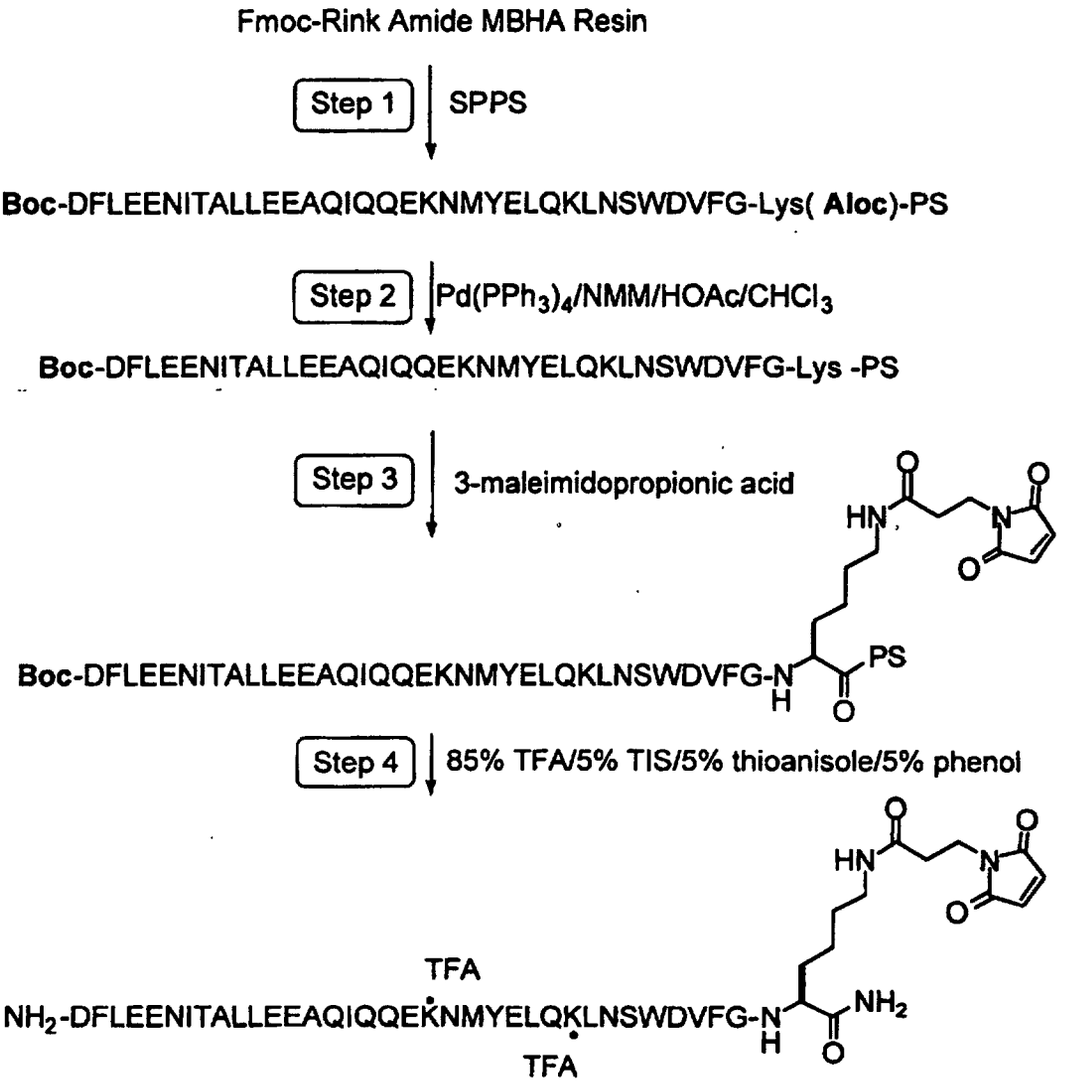
Example 29

Preparation of a Modified anti-SIV peptide

[0155] In this example, the peptide SEQ ID NO:72 is synthesized and modified to include a linker and maleimide group according to the synthesis scheme set forth below. As reported in U.S. Pat. Nos. 6,013,236 and 6,020,459, SEQ ID NO:72 exhibits potent antiviral activity as a crude peptide against simian immunodeficiency virus (SIV).

[0156] Solid phase peptide synthesis of the modified peptide on a 100 μmole scale is performed using manual solid-phase synthesis, a Symphony Peptide Synthesizer and Fmoc protected Rink Amide MBHA. The following protected amino acids are sequentially added to resin: Fmoc-Lys(Aloc)-OH, Fmoc-Gly-OH, Fmoc-Phe-OH, Fmoc-Val-OH, Fmoc-Asp(tBu)-OH, Fmoc-Trp(Boc)-OH, Fmoc-Ser(tBu)-OH, Fmoc-Asn(Trt)-OH, Fmoc-Leu-OH, Fmoc-Lys(Boc)-OH, Fmoc-Gln(Trt)-OH, Fmoc-Leu-OH, Fmoc-Glu(tBu)-OH, Fmoc-Tyr(tBu)-OH, Fmoc-Met-OH, Fmoc-Asn(Trt)-OH, Fmoc-Lys

(Boc)-OH, Fmoc-Glu(tBu)-OH, Fmoc-Gln(Trt)-OH, Fmoc-Gln(Trt)-OH, Fmoc-Ile-OH, Fmoc-Gln(Trt)-OH, Fmoc-Ala-OH, Fmoc-Glu(tBu)-OH, Fmoc-Glu(tBu)-OH, Fmoc-Leu-OH, Fmoc-Leu-OH, Fmoc-Ala-OH, Fmoc-Thr(tBu)-OH, Fmoc-Ile-OH, Fmoc-Asn(Trt)-OH, Fmoc-Glu(tBu)-OH, Fmoc-Glu(tBu)-OH, Fmoc-Leu-OH, Fmoc-Phe-OH, Fmoc-Asp (tBu)-OH. They are dissolved in *N,N*-dimethylformamide (DMF) and, according to the sequence, activated using O-benzotriazol-1-yl-*N, N, N', N'*-tetramethyl-uronium hexafluorophosphate (HBTU) and Diisopropylethylamine (DIEA). Removal of the Fmoc protecting group is achieved using a solution of 20% (V/V) piperidine in *N,N*-dimethylformamide (DMF) for 20 minutes (step 1).The selective deprotection of the Lys (Aloc) group is performed manually and accomplished by treating the resin with a solution of 3 eq of $Pd(PPh_3)_4$ dissolved in 5 mL of $CHCl_3$:NMM:HOAc (18:1:0.5) for 2 h (Step 2). The resin is then washed with $CHCl_3$ (6 x 5 mL), 20% HOAc in DCM (6 x 5 mL), DCM (6 x 5 mL), and DMF (6 x 5 mL). The synthesis is then re-automated for the addition of the 3-maleimidopropionic acid (Step 3). Between every coupling, the resin is washed 3 times with *N,N*-dimethylformamide (DMF) and 3 times with isopropanol. The peptide is cleaved from the resin using 85% TFA/5% TIS/5% thioanisole and 5% phenol, followed by precipitation by dry-ice cold Et_2O (Step 4). The product is purified by preparative reversed phased HPLC using a Varian (Rainin) preparative binary HPLC system: gradient elution of 30-55% B (0.045% TFA in H_2O (A) and 0.045% TFA in CH_3CN (B)) over 180 min at 9.5 mL/min using a Phenomenex Luna 10 μ phenyl-hexyl, 21 mm x 25 cm column and UV detector (Varian Dynamax UVD II) at λ 214 and 254 nm to afford the desired modified peptide (i.e., DAC) in >95% purity, as determined by RP-HPLC.

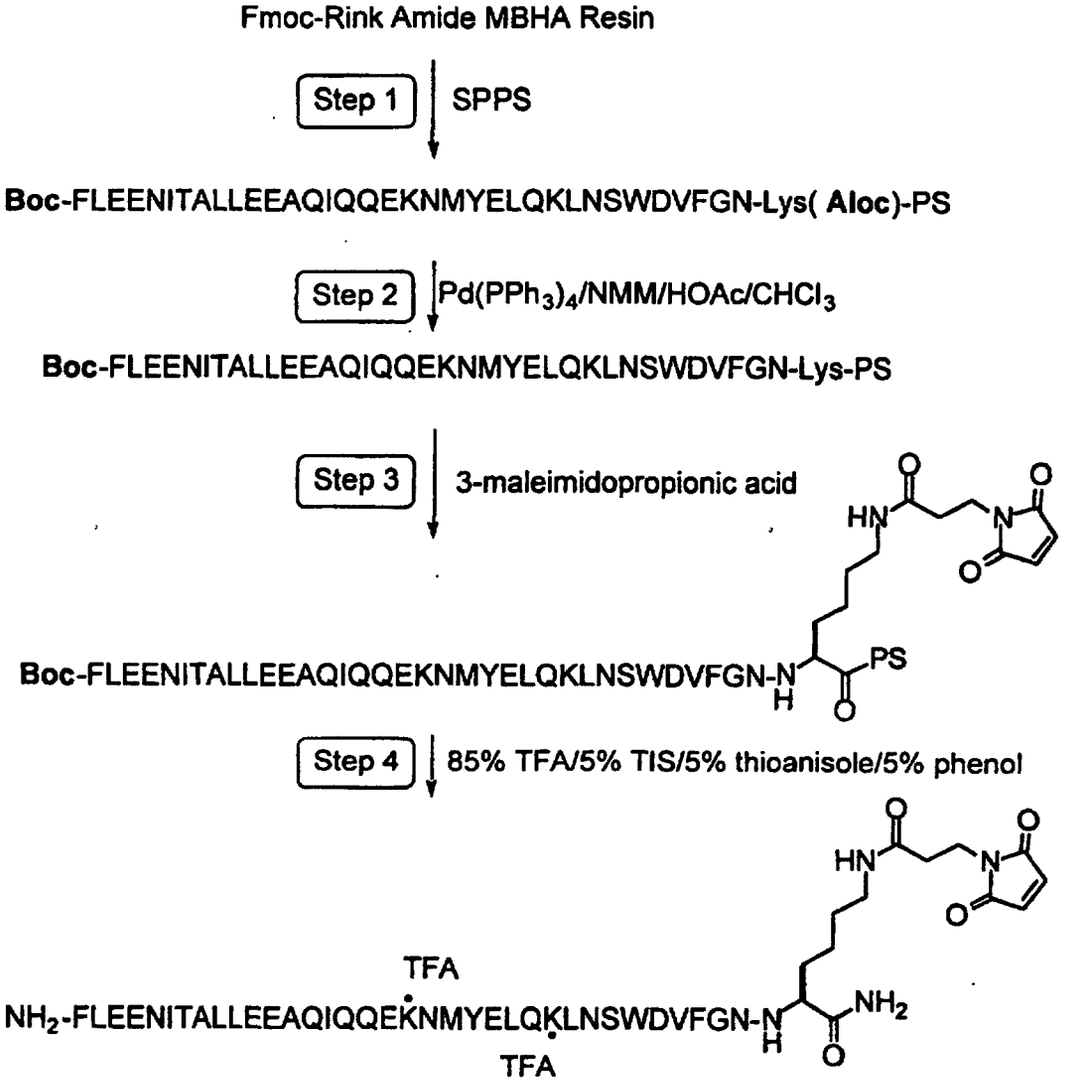


Example 30

Preparation of a Modified anti-SIV peptide

[0157] In this example, the peptide SEQ ID NO:73 is synthesized and modified to include a linker and maleimide group according to the synthesis scheme set forth below. As reported in U.S. Pat. Nos. 6,013,236 and 6,020,459, SEQ ID NO:73 exhibits potent antiviral activity as a crude peptide against simian immunodeficiency virus (SIV).

