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(54) **Method for producing l-amino acid using bacteria belonging to the genus escherichia**

Verfahren zur Produktion von L-Aminosäuren mittels Bakterien der Gattung Escherichia

Procédé de production des acides L-aminés au moyen de bactéries du genre Escherichia

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DescriptionTechnical Field

5 **[0001]** The present invention relates to biotechnology, specifically to a method for producing L-amino acids by fermentation and more specifically to genes derived from bacteria *Escherichia coli*. The genes are useful for improvement of L-amino acid productivity, for example, L-threonine, L-valine, L-proline, L-leucine, L-methionine and L-arginine.

Background art

10 **[0002]** Conventionally the L-amino acids have been industrially produced by method of fermentation utilizing strains of microorganisms obtained from natural sources or mutant of the same especially modified to enhance L-amino acid productivity.

15 **[0003]** There have been disclosed many techniques to enhance L-amino acid productivity, for example, by transformation of microorganism by recombinant DNA (see, for example, US patent No. 4,278,765). These techniques based on the increasing of activities of the enzymes involved into amino acid biosynthesis and/or desensitizing the target enzymes to the feedback inhibition by produced L-amino acid (see, for example, Japanese Laid-open application No56-18596 (1981), WO 95/16042 or US patent Nos. 5,661,012 and 6,040,160).

20 **[0004]** On the other hand, increased L-amino acid excretion can enhance the productivity of strain producing L-amino acid. Strain of bacterium belonging to the genus *Corynebacterium* having increased expression of L-lysine excretion gene (*lysE* gene) is disclosed (WO 9723597A2). In addition, genes coding for the efflux proteins suitable for secretion of L-cysteine, L-cystine, N-acetylserine or thiazolidine derivatives are also disclosed (USA Patent No. 5,972,663). At present several *Escherichia coli* genes coding for putative membrane proteins enhancing L-amino acid production are disclosed. Additional copy of *rhtB* gene makes a bacterium more resistant to L-homoserine and enhances production
25 of L-homoserine, L-threonine, L-alanine, L-valine and L-isoleucine (European patent application publication EP994190A2). Additional copy of *rhtC* gene makes a bacterium more resistant to L-homoserine and L-threonine and enhances production of L-homoserine, L-threonine and L-leucine (European patent application publication EP1013765A1). Additional copy of *yahN*, *yeaS*, *yfiK* and *yggA* genes enhance production of L-glutamic acid, L-lysine,
30 L-threonine L-alanine, L-histidine, L-proline, L-arginine L-valine and L-isoleucine (European patent application publication EP1016710A2). And though complete genome sequence of *Escherichia coli* strain K-12 is described (Blattner F.R., Plunkett G., Bloch C.A. et al., Science, 227, 1453-1474, 1997; ftp://ftp.genetics.wisc.edu/pub/sequence/ecolim52.seq.gz), there are many ORFs, the function of which still remains unknown.

Disclosure of the invention

35 **[0005]** An object of present invention is to enhance the productivity of L-amino acid producing strains and to provide a method for producing L-amino acid, for example, L-threonine, L-valine, L-proline, L-leucine or L-methionine or L-arginine, using the strains.

40 **[0006]** This aim was achieved by identifying genes coding for proteins, which are not involved into biosynthetic pathway of target L-amino acid but enhance its production. An example of such protein could be a membrane protein having L-amino acid excretion activity. Based on the analysis of complete genome sequence of *Escherichia coli*, proteins with 4 or more putative transmembrane segments (TMS) were selected. As a result of diligent screening, the present inventors have identified several genes among them, that is, b2682, b2683, b1242 and b3434, and thoroughly studied it. The genes b2682 and b2683 have been known as putative CDS which may encode functionally unknown proteins (nucleotide numbers 92 to 829 and 819 to 1154 in the sequence of GenBank accession AE000353 U00096, respectively). The gene
45 b2683 is also known as *ygaH*. The gene b1242 has been known as putative CDS which may encode functionally unknown protein (numbers 8432 to 9079 in the sequence of GenBank accession AE000222 U00096). The gene b1242 is also known as *ychE*. The gene b3434 also has been known as putative CDS which may encode functionally unknown protein (numbers 1463 to 2056 in the sequence of GenBank accession AE000420 U00096). The gene b3434 is also known as
50 *yhgN*.

[0007] The present inventors have found that by enhancing the activity of the protein encoded by b1242 gene the productivity of L-amino acid producing strain is enhanced. Thus the present invention has been completed.

[0008] The present inventions are as follows:

55 1) An L-amino acid producing bacterium belonging to the genus *Escherichia*, wherein the bacterium has been modified so that the L-amino acid production by the bacterium should be enhanced by enhancing activities of proteins as defined in the following (E) or (F) in a cell of the bacterium:

(E) a protein which comprises the amino acid sequence shown in SEQ ID NO:11 in Sequence listing;
 (F) a protein which comprises an amino acid sequence including deletion, substitution, insertion or addition of 1 to 22 amino acids in the amino acid sequence shown in SEQ ID NO:11 in Sequence listing, and which has an activity of making bacterium having enhanced resistance to the L-amino acids and/or its analogs;
 (hereinafter, the proteins as defined in the above (E) or (F) are sometimes referred to as "proteins of the present invention" and the bacterium belonging to the genus *Escherichia* which is enhanced the activities of the proteins (E) or (F) is sometimes referred to as "a bacterium of the present invention");

wherein the activities of the proteins as defined in (E) or (F) are enhanced by transformation of the bacterium with a DNA coding for the proteins as defined in (E) or (F), or by alteration of promoter sequence of the DNA on the chromosome of the bacterium.

2) The bacterium according to the above bacterium, wherein the transformation is performed with a multicopy vector.

3) A method for producing L-amino acid, which comprises cultivating the bacterium according to the above bacterium in a culture medium and collecting from the culture medium L-amino acid to be produced and accumulated.

4) The method according to the above method, wherein L-amino acid is L-threonine.

5) The method according to the above method, wherein the bacterium has been modified so that the bacterium should have enhanced expression of threonine operon.

6) The method according to the above method, wherein L-amino acid is L-valine.

7) The method according to the above method,

wherein the bacterium has been modified so that the bacterium should have enhanced expression of *ilv* operon.

[0009] The method for producing L-amino acid includes production of L-threonine using L-threonine producing bacterium wherein activities of the proteins of the present invention such as that comprising amino acid sequence shown in SEQ ID NO:11 are enhanced. Also a method for producing L-amino acid includes production of

L-valine using L-valine producing bacterium wherein activities of the proteins of the present invention such as that comprising amino acid sequence shown in SEQ ID NO:11 are enhanced.

[0010] The present invention will be explained in detail below.

[0011] The bacterium of the present invention is an L-amino acid producing bacterium belonging to the genus *Escherichia*, wherein the bacterium has been modified so that the L-amino acid production by the bacterium should be enhanced by enhancing activities of the proteins of the present invention in a cell of the bacterium.

[0012] In the present invention, "L-amino acid producing bacterium" means a bacterium which has an ability to accumulate L-amino acid in a medium, when the bacterium is cultured in the medium. The L-amino acid producing ability may be possessed by the bacterium as a property of a wild strain of the bacterium or may be imparted or enhanced by breeding.

[0013] The bacterium of the present invention is L-amino acid producing bacterium belonging to the genus *Escherichia* having enhanced activities of proteins, which enhance the productivity of the target L-amino acid. Concretely the bacterium of present invention is L-amino acid producing bacterium belonging to the genus *Escherichia* which has enhanced activity of at least one or two of the proteins of the present invention.

[0014] The term "enhancing an activity of a protein " means that the activity per cell has become higher than that of a non-modified strain, for example, a wild-type bacterium belonging to the genus *Escherichia*. For example, there can be mentioned a case where number of the protein molecules per cell increases, a case where specific activity per the protein molecule increases and so forth. Further, as a wild-type bacterium belonging to the genus *Escherichia* that serves as an object for comparison, for example, the wild type strain of *Escherichia coli* can be mentioned.

[0015] The proteins of the present invention include ones as defined in the following (E) or (F):

(E) a protein which comprises the amino acid sequence shown in SEQ ID NO:11 in Sequence listing;

(F) a protein which comprises an amino acid sequence including deletion, substitution, insertion or addition of 1 to 22 amino acids in the amino acid sequence shown in SEQ ID NO:11 in Sequence listing, and which has an activity of making bacterium having enhanced resistance to the L-amino acids and/or its analogs;

[0016] Enhanced resistance to L-amino acids and/or its analogs means ability for bacterium to grow on a minimal medium containing L-amino acid or its analog in concentration under which the unmodified strain or the wild type strain, or the parental strain of the bacterium cannot grow, or ability for bacterium to grow faster on a medium containing L-amino acid or its analog than the unmodified strain or the wild type strain, or the parental strain of the bacterium.

[0017] More concretely, it can be said that *E. coli* strain has enhanced resistance to the L-amino acid or its analog if the strain forms a colony which is larger than that of the unmodified strain or wild type strain of *E. coli* after 2 - 4 days incubation at 37°C on a plate with solid Adams medium at 37°C when the strain is cultivated on an agar medium containing the L-amino acid or its analog under an appropriate condition. The term "an appropriate condition" refers to temperature,

pH, air supply or optional presence of essential nutrients or the like for the *E. coli* strain which is to be cultivated.

[0018] L-amino acid analogs are exemplified by 3,4-dihydroproline, DL-thiaiso-leucine, DL-o-methylserine, 4-azaleucine, norleucine, L-o-fluorophenylalanine and DL-o-fluorophenylalanine, homoserine, 6-diazo-5-oxo-L-norleucine and DL- β -hydroxy-norvaline.

[0019] Above mentioned concentration of L-amino acid or its analog, under which the unmodified strain or the wild type strain of the bacterium cannot grow, varies very significantly (from 0.5 μ g/ml for DL-thiaiso-leucine to 9600 μ g/ml for DL-o-methylserine) depending on the structure of used compound. For example, such concentration is generally 7 to 70 μ g/ml, preferably 20 to 25 μ g/ml in case of 3,4-dihydroproline; generally 0.5 to 5 μ g/ml, preferably 0.9 to 1.1 in case of DL-thiaiso-leucine; generally 1100 to 9600 μ g/ml, preferably 3000 to 3500 in case of DL-o-methylserine; generally 15 to 150 μ g/ml, preferably 40 to 50 μ g/ml in case of 4-azaleucine; generally 150 to 1500 μ g/ml, preferably 450 to 550 μ g/ml in case of norleucine; generally 0.6 to 6 μ g/ml, preferably 1.5 to 2 μ g/ml in case of L-o-fluorophenylalanine; generally 2 to 20 μ g/ml, preferably 5 to 7 μ g/ml in case of DL-o-fluorophenylalanine; and generally 330 to 3300 μ g/ml, preferably 900 to 1100 μ g/ml in case of homoserine, generally 5 to 50 μ g/ml, preferably 12 to 18 in case of 6-diazo-5-oxo-L-norleucine, and generally 25 to 250 μ g/ml, preferably 70 to 90 μ g/ml in case of DL- β -hydroxy-norvaline

[0020] In the bacterium of the present invention, the activities of the proteins of the present invention are enhanced by transformation of the bacterium with DNA coding for protein as defined in (E) or (F) or by alteration of promoter sequence of the DNA on the chromosome of the bacterium. The DNA, which is used for modification of the bacterium of the present invention, is represented by the b1242 gene. The b1242 gene can be obtained by, for example, PCR using primers having nucleotide sequence shown in SEQ ID No: 9 and 10.

[0021] Analysis of complete genome sequence of *Escherichia coli* allowed to select the genes coding for proteins having 4 or more putative TMS. Proteins with known function and transporters described by Paulsen I.T., Sliwinski M.I., Saier M.H. (J.Hol.Biol., 1998, 277, 573) and Linton K.J., Higgins C.F. (Molecular Microbiology, 1998, 28(1), 5) were excluded from the group to be screened. As a result of diligent screening among the rest of genes, several genes coding for putative membrane exporters were chosen. And it was found that the overexpression of b2682 and b2683 genes, or b1242 or b3434 gene enhances the L-amino acid production by L-amino acid producing strain.

[0022] The DNA coding for substantially the same protein as the protein defined in (E) may be obtained by, for example, modification of nucleotide sequence coding for the protein defined in (E) using site-directed mutagenesis so that one or more amino acid residue will be deleted, substituted, inserted or added. Such modified DNA can be obtained by conventional methods using treatment with reagents and conditions generating mutations. Such treatment includes treatment the DNA coding for proteins of present invention with hydroxylamine or treatment the bacterium harboring the DNA with UV irradiation or reagent such as N-methyl-N'-nitro-N-nitrosoguanidine or nitrous acid.

[0023] The DNA includes variants which can be found in the different strains and variants of bacteria belonging to the genus *Escherichia* according to natural diversity. The DNA coding for such variants can be obtained by isolating the DNA, which hybridizes with b1242 gene or part of the genes under the stringent conditions, and which codes the protein enhancing L-amino acid production. The term "stringent conditions" referred to herein is a condition under which so-called specific hybrid is formed, and non-specific hybrid is not formed. For example, the stringent conditions includes a condition under which DNAs having high homology, for instance DNAs having homology no less than 70% to each other, are hybridized. Alternatively, the stringent conditions are exemplified by conditions which comprise ordinary condition of washing in Southern hybridization, e.g., 60°C, 1 x SSC, 0.1% SDS, preferably 0.1 x SSC, 0.1% SDS. As a probe for the DNA which codes for variants and hybridizes with b1242 gene, a partial sequence of the nucleotide sequence of SEQ ID NO: 3 or SEQ ID NO: 5 respectively can also be used. Such a probe may be prepared by PCR using oligonucleotides produced based on the nucleotide sequence of SEQ ID NO: 11 as primers, and a DNA fragment containing the nucleotide sequence of SEQ ID NO: 11 as a template. When a DNA fragment in a length of about 300 bp is used as the probe, the conditions of washing for the hybridization consist of, for example, 50°C, 2 x SSC, and 0.1% SDS.

[0024] Transformation of bacterium with DNA coding for protein means introduction of the DNA into bacterium cell for example by conventional methods to increase expression of the gene coding for the protein of present invention and to enhance the activity of the protein in the bacterial cell.

[0025] Techniques for enhancement of gene expression includes methods increasing the gene copy number. Introduction of a gene into a vector that is able to function in a bacterium belonging to the genus *Escherichia* increases copy number of the gene. For such purposes multi-copy vectors can be preferably used. The multi-copy vector is exemplified by pBR322, pMW119, pUC19, pET22b or the like.

[0026] Besides, enhancement of gene expression can be achieved by introduction of multiple copies of the gene into bacterial chromosome by, for example, method of homologous recombination or the like.

[0027] In case that expression of two or more genes is enhanced, the genes may be harbored together on the same plasmid or separately on different plasmids. It is also acceptable that one of the genes is harbored on a chromosome, and the other gene is harbored on a plasmid.

[0028] On the other hand, enhancement of gene expression can be achieved by alteration of promoter sequence of the gene. Alteration of promoter sequence of a gene includes introducing mutation in the inherent promoter sequence

of the gene so that the expression of the gene is enhanced (WO00/18935) and locating the DNA of the present invention under control of a potent promoter. For example, *lac* promoter, *trp* promoter, *trc* promoter, P_L promoter of lambda phage are known as potent promoters. Using the potent promoter can be combined with multiplication of gene copies.