[0158] Solid phase peptide synthesis of the modified peptide on a 100 μmole scale is performed using manual solid-phase synthesis, a Symphony Peptide Synthesizer and Fmoc protected Rink Amide MBHA. The following protected amino acids are sequentially added to resin: Fmoc-Lys(Aloc)-OH, Fmoc-Asn(Trt)-OH, Fmoc-Gly-OH, Fmoc-Phe-OH, Fmoc-Val-OH, Fmoc-Asp(tBu)-OH, Fmoc-Trp(Boc)-OH, Fmoc-Ser(tBu)-OH, Fmoc-Asn(Trt)-OH, Fmoc-Leu-OH, Fmoc-Lys(Boc)-OH, Fmoc-Gln(Trt)-OH, Fmoc-Leu-OH, Fmoc-Glu(tBu)-OH, Fmoc-Tyr(tBu)-OH, Fmoc-Met-OH, Fmoc-Asn(Trt)-OH, Fmoc-Lys(Boc)-OH, Fmoc-Glu(tBu)-OH, Fmoc-Gln(Trt)-OH, Fmoc-Gln(Trt)-OH, Fmoc-Ile-OH, Fmoc-Gln(Trt)-OH, Fmoc-Ala-OH, Fmoc-Glu(tBu)-OH, Fmoc-Glu(tBu)-OH, Fmoc-Leu-OH, Fmoc-Leu-OH, Fmoc-Ala-OH, Fmoc-Thr(tBu)-OH, Fmoc-Ile-OH, Fmoc-Asn(Trt)-OH, Fmoc-Glu(tBu)-OH, Fmoc-Glu(tBu)-OH, Fmoc-Leu-OH, Fmoc-Phe-OH. They are dissolved in *N,N*-dimethylformamide (DMF) and, according to the sequence, activated using O-benzotriazol-1-yl-*N, N, N′, N′*-tetramethyl-uronium hexafluorophosphate (HBTU) and Diisopropylethylamine (DIEA). Removal of the Fmoc protecting group is achieved using a solution of 20% (V/V) piperidine in *N,N*-dimethylformamide (DMF) for 20 minutes (step 1).The selective deprotection of the Lys (Aloc) group is performed manually and accomplished by treating the resin with a solution of 3 eq of $Pd(PPh_3)_4$ dissolved in 5 mL of $CHCl_3$:NMM: HOAc (18:1:0.5) for 2 h (Step 2). The resin is then washed with $CHCl_3$ (6 x 5 mL), 20% HOAc in DCM (6 x 5 mL), DCM (6 x 5 mL), and DMF (6 x 5 mL). The synthesis is then re-automated for the addition of the 3-maleimidopropionic acid (Step 3). Between every coupling, the resin is washed 3 times with *N,N*-dimethylformamide (DMF) and 3 times with isopropanol. The peptide is cleaved from the resin using 85% TFA/5% TIS/5% thioanisole and 5% phenol, followed by precipitation by dry-ice cold Et_2O (Step 4). The product is purified by preparative reversed phased HPLC using a Varian (Rainin) preparative binary HPLC system: gradient elution of 30-55% B (0.045% TFA in H_2O (A) and 0.045% TFA in CH_3CN (B)) over 180 min at 9.5 mL/min using a Phenomenex Luna 10 μ phenyl-hexyl, 21 mm x 25 cm column and UV detector (Varian Dynamax UVD II) at λ 214 and 254 nm to afford the desired modified peptide (i.e., DAC) in >95% purity, as determined by RP-HPLC.



[0159] While certain embodiments of the invention have been described and exemplified, those having ordinary skill in the art will understand that the invention is not intended to be limited to the specifics of any of these embodiments, but is rather defined by the accompanying claims.

TABLE 2

DP178 CARBOXY TRUNCATIONS
YTS
YTSL
YTSLI
YTSLIH
YTSLIHS
YTSLIHSL
YTSLIHSLI
YTSLIHSLIE
YTSLIHSLIEE
YTSLIHSLIEES
YTSLIHSLIEESQ
YTSLIHSLIEESQN
YTSLIHSLIEESQNQ
YTSLJHSLJEESQNQQ
YTSLIHSLIEESQNQQE
YTSLIHSLIEESQNQQEK
YTSLIHSLIEESQNQQEKN
YTSLIHSLIEESQNQQEKNE
YTSLIHSLIEESQNQQEKNEQ
YTSLIHSLIEESQNQQEKNEQE
YTSLIHSLIEESQNQQEKNEQEL
YTSLIHSLIEESQNQQEKNEQELL
YTSLIHSLIEESQNQQEKNEQELLE
YTSLIHSLIEESQNQQEKNEQELLEL
YTSLIHSLIEESQNQQEKNEQELLELD
YTSLIHSLIEESQNQQEKNEQELLELDK

YTSLIHSLIEESQNQQEKNEQELLELDKW

YTSLIHSLIEESQNQQEKNEQELLELDKWA

YTSLIHSLIEESQNQQEKNEQELLELDKWAS

YTSLIHSLIEESQNQQEKNEQELLELDKWASL

YTSLIHSLIEESQNQQEKNEQELLELDKWASLW

YTSLIHSLIEESQNQQEKNEQELLELDKWASLWN

YTSLIHSLIEESQNQQEKNEQELLELDKWASLWNW

YTSLIHSLIEESQNQQEKNEQELLELDKWASLWNWF

The one letter amino acid code of Table 1 is used.

TABLE 3

DP178 AMINO TRUNCATIONS
NWF
WNWF
LWNWF
SLWNWF
ASLWNWF
WASLWNWF
KWASLWNWF
DKWASLWNWF
LDKWASLWNWF
ELDKWASLWNWF
LELDKWASLWNWF
LLELDKWASLWNWF
ELLELDKWASLWNWF
QELLELDKWASLWNWF
EQELLELDKWASLWNWF
NEQELLELDKWASLWNWF
KNEQELLELDKWASLWNWF
EKNEQELLELDKWASLWNWF
QEKNEQELLELDKWASLWNWF
QQEKNEQELLELDKWASLWNWF
NQQEKNEQELLELDKWASLWNWF
QNQQEKNEQELLELDKWASLWNWF
SQNQQEKNEQELLELDKWASLWNWF
ESQNQQEKNEQELLELDKWASLWNWF
EESQNQQEKNEQELLELDKWASLWNWF

IEESQNQQEKNEQELLELDKWASLWNWF

LIEESQNQQEKNEQELLELDKWASLWNWF

SLIEESQNQQEKNEQELLELDKWASLWNWF

HSLIEESQNQQEKNEQELLELDKWASLWNWF

IHSLIEESQNQQEKNEQELLELDKWASLWNWF

LIHSLIEESQNQQEKNEQELLELDKWASLWNWF

SLIHSLIEESQNQQEKNEQELLELDKWASLWNWF

TSLIHSLIEESQNQQEKNEQELLELDKWASLWNWF

YTSLIHSLIEESQNQQEKNEQELLELDKWASLWNWF

The one letter amino acid code of Table 1 is used.

TABLE 4

DP107 CARBOXY TRUNCATIONS
NNL
NNLL
NNLLR
NNLLRA
NNLLRAI
NNLLRAIE
NNLLRAIEA
NNLLRAIEAQ
NNLLRAIEAQQ
NNLLRAIEAQQH
NNLLRAIEAQQHL
NNLLRAIEAQQHLL
NNLLRAIEAQQHLLQ
NNLLRAIEAQQHLLQL
NNLLRAIEAQQHLLQLT
NNLLRAIEAQQHLLQLTV
NNLLRAIEAQQHLLQLTVW
NNLLRAIEAQQHLLQLTVWQ
NNLLRAIEAQQHLLQLTVWQI
NNLLRAIEAQQHLLQLTVWQIK
NNLLRAIEAQQHLLQLTVWQIKQ
NNLLRAIEAQQHLLQLTVWQIKQL
NNLLRAIEAQQHLLQLTVWQIKQLQ
NNLLRAIEAQQHLLQLTVWQIKQLQA
NNLLRAIEAQQHLLQLTVWQIKQLQAR
NNLLRAIEAQQHLLQLTVWQIKQLQARI

NNLLRAIEAQQHLLQLTVWQIKQLQARIL

NNLLRAIEAQQHLLQLTVWQIKQLQARILA

NNLLRAIEAQQHLLQLTVWQIKQLQARILAV

NNLLRAIEAQQHLLQLTVWQIKQLQARILAVE

NNLLRAIEAQQHLLQLTVWQIKQLQARILAVER

NNLLRAIEAQQHLLQLTVWQIKQLQARILAVERY

NNLLRAIEAQQHLLQLTVWQIKQLQARILAVERYL

NNLLRAIEAQQHLLQLTVWQIKQLQARILAVERYLK

NNLLRAIEAQQHLLQLTVWQIKQLQARILAVERYLKD

NNLLRAIEAQQHLLQLTVWQIKQLQARILAVERYLKDQ

The one letter amino acid code of Table 1 is used.

TABLE 5

DP107 AMINO TRUNCATIONS
KDQ
LKDQ
YLKDQ
RYLKDQ
ERYLKDQ
VERYLKDQ
AVERYLKDQ
LAVERYLKDQ
ILAVERYLKDQ
RILAVERYLKDQ
ARILAVERYLKDQ
QARILAVERYLKDQ
LQARILAVERYLKDQ
QLQARILAVERYLKDQ
KQLQARILAVERYLKDQ
IKQLQARILAVERYLKDQ
QIKQLQARILAVERYLKDQ
WQIKQLQARILAVERYLKDQ
VWQIKQLQARILAVERYLKDQ
TVWQIKQLQARILAVERYLKDQ
LTVWQIKQLQARILAVERYLKDQ
QLTVWQIKQLQARILAVERYLKDQ
LQLTVWQIKQLQARILAVERYLKDQ
LLQLTVWQIKQLQARILAVERYLKDQ
HLLQLTVWQIKQLQARILAVERYLKDQ
QHLLQLTVWQIKQLQARILAVERYLKDQ

QQHLLQLTVWQIKQLQARILAVERYLKDQ

AQQHLLQLTVWQIKQLQARILAVERYLKDQ

EAQQHLLQLTVWQIKQLQARILAVERYLKDQ

IEAQQHLLQLTVWQIKQLQARILAVERYLKDQ

AIEAQQHLLQLTVWQIKQLQARILAVERYLKDQ

RAIEAQQHLLQLTVWQIKQLQARILAVERYLKDQ

LRAIEAQQHLLQLTVWQIKQLQARILAVERYLKDQ

LLRAIEAQQHLLQLTVWQIKQLQARILAVERYLKDQ

NLLRAIEAQQHLLQLTVWQIKQLQARILAVERYLKDQ

NNLLRAIEAQQHLLQLTVWQIKQLQARILAVERYLKDQ

The one letter amino acid code of Table 1 is used.

TABLE 6

HIV-2_{NIHZ} DP178 analog carboxy truncations.

LEA
LEAN
LEANI
LEANIS
LEANISQ
LEANISQS
LEANISQSL
LEANISQSLE
LEANISQSLEQ
LEANISQSLEQA
LEANISQSLEQAQ
LEANISQSLEQAQI
LEANISQSLEQAQIQ
LEANISQSLEQAQIQQ
LEANISQSLEQAQIQQE
LEANISQSLEQAQIQQEK
LEANISQSLEQAQIQQEKN
LEANISQSLEQAQIQQEKNM
LEANISQSLEQAQIQQEKNMY
LEANISQSLEQAQIQQEKNMYE
LEANISQSLEQAQIQQEKNMYEL
LEANISQSLEQAQIQQEKNMYELQ
LEANISQSLEQAQIQQEKNMYELQK
LEANISQSLEQAQIQQEKNMYELQKL
LEANISQSLEQAQIQQEKNMYELQKLN
LEANISQSLEQAQIQQEKNMYELQKLNS

LEANISQSLEQAQIQQEKNMYELQKLNSW

LEANISQSLEQAQIQQEKNMYELQKLNSWD

LEANISQSLEQAQIQQEKNMYELQKLNSWDV

LEANISQSLEQAQIQQEKNMYELQKLNSWDVF

LEANISQSLEQAQIQQEKNMYELQKLNSWDVFT

LEANISQSLEQAQIQQEKNMYELQKLNSWDVFTN

LEANISQSLEQAQIQQEKNMYELQKLNSWDVFTNW

LEANISQSLEQAQIQQEKNMYELQKLNSWDVFTNWL

The one letter amino acid code of Table 1 is used.

TABLE 7

HIV-2_{NIH2} DP178 analog amino truncations.

NWL
TNWL
FTNWL
VFTNWL
DVFTNWL
WDVFTNWL
SWDVFTNWL
NSWDVFTNWL
LNSWDVFTNWL
KLNSWDVFTNWL
QKLNSWDVFTNWL
LQKLNSWDVFTNWL
ELQKLNSWDVFTNWL
YELQKLNSWDVFTNWL
MYELQKLNSWDVFTNWL
NMYELQKLNSWDVFTNWL
KNMYELQKLNSWDVFTNWL
EKNMYELQKLNSWDVFTNWL
QEKNMYELQKLNSWDVFTNWL
QQEKNMYELQKLNSWDVFTNWL
IQQEKNMYELQKLNSWDVFTNWL
QIQQEKNMYELQKLNSWDVFTNWL
AQIQQEKNMYELQKLNSWDVFTNWL
QAQIQQEKNMYELQKLNSWDVFTNWL
EQAQIQQEKNMYELQKLNSWDVFTNWL
LEQAQIQQEKNMYELQKLNSWDVFTNWL

SLEQAQIQQEKNMYELQKLNSWDVFTNWL

QSLEQAQIQQEKNMYELQKLNSWDVFTNWL

SQSLEQAQIQQEKNMYELQKLNSWDVFTNWL

ISQSLEQAQIQQEKNMYELQKLNSWDVFTNWL

NISQSLEQAQIQQEKNMYELQKLNSWDVFTNWL

ANISQSLEQAQIQQEKNMYELQKLNSWDVFTNWL

EANISQSLEQAQIQQEKNMYELQKLNSWDVFTNWL

LEANISQSLEQAQIQQEKNMYELQKLNSWDVFTNWL

The one letter amino acid code of Table 1 is used.