[0029] The bacterium of the present invention can be obtained by introduction of the aforementioned DNAs into bacterium inherently having ability to produce L-amino acid. Alternatively, the bacterium of present invention can be obtained by imparting ability to produce L-amino acid to the bacterium already harboring the DNAs. For the parent strain which is to be enhanced in activities of the proteins of the present invention, L-threonine producing bacteria belonging to the genus *Escherichia* such as strains VL2054 (VKPM B-8067), VNIIGenetika 472T23 (US patent No.5,631,157), VKPM B-3996 (US patent Nos. 5,175,107 and 5,976,843), KCCM-10132 (WO009660A1), KCCM-10133 (WO009661A1) or the like can be employed. Also for the parent strain which is to be enhanced in activities of the proteins of the present invention, L-valine producing bacteria belonging to the genus *Escherichia* such as H-81 (VKPM B- 8066), NRRL B-12287 and NRRL B-12288 (US patent No. 4,391,907), VKPM B-4411 (US patent No. 5,658,766), VKPM B-7707 (European patent application publication EP1016710A2) or the like is employed. Besides, for the parent strain which is to be enhanced in activities of the proteins of the present invention, L-proline producing bacteria belonging to the genus *Escherichia* such as NRRL B-12403 and NRRL B-12404 (GB2075056), VKPM B-8012 (Russian patent publication 2207371), plasmid mutants described in the patent DE3127361, plasmid mutants described by Bloom F.R. *et al.* (The 15th Miami winter symposium, 1983, p.34) or the like are employed. Also, for the parent strain which is to be enhanced in activities of the proteins of the present invention, L-leucine producing bacteria belonging to the genus *Escherichia* such as H-9070 (FERM BP-4704) and H-9072 (FERM BP-4706) (US5744331), VKPM B-7386 and VKPM B-7388 (RU2140450), W1485atpA401/pMWdAR6, W14851ip2/pMWdAR6 and AJ12631/pMWdAR6 (EP0872547) or the like are employed. And, for the parent strain which is to be enhanced in activities of the proteins of the present invention, L-methionine producing bacteria belonging to the genus *Escherichia* such as AJ11539 (NRRL B-12399), AJ11540 (NRRL B-12400), AJ11541 (NRRL B-12401), AJ 11542 (NRRL B-12402) (GB2075055) or the like are employed as well.

[0030] Further, for the parent strain which is to be enhanced in activity of the proteins of the present invention, L-arginine producing bacteria belonging to the genus *Escherichia* such as strains AJ11531 and AJ11538 (JP56106598A2), AJ11593 (FERM P-5616) and AJ11594 (FERM P-5617) (Japanese Patent Laid-open No. 57-5693) or the like can be employed.

[0031] The bacterium of the present invention may be further enhanced expression of one or more genes which are involved in L-amino acid biosynthesis. Such genes are exemplified by threonine operon, which preferably comprises a gene encoding aspartate kinase - homoserine dehydrogenase of which feedback inhibition by L-threonine is desensitized (Japanese Patent Publication No. 1-29559), for L-threonine producing bacteria. Such genes are exemplified by *ilv* operon, i.e. *ilvGMEDA* operon, which does not preferably express threonine deaminase and of which attenuation is suppressed (Japanese Patent Laid-Open Publication No. 8-47397), for L-valine producing bacteria. Such genes are exemplified by genes for L-proline biosynthesis, which are preferably represented by gene *proB* encoding for glutamate kinase of which feedback inhibition by L-proline is desensitized (DE3127361), for L-proline producing bacteria. Also, such genes are exemplified by leucine operon, i.e. *leu* operon, which preferably comprises a gene coding for isopropylmalate synthase of which feedback inhibition by L-leucine is desensitized (Russian patent publication 2201454), for L-leucine producing bacteria. Also, such genes are exemplified by methionine regulon, for L-methionine producing bacteria. The methionine regulon may have mutated genes coding for proteins lowered in activity in repressing the amino acid biosynthesis. Such gene is exemplified by variation type *metJ* gene coding for a L-methionine biosynthesis-relating repressor protein from *E. coli* of which activity in repressing methionine biosynthesis is lowered (JP 2000-157267 A2). Further, such gene is exemplified by arginine regulon, which preferably comprises a gene encoding N-acetylglutamate synthase of which feedback inhibition by L-arginine is desensitized (Rajagopal B.S. *et al.*, Appl. Environ. Microbiol., 1998, v.64, No.5, p. 1805-1811).

[0032] The method of the present invention includes method for producing L-threonine, comprising steps of cultivating the bacterium of the present invention in a culture medium, to allow L-threonine to be produced and accumulated in the culture medium, and collecting L-threonine from the culture medium. Also the method of present invention includes method for producing L-valine, comprising steps of cultivating the bacterium of the present invention in a culture medium, to allow L-valine to be produced and accumulated in the culture medium, and collecting L-valine from the culture medium.

[0033] In the present invention, the cultivation, the collection and purification of L-amino acid from the medium and the like may be performed in a manner similar to the conventional fermentation method wherein an amino acid is produced using a microorganism. A medium used for culture may be either a synthetic medium or a natural medium, so long as the medium includes a carbon source and a nitrogen source and minerals and, if necessary, appropriate amounts of nutrients which the microorganism requires for growth. The carbon source may include various carbohydrates such as glucose and sucrose, and various organic acids. Depending on the mode of assimilation of the used microorganism, alcohol including ethanol and glycerol may be used. As the nitrogen source, various ammonium salts such as ammonia and ammonium sulfate, other nitrogen compounds such as amines, a natural nitrogen source such as peptone, soybean-hydrolysate and digested fermentative microorganism are used. As minerals, potassium monophosphate, magnesium

sulfate, sodium chloride, ferrous sulfate, manganese sulfate, calcium chloride, and the like are used.

[0034] The cultivation is performed preferably under aerobic conditions such as a shaking culture, and stirring culture with aeration, at a temperature of 20 to 40 °C, preferably 30 to 38 °C. The pH of the culture is usually between 5 and 9, preferably between 6.5 and 7.2. The pH of the culture can be adjusted with ammonia, calcium carbonate, various acids, various bases, and buffers. Usually, a 1 to 5-day cultivation leads to the accumulation of the target L-amino acid in the liquid medium.

[0035] After cultivation, solids such as cells can be removed from the liquid medium by centrifugation or membrane filtration, and then the target L-amino acid can be collected and purified by ion-exchange, concentration and crystallization methods.

Brief description of the drawing

[0036] Figure 1 shows the construction of plasmid pΔlacZ.

Best Mode for Carrying out the Invention

[0037] The present invention will be more concretely explained below with reference to Examples. In the Examples an amino acid is of L-configuration unless otherwise noted.

Example 1: Cloning of the b2682, b2683, b1242, and b3434 genes on the plasmid pΔlacZ

[0038] For cloning of the b2682 and b2683 genes vector pΔlacZ was used. Vector pΔlacZ is a derivative of the vector pET-22b(+) (Novagen, Madison, WI, USA). pET-22b(+) was treated by *Bgl*II and *Xba*I and ligated with polymerase chain reaction (PCR) fragment of plasmid pMB9-*lac* (Fuller F., Gene, 19, 43-54, 1982) treated with the same restrictases and carried P_{lac} UV5 promoter. For amplifying the P_{lac} UV5 promoter fragment by PCR primers having sequence depicted in SEQ ID Nos: 7 and 8 were used. The resulted plasmid was supplemented with structural part of *lacZ* gene (237 bp without promoter) by cloning *Sal*I-*Bam*HI fragment of the plasmid pJEL250 (Dymakova E. et al., Genetika (rus), 35, 2, 181-186, 1999). Scheme for obtaining vector pΔlacZ is shown in Figure 1.

[0039] The initial material for cloning of *E. coli* b2682 and b2683 putative reading frames (b2682 and b2683 genes) was the PCR fragment, which was obtained using DNA from *E. coli* strain TG1 as a template. For synthesis of this fragment two primers having sequence depicted in SEQ ID Nos: 1 and 2 were used. PCR was carried out on "Perkin Elmer GeneAmp PCR System 2400" under the following conditions: 40 sec. at 95 °C, 40 sec. at 47 °C, 40 sec. at 72 °C, 30 cycles. Thus, the 1158 bp linear DNA fragment contained b2682 and b2683 genes was obtained. This PCR fragment was treated by *Xba*I and *Bam*HI restrictases and inserted into multicopy vector pΔlacZ previously treated by the same restrictases.

[0040] Resulted plasmid with the PCR fragment was named pYGAZH and carried both gene b2682 and b2683 under the control of the lactose promoter (P_{lac} UV5).

[0041] Similarly, the initial material for cloning of *E. coli* b1242 putative reading frame (b1242 gene) was the PCR fragment, which was obtained using DNA from *E. coli* strain TG1 as a template. For synthesis of this fragment two primers having sequence depicted in SEQ ID Nos: 9 and 10 were used. Resulted plasmid with the PCR fragment was named pYCHE and carried b1242 gene under the control of the lactose promoter (P_{lac} UV5). The initial material for cloning of *E. coli* b3434 putative reading frame (b3434 gene) was the PCR fragment, which was obtained using DNA from *E. coli* strain TG1 as a template. For synthesis of this fragment two primers having sequence depicted in SEQ ID Nos: 13 and 14 were used. Resulted plasmid with the PCR fragment was named pYHGN and carried b3434 gene under the control of the lactose promoter (P_{lac} UV5).

Example 2: The influence of the amplified b2682 and b2683 genes on resistance of *E. coli* strain TG1 to amino acids and its analogs

[0042] *E. coli* strain TG1(pYGAZH), TG1(pYCHE), TG1(pYHGN) and TG1 strain having a vector without insertion (control strain) were grown overnight on LB medium supplemented with ampicilline (100 μg/ml). The night cultures of all strains were diluted at 25 times in fresh LB medium supplemented with ampicilline (100 μg/ml) and IPTG (0.5 mM) and were incubated 2 hours at 37 °C with aeration. The log phase cultures were diluted in 0,9% solution of NaCl and about 1000 cells were seeded on plates with solid Adams medium supplemented with ampicilline (100 μg/ml), IPTG (0.5 mM) and amino acid or its analog. After 2 - 4 days incubation at 37 °C the differences in colony size or colony number between the TG1 strain with hybrid plasmid and control TG1 strain were registered. The results of experiments are presented in Table 1.

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Table 1

	Inhibitors	Concentration in media, µg/ml	Effect on the growth of TG1 strain having plasmid		
			pYGAZH	pYCHE	pYHGN
5	Proline	30000	No	No	No
	3,4-Dihydroproline	23	R	No	No
10	Isoleucine	18000	No	No	No
	DL-Thiaisoleucine	1	R	No	No
	o-Methylthreonine	6	No	No	No
	L-Serine	2800	No	No	No
15	DL-Serine	3600	No	No	No
	DL-Serine hydroxamate	140	No	No	No
	DL-o-Methylserine	3200	R	R	R
20	4-Azaleucine	45	R	No	No
	6-Diazo-5-oxo-L-norleucine	15	No	No	R
	Valine	7	R	No	No
	Methionine	38000	No	No	No
25	Norleucine	500	R	No	No
	Cysteine	1600	No	No	No
	Homoserine	1000	No	R	No
30	DL-β-Hydroxy-norvaline	80	No	No	R
	L-Aspartic acid β-hydroxamate	100	No	No	No
	Arginine	4300	No	No	No
	Lysine	5000	No	No	No
35	S-(2-Aminoethyl)cysteine	0.75	No	No	S
	Histidine	3000	No	No	No
	L-Histidine hydroxamate	200	No	No	No
40	DL-1,2,4-Triazole-3-alanine	80	No	No	No
	Phenylalanine	13000	No	No	No
	p-Fluorophenylalanine	6	No	No	No
	L-o-Fluorophenylalanine	1.7	R	No	No
45	DL-o-Fluorophenylalanine	6	R	No	No
	Tryptophan	12500	No	No	No
	DL-4-Fluorotryptophan	0.1	No	No	No
50	4-Methyltryptophan	0.25	No	No	No
	7-Methyltryptophan	100	No	No	No
	DL-a-Methyltryptophan	400	No	No	No

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(continued)

Inhibitors	Concentration in media, $\mu\text{g/ml}$	Effect on the growth of TG1 strain having plasmid		
		pYGAZH	pYCHE	pYHGN
m-Fluoro-DL-tyrosine	0.5	No	No	No
No - no differences compare to the control strain R - more colonies or colony size S - less colonies or colony size compare to the control strain				

Example 3: Production of threonine by a strain having plasmid pYGAZH (not belonging to the scope of the present invention)

[0043] The threonine producing strain VL2054 was transformed by the plasmid pYGAZH carried the b2682 and b2683 genes under the control of P_{lac} UV5 promoter. Obtained strain was named VL2054(pYGAZH). The strain VL2054 is derivative of the strain VKPM B-3996 and carried on its chromosome:

- a) the integrated threonine operon under the control of P_R promoter
- b) wild type *rhtA* gene
- c) the inactivated chromosomal gene encoding transhydrogenase (*tdh* gene) and inactivated kanamycin resistant gene (*kan*) gene in the Tn5 (*tdh::Tn5*, *Kan^S*)
- d) mutation *ilvA₄₄₂*

[0044] The strain VL2054 has been deposited in the Russian National Collection of Industrial Microorganisms (VKPM) (Russia 113545, Moscow, 1 Dorozhny proezd, 1) on January 30, 2001 under accession number VKPM B-8067, and transferred from the original deposit to international deposit based on Budapest Treaty on February 1, 2002.

[0045] The 5 colonies of each strain VL2054, strain VL2054($\Delta lacZ$) as a control strain contained plasmid without insertion and VL2054(pYGAZH) were suspended in 2 ml of minimal medium ($(\text{NH}_4)_2\text{SO}_4$ - 11 g/l; NaCl - 0.4 g/l; MgSO_4 - 0.4 g/l; K_2HPO_4 - 1 g/l; FeSO_4 - 10 mg/l; MnSO_4 - 10 mg/l; thiamin - 0.1 mg/l; yeast extract - 0.5 g/l; glucose - 40 g/l; ampicilline - 300 mg/l if necessary) in 20-ml test tubes and were incubated overnight with aeration at 32 °C. The 0.2 ml of each night culture was transferred to the three 20-ml test tubes with 2 ml of fresh medium for fermentation with or without IPTG and cultivated at 32 °C for 48 or 72 hours with rotary shaker.

Fermentation medium composition:

$(\text{NH}_4)_2\text{SO}_4$	22 g/l
NaCl	0.8 g/l
MgSO_4	0.8 g/l
K_2HPO_4	2 g/l
FeSO_4	20 mg/l
MnSO_4	20 mg/l
Thiamin	0.2 mg/l
Yeast extract	1 g/l
CaCO_3	30 g/l
Glucose	80 g/l
Ampicilline	300 mg/l, if necessary
IPTG	0.5 mM, if necessary

[0046] After cultivation the plasmid stability and optical absorbance of the medium at 540 nm were determined by conventional methods. Accumulated amount of threonine in the medium was determined by thin layer chromatography (TLC). Liquid phase composition for TLC was as follows: isopropanol - 50 ml, acetone - 50 ml, NH_4OH (30 %) - 12 ml, H_2O - 8 ml. The results are shown in Table 2. As it is seen, the hybrid plasmid pYGAZH improved the threonine accumulation by the threonine producing strain VL2054.