TABLE 8

RESPIRATORY SYNCYTIAL VIRUS (RSV) DP107 F2 REGION ANALOG CARBOXY TRUNCATIONS
YTS
YTSV
YTSVI
YTSVIT
YTSVITI
YTSVITIE
YTSVITIEL
YTSVITIELS
YTSVITIELSN
YTSVITIELSNI
YTSVITIELSNIK
YTSVITIELSNIKE
YTSVITIELSNIKEN
YTSVITIELSNIKENK
YTSVITIELSNIKENKC
YTSVITIELSNIKENKCN
YTSVITIELSNIKENKCNG
YTSVITIELSNIKENKCNGT
YTSVITIELSNIKENKCNGTD
YTSVITIELSNIKENKCNGTDA
YTSVITIELSNIKENKCNGTDAK
YTSVITIELSNIKENKCNGTDAKV
YTSVITIELSNIKENKCNGTDAKVK
YTSVITIELSNIKENKCNGTDAKVKL
YTSVITIELSNIKENKCNGTDAKVKLI

YTSVITIELSNIKENKCNGTDAKVKLIK

YTSVITIELSNIKENKCNGTDAKVKLIKQ

YTSVITIELSNIKENKCNGTDAKVKLIKQE

YTSVITIELSNIKENKCNGTDAKVKLIKQEL

YTSVITIELSNIKENKCNGTDAKVKLIKQELD

YTSVITIELSNIKENKCNGTDAKVKLIKQELDK

YTSVITIELSNIKENKCNGTDAKVKLIKQELDKY

YTSVITIELSNIKENKCNGTDAKVKLIKQELDKYK

YTSVITIELSNIKENKCNGTDAKVKLIKQELDKYKN

YTSVITIELSNIKENKCNGTDAKVKLIKQELDKYKNA

YTSVITIELSNIKENKCNGTDAKVKLIKQELDKYKNAV

YTSVITIELSNIKENKCNGTDAKVKLIKQELDKYKNAVT

YTSVITIELSNIKENKCNGTDAKVKLIKQELDKYKNAVTE

YTSVITIELSNIKENKCNGTDAKVKLIKQELDKYKNAVTEL

YTSVITIELSNIKENKCNGTDAKVKLIKQELDKYKNAVTELQ

YTSVITIELSNIKENKCNGTDAKVKLIKQELDKYKNAVTELQL

YTSVITIELSNIKENKCNGTDAKVKLIKQELDKYKNAVTELQLL

YTSVITIELSNIKENKCNGTDAKVKLIKQELDKYKNAVTELQLLM

YTSVITIELSNIKENKCNGTDAKVKLIKQELDKYKNAVTELQLLMQ

YTSVITIELSNIKENKCNGTDAKVKLIKQELDKYKNAVTELQLLMQS

YTSVITIELSNIKENKCNGTDAKVKLIKQELDKYKNAVTELQLLMQST

The one letter amino acid code of Table 1 is used.

TABLE 9

RESPIRATORY SYNCYTIAL VIRUS (RSV) DP107 F2 REGION ANALOG AMINO TRUNCATIONS
QST
MQST
LMQST
LLMQST
QLLMQST
LQLLMQST
ELQLLMQST
TELQLLMQST
VTELQLLMQST
AVTELQLLMQST
NAVTELQLLMQST
KNAVTELQLLMQST
YKNAVTELQLLMQST
KYKNAVTELQLLMQST
DKYKNAVTELQLLMQST
LDKYKNAVTELQLLMQST
ELDKYKNAVTELQLLMQST
QELDKYKNAVTELQLLMQST
KQELDKYKNAVTELQLLMQST
IKQELDKYKNAVTELQLLMQST
LIKQELDKYKNAVTELQLLMQST
KLIKQELDKYKNAVTELQLLMQST
VKLIKQELDKYKNAVTELQLLMQST
KVKLIKQELDKYKNAVTELQLLMQST
AKVKLIKQELDKYKNAVTELQLLMQST

DAKVKLIKQELDKYKNAVTELQLLMQST

TDAKVKLIKQELDKYKNAVTELQLLMQST

GTDAKVKLIKQELDKYKNAVTELQLLMQST

NGTDAKVKLIKQELDKYKNAVTELQLLMQST

CNGTDAKVKLIKQELDKYKNAVTELQLLMQST

KCNGTDAKVKLIKQELDKYKNAVTELQLLMQST

NKCNGTDAKVKLIKQELDKYKNAVTELQLLMQST

KENKCNGTDAKVKLIKQELDKYKNAVTELQLLMQST

IKENKCNGTDAKVKLIKQELDKYKNAVTELQLLMQST

NIKENKCNGTDAKVKLIKQELDKYKNAVTELQLLMQST

SNIKENKCNGTDAKVKLIKQELDKYKNAVTELQLLMQST

LSNIKENKCNGTDAKVKLIKQELDKYKNAVTELQLLMQST

ELSNIKENKCNGTDAKVKLIKQELDKYKNAVTELQLLMQST

IELSNIKENKCNGTDAKVKLIKQELDKYKNAVTELQLLMQST

TIELSNIKENKCNGTDAKVKLIKQELDKYKNAVTELQLLMQST

ITIELSNIKENKCNGTDAKVKLIKQELDKYKNAVTELQLLMQST

VITIELSNIKENKCNGTDAKVKLIKQELDKYKNAVTELQLLMQST

SVITIELSNIKENKCNGTDAKVKLIKQELDKYKNAVTELQLLMQST

TSVITIELSNIKENKCNGTDAKVKLIKQELDKYKNAVTELQLLMQST

The one letter amino acid code of Table 1 is used.

TABLE 10

RESPIRATORY SYNCYTIAL VIRUS (RSV) F1 DP178 REGION ANALOG CARBOXY TRUNCATIONS
FYD
FYDP
FYDPL
FYDPLV
FYDPLVF
FYDPLVFP
FYDPLVFPS
FYDPLVFPSD
FYDPLVFPSDE
FYDPLVFPSDEF
FYDPLVFPSDEFD
FYDPLVFPSDEFDA
FYDPLVFPSDEFDAS
FYDPLVFPSDEFDASI
FYDPLVFPSDEFDASIS
FYDPLVFPSDEFDASISQ
FYDPLVFPSDEFDASISQV
FYDPLVFPSDEFDASISQVN
FYDPLVFPSDEFDASISQVNE
FYDPLVFPSDEFDASISQVNEK
FYDPLVFPSDEFDASISQVNEKI
FYDPLVFPSDEFDASISQVNEKIN
FYDPLVFPSDEFDASISQVNEKINQ
FYDPLVFPSDEFDASISQVNEKINQS
FYDPLVFPSDEFDASISQVNEKINQSL

FYDPLVFPSDEFDASISQVNEKINQSLA

FYDPLVFPSDEFDASISQVNEKINQSLAF

FYDPLVFPSDEFDASISQVNEKINQSLAFI

FYDPLVFPSDEFDASISQVNEKINQSLAFIR

FYDPLVFPSDEFDASISQVNEKINQSLAFIRK

FYDPLVFPSDEFDASISQVNEKINQSLAFIRKS

FYDPLVFPSDEFDASISQVNEKINQSLAFIRKSD

FYDPLVFPSDEFDASISQVNEKINQSLAFIRKSDE

FYDPLVFPSDEFDASISQVNEKINQSLAFIRKSDEL

FYDPLVFPSDEFDASISQVNEKINQSLAFIRKSDELL

The one letter amino acid code of Table 1 is used.

TABLE 11

RESPIRATORY SYNCYTIAL VIRUS (RSV) F1 DP178 REGION ANALOG AMINO TRUNCATIONS
DELL
SDELL
KSDELL
RKSDELL
IRKSDELL
FIRKSDELL
AFIRKSDELL
LAFIRKSDELL
SLAFIRKSDELL
QSLAFIRKSDELL
NQSLAFIRKSDELL
INQSLAFIRKSDELL
KINQSLAFIRKSDELL
EKINQSLAFIRKSDELL
NEKINQSLAFIRKSDELL
VNEKINQSLAFIRKSDELL
QVNEKINQSLAFIRKSDELL
SQVNEKINQSLAFIRKSDELL
ISQVNEKINQSLAFIRKSDELL
SISQVNEKINQSLAFIRKSDELL
ASISQVNEKINQSLAFIRKSDELL
DASISQVNEKINQSLAFIRKSDELL
FDASISQVNEKINQSLAFIRKSDELL
EFDASISQVNEKINQSLAFIRKSDELL
DEFDASISQVNEKINQSLAFIRKSDELL

SDEFDASISQVNEKINQSLAFIRKSDELL

PSDEFDASISQVNEKINQSLAFIRKSDELL

FPSDEFDASISQVNEKINQSLAFIRKSDELL

VFPSDEFDASISQVNEKINQSLAFIRKSDELL

LVFPSDEFDASISQVNEKINQSLAFIRKSDELL

PLVFPSDEFDASISQVNEKINQSLAFIRKSDELL

DPLVFPSDEFDASISQVNEKINQSLAFIRKSDELL

YDPLVFPSDEFDASISQVNEKINQSLAFIRKSDELL

The one letter amino acid code of Table 1 is used.

TABLE 12

HUMAN PARAINFLUENZA VIRUS 3 (HPV3) F1 REGION DP178 ANALOG CARBOXY TRUNCATIONS

ITL
ITLN
ITLNN
ITLNNS
ITLNNSV
ITLNNSVA
ITLNNSVAL
ITLNNSVALD
ITLNNSVALDP
ITLNNSVALDPI
ITLNNSVALDPID
ITLNNSVALDPIDI
ITLNNSVALDPIDIS
ITLNNSVALDPIDISI
ITLNNSVALDPIDISIE
ITLNNSVALDPIDISIEL
ITLNNSVALDPIDISIELN
ITLNNSVALDPIDISIELNK
ITLNNSVALDPIDISIELNKA
ITLNNSVALDPIDISIELNKAK
ITLNNSVALDPIDISIELNKAKS
ITLNNSVALDPIDISIELNKAKSD
ITLNNSVALDPIDISIELNKAKSDL
ITLNNSVALDPIDISIELNKAKSDLE
ITLNNSVALDPIDISIELNKAKSDLEE

ITLNNSVALDPIDISIELNKAKSDLEES

ITLNNSVALDPIDISIELNKAKSDLEESK

ITLNNSVALDPIDISIELNKAKSDLEESKE

ITLNNSVALDPIDISIELNKAKSDLEESKEW

ITLNNSVALDPIDISIELNKAKSDLEESKEWI

ITLNNSVALDPIDISIELNKAKSDLEESKEWIR

ITLNNSVALDPIDISIELNKAKSDLEESKEWIRR

ITLNNSVALDPIDISIELNKAKSDLEESKEWIRRS

The one letter amino acid code of Table 1 is used.

TABLE 13

HUMAN PARAINFLUENZA VIRUS 3 (HPV3) F1 REGION DP178 ANALOG AMINO TRUNCATIONS

RRS
IRRS
WIRRS
EWIRRS
KEWIRRS
SKEWIRRS
ESKEWIRRS
EESKEWIRRS
LEESKEWIRRS
DLEESKEWIRRS
SDLEESKEWIRRS
KSDLEESKEWIRRS
AKSDLEESKEWIRRS
KAKSDLEESKEWIRRS
NKAKSDLEESKEWIRRS
LNKAKSDLEESKEWIRRS
ELNKAKSDLEESKEWIRRS
IELNKAKSDLEESKEWIRRS
SIELNKAKSDLEESKEWIRRS
ISIELNKAKSDLEESKEWIRRS
DISIELNKAKSDLEESKEWIRRS
IDISIELNKAKSDLEESKEWIRRS
PIDISIELNKAKSDLEESKEWIRRS
DPIDISIELNKAKSDLEESKEWIRRS
LDPIDISIELNKAKSDLEESKEWIRRS

ALDPIDISIELNKAKSDLEESKEWIRRS

VALDPIDISIELNKAKSDLEESKEWIRRS

SVALDPIDISIELNKAKSDLEESKEWIRRS

NSVALDPIDISIELNKAKSDLEESKEWIRRS

NNSVALDPIDISIELNKAKSDLEESKEWIRRS

LNNSVALDPIDISIELNKAKSDLEESKEWIRRS

TLNNSVALDPIDISIELNKAKSDLEESKEWIRRS

The one letter amino acid code of Table 1 is used.

TABLE 14

HUMAN PARAINFLUENZA VIRUS 3 (HPV3) F1 REGION
DP107 ANALOG CARBOXY TRUNCATIONS

ALG
ALGV
ALGVA
ALGVAT
ALGVATS
ALGVATSA
ALGVATSAQ
ALGVATSAQI
ALGVATSAQIT
ALGVATSAQITA
ALGVATSAQITAA
ALGVATSAQITAAV
ALGVATSAQITAAVA
ALGVATSAQITAAVAL
ALGVATSAQITAAVALV
ALGVATSAQITAAVALVE
ALGVATSAQITAAVALVEA
ALGVATSAQITAAVALVEAK
ALGVATSAQITAAVALVEAKQ
ALGVATSAQITAAVALVEAKQA
ALGVATSAQITAAVALVEAKQAR
ALGVATSAQITAAVALVEAKQARS
ALGVATSAQITAAVALVEAKQARSD
ALGVATSAQITAAVALVEAKQARSDI
ALGVATSAQITAAVALVEAKQARSDIE

ALGVATSAQITAAVALVEAKQARSDIEK

ALGVATSAQITAAVALVEAKQARSDIEKL

ALGVATSAQITAAVALVEAKQARSDIEKLK

ALGVATSAQITAAVALVEAKQARSDIEKLKE

ALGVATSAQITAAVALVEAKQARSDIEKLKEA

ALGVATSAQITAAVALVEAKQARSDIEKLKEAI

ALGVATSAQITAAVALVEAKQARSDIEKLKEAIR

The one letter amino acid code of Table 1 is used.