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Table 2

VL2054 with plasmid	IPTG	48 hours			72 hours		
		OD ₅₄₀	Thr, g/l	Thr/OD	OD ₅₄₀	Thr, g/l	Thr/OD
no	-	19	5.2	0.27	26	9.1	0.35
	+	21	4.1	0.20	29	7.8	0.27
pΔlacZ	-	20	6.4	0.32	24	9.1	0.40
	+	15	3.5	0.23	24	7.2	0.30
pYGAZH	-	17	5.7	0.34	24	9.7	0.40
	+	21	9.8	0.47	23	15.5	0.67

Example 4: Production of valine by a strain having plasmid pYGAZH (not belonging to the scope of the present invention)

[0047] The valine producing strain H-81 was transformed by the plasmid pYGAZH carried the b2682 and b2683 genes under the control of P_{lac} UV5 promoter. The strain H-81 has been deposited in the Russian National Collection of Industrial Microorganisms (VKPM) (Russia 113545, Moscow, 1 Dorozhny proezd, 1) on January 30, 2001 under accession number VKPM B-8066, and transferred from the original deposit to international deposit based on Budapest Treaty on February 1, 2002.

[0048] The 5 colonies of each strain H-81, H-81(pΔlacZ) as a control strain contained plasmid without insertion and H-81(pYGAZH) were suspended in 2 ml of minimal medium ((NH₄)₂SO₄ - 18 g/l, K₂HPO₄ - 1.8 g/l, MgSO₄ - 1.2 g/l, thiamin - 0.1 mg/l, yeast extract - 0.5 g/l, glucose - 60 g/l, ampicilline - 300 mg/l, if necessary) in 20-ml test tubes and were incubated overnight with aeration at 32 °C. The 0.2 ml of each night culture was transferred to the three 20-ml test tubes with 2 ml of fresh medium for fermentation with or without IPTG and cultivated at 32 °C for 48 or 72 hours with rotary shaker.

Fermentation medium composition:

(NH ₄) ₂ SO ₄	18 g/l,
K ₂ HPO ₄	1.8 g/l,
MgSO ₄	1.2 g/l,
CaCO ₃	20 g/l,
Thiamin	0.1 mg/l,
Glucose	60 g/l,
Ampicilline	300 mg/l, if necessary
IPTG	0.5 mM, if necessary

[0049] After cultivation the plasmid stability and optical absorbance of the medium at 540 nm were determined by conventional methods. Accumulated amount of valine in the medium was determined by TLC. Liquid phase composition for TLC was as follows: isopropanol - 80 ml, ethylacetate - 80 ml, NH₄OH (30 %) - 15 ml, H₂O - 45 ml. The results are shown in Table 3. As it is seen, the hybrid plasmid pYGAZH improved the valine accumulation by the valine producing strain H-81.

Table 3

H-81 with plasmid	IPTG	48 hours			72 hours		
		OD ₅₄₀	Val, g/l	Val/O D	OD ₅₄₀	Val, g/l	Val/O D
No	-	34	11,6	0,34	32	10,3	0,32
	+	34	11,7	0,34	30	10,1	0,34
pΔlacZ	-	34	10,5	0,31	30	10,0	0,33
	+	20	7,8	0,39	25	9,0	0,36
pYGAZH	-	29	10,5	0,36	31	12,8	0,41
	+	22	10,8	0,49	23	12,3	0,53

Reference Example 1 : Production of L-proline by an *ilvA* deficient L-proline producer

[0050] The cells of wild type strain *E. coli* K12 (VKPM B-7) was treated with a mutagen, N-methyl-N'-nitro-N-nitrosoguanidine (0.1 mg/ml), for 20 min at 37°C, washed and plated on minimal agar medium M9 supplemented with 1.25 mg/ml tryptone, 10 mg/ml L-proline and 0.05 mg/ml 2,3,5-triphenyltetrazolium chloride. Most colonies arisen after 3 day of incubation at 37°C were colored red. A few colonies, which could not oxidize L-proline, were white. One of such colonies was used as a parent for obtaining mutants resistant to proline analogs (3,4-dehydroxyproline and azetidine-2-carboxylate) which were added into M9 agar medium in concentration of 2 mg/ml each.

[0051] Some of mutants arisen could produce L-proline. The best L-proline producer 702 was treated with a P1 bacteriophage grown on cells of the strain TG1 in which the gene *ilvA* was disrupted by the insertion of chloramphenicol (Cm) resistance (Cm^r) gene. One of obtained Cm resistant transductant, 702ilvA, which turned to be L-isoleucine auxotroph, was much more effective L-proline producer than the L-isoleucine prototrophic parent strain 702 (Table 4). The fermentation medium contained 60 g/l glucose, 25 g/l ammonium sulfate, 2 g/l KH₂PO₄, 1 g/l MgSO₄, 0.1 mg/l thiamine, 50 mg/l L-isoleucine and 25 g/l chalk (pH 7.2). Glucose and chalk were sterilized separately. 2 ml of the medium was placed into test tubes, and inoculated with one loop of the tested microorganisms, and the cultivation was carried out at 37°C for 2 days with shaking.

Table 4

Strain	Phenotype	Accumulation of L-proline (g/l)
K12 (VKPM B-7)	Wild type	<0.1
702 (VKPM B-8011)	Defective L-proline degradation, resistance to proline analogs	0.5
702ilvA (VKPM B-8012)	Defective L-proline degradation, resistance to proline analogs, L-isoleucine auxotroph, Cm ^r	8.0

[0052] The strains 702 and 702ilvA have been deposited in the Russian National Collection of Industrial Microorganisms (VKPM) under the accession number VKPM B-8011 and VKPM B-8012, respectively, since July 25, 2000.

Example 5: Production of proline by a strain having plasmid pYGZAH (not belonging to the scope of the present invention)

[0053] The proline producing strain *E. coli* 702ilvA was transformed by the plasmid pYGZAH carried the b2682 and b2683 genes under the control of P_{lac} UV5 promoter.

[0054] The 5 colonies of each strain 702ilvA, 702ilvA(pΔlacZ) as a control strain contained plasmid without insertion and 702ilvA(pYGZAH) were suspended in 2 ml of minimal medium ((NH₄)₂SO₄ - 18 g/l, K₂HPO₄ - 1.8 g/l, MgSO₄ - 1.2 g/l, thiamin - 0.1 mg/l, yeast extract - 0.5 g/l, glucose - 60 g/l, isoleucine - 50 mg/l, ampicilline - 300 mg/l, if necessary) in 20-ml test tubes and were incubated overnight with aeration at 32 °C. The 0.2 ml of each night culture was transferred to the three 20-ml test tubes with 2 ml of fresh medium for fermentation with or without IPTG and cultivated at 32 °C for 40 hours with rotary shaker.

Fermentation medium composition:

(NH ₄) ₂ SO ₄	18 g/l,
K ₂ HPO ₄	1.8 g/l,
MgSO ₄	1.2 g/l,
CaCO ₃	20 g/l,
Thiamin	0.1 mg/l,
Glucose	60 g/l,
Isoleucine	50 mg/l
Ampicilline	300 mg/l, if necessary
IPTG	0.5 mM, if necessary

[0055] After cultivation the plasmid stability and optical absorbance of the medium at 540 nm were determined by conventional methods. Accumulated amount of proline in the medium was determined by TLC. Liquid phase composition for TLC was as follows: ethanol - 80 ml, NH₄OH (30 %) - 5 ml, H₂O - 25 ml. The results are shown in Table 5. As it is seen, the hybrid plasmid pYGZAH improved the proline accumulation by the proline producing strain 702ilvA.

Table 5

702ilvA with plasmid	IPTG	40 hours		
		OD ₅₄₀	Pro, g/l	Pro/O D
No	-	25	4,0	0,16
	+	23	4,1	0,18
pΔlacZ	-	24	5,3	0,22
	+	22	5,0	0,23
pYGAZH	-	21	5,0	0,24
	+	23	10,6	0,46

15 Reference Example 2: Production of L-leucine by an *ilvE* deficient L-leucine producer

[0056] The cells of wild type strain *E. coli* K12 (VKPM B-7) was treated with a mutagen, N-methyl-N'-nitro-N-nitrosoguanidine (0.05 mg/ml), for 20 min at 37°C, washed 4 times with physiological solution and plated on minimal agar medium M9 supplemented with 4.0 mg/ml DL-4-azaleucine. The plates were incubated for 5 days at 37°C. Colonies
 20 appeared on the plates were picked up and purified by streaking on the L-agar plates. One of the obtained mutant resistant to DL-4-azaleucine was used for induction of double L-isoleucine and L-valine auxotrophy. The numerous amount of double auxotrophs, requiring L-isoleucine and L-valine for growth, were obtained. It was shown that double L-isoleucine and L-valine auxotrophy was caused by mutation in the *ilvE* gene. Among the obtained double auxotrophs, the best L-leucine producer, strain 505 producing 1.8 g/l of L-leucine, has been selected. The fermentation medium
 25 contained 60 g/l glucose, 25 g/l ammonium sulfate, 2 g/l KH₂PO₄, 1 g/l MgSO₄, 0.1 mg/l thiamine, 100 mg/l L-isoleucine, 100 mg/l L-valine and 25 g/l chalk (pH 7.2). Glucose and chalk were sterilized separately. 2 ml of the medium was placed into test tubes, and inoculated with one loop of the tested microorganisms, and the cultivation was carried out at 37°C for 2 days with shaking.

[0057] The strain *E. coli* 505 has been deposited in the Russian National Collection of Industrial Microorganisms (VKPM) (Russia 113545, Moscow, 1 Dorozhny proezd, 1) on May 14, 2001 under accession number VKPM B- 8124, and transferred from the original deposit to international deposit based on Budapest Treaty on February 1, 2002.

Example 6: Production of leucine by a strain having plasmid pYGAZH (not belonging to the scope of the present invention)

35 [0058] The leucine producing strain *E. coli* 505 was transformed by the plasmid pYGAZH carried the b2682 and b2683 genes under the control of P_{lac} UV5 promoter.

[0059] The 20 colonies of each strain 505, 505(pΔlacZ) as a control strain contained plasmid without insertion and 505(pYGAZH) were transferred by one loop of culture in 20-ml test tubes with L-broth with or without ampicilline and were incubated overnight with aeration at 32 °C. The 0.1 ml of each night culture was transferred into the 20-ml test
 40 tubes (inner diameter 22 mm), suspended in 2 ml of medium for fermentation with or without IPTG and cultivated at 32 °C for 72 hours with rotary shaker.

Fermentation medium composition:

(NH ₄) ₂ SO ₄	15 g/l,
K ₂ HPO ₄	1.5 g/l,
MgSO ₄ x 7H ₂ O	1.0 g/l,
CaCO ₃	20 g/l (sterilized separately),
Thiamin	0.1 mg/l,
Glucose	60 g/l (sterilized separately),
Isoleucine	0.3 g/l
Valine	0.3 g/l
Ampicilline	150 mg/l, if necessary
IPTG	0.5 mM, if necessary

55 [0060] After cultivation the plasmid stability was determined by conventional method. Accumulated amount of leucine in the medium was determined by TLC. Liquid phase composition for TLC was as follows: isopropanol - 80 ml, ethylacetate

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- 80 ml, NH₄OH (30 %) - 25 ml, H₂O - 50 ml. The results are shown in Table 6. As it is seen, the hybrid plasmid pYGAZH improved the leucine accumulation by the leucine producing strain 505.

Table 6

505 with plasmid	IPTG	72 hours
		Leu, g/l
No	-	1,8
	+	2,0
pΔlacZ	-	1,8
	+	2,0
pYGAZH	-	2,0
	+	2,8

Reference Example 3: Production of L-methionine by L-methionine producer resistant to norleucine

[0061] The plasmidless threonine and leucine deficient strain *E. coli* C600 was used as a parental strain. At first, the Leu⁺ variants of *E. coli* C600 strain was obtained by transduction of phage P1 grown on *E. coli* K-12 strain. Then, after treatment with N-methyl-N'-nitro-N-nitrosoguanidine (NTG) the mutant strain 44 resistant to 8 g/l of L-homoserine has been obtained. The strain 44 is L-threonine-deficient, resistant to high concentrations of L-homoserine. The strain 44 has been deposited in Russian National Collection of Industrial Microorganisms (VKPM) under the accession number VKPM B-2175.

[0062] Then, the strains, which are the mutants resistant to a methionine analog, norleucine, was induced from strain 44 by mutagenesis using NTG. The cells of night culture grown in L-broth were spun down and resuspended in physiological solution (0.9% NaCl) containing 50 μg/ml of NTG. After 30 min of exposure with NTG at 37° C the cells were spun down, washed 4 times with physiological solution and plated on the minimal agar medium M9, containing 0.5 mg/ml of threonine and 2.5 mg/ml or 5.0 mg/ml of norleucine. The plates were incubated for 5 days at 37°C. Colonies appeared on the plates were picked up and purified by streaking on the L-agar plates. The best L-methionine producer among them was strain 218. Test-tube cultivation of the novel strain 218 carried out at 32°C for 3 days with shaking leads to accumulation in the culture medium about 1 g/l of L-methionine. As a fermentation medium was used minimal medium M9 containing glucose (4%), ammonia sulfate (2.5%), threonine (0.5 g/l), calcium carbonate (25 g/l). Glucose and chalk were sterilized separately.

[0063] The strain 218 has been deposited in Russian National Collection of Industrial Microorganisms (VKPM) under the accession number VKPM B-8125 since May 14, 2001, and transferred from the original deposit to international deposit based on Budapest Treaty on February 1, 2002.

[0064] Further, the phage P1 mediated deletion of *ppc* gene has been introduced into strain 218 followed by integration of *pycA* gene from *Bacillus subtilis* (Russian patent publication 2207376). Resulted strain 218pycA lost resistance to norleucine. Therefore, resistance to norleucine has been imparted to the strain again as described above. The best L-methionine producer among obtained strains was strain *E. coli* 73 which produced about 1 g/l of L-methionine under condition described above.

[0065] The strain *E. coli* 73 has been deposited in the Russian National Collection of Industrial Microorganisms (VKPM) (Russia 113545 Moscow 1 Dorozhny proezd, 1) on May 14, 2001 under accession number VKPM B-8126, and transferred from the original deposit to international deposit based on Budapest Treaty on February 1, 2002.

Example 7: Production of methionine by a strain having plasmid pYGAZH (not belonging to the scope of the present invention)

[0066] The methionine producing strain *E. coli* 73 was transformed by the plasmid pYGAZH carried the b2682 and b2683 genes under the control of P_{lac} UV5 promoter.

[0067] The 5 colonies of each strain 73, 73(pΔlacZ) as a control strain contained plasmid without insertion and 73 (pYGAZH) were suspended in 2 ml of minimal medium ((NH₄)₂SO₄ - 18 g/l, K₂HPO₄ - 1.8 g/l, MgSO₄ - 1.2 g/l, thiamin - 0.1 mg/l, yeast extract - 10 g/l, glucose - 60 g/l, threonine - 400 mg/l, ampicilline - 300 mg/l, if necessary) in 20-ml test tubes and were incubated overnight with aeration at 32 °C. The 0.2 ml of each night culture was transferred to the three 20-ml test tubes with 2 ml of fresh medium for fermentation with or without IPTG and cultivated at 32 °C for 48 hours with rotary shaker.