TABLE 15

HUMAN PARAINFLUENZA VIRUS 3 (HPV3) F1 REGION
DP107 ANALOG AMINO TRUNCATIONS

IRD
AIRD
EAIRD
KEAIRD
LKEAIRD
KLKEAIRD
EKLKEAIRD
IEKLKEAIRD
DIEKLKEAIRD
SDIEKLKEAIRD
RSDIEKLKEAIRD
ARSDIEKLKEAIRD
QARSDIEKLKEAIRD
KQARSDIEKLKEAIRD
AKQARSDIEKLKEAIRD
EAKQARSDIEKLKEAIRD
VEAKQARSDIEKLKEAIRD
LVEAKQARSDIEKLKEAIRD
ALVEAKQARSDIEKLKEAIRD
VALVEAKQARSDIEKLKEAIRD
AVALVEAKQARSDIEKLKEAIRD
AAVALVEAKQARSDIEKLKEAIRD
TAAVALVEAKQARSDIEKLKEAIRD
ITAAVALVEAKQARSDIEKLKEAIRD
QITAAVALVEAKQARSDIEKLKEAIRD

AQITAAVALVEAKQARSDIEKLKEAIRD

SAQITAAVALVEAKQARSDIEKLKEAIRD

TSAQITAAVALVEAKQARSDIEKLKEAIRD

ATSAQITAAVALVEAKQARSDIEKLKEAIRD

VATSAQITAAVALVEAKQARSDIEKLKEAIRD

GVATSAQITAAVALVEAKQARSDIEKLKEAIRD

LGVATSAQITAAVALVEAKQARSDIEKLKEAIRD

The one letter amino acid code of Table 1 is used.

TABLE 16

ANTI-RESPIRATORY SYNCYTIAL VIRUS (RSV) PEPTIDES
TSVITIELSNIKENKCNGTDAKVKLIKQELDKYKN
SVITIELSNIKENKCNGTDAKVKLIKQELDKYKNA
VITIELSNIKENKCNGTDAKVKLIKQELDKYKNAV
VAVSKVLHLEGEVNKIALLSTNKAVVSLSNGVS
AVSKVLHLEGEVNKIALLSTNKAVVSLSNGVSV
VSKVLHLEGEVNKIALLSTNKAVVSLSNGVSVL
SKVLHLEGEVNKIALLSTNKAVVSLSNGVSVLT
KVLHLEGEVNKIALLSTNKAVVSLSNGVSVLTS
LEGEVNKIALLSTNKAVVSLSNGVSVLTSKVLD
GEVNKIALLSTNKAVVSLSNGVSVLTSKVLDLK
EVNKIALLSTNKAVVSLSNGVSVLTSKVLDLKN
VNKIALLSTNKAVVSLSNGVSVLTSKVLDLKNY
NKIALLSTNKAVVSLSNGVSVLTSKVLDLKNYI
KIALLSTNKAVVSLSNGVSVLTSKVLDLKNYID
IALLSTNKAVVSLSNGVSVLTSKVLDLKNYIDK
ALLSTNKAVVSLSNGVSVLTSKVLDLKNYIDKQ
VAVSKVLHLEGEVNKIALLSTNKAVVSLSNGVS
AVSKVLHLEGEVNKIALLSTNKAVVSLSNGVSV
VSKVLHLEGEVNKIALLSTNKAVVSLSNGVSVL
SKVLHLEGEVNKIALLSTNKAVVSLSNGVSVLT
KVLHLEGEVNKIALLSTNKAVVSLSNGVSVLTS
LEGEVNKIALLSTNKAVVSLSNGVSVLTSKVLD
GEVNKIALLSTNKAVVSLSNGVSVLTSKVLDLK
EVNKIALLSTNKAVVSLSNGVSVLTSKVLDLKN
VNKIALLSTNKAVVSLSNGVSVLTSKVLDLKNY
NKIALLSTNKAVVSLSNGVSVLTSKVLDLKNYI

KIALLSTNKAVVSLSNGVSVLTSKVLDLKNYID

IALLSTNKAVVSLSNGVSVLTSKVLDLKNYIDK

ALLSTNKAVVSLSNGVSVLTSKVLDLKNYIDKQ

The one letter amino acid code of Table 1 is used.

TABLE 17

ANTI-HUMAN PARAINFLUENZA VIRUS 3 (HPV3) PEPTIDES
TLNNSVALDPIDISIELNKAKSDLEESKEWIRRSN
LNNSVALDPIDISIELNKAKSDLEESKEWIRRSNQ
NNSVALDPIDISIELNKAKSDLEESKEWIRRSNQK
NSVALDPIDISIELNKAKSDLEESKEWIRRSNQKL
SVALDPIDISIELNKAKSDLEESKEWIRRSNQKLD
VALDPIDISIELNKAKSDLEESKEWIRRSNQKLDS
ALDPIDISIELNKAKSDLEESKEWIRRSNQKLDSI
LDPIDISIELNKAKSDLEESKEWIRRSNQKLDSIG
DPIDISIELNKAKSDLEESKEWIRRSNQKLDSIGN
PIDISIELNKAKSDLEESKEWIRRSNQKLDSIGNW
IDISIELNKAKSDLEESKEWIRRSNQKLDSIGNWH
DISIELNKAKSDLEESKEWIRRSNQKLDSIGNWHQ
ISIELNKAKSDLEESKEWIRRSNQKLDSIGNWHQS
SIELNKAKSDLEESKEWIRRSNQKLDSIGNWHQSS
IELNKAKSDLEESKEWIRRSNQKLDSIGNWHQSST
ELNKAKSDLEESKEWIRRSNQKLDSIGNWHQSSTT
TAAVALVEAKQARSDIEKLKEAIRDTNKAVQSVQS
AVALVEAKQARSDIEKLKEAIRDTNKAVQSVQSSI
LVEAKQARSDIEKLKEAIRDTNKAVQSVQSSIGNL
VEAKQARSDIEKLKEAIRDTNKAVQSVQSSIGNLI
EAKQARSDIEKLKEAIRDTNKAVQSVQSSIGNLIV
AKQARSDIEKLKEAIRDTNKAVQSVQSSIGNLIVA
KQARSDIEKLKEAIRDTNKAVQSVQSSIGNLIVAI
QARSDIEKLKEAIRDTNKAVQSVQSSIGNLIVAIK
ARSDIEKLKEAIRDTNKAVQSVQSSIGNLIVAIKS
RSDIEKLKEAIRDTNKAVQSVQSSIGNLIVAIKSV

SDIEKLKEAIRDTNKAVQSVQSSIGNLIVAIKSVQ

KLKEAIRDTNKAVQSVQSSIGNLIVAIKSVQDYVN

LKEAIRDTNKAVQSVQSSIGNLIVAIKSVQDYVNK

AIRDTNKAVQSVQSSIGNLIVAIKSVQDYVNKEIV

The one letter amino acid code of Table 1 is used.

TABLE 18

ANTI-SIMIAN IMMUNODEFICIENCY VIRUS (SIV) PEPTIDES

WQEWERKVDFLEENITALLEEAQIQQEKNMYELQK

QEWERKVDFLEENITALLEEAQIQQEKNMYELQKL

EWERKVDFLEENITALLEEAQIQQEKNMYELQKLN

WERKVDFLEENITALLEEAQIQQEKNMYELQKLNS

ERKVDFLEENITALLEEAQIQQEKNMYELQKLNSW

RKVDFLEENITALLEEAQIQQEKNMYELQKLNSWD

KVDFLEENITALLEEAQIQQEKNMYELQKLNSWDV

VDFLEENITALLEEAQIQQEKNMYELQKLNSWDVF

DFLEENITALLEEAQIQQEKNMYELQKLNSWDVFG

FLEENITALLEEAQIQQEKNMYELQKLNSWDVFGN

The one letter amino acid code of Table 1 is used.

TABLE 19

ANTI-MEASLES VIRUS (MEV) PEPTIDES
LHRIDLGPPISLERLDVGTNLGNAIAKLEAKELL
HRIDLGPPISLERLDVGTNLGNAIAKLEAKELLE
RIDLGPPISLERLDVGTNLGNAIAKLEAKELLES
IDLGPPISLERLDVGTNLGNAIAKLEAKELLESS
DLGPPISLERLDVGTNLGNAIAKLEAKELLESSD
LGPPISLERLDVGTNLGNAIAKLEAKELLESSDQ
GPPISLERLDVGTNLGNAIAKLEAKELLESSDQI
PPISLERLDVGTNLGNAIAKLEAKELLESSDQIL
PISLERLDVGTNLGNAIAKLEAKELLESSDQILR
SLERLDVGTNLGNAIAKLEAKELLESSDQILRSM
LERLDVGTNLGNAIAKLEAKELLESSDQILRSMK

The one letter amino acid code of Table 1 is used.

SEQUENCE LISTING

<110> ConjuChem, Inc.

<120> LONG LASTING FUSION PEPTIDE INHIBITORS OF VIRAL INFECTION

<130> REDC-1510

<140>
<141>

<150> US 60/134,406
<151> 1999-05-17

<150> US 60/153,406
<151> 1999-09-10

<160> 86

<170> PatentIn Ver. 2.1

<210> 1
<211> 36
<212> PRT
<213> Artificial Sequence

<220>
<223> Description of Artificial Sequence: synthetic peptide

<400> 1
Tyr Thr Ser Leu Ile His Ser Leu Ile Glu Glu Ser Gln Asn Gln Gln
1 5 10 15

Glu Lys Asn Glu Gln Glu Leu Leu Glu Leu Asp Lys Trp Ala Ser Leu
20 25 30

Trp Asn Trp Phe
35

<210> 2
<211> 38
<212> PRT
<213> Artificial Sequence

<220>

<223> Description of Artificial Sequence: synthetic peptide

<400> 2

Asn Asn Leu Leu Arg Ala Ile Glu Ala Gln Gln His Leu Leu Gln Leu
1 5 10 15

Thr Val Trp Gln Ile Lys Gln Leu Gln Ala Arg Ile Leu Ala Val Glu
20 25 30

Arg Tyr Leu Lys Asp Gln
35

<210> 3
<211> 36
<212> PRT
<213> Artificial Sequence

<220>
<223> Description of Artificial Sequence: synthetic peptide

<400> 3

Tyr Thr Asn Thr Ile Tyr Thr Leu Leu Glu Glu Ser Gln Asn Gln Gln
1 5 10 15

Glu Lys Asn Glu Gln Glu Leu Leu Glu Leu Asp Lys Trp Ala Ser Leu
20 25 30

Trp Asn Trp Phe
35

<210> 4
<211> 36
<212> PRT
<213> Artificial Sequence

<220>
<223> Description of Artificial Sequence: synthetic peptide

<400> 4

Tyr Thr Gly Ile Ile Tyr Asn Leu Leu Glu Glu Ser Gln Asn Gln Gln
1 5 10 15

Glu Lys Asn Glu Gln Glu Leu Leu Glu Leu Asp Lys Trp Ala Asn Leu

20 25 30

Trp Asn Trp Phe
35

<210> 5
<211> 36
<212> PRT
<213> Artificial Sequence

<220>
<223> Description of Artificial Sequence: synthetic peptide

<400> 5
Tyr Thr Ser Leu Ile Tyr Ser Leu Leu Glu Lys Ser Gln Thr Gln Gln
1 5 10 15

Glu Lys Asn Glu Gln Glu Leu Leu Glu Leu Asp Lys Trp Ala Ser Leu
20 25 30

Trp Asn Trp Phe
35

<210> 6
<211> 36
<212> PRT
<213> Artificial Sequence

<220>
<223> Description of Artificial Sequence: synthetic peptide

<400> 6
Leu Glu Ala Asn Ile Ser Lys Ser Leu Glu Gln Ala Gln Ile Gln Gln
1 5 10 15

Glu Lys Asn Met Tyr Glu Leu Gln Lys Leu Asn Ser Trp Asp Ile Phe
20 25 30

Gly Asn Trp Phe
35

<210> 7
<211> 36

<212> PRT
<213> Artificial Sequence

<220>
<223> Description of Artificial Sequence: synthetic peptide

<400> 7
Leu Glu Ala Asn Ile Ser Gln Ser Leu Glu Gln Ala Gln Ile Gln Gln
1 5 10 15

Glu Lys Asn Met Tyr Glu Leu Gln Lys Leu Asn Ser Trp Asp Val Phe
20 25 30

Thr Asn Trp Leu
35

<210> 8
<211> 41
<212> PRT
<213> Artificial Sequence

<220>
<223> Description of Artificial Sequence: synthetic peptide

<400> 8
Cys Gly Gly Asn Asn Leu Leu Arg Ala Ile Glu Ala Gln Gln His Leu
1 5 10 15

Leu Gln Leu Thr Val Trp Gly Ile Lys Gln Leu Gln Ala Arg Ile Leu
20 25 30

Ala Val Glu Arg Tyr Leu Lys Asp Gln
35 40

<210> 9
<211> 38
<212> PRT
<213> Artificial Sequence

<220>
<223> Description of Artificial Sequence: synthetic peptide

<400> 9

Gln Gln Leu Leu Asp Val Val Lys Arg Gln Gln Glu Met Leu Arg Leu
1 5 10 15

Thr Val Trp Gly Thr Lys Asn Leu Gln Ala Arg Val Thr Ala Ile Glu
20 25 30

Lys Tyr Leu Lys Asp Gln
35

<210> 10
<211> 46
<212> PRT
<213> Artificial Sequence

<220>
<223> Description of Artificial Sequence: synthetic peptide

<400> 10
Tyr Thr Ser Val Ile Thr Ile Glu Leu Ser Asn Ile Lys Glu Asn Lys
1 5 10 15

Cys Asn Gly Ala Lys Val Lys Leu Ile Lys Gln Glu Leu Asp Lys Tyr
20 25 30

Lys Asn Ala Val Thr Glu Leu Gln Leu Leu Met Gln Ser Thr
35 40 45

<210> 11
<211> 54
<212> PRT
<213> Artificial Sequence

<220>
<223> Description of Artificial Sequence: synthetic peptide

<400> 11
Ala Ser Gly Val Ala Val Ser Lys Val Leu His Leu Glu Gly Glu Val
1 5 10 15

Asn Lys Ile Ala Leu Leu Ser Thr Asn Lys Ala Val Val Ser Leu Ser
20 25 30

Asn Gly Val Ser Val Leu Thr Ser Lys Val Leu Asp Leu Lys Asn Tyr
35 40 45

Ile Asp Lys Gln Leu Leu
50

<210> 12
<211> 53
<212> PRT
<213> Artificial Sequence

<220>
<223> Description of Artificial Sequence: synthetic peptide

<400> 12
Gly Glu Pro Ile Ile Asn Phe Tyr Asp Pro Leu Val Phe Pro Ser Asp
1 5 10 15

Glu Phe Asp Ala Ser Ile Ser Gln Val Asn Glu Lys Ile Asn Gln Ser
20 25 30

Leu Ala Phe Ile Arg Lys Ser Asp Glu Leu Leu His Asn Val Asn Ala
35 40 45

Gly Lys Ser Thr Thr
50

<210> 13
<211> 48
<212> PRT
<213> Artificial Sequence

<220>
<223> Description of Artificial Sequence: synthetic peptide

<400> 13
Tyr Thr Ser Val Ile Thr Ile Glu Leu Ser Asn Ile Lys Glu Asn Lys
1 5 10 15

Cys Asn Gly Thr Asp Ala Lys Val Lys Leu Ile Lys Gln Glu Leu Asp
20 25 30

Lys Tyr Lys Asn Ala Val Thr Glu Leu Gln Leu Leu Met Gln Ser Thr
35 40 45

<210> 14
<211> 34
<212> PRT
<213> Artificial Sequence

<220>
<223> Description of Artificial Sequence: synthetic peptide

<400> 14
Tyr Thr Ser Val Ile Thr Ile Glu Leu Ser Asn Ile Lys Glu Asn Lys
1 5 10 15

Cys Asn Gly Asp Ala Lys Val Lys Leu Ile Lys Gln Glu Leu Asp Lys
20 25 30

Tyr Lys

<210> 15
<211> 34
<212> PRT
<213> Artificial Sequence

<220>
<223> Description of Artificial Sequence: synthetic peptide

<400> 15
Thr Ser Val Ile Thr Ile Glu Leu Ser Asn Ile Lys Glu Asn Lys Cys
1 5 10 15

Asn Gly Asp Ala Lys Val Lys Leu Ile Lys Gln Glu Leu Asp Lys Tyr
20 25 30

Lys Asn

<210> 16
<211> 34
<212> PRT
<213> Artificial Sequence

<220>
<223> Description of Artificial Sequence: synthetic peptide

<400> 16
Val Ile Thr Ile Glu Leu Ser Asn Ile Lys Glu Asn Lys Cys Asn Gly
1 5 10 15

Asp Ala Lys Val Lys Leu Ile Lys Gln Glu Leu Asp Lys Tyr Lys Asn
20 25 30

Ala Val

<210> 17
<211> 34
<212> PRT
<213> Artificial Sequence

<220>
<223> Description of Artificial Sequence: synthetic peptide

<400> 17
Val Ile Thr Ile Glu Leu Ser Asn Ile Lys Glu Asn Lys Met Asn Gly
1 5 10 15

Asp Ala Lys Val Lys Leu Ile Lys Gln Glu Leu Asp Lys Tyr Lys Asn
20 25 30

Ala Val

<210> 18
<211> 33
<212> PRT
<213> Artificial Sequence

<220>
<223> Description of Artificial Sequence: synthetic peptide

<400> 18
Val Ala Val Ser Lys Val Leu His Leu Glu Gly Glu Val Asn Lys Ile
1 5 10 15

Ala Leu Leu Ser Thr Asn Lys Ala Val Val Ser Leu Ser Asn Gly Val
20 25 30

Ser

<210> 19
<211> 33
<212> PRT
<213> Artificial Sequence

<220>
<223> Description of Artificial Sequence: synthetic peptide

<400> 19
Ala Val Ser Lys Val Leu His Leu Glu Gly Glu Val Asn Lys Ile Ala
1 5 10 15

Leu Leu Ser Thr Asn Lys Ala Val Val Ser Leu Ser Asn Gly Val Ser
20 25 30

Val

<210> 20
<211> 33
<212> PRT
<213> Artificial Sequence

<220>
<223> Description of Artificial Sequence: synthetic peptide

<400> 20
Val Ser Lys Val Leu His Leu Glu Gly Glu Val Asn Lys Ile Ala Leu
1 5 10 15

Leu Ser Thr Asn Lys Ala Val Val Ser Leu Ser Asn Gly Val Ser Val
20 25 30

Leu

<210> 21

<211> 33
<212> PRT
<213> Artificial Sequence

<220>
<223> Description of Artificial Sequence: synthetic peptide

<400> 21
Ser Lys Val Leu His Leu Glu Gly Glu Val Asn Lys Ile Ala Leu Leu
1 5 10 15

Ser Thr Asn Lys Ala Val Val Ser Leu Ser Asn Gly Val Ser Val Leu
20 25 30

Thr

<210> 22
<211> 33
<212> PRT
<213> Artificial Sequence

<220>
<223> Description of Artificial Sequence: synthetic peptide

<400> 22
Lys Val Leu His Leu Glu Gly Glu Val Asn Lys Ile Ala Leu Leu Ser
1 5 10 15

Thr Asn Lys Ala Val Val Ser Leu Ser Asn Gly Val Ser Val Leu Thr
20 25 30

Ser

<210> 23
<211> 33
<212> PRT
<213> Artificial Sequence

<220>
<223> Description of Artificial Sequence: synthetic peptide

<400> 23
Leu Glu Gly Glu Val Asn Lys Ile Ala Leu Leu Ser Thr Asn Lys Ala
1 5 10 15

Val Val Ser Leu Ser Asn Gly Val Ser Val Leu Thr Ser Lys Val Leu
20 25 30

Asp

<210> 24
<211> 33
<212> PRT
<213> Artificial Sequence

<220>
<223> Description of Artificial Sequence: synthetic peptide

<400> 24
Gly Glu Val Asn Lys Ile Ala Leu Leu Ser Thr Asn Lys Ala Val Val
1 5 10 15

Ser Leu Ser Asn Gly Val Ser Val Leu Thr Ser Lys Val Leu Asp Leu
20 25 30

Lys

<210> 25
<211> 33
<212> PRT
<213> Artificial Sequence

<220>
<223> Description of Artificial Sequence: synthetic peptide

<400> 25
Glu Val Asn Lys Ile Ala Leu Leu Ser Thr Asn Lys Ala Val Val Ser
1 5 10 15

Leu Ser Asn Gly Val Ser Val Leu Thr Ser Lys Val Leu Asp Leu Lys
20 25 30

Asn

<210> 26
<211> 33
<212> PRT
<213> Artificial Sequence

<220>
<223> Description of Artificial Sequence: synthetic peptide

<400> 26
Val Asn Lys Ile Ala Leu Leu Ser Thr Asn Lys Ala Val Val Ser Leu
1 5 10 15

Ser Asn Gly Val Ser Val Leu Thr Ser Lys Val Leu Asp Leu Lys Asn
20 25 30

Tyr

<210> 27
<211> 33
<212> PRT
<213> Artificial Sequence

<220>
<223> Description of Artificial Sequence: synthetic peptide

<400> 27
Asn Lys Ile Ala Leu Leu Ser Thr Asn Lys Ala Val Val Ser Leu Ser
1 5 10 15

Asn Gly Val Ser Val Leu Thr Ser Lys Val Leu Asp Leu Lys Asn Tyr
20 25 30

Ile

<210> 28
<211> 33
<212> PRT
<213> Artificial Sequence

<220>
<223> Description of Artificial Sequence: synthetic peptide

<400> 28
Lys Ile Ala Leu Leu Ser Thr Asn Lys Ala Val Val Ser Leu Ser Asn
1 5 10 15

Gly Val Ser Val Leu Thr Ser Lys Val Leu Asp Leu Lys Asn Tyr Ile
20 25 30

Asp

<210> 29
<211> 33
<212> PRT
<213> Artificial Sequence

<220>
<223> Description of Artificial Sequence: synthetic peptide

<400> 29
Ile Ala Leu Leu Ser Thr Asn Lys Ala Val Val Ser Leu Ser Asn Gly
1 5 10 15

Val Ser Val Leu Thr Ser Lys Val Leu Asp Leu Lys Asn Tyr Ile Asp
20 25 30

Lys

<210> 30
<211> 33
<212> PRT
<213> Artificial Sequence

<220>
<223> Description of Artificial Sequence: synthetic peptide

<400> 30
Ala Leu Leu Ser Thr Asn Lys Ala Val Val Ser Leu Ser Asn Gly Val
1 5 10 15

Ser Val Leu Thr Ser Lys Val Leu Asp Leu Lys Asn Tyr Ile Asp Lys
20 25 30

Gln

<210> 31
<211> 70
<212> PRT
<213> Artificial Sequence

<220>
<223> Description of Artificial Sequence: synthetic peptide

<400> 31
Gly Thr Ile Ala Leu Gly Val Ala Thr Ser Ala Gln Ile Thr Ala Ala
1 5 10 15

Val Ala Leu Val Glu Ala Lys Gln Ala Arg Ser Asp Ile Glu Lys Leu
20 25 30

Lys Glu Ala Ile Arg Asp Thr Asn Lys Ala Val Gln Ser Val Gln Ser
35 40 45

Ser Ile Gly Asn Leu Ile Val Ala Ile Lys Ser Val Gln Asp Tyr Val
50 55 60

Asn Lys Glu Ile Val Pro
65 70

<210> 32
<211> 56
<212> PRT
<213> Artificial Sequence

<220>
<223> Description of Artificial Sequence: synthetic peptide

<400> 32
Tyr Thr Pro Asn Asp Ile Thr Leu Asn Asn Ser Val Ala Leu Asp Pro
1 5 10 15

Ile Asp Ile Ser Ile Glu Leu Asn Lys Ala Lys Ser Asp Leu Glu Glu
20 25 30

Ser Lys Glu Trp Ile Arg Arg Ser Asn Gln Lys Leu Asp Ser Ile Gly
35 40 45

Asn Trp His Gln Ser Ser Thr Thr
50 55

<210> 33
<211> 35
<212> PRT
<213> Artificial Sequence

<220>
<223> Description of Artificial Sequence: synthetic peptide

<400> 33
Thr Leu Asn Asn Ser Val Ala Leu Asp Pro Ile Asp Ile Ser Ile Glu
1 5 10 15

Leu Asn Lys Ala Lys Ser Asp Leu Glu Glu Ser Lys Glu Trp Ile Arg
20 25 30

Arg Ser Asn
35

<210> 34
<211> 35
<212> PRT
<213> Artificial Sequence

<220>
<223> Description of Artificial Sequence: synthetic peptide

<400> 34
Leu Asn Asn Ser Val Ala Leu Asp Pro Ile Asp Ile Ser Ile Glu Leu
1 5 10 15

Asn Lys Ala Lys Ser Asp Leu Glu Glu Ser Lys Glu Trp Ile Arg Arg
20 25 30

Ser Asn Gln
35

<210> 35
<211> 35
<212> PRT
<213> Artificial Sequence

<220>
<223> Description of Artificial Sequence: synthetic peptide

<400> 35
Asn Asn Ser Val Ala Leu Asp Pro Ile Asp Ile Ser Ile Glu Leu Asn
1 5 10 15