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Fermentation medium composition:

$(\text{NH}_4)_2\text{SO}_4$	18 g/l,
K_2HPO_4	1.8 g/l,
MgSO_4	1.2 g/l,
CaCO_3	20 g/l,
Thiamin	0.1 mg/l,
Glucose	60 g/l,
Threonine	400 mg/l,
Yeast extract	1.0 g/l,
Ampicilline	300 mg/l, if necessary
IPTG	0.5 mM, if necessary

[0068] After cultivation the plasmid stability and optical absorbance of the medium at 540 nm were determined by conventional methods. Accumulated amount of methionine in the medium was determined by TLC. Liquid phase composition for TLC was as follows: isopropanol - 80 ml, ethylacetate - 80 ml, NH_4OH (30 %) - 15 ml, H_2O - 45 ml. The results are shown in Table 7. As it is seen, the hybrid plasmid pYGAZH improved the methionine accumulation by the methionine producing strain 73.

Table 7

73 with plasmid	IPTG	48 hours		
		OD ₅₄₀	Met, g/l	Met/O D
No	-	45	0,7	0,016
	+	42	1,1	0,026
pΔlacZ	-	45	1,0	0,022
pYGAZH	-	48	0,9	0,019
	+	46	1,3	0,028

Example 8: Production of threonine by a strain having plasmid pYCHE

[0069] The threonine producing strain VL2054 was transformed by the plasmid pYCHE carried the b1242 gene under the control of P_{lac} UV5 promoter. Obtained strain was named VL2054(pYCHE).

[0070] The 5 colonies of each strain VL2054, strain VL2054(pΔ1acZ) as a control strain contained plasmid without insertion and VL2054(pYCHE) were suspended in 2 ml of minimal medium ($(\text{NH}_4)_2\text{SO}_4$ - 11 g/l; NaCl - 0.4 g/l; MgSO_4 - 0.4 g/l; K_2HPO_4 - 1 g/l; FeSO_4 - 10 mg/l; MnSO_4 - 10 mg/l; thiamin - 0.1 mg/l; yeast extract - 0.5 g/l; glucose - 40 g/l; ampicilline - 300 mg/l if necessary) in 20-ml test tubes and were incubated overnight with aeration at 32 °C. The 0.2 ml of each night culture was transferred to the three 20-ml test tubes with 2 ml of fresh medium for fermentation with or without IPTG and cultivated at 32 °C for 45 hours with rotary shaker. Fermentation medium composition:

$(\text{NH}_4)_2\text{SO}_4$	22 g/l
NaCl	0.8 g/l
MgSO_4	0.8 g/l
K_2HPO_4	2 g/l
FeSO_4	20 mg/l
MnSO_4	20 mg/l
Thiamin	0.2 mg/l
Yeast extract	1 g/l
CaCO_3	30 g/l
Glucose	80 g/l
Ampicilline	300 mg/l, if necessary
IPTG	0.5 mM, if necessary

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[0071] After cultivation the plasmid stability and optical absorbance of the medium at 540 nm were determined by conventional methods. Accumulated amount of threonine in the medium was determined by thin layer chromatography (TLC). Liquid phase composition for TLC was as follows: isopropanol - 50 ml, acetone - 50 ml, NH₄OH (30 %) - 12 ml, H₂O - 8 ml. The results are shown in Table 8. As it is seen, the hybrid plasmid pYCHE improved the threonine accumulation by the threonine producing strain VL2054.

Table 8

VL2054 with plasmid	IPTG	OD ₅₄₀	Thr, g/l	Thr/OD
no	-	21	4.8	0.23
	+	20	4.7	0.24
pΔlacZ	-	16	4.6	0.29
	+	13	3.0	0.23
pYCHE	-	20	6.2	0.31
	+	20	7.0	0.35

Example 9: Production of valine by a strain having plasmid pYCHE

[0072] The valine producing strain H-81 was transformed by the plasmid pYCHE carried the b1242 gene under the control of P_{lac} UV5 promoter.

[0073] The 5 colonies of each strain H-81, H-81(pΔlacZ) as a control strain contained plasmid without insertion and H-81(pYCHE) were suspended in 2 ml of minimal medium ((NH₄)₂SO₄ - 18 g/l, K₂HPO₄ - 1.8 g/l, MgSO₄ - 1.2 g/l, thiamin - 0.1 mg/l, yeast extract - 0.5 g/l, glucose - 60 g/l, ampicilline - 300 mg/l, if necessary) in 20-ml test tubes and were incubated overnight with aeration at 32 °C. The 0.2 ml of each night culture was transferred to the three 20-ml test tubes with 2 ml of fresh medium for fermentation with or without IPTG and cultivated at 32 °C for 45 hours with rotary shaker.

Fermentation medium composition:

(NH ₄) ₂ SO ₄	18 g/l.
K ₂ HPO ₄	1.8 g/l.
MgSO ₄	1.2 g/l,
CaCO ₃	20 g/l,
Thiamin	0.1 mg/l,
Glucose	60 g/l,
Ampicilline	300 mg/l, if necessary
IPTG	0.5 mM, if necessary

[0074] After cultivation the plasmid stability and optical absorbance of the medium at 540 nm were determined by conventional methods. Accumulated amount of valine in the medium was determined by TLC. Liquid phase composition for TLC was as follows: isopropanol - 80 ml, ethylacetate - 80 ml, NH₄OH (30 %) - 15 ml, H₂O - 45 ml. The results are shown in Table 9. As it is seen, the hybrid plasmid pYCHE improved the valine accumulation by the valine producing strain H-81.

Table 9

H-81 with plasmid	IPTG	OD ₅₄₀	Val, g/l	Vai/OD
no	-	34	11.6	0.34
	+	34	11.7	0.34
pΔlacZ	-	34	10.5	0.31
	+	20	7.8	0.39
pYCHE	-	32	14.0	0.44
	+	30	13.9	0.46

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Example 10: Production of arginine by a strain having plasmid pYHGN (not belonging to the scope of the present invention)

[0075] The arginine producing strain 382 was transformed by the plasmid pYHGN carried the b3434 gene under the control of P_{lac} UV5 promoter. The strain 382 has been deposited in the Russian National Collection of Industrial Microorganisms (VKPM) (Russia 113545, Moscow, 1 Dorozhny proezd, 1) on April 10, 2000 under accession number VKPM B - 7926.

[0076] The 5 colonies of each strain 382, 382(pΔlacZ) as a control strain contained plasmid without insertion and 382 (pYHGN) were suspended in 2 ml of minimal medium ((NH₄)₂SO₄ - 25.0 g/l, K₂HPO₄ - 2.0 g/l, MgSO₄ 7H₂O- 1.0 g/l, thiamin - 0.2 mg/l, yeast extract - 5 g/l, glucose - 60 g/l, ampicilline - 100 mg/l, if necessary) in 20-ml test tubes and were incubated overnight with aeration at 32 °C. The 0.2 ml of each night culture was transferred to the three 20-ml test tubes with 2 ml of fresh medium for fermentation with or without IPTG and cultivated at 32 °C for 72 hours with rotary shaker.