Lys Ala Lys Ser Asp Leu Glu Glu Ser Lys Glu Trp Ile Arg Arg Ser
20 25 30

Asn Gln Lys
35

<210> 36
<211> 35
<212> PRT
<213> Artificial Sequence

<220>
<223> Description of Artificial Sequence: synthetic peptide

<400> 36
Asn Ser Val Ala Leu Asp Pro Ile Asp Ile Ser Ile Glu Leu Asn Lys
1 5 10 15

Ala Lys Ser Asp Leu Glu Glu Ser Lys Glu Trp Ile Arg Arg Ser Asn
20 25 30

Gln Lys Leu
35

<210> 37
<211> 35
<212> PRT
<213> Artificial Sequence

<220>
<223> Description of Artificial Sequence: synthetic peptide

<400> 37
Ser Val Ala Leu Asp Pro Ile Asp Ile Ser Ile Glu Leu Asn Lys Ala
1 5 10 15

Lys Ser Asp Leu Glu Glu Ser Lys Glu Trp Ile Arg Arg Ser Asn Gln
20 25 30

Lys Leu Asp
35

<210> 38
<211> 35
<212> PRT
<213> Artificial Sequence

<220>
<223> Description of Artificial Sequence: synthetic peptide

<400> 38
Val Ala Leu Asp Pro Ile Asp Ile Ser Ile Glu Leu Asn Lys Ala Lys
1 5 10 15

Ser Asp Leu Glu Glu Ser Lys Glu Trp Ile Arg Arg Ser Asn Gln Lys
20 25 30

Leu Asp Ser
35

<210> 39
<211> 35
<212> PRT
<213> Artificial Sequence

<220>
<223> Description of Artificial Sequence: synthetic peptide

<400> 39
Ala Leu Asp Pro Ile Asp Ile Ser Ile Glu Leu Asn Lys Ala Lys Ser
1 5 10 15

Asp Leu Glu Glu Ser Lys Glu Trp Ile Arg Arg Ser Asn Gln Lys Leu
20 25 30

Asp Ser Ile
35

<210> 40
<211> 35
<212> PRT
<213> Artificial Sequence

<220>
<223> Description of Artificial Sequence: synthetic peptide

<400> 40
Leu Asp Pro Ile Asp Ile Ser Ile Glu Leu Asn Lys Ala Lys Ser Asp
1 5 10 15

Leu Glu Glu Ser Lys Glu Trp Ile Arg Arg Ser Asn Gln Lys Leu Asp
20 25 30

Ser Ile Gly
35

<210> 41
<211> 35
<212> PRT
<213> Artificial Sequence

<220>
<223> Description of Artificial Sequence: synthetic peptide

<400> 41
Asp Pro Ile Asp Ile Ser Ile Glu Leu Asn Lys Ala Lys Ser Asp Leu
1 5 10 15

Glu Glu Ser Lys Glu Trp Ile Arg Arg Ser Asn Gln Lys Leu Asp Ser
20 25 30

Ile Gly Asn
35

<210> 42
<211> 35
<212> PRT
<213> Artificial Sequence

<220>
<223> Description of Artificial Sequence: synthetic peptide

<400> 42
Pro Ile Asp Ile Ser Ile Glu Leu Asn Lys Ala Lys Ser Asp Leu Glu
1 5 10 15

Glu Ser Lys Glu Trp Ile Arg Arg Ser Asn Gln Lys Leu Asp Ser Ile
20 25 30

Gly Asn Trp
35

<210> 43
<211> 35
<212> PRT
<213> Artificial Sequence

<220>
<223> Description of Artificial Sequence: synthetic peptide

<400> 43
Ile Asp Ile Ser Ile Glu Leu Asn Lys Ala Lys Ser Asp Leu Glu Glu
1 5 10 15

Ser Lys Glu Trp Ile Arg Arg Ser Asn Gln Lys Leu Asp Ser Ile Gly
20 25 30

Asn Trp His
35

<210> 44
<211> 35
<212> PRT
<213> Artificial Sequence

<220>
<223> Description of Artificial Sequence: synthetic peptide

<400> 44
Asp Ile Ser Ile Glu Leu Asn Lys Ala Lys Ser Asp Leu Glu Glu Ser
1 5 10 15

Lys Glu Trp Ile Arg Arg Ser Asn Gln Lys Leu Asp Ser Ile Gly Asn
20 25 30

Trp His Gln
35

<210> 45
<211> 35
<212> PRT
<213> Artificial Sequence

<220>
<223> Description of Artificial Sequence: synthetic peptide

<400> 45
Ile Ser Ile Glu Leu Asn Lys Ala Lys Ser Asp Leu Glu Glu Ser Lys
1 5 10 15

Glu Trp Ile Arg Arg Ser Asn Gln Lys Leu Asp Ser Ile Gly Asn Trp
20 25 30

His Gln Ser
35

<210> 46
<211> 35
<212> PRT
<213> Artificial Sequence

<220>
<223> Description of Artificial Sequence: synthetic peptide

<400> 46
Ser Ile Glu Leu Asn Lys Ala Lys Ser Asp Leu Glu Glu Ser Lys Glu
1 5 10 15

Trp Ile Arg Arg Ser Asn Gln Lys Leu Asp Ser Ile Gly Asn Trp His
20 25 30

Gln Ser Ser
35

<210> 47
<211> 35
<212> PRT
<213> Artificial Sequence

<220>
<223> Description of Artificial Sequence: synthetic peptide

<400> 47
Ile Glu Leu Asn Lys Ala Lys Ser Asp Leu Glu Glu Ser Lys Glu Trp
1 5 10 15

Ile Arg Arg Ser Asn Gln Lys Leu Asp Ser Ile Gly Asn Trp His Gln
20 25 30

Ser Ser Thr
35

<210> 48
<211> 35
<212> PRT
<213> Artificial Sequence

<220>
<223> Description of Artificial Sequence: synthetic peptide

<400> 48
Glu Leu Asn Lys Ala Lys Ser Asp Leu Glu Glu Ser Lys Glu Trp Ile
1 5 10 15

Arg Arg Ser Asn Gln Lys Leu Asp Ser Ile Gly Asn Trp His Gln Ser
20 25 30

Ser Thr Thr
35

<210> 49
<211> 35
<212> PRT
<213> Artificial Sequence

<220>
<223> Description of Artificial Sequence: synthetic peptide

<400> 49
Thr Ala Ala Val Ala Leu Val Glu Ala Lys Gln Ala Arg Ser Asp Ile
1 5 10 15

Glu Lys Leu Lys Glu Ala Ile Arg Asp Thr Asn Lys Ala Val Gln Ser
20 25 30

Val Gln Ser
35

<210> 50
<211> 35
<212> PRT
<213> Artificial Sequence

<220>
<223> Description of Artificial Sequence: synthetic peptide

<400> 50
Ala Val Ala Leu Val Glu Ala Lys Gln Ala Arg Ser Asp Ile Glu Lys
1 5 10 15

Leu Lys Glu Ala Ile Arg Asp Thr Asn Lys Ala Val Gln Ser Val Gln
20 25 30

Ser Ser Ile
35

<210> 51
<211> 35
<212> PRT
<213> Artificial Sequence

<220>
<223> Description of Artificial Sequence: synthetic peptide

<400> 51
Leu Val Glu Ala Lys Gln Ala Arg Ser Asp Ile Glu Lys Leu Lys Glu
1 5 10 15

Ala Ile Arg Asp Thr Asn Lys Ala Val Gln Ser Val Gln Ser Ser Ile
20 25 30

Gly Asn Leu
35

<210> 52
<211> 35
<212> PRT
<213> Artificial Sequence

<220>
<223> Description of Artificial Sequence: synthetic peptide

<400> 52
Val Glu Ala Lys Gln Ala Arg Ser Asp Ile Glu Lys Leu Lys Glu Ala
1 5 10 15

Ile Arg Asp Thr Asn Lys Ala Val Gln Ser Val Gln Ser Ser Ile Gly
20 25 30

Asn Leu Ile
35

<210> 53
<211> 35
<212> PRT
<213> Artificial Sequence

<220>
<223> Description of Artificial Sequence: synthetic peptide

<400> 53
Glu Ala Lys Gln Ala Arg Ser Asp Ile Glu Lys Leu Lys Glu Ala Ile
1 5 10 15

Arg Asp Thr Asn Lys Ala Val Gln Ser Val Gln Ser Ser Ile Gly Asn
20 25 30

Leu Ile Val
35

<210> 54
<211> 35
<212> PRT
<213> Artificial Sequence

<220>
<223> Description of Artificial Sequence: synthetic peptide

<400> 54
Ala Lys Gln Ala Arg Ser Asp Ile Glu Lys Leu Lys Glu Ala Ile Arg
1 5 10 15

Asp Thr Asn Lys Ala Val Gln Ser Val Gln Ser Ser Ile Gly Asn Leu
20 25 30

Ile Val Ala
35

<210> 55
<211> 35
<212> PRT
<213> Artificial Sequence

<220>
<223> Description of Artificial Sequence: synthetic peptide

<400> 55
Lys Gln Ala Arg Ser Asp Ile Glu Lys Leu Lys Glu Ala Ile Arg Asp
1 5 10 15

Thr Asn Lys Ala Val Gln Ser Val Gln Ser Ser Ile Gly Asn Leu Ile
20 25 30

Val Ala Ile
35

<210> 56
<211> 35
<212> PRT
<213> Artificial Sequence

<220>
<223> Description of Artificial Sequence: synthetic peptide

<400> 56
Gln Ala Arg Ser Asp Ile Glu Lys Leu Lys Glu Ala Ile Arg Asp Thr
1 5 10 15

Asn Lys Ala Val Gln Ser Val Gln Ser Ser Ile Gly Asn Leu Ile Val
20 25 30

Ala Ile Lys
35

<210> 57
<211> 35
<212> PRT
<213> Artificial Sequence

<220>
<223> Description of Artificial Sequence: synthetic peptide

<400> 57
Ala Arg Ser Asp Ile Glu Lys Leu Lys Glu Ala Ile Arg Asp Thr Asn
1 5 10 15

Lys Ala Val Gln Ser Val Gln Ser Ser Ile Gly Asn Leu Ile Val Ala
20 25 30

Ile Lys Ser
35

<210> 58
<211> 35
<212> PRT
<213> Artificial Sequence

<220>
<223> Description of Artificial Sequence: synthetic peptide

<400> 58
Arg Ser Asp Ile Glu Lys Leu Lys Glu Ala Ile Arg Asp Thr Asn Lys
1 5 10 15

Ala Val Gln Ser Val Gln Ser Ser Ile Gly Asn Leu Ile Val Ala Ile
20 25 30

Lys Ser Val
35

<210> 59
<211> 35
<212> PRT
<213> Artificial Sequence

<220>
<223> Description of Artificial Sequence: synthetic peptide

<400> 59
Ser Asp Ile Glu Lys Leu Lys Glu Ala Ile Arg Asp Thr Asn Lys Ala
1 5 10 15

Val Gln Ser Val Gln Ser Ser Ile Gly Asn Leu Ile Val Ala Ile Lys
20 25 30

Ser Val Gln
35

<210> 60
<211> 35
<212> PRT
<213> Artificial Sequence

<220>
<223> Description of Artificial Sequence: synthetic peptide

<400> 60
Lys Leu Lys Glu Ala Ile Arg Asp Thr Asn Lys Ala Val Gln Ser Val
1 5 10 15

Gln Ser Ser Ile Gly Asn Leu Ile Val Ala Ile Lys Ser Val Gln Asp
20 25 30

Tyr Val Asn
35

<210> 61
<211> 35
<212> PRT
<213> Artificial Sequence

<220>
<223> Description of Artificial Sequence: synthetic peptide

<400> 61
Leu Lys Glu Ala Ile Arg Asp Thr Asn Lys Ala Val Gln Ser Val Gln
1 5 10 15

Ser Ser Ile Gly Asn Leu Ile Val Ala Ile Lys Ser Val Gln Asp Tyr
20 25 30

Val Asn Lys
35

<210> 62
<211> 35
<212> PRT
<213> Artificial Sequence

<220>
<223> Description of Artificial Sequence: synthetic peptide

<400> 62
Ala Ile Arg Asp Thr Asn Lys Ala Val Gln Ser Val Gln Ser Ser Ile
1 5 10 15

Gly Asn Leu Ile Val Ala Ile Lys Ser Val Gln Asp Tyr Val Asn Lys
20 25 30

Glu Ile Val
35

<210> 63
<211> 47
<212> PRT
<213> Artificial Sequence

<220>
<223> Description of Artificial Sequence: synthetic peptide

<400> 63
Thr Trp Gln Glu Trp Glu Arg Lys Val Asp Phe Leu Glu Glu Asn Ile
1 5 10 15

Thr Ala Leu Leu Glu Glu Ala Gln Ile Gln Gln Glu Lys Asn Met Tyr
20 25 30

Glu Leu Gln Lys Leu Asn Ser Trp Asp Val Phe Gly Asn Trp Phe
35 40 45

<210> 64
<211> 35
<212> PRT
<213> Artificial Sequence

<220>
<223> Description of Artificial Sequence: synthetic peptide

<400> 64
Trp Gln Glu Trp Glu Arg Lys Val Asp Phe Leu Glu Glu Asn Ile Thr
1 5 10 15

Ala Leu Leu Glu Glu Ala Gln Ile Gln Gln Glu Lys Asn Met Tyr Glu
20 25 30

Leu Gln Lys
35

<210> 65
<211> 35
<212> PRT
<213> Artificial Sequence

<220>
<223> Description of Artificial Sequence: synthetic peptide

<400> 65
Gln Glu Trp Glu Arg Lys Val Asp Phe Leu Glu Glu Asn Ile Thr Ala
1 5 10 15

Leu Leu Glu Glu Ala Gln Ile Gln Gln Glu Lys Asn Met Tyr Glu Leu
20 25 30

Gln Lys Leu
35

<210> 66
<211> 35
<212> PRT
<213> Artificial Sequence

<220>
<223> Description of Artificial Sequence: synthetic peptide

<400> 66
Glu Trp Glu Arg Lys Val Asp Phe Leu Glu Glu Asn Ile Thr Ala Leu
1 5 10 15

Leu Glu Glu Ala Gln Ile Gln Gln Glu Lys Asn Met Tyr Glu Leu Gln
20 25 30

Lys Leu Asn
35

<210> 67
<211> 35
<212> PRT
<213> Artificial Sequence

<220>
<223> Description of Artificial Sequence: synthetic peptide

<400> 67
Trp Glu Arg Lys Val Asp Phe Leu Glu Glu Asn Ile Thr Ala Leu Leu
1 5 10 15

Glu Glu Ala Gln Ile Gln Gln Glu Lys Asn Met Tyr Glu Leu Gln Lys
20 25 30

Leu Asn Ser
35

<210> 68
<211> 35
<212> PRT
<213> Artificial Sequence

<220>
<223> Description of Artificial Sequence: synthetic peptide

<400> 68
Glu Arg Lys Val Asp Phe Leu Glu Glu Asn Ile Thr Ala Leu Leu Glu
1 5 10 15

Glu Ala Gln Ile Gln Gln Glu Lys Asn Met Tyr Glu Leu Gln Lys Leu
20 25 30

Asn Ser Trp
35

<210> 69
<211> 35
<212> PRT
<213> Artificial Sequence

<220>
<223> Description of Artificial Sequence: synthetic peptide

<400> 69
Arg Lys Val Asp Phe Leu Glu Glu Asn Ile Thr Ala Leu Leu Glu Glu
1 5 10 15

Ala Gln Ile Gln Gln Glu Lys Asn Met Tyr Glu Leu Gln Lys Leu Asn
20 25 30

Ser Trp Asp
35

<210> 70
<211> 35
<212> PRT
<213> Artificial Sequence

<220>
<223> Description of Artificial Sequence: synthetic peptide

<400> 70
Lys Val Asp Phe Leu Glu Glu Asn Ile Thr Ala Leu Leu Glu Glu Ala
1 5 10 15

Gln Ile Gln Gln Glu Lys Asn Met Tyr Glu Leu Gln Lys Leu Asn Ser
20 25 30

Trp Asp Val
35

<210> 71
<211> 35
<212> PRT
<213> Artificial Sequence

<220>
<223> Description of Artificial Sequence: synthetic peptide

<400> 71
Val Asp Phe Leu Glu Glu Asn Ile Thr Ala Leu Leu Glu Glu Ala Gln
1 5 10 15

Ile Gln Gln Glu Lys Asn Met Tyr Glu Leu Gln Lys Leu Asn Ser Trp
20 25 30

Asp Val Phe
35

<210> 72
<211> 35
<212> PRT
<213> Artificial Sequence

<220>
<223> Description of Artificial Sequence: synthetic peptide

<400> 72
Asp Phe Leu Glu Glu Asn Ile Thr Ala Leu Leu Glu Glu Ala Gln Ile
1 5 10 15

Gln Gln Glu Lys Asn Met Tyr Glu Leu Gln Lys Leu Asn Ser Trp Asp
20 25 30

Val Phe Gly
35

<210> 73
<211> 35
<212> PRT
<213> Artificial Sequence

<220>
<223> Description of Artificial Sequence: synthetic peptide

<400> 73
Phe Leu Glu Glu Asn Ile Thr Ala Leu Leu Glu Glu Ala Gln Ile Gln
1 5 10 15

Gln Glu Lys Asn Met Tyr Glu Leu Gln Lys Leu Asn Ser Trp Asp Val
20 25 30

Phe Gly Asn
35

<210> 74
<211> 35
<212> PRT
<213> Artificial Sequence

<220>
<223> Description of Artificial Sequence: synthetic peptide

<400> 74
Pro Asp Ala Val Tyr Leu His Arg Ile Asp Leu Gly Pro Pro Ile Ser
1 5 10 15

Leu Glu Arg Leu Asp Val Gly Thr Asn Leu Gly Asn Ala Ile Ala Lys
20 25 30

Leu Glu Asp
35

<210> 75
<211> 34
<212> PRT
<213> Artificial Sequence

<220>
<223> Description of Artificial Sequence: synthetic peptide

<400> 75
Leu Glu Arg Leu Asp Val Gly Thr Asn Leu Gly Asn Ala Ile Ala Lys
1 5 10 15

Leu Glu Ala Lys Glu Leu Leu Glu Ser Ser Asp Gln Ile Leu Arg Ser
20 25 30

Met Lys

<210> 76
<211> 34
<212> PRT
<213> Artificial Sequence

<220>
<223> Description of Artificial Sequence: synthetic peptide

<400> 76
Leu His Arg Ile Asp Leu Gly Pro Pro Ile Ser Leu Glu Arg Leu Asp
1 5 10 15

Val Gly Thr Asn Leu Gly Asn Ala Ile Ala Lys Leu Glu Ala Lys Glu
20 25 30

Leu Leu

<210> 77
<211> 34
<212> PRT
<213> Artificial Sequence

<220>
<223> Description of Artificial Sequence: synthetic peptide

<400> 77
His Arg Ile Asp Leu Gly Pro Pro Ile Ser Leu Glu Arg Leu Asp Val
1 5 10 15

Gly Thr Asn Leu Gly Asn Ala Ile Ala Lys Leu Glu Ala Lys Glu Leu
20 25 30

Leu Glu

<210> 78
<211> 34
<212> PRT
<213> Artificial Sequence

<220>
<223> Description of Artificial Sequence: synthetic peptide

<400> 78
Arg Ile Asp Leu Gly Pro Pro Ile Ser Leu Glu Arg Leu Asp Val Gly
1 5 10 15

Thr Asn Leu Gly Asn Ala Ile Ala Lys Leu Glu Ala Lys Glu Leu Leu
20 25 30

Glu Ser

<210> 79
<211> 34
<212> PRT
<213> Artificial Sequence

<220>
<223> Description of Artificial Sequence: synthetic peptide

<400> 79
Ile Asp Leu Gly Pro Pro Ile Ser Leu Glu Arg Leu Asp Val Gly Thr
1 5 10 15

Asn Leu Gly Asn Ala Ile Ala Lys Leu Glu Ala Lys Glu Leu Leu Glu
20 25 30

Ser Ser

<210> 80
<211> 34
<212> PRT
<213> Artificial Sequence

<220>
<223> Description of Artificial Sequence: synthetic peptide

<400> 80
Asp Leu Gly Pro Pro Ile Ser Leu Glu Arg Leu Asp Val Gly Thr Asn
1 5 10 15

Leu Gly Asn Ala Ile Ala Lys Leu Glu Ala Lys Glu Leu Leu Glu Ser
20 25 30

Ser Asp

<210> 81
<211> 34
<212> PRT
<213> Artificial Sequence

<220>
<223> Description of Artificial Sequence: synthetic peptide

<400> 81
Leu Gly Pro Pro Ile Ser Leu Glu Arg Leu Asp Val Gly Thr Asn Leu
1 5 10 15

Gly Asn Ala Ile Ala Lys Leu Glu Ala Lys Glu Leu Leu Glu Ser Ser
20 25 30

Asp Gln

<210> 82
<211> 34
<212> PRT
<213> Artificial Sequence

<220>
<223> Description of Artificial Sequence: synthetic peptide

<400> 82
Gly Pro Pro Ile Ser Leu Glu Arg Leu Asp Val Gly Thr Asn Leu Gly
1 5 10 15

Asn Ala Ile Ala Lys Leu Glu Ala Lys Glu Leu Leu Glu Ser Ser Asp
20 25 30

Gln Ile

<210> 83
<211> 34
<212> PRT
<213> Artificial Sequence

<220>
<223> Description of Artificial Sequence: synthetic peptide

<400> 83
Pro Pro Ile Ser Leu Glu Arg Leu Asp Val Gly Thr Asn Leu Gly Asn
1 5 10 15

Ala Ile Ala Lys Leu Glu Ala Lys Glu Leu Leu Glu Ser Ser Asp Gln
20 25 30

Ile Leu

<210> 84
<211> 34
<212> PRT
<213> Artificial Sequence

<220>
<223> Description of Artificial Sequence: synthetic peptide

<400> 84
Pro Ile Ser Leu Glu Arg Leu Asp Val Gly Thr Asn Leu Gly Asn Ala
1 5 10 15

Ile Ala Lys Leu Glu Ala Lys Glu Leu Leu Glu Ser Ser Asp Gln Ile
20 25 30

Leu Arg

<210> 85
<211> 34
<212> PRT
<213> Artificial Sequence

<220>
<223> Description of Artificial Sequence: synthetic peptide

<400> 85
Ser Leu Glu Arg Leu Asp Val Gly Thr Asn Leu Gly Asn Ala Ile Ala
1 5 10 15

Lys Leu Glu Ala Lys Glu Leu Leu Glu Ser Ser Asp Gln Ile Leu Arg
20 25 30

Ser Met

<210> 86
<211> 34
<212> PRT
<213> Artificial Sequence

<220>
<223> Description of Artificial Sequence: synthetic peptide

<400> 86
Leu Glu Arg Leu Asp Val Gly Thr Asn Leu Gly Asn Ala Ile Ala Lys
1 5 10 15

Leu Glu Ala Lys Glu Leu Leu Glu Ser Ser Asp Gln Ile Leu Arg Ser
20 25 30

Met Lys

Claims

1. A modified anti-viral peptide comprising:

   a peptide that exhibits anti-viral activity, and
   a maleimide group which is reactive with a thiol group on blood components to form stable covalent bonds.

2. The modified peptide of claim 1 wherein the blood component is serum albumin.

3. The modified peptide of claim 1 or claim 2 wherein said peptide is DP178 or DP107 or analogs thereof.

4. The modified peptide of any one of claims 1,2 or 3 wherein said peptide exhibits anti-viral activity against human immunodeficiency virus (HIV).

5. The modified peptide of claim 4 wherein said peptide is selected from the group consisting of SEQ ID NO: 1 to SEQ ID NO: 9.

6. The modified peptide of claim 4 wherein said peptide is DP178 or DP107.

7. The modified peptide of claim 1 or claim 2 wherein said peptide exhibits anti-viral activity against human respiratory

syncytial virus (RSV).

8. The modified peptide of claim 7 wherein said peptide is selected from the group consisting of SEQ ID NO: 10 to SEQ ID NO: 30.

9. The modified peptide of claim 7 wherein said peptide is selected from the group consisting of SEQ ID NO: 14 to SEQ ID NO: 17 and SEQ ID NO: 29.

10. The modified peptide of claim 1 or claim 2 wherein said peptide exhibits anti-viral activity against human parainfluenza virus (HPIV).

11. The modified peptide of claim 10 wherein said peptide is selected from the group consisting of SEQ ID NO: 31 to SEQ ID NO: 62.

12. The modified peptide of claim 10 wherein said peptide is selected from the group consisting of SEQ ID NO: 35, SEQ ID NO: 38 to SEQ ID NO: 42, SEQ ID NO: 52 and SEQ ID NO: 58.

13. The modified peptide of claim 1 or claim 2 wherein said peptide exhibits anti-viral activity against measles virus (MeV).

14. The modified peptide of claim 13 wherein said peptide is selected from the group consisting of SEQ ID NO: 74 to SEQ ID NO: 86.

15. The modified peptide of claim 13 wherein said peptide is selected from the group consisting of SEQ ID NO: 77, SEQ ID NO: 79, SEQ ID NO: 81 and SEQ ID NO: 84.

16. The modified peptide of claim 1 or claim 2 wherein said peptide exhibits anti-viral activity against simian immunodeficiency virus (SIV).

17. The modified peptide of claim 16 wherein said peptide is selected from the group consisting of SEQ ID NO: 63 to SEQ ID NO: 73.

18. A composition for use in the prevention and/or treatment of acquired immune deficiency syndrome (AIDS) comprising a peptide that exhibits anti-viral activity against human immunodeficiency virus (HTV), modified with a maleimide group which is reactive with a thiol group on blood components to form stable covalent bonds.