Fermentation medium composition:

(NH ₄) ₂ SO ₄	25 g/l,
K ₂ HPO ₄	2.0 g/l,
MgSO ₄ 7H ₂ O	1.0 g/l,
Thiamin	0.2 mg/l,
Yeast extract	5 g/l
Glucose	60 g/l,
CaCO ₃	20 g/l
Ampicilline	100 mg/l, if necessary
IPTG	0.5 mM, if necessary

[0077] After cultivation the plasmid stability and optical absorbance of the medium at 540 nm were determined by conventional methods. Accumulated amount of arginine in the medium was determined by TLC. Liquid phase composition for TLC was as follows: isopropanol - 80 ml, ethylacetate - 40 ml, NH₄OH (30 %) - 25 ml, H₂O - 50 ml. The results are shown in Table 10. As it is seen, the hybrid plasmid pYHGN improved the arginine accumulation by the arginine producing strain 382.

Table 10

<i>E. coli</i> 382 with plasmid	IPTG	OD ₅₄₀	Arg. g/l	Arg/OD
No	-	20	8.5	0.43
	+	22	6.7	0.31
pΔlacZ	-	28	6.3	0.23
	+	26	5.4	0.21
pYHGN	-	24	5.8	0.24
	+	26	9.3	0.36

Example 11: Production of proline by a strain having plasmid pYHGN (not belonging to the scope of the present invention)

[0078] The proline producing strain *E. coli* 702ilvA was transformed by the plasmid pYHGN carried the b3434 gene under the control of P_{lac} UV5 promoter.

[0079] The 5 colonies of each strain 702ilvA, 702ilvA(pΔlacZ) as a control strain contained plasmid without insertion and 702ilvA(pYHGN) were suspended in 2 ml of minimal medium ((NH₄)₂SO₄ - 18 g/l, K₂HPO₄ - 1.8 g/l, MgSO₄ - 1.2 g/l, thiamin - 0.1 mg/l, yeast extract - 0.5 g/l, glucose - 60 g/l, isoleucine - 50 mg/l, ampicilline - 300 mg/l, if necessary) in 20-ml test tubes and were incubated overnight with aeration at 32 °C. The 0.2 ml of each night culture was transferred to the three 20-ml test tubes with 2 ml of fresh medium for fermentation with or without IPTG and cultivated at 32 °C for 40 hours with rotary shaker.

Fermentation medium composition:

(NH ₄) ₂ SO ₄	18 g/l,
K ₂ HPO ₄	1.8 g/l,

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(continued)

MgSO ₄	1.2 g/l,
CaCO ₃	20 g/l,
Thiamin	0.1 mg/l,
Glucose	60 g/l,
Isoleucine	50 mg/l
Ampicilline	300 mg/l, if necessary
IPTG	0.5 mM, if necessary

[0080] After cultivation the plasmid stability and optical absorbance of the medium at 540 nm were determined by conventional methods. Accumulated amount of proline in the medium was determined by TLC. Liquid phase composition for TLC was as follows: ethanol - 80 ml, NH₄OH (30 %) - 5 ml, H₂O - 25 ml. The results are shown in Table 11. As it is seen, the hybrid plasmid pYHGN improved the proline accumulation by the proline producing strain 702ilvA.

Table 11

702ilvA with plasmid	IPTG	40 hours		
		OD ₅₄₀	Pro, g/l	Pro/ OD
No	-	25	4,0	0,16
	+	23	4,1	0,18
pΔlacZ	-	24	5,3	0,22
	+	22	5,0	0,23
pYHGN	-	24	5,9	0,25
	+	17	7,1	0,42

SEQUENCE LISTING

[0081]

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Phe Leu Ile Ala Ile Lys Met Ile Phe Pro Ser Ala Ser Gly Asn Ser
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35

Pro Leu Val Ala Gly Pro Thr Ile Leu Ala Thr Leu Met Leu Leu Ser
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45

Leu Ile Leu Val Met Met Ala Thr Gln Met Phe Leu Asp Gly Ile Arg
 180 185 190

Met Trp Met Lys Gly
 195

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Claims

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1. An L-amino acid producing bacterium belonging to the genus *Escherichia*, wherein the bacterium has been modified so that the L-amino acid production by said bacterium is enhanced by enhancing activities of proteins as defined in the following (E) or (F) in a cell of said bacterium:

(E) a protein which comprises the amino acid sequence shown in SEQ ID NO:11 in Sequence listing;
(F) a protein which comprises an amino acid sequence including deletion, substitution, insertion or addition of 1-22 amino acids in the amino acid sequence shown in SEQ ID NO:11 in Sequence listing, and which has an activity of making bacterium having enhanced resistance to L-amino acids and/or its analogs;

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the activities of proteins being enhanced by transformation of said bacterium with DNA coding for protein as defined in (E) or (F), or by alteration of promoter sequence of said DNA on the chromosome of the bacterium.

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2. The bacterium according to the claim 1, wherein the transformation is performed with a multicopy vector.
3. A method for producing L-amino acid, which comprises cultivating the bacterium according to any of claim 1 or 2 in a culture medium and collecting from the culture medium L-amino acid to be produced and accumulated.
4. The method according to claim 3, wherein L-amino acid is L-threonine.
5. The method according to claim 4, wherein the bacterium has been modified so that the bacterium has enhanced expression of threonine operon.
6. The method according to claim 3, wherein L-amino acid is L-valine.
7. The method according to claim 6, wherein the bacterium has been modified so that the bacterium has enhanced expression of *ilv* operon.

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Patentansprüche

1. L-Aminosäure produzierendes Bakterium der Gattung Escherichia, welches so modifiziert worden ist, daß die L-Aminosäure-Produktion durch das Bakterium durch Verstärken der Aktivitäten von Proteinen nach (E) oder (F) in einer Zelle des Bakteriums erhöht ist:

30

(E) ein Protein, das die in Sequenz ID NO:11 des Sequenzprotokolls gezeigte Aminosäuresequenz aufweist;
(F) ein Protein, das eine Aminosäuresequenz, einschließlich Deletion, Substitution, Insertion oder Addition von 1 bis 22 Aminosäuren in der in Sequenz ID NO:11 des Sequenzprotokolls gezeigten Aminosäuresequenz aufweist und das die Aktivität hat, einem Bakterium erhöhte Resistenz gegen L-Aminosäuren und/oder deren Analoga zu verleihen;

35

wobei die Aktivitäten des Proteins durch Transformation des Bakteriums mit DNA, die für ein Protein nach (E) oder (F) kodiert, oder durch Veränderung der Promotorsequenz der DNA auf dem Chromosom des Bakteriums erhöht sind.

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2. Bakterium nach Anspruch 1, worin die Transformation mit einem Mehrkopien-Vektor durchgeführt wird.
3. Verfahren zur Produktion einer L-Aminosäure, welches das Kultivieren des Bakteriums nach Anspruch 1 oder 2 in einem Kulturmedium und das Gewinnen der zu produzierenden und anzuhäufenden L-Aminosäure aus dem Kulturmedium umfaßt.
4. Verfahren nach Anspruch 3, wobei die L-Aminosäure L-Threonin ist.
5. Verfahren nach Anspruch 4, wobei das Bakterium so modifiziert worden ist, daß es eine erhöhte Expression des Threonin-Operons aufweist.
6. Verfahren nach Anspruch 3, wobei die L-Aminosäure L-Valin ist.
7. Verfahren nach Anspruch 6, wobei das Bakterium so modifiziert worden ist, daß das Bakterium eine erhöhte Expression des *ilv*-Operons aufweist.

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Revendications

- 5 1. Bactérie productrice de L-aminoacide appartenant au genre *Escherichia*, où la bactérie a été modifiée de telle manière que la production de L-aminoacide par ladite bactérie est augmentée par augmentation des activités de protéines telles que définies en (E) ou (F) suivants dans une cellule de ladite bactérie :

(E) une protéine qui comprend la séquence d'aminoacides montrée dans SEQ ID NO : 11 dans le listage de séquences ;

10 (F) une