19. The composition of claim 18 wherein said peptide is DP178 or DP107 or analogs thereof.

20. A composition for use in the prevention and/or treatment of human respiratory syncytial virus (RSV) infection comprising a peptide that exhibits anti-viral activity against RSV, modified with a maleimide group which is reactive with a thiol group on blood components to form stable covalent bonds.

21. The composition of claim 20 wherein said peptide is selected from the group consisting of SEQ ID NO: 14 to SEQ ID NO: 17 and SEQ ID NO: 29.

22. A composition for use in the prevention and/or treatment of human parainfluenza virus (HPIV) infection comprising a peptide that exhibits anti-viral activity against human parainfluenza (HPIV), modified with a maleimide group which is reactive with a thiol group on blood components to form stable covalent bonds.

23. The composition of claim 22 wherein said peptide is selected from the group consisting of SEQ ID NO: 35, SEQ ID NO: 38 to SEQ ID NO: 42, SEQ ID NO:52 and SEQ ID NO: 58.

24. A composition for use in the prevention and/or treatment of measles virus (MeV) infection comprising a peptide that exhibits anti-viral activity against measles virus (MeV), modified with a maleimide group which is reactive with a thiol group on blood components to form stable covalent bonds.

25. The composition of claim 24 wherein said peptide is selected from the group consisting of SEQ ID NO: 77, SEQ ID NO: 79, SEQ ID NO: 81 and SEQ ID NO: 84.

26. The composition of any one of claims 18 to 25 wherein the blood component is serum albumin.

27. The modified peptide or composition or any one of the preceding claims wherein the maleimide group is coupled to the peptide via a linking group.

Patentansprüche

1. Ein modifiziertes antivirales Peptid, das folgendes enthält:

ein Peptid, das eine antivirale Wirkung zeigt, und
eine Maleimidgruppe, die mit einer Thiolgruppe an Blutkomponenten unter Bildung stabiler kovalenter Bindungen reaktiv ist.

2. Das modifizierte Peptid nach Anspruch 1, wobei die Blutkomponente Serumalbumin ist.

3. Das modifizierte Peptid nach einem der Ansprüche 1 oder 2, wobei das Peptid DP178 oder DP107 oder Analoge hiervon ist.

4. Das modifizierte Peptid nach einem der Ansprüche 1, 2 oder 3, wobei das Peptid eine antivirale Wirkung gegen das humane Immundefizienz-Virus (HIV) aufweist.

5. Das modifizierte Peptid nach Anspruch 4, wobei das Peptid aus der Gruppe ausgewählt ist, bestehend aus der SEQ ID NR. 1 bis SEQ ID NR. 9,

6. Das modifizierte Peptid nach Anspruch 4, wobei das Peptid DP 178 oder DP 107 ist.

7. Das modifizierte Peptid nach Anspruch 1 oder 2, wobei das Peptid eine antivirale Wirkung gegen das humane Respiratory Syncytial Virus (RSV) zeigt.

8. Das modifizierte Peptid nach Anspruch 7, wobei das Peptid aus der Gruppe ausgewählt ist, bestehend aus der SEQ ID NR. 10 bis SEQ ID NR. 30.

9. Das modifizierte Peptid nach Anspruch 7, wobei das Peptid aus der Gruppe ausgewählt ist, bestehend aus der SEQ ID NR. 14 bis SEQ ID NR. 17 und SEQ ID NR. 29.

10. Das modifizierte Peptid nach Anspruch 1 oder 2, wobei das Peptid eine antivirale Wirkung gegen das humane Parainfluenza-Virus (GPIV) zeigt.

11. Das modifizierte Peptid nach Anspruch 10, wobei das Peptid aus der Gruppe ausgewählt ist, bestehend aus der SEQ ID NR. 31 bis SEQ ID NR. 62

12. Das modifizierte Peptid nach Anspruch 10, wobei das Peptid aus der Gruppe ausgewählt ist, bestehend aus der SEQ ID NR. 35, SEQ ID NR. 38 bis SEQ ID NR. 42, SEQ ID NR. 52 und SEQ ID NR. 58.

13. Das modifizierte Peptid nach Anspruch 1 oder 2, wobei das Peptid eine antivirale Wirkung gegen das Masernvirus (MeV) zeigt.

14. Das modifizierte Peptid nach Anspruch 13, wobei das Peptid aus der Gruppe ausgewählt ist, bestehend aus der SEQ ID NR. 74 bis SEQ ID NR. 86.

15. Das modifizierte Peptid nach Anspruch 13, wobei das Peptid aus der Gruppe ausgewählt ist, bestehend aus der SEQ ID NR. 77, SEQ ID NR. 79, SEQ ID NR, 81 und SEQ ID NR. 84.

16. Das modifizierte Peptid nach Anspruch 1 oder 2, wobei das Peptid eine antivirale Wirkung gegen das Simian-Immundefizienz-Virus (SIV) zeigt.

17. Das modifizierte Peptid nach Anspruch 16, wobei das Peptid aus der Gruppe ausgewählt ist, bestehend aus der

SEQ ID NR. 63 bis SEQ ID NR. 73.

18. Eine zusammensetzung zur Verwendung bei der Prävention und/oder Behandlung des erworbenen Immundefekt-Syndroms (AIDS), die ein Peptid enthält, das eine antivirale Wirkung gegen das humane Immunodefizienz-Virus (HIV) zeigt, das mit einer Maleimidgruppe modifiziert ist, die mit einer Thiolgruppe an Blutkomponenten unter Bildung stabiler kovalenter Bindungen reaktiv ist.

19. Zusammensetzung nach Anspruch 18, wobei das Peptid DP178 oder DP107 oder Analoge davon ist.

20. Eine Zusammensetzung zur Verwendung bei der Prävention und/oder Behandlung der humanen Respiratory Syncytial Virus (RSV)-Infektion, die ein Peptid enthält, das eine antivirale Wirkung gegen das RSV aufweist, das mit einer Maleimidgruppe modifiziert ist, die mit einer Thiolgruppe an Blutkomponenten unter Bildung stabiler kovalenter Bindungen reaktiv ist.

21. Das modifizierte Peptid nach Anspruch 20, wobei das Peptid aus der Gruppe ausgewählt ist, bestehend aus der SEQ ID NR. 14 bis SEQ ID NR. 17 und SEQ ID NR, 29.

22. Eine Zusammensetzung zur Verwendung bei der Prävention und/oder Behandlung der humanen Parainfluenza-Virus (HPIV)-Infektion, die ein Peptid enthält, das eine antivirale Wirkung gegen humane Parainfluenza (HPIV) aufweist, das mit einer Maleimidgruppe modifiziert ist, die mit einer Thiolgruppe an Blutkomponenten unter Bildung stabiler kovalenter Bindungen reaktiv ist.

23. Die Zusammensetzung nach Anspruch 22, wobei das Peptid aus der Gruppe ausgewählt ist, bestehend aus der SEQ ID NR. 35, SEQ ID NR, 38 bis SEQ ID NR, 42, SEQ ID NR. 52 und SEQ ID NR. 58.

24. Eine Zusammensetzung zur Verwendung bei der Prävention und/oder Behandlung der humanen Masern-Virus (MeV) - Infektion, die ein Peptid enthält, das eine antivirale Wirkung gegen das Masern-Virus (MeV) aufweist und mit einer Maleimidgruppe modifiziert ist, die mit einer Thiolgruppe an Blutkomponenten unter Bildung stabiler kovalenter Bindungen reaktiv ist.

25. Zusammensetzung nach Anspruch 24, wobei das Peptid aus der Gruppe ausgewählt ist, bestehend aus der SEQ ID NR. 77, SEQ ID NR. 79, SEQ ID NR. 81 und SEQ ID NR. 84.

26. Zusammensetzung nach einem der Ansprüche 18 bis 25, wobei die Blutkomponente Serumalbumin ist.

27. Das modifizierte Peptid oder die Zusammensetzung nach einem der vorstehenden Ansprüche, wobei die Maleimidgruppe über eine Verknüpfungsgruppe an das Peptid gekoppelt ist.

Revendications

1. Peptide antiviral modifié comprenant :

un peptide qui présente une activité antivirale, et
un groupe maléimide qui peut réagir avec un groupe thiol sur des composants du sang pour former des liaisons covalentes stables.

2. Peptide modifié de la revendication 1 dans laquelle le composant du sang est la sérum-albumine.

3. Peptide modifié revendication 1 ou 2 dans laquelle ledit peptide est DP178 ou DP107 ou des analogues de ceux-ci.

4. Peptide modifié selon l'une quelconque des revendications 1, 2 ou 3 dans laquelle ledit peptide présente une activité antivirale contre le virus de l'immunodéficience humaine (HIV).

5. Peptide modifié de la revendication 4 dans laquelle ledit peptide est choisi dans le groupe constitué de SEQ ID NO : 1 à SEQ ID NO : 9.

6. Peptide modifié de la revendication 4 dans laquelle ledit peptide est DP178 ou DP107.

7. Peptide modifié de la revendication 1 ou 2 dans laquelle ledit peptide présente une activité antivirale contre le virus syncytial respiratoire humain (RSV).

8. Peptide modifié de la revendication 7 dans laquelle ledit peptide est choisi dans le groupe constitué de SEQ ID NO : 10 à SEQ ID NO : 30.

9. Peptide modifié de la revendication 7 dans laquelle ledit peptide est choisi dans le groupe constitué de SEQ ID NO : 14 à SEQ ID NO : 17 et SEQ ID NO : 29.

10. Peptide modifié de la revendication 1 ou 2 dans laquelle ledit peptide présente une activité antivirale contre le virus parainfluenza humain (HPIV).

11. Peptide modifié de la revendication 10 dans laquelle ledit peptide est choisi dans le groupe constitué de SEQ ID NO : 31 à SEQ ID NO : 62.

12. Peptide modifié de la revendication 10 dans laquelle ledit peptide est choisi dans le groupe constitué de SEQ ID NO : 35, SEQ ID NO : 38 à SEQ ID NO : 42, SEQ ID NO : 52 et SEQ ID NO : 58

13. Peptide modifié de la revendication 1 ou 2 dans laquelle ledit peptide présente une activité antivirale contre le virus de la rougeole (MeV).

14. Peptide modifié de la revendication 13 dans laquelle ledit peptide est choisi dans le groupe constitué de SEQ ID NO : 74 à SEQ ID NO : 86.

15. Peptide modifié de la revendication 13 dans laquelle ledit peptide est choisi dans le groupe constitué de SEQ ID NO : 77, SEQ ID NO : 79, SEQ ID NO : 81 et SEQ ID NO : 84.

16. Peptide modifié de la revendication 1 dans laquelle ledit peptide présente une activité antivirale contre le virus de l'immunodéficience simienne (SIV).

17. Peptide modifié de la revendication 16 dans laquelle ledit peptide est choisi dans le groupe constitué de SEQ ID NO : 63 à SEQ ID NO : 73.

18. Composition utilisable dans la prévention et/ou le traitement du syndrome d'immunodéficience acquise (SIDA) comprenant un peptide qui présente une activité antivirale contre le virus de l'immunodéficience humaine (HIV), modifié avec un groupe maléimide qui peut réagir avec un groupe thiol sur des composants du sang pour former des liaisons covalentes stables.

19. Composition de la revendication 18 dans laquelle ledit peptide est DP178 ou DP107 ou des analogues de ceux-ci.

20. Composition utilisable dans la prévention et/ou le traitement d'une infection par le virus syncytial respiratoire humain (RSV) comprenant un peptide qui présente une activité antivirale contre RSV, modifié avec un groupe maléimide qui peut réagir avec un groupe thiol sur des composants du sang pour former des liaisons covalentes stables.

21. Composition de la revendication 20 dans laquelle ledit peptide est choisi dans le groupe constitué de SEQ ID NO : 14 à SEQ ID NO : 17 et SEQ ID NO : 29.

22. Composition utilisable dans la prévention et/ou le traitement d'une infection par le virus parainfluenza humain (HPIV) comprenant un peptide qui présente une activité antivirale contre le virus parainfluenza humain (HPIV), modifié avec un groupe maléimide qui peut réagir avec un groupe thiol sur des composants du sang pour former des liaisons covalentes stables.

23. Composition de la revendication 22 dans laquelle ledit peptide est choisi dans le groupe constitué de SEQ ID NO : 35, SEQ ID NO : 38 à SEQ ID NO : 42, SEQ ID NO : 52 et SEQ ID NO : 58.

24. Composition utilisable dans la prévention et/ou le traitement d'une infection par le virus de la rougeole (MeV) comprenant un peptide qui présente une activité antivirale contre le virus de la rougeole (MeV), modifié avec un

groupe maléimide qui peut réagir avec un groupe thiol sur des composants du sang pour former des liaisons covalentes stables.

25. Composition de la revendication 24 dans laquelle ledit peptide est choisi dans le groupe constitué de SEQ ID NO : 77, SEQ ID NO : 79, SEQ ID NO : 81 et SEQ ID NO : 84.

26. Composition selon l'une quelconque des revendications 18 à 25 dans laquelle le composant du sang est la sérum-albumine.

27. Peptide modifié ou composition selon l'une quelconque des revendications précédentes dans laquelle le groupe maléimide est couplé au peptide via un groupe de liaison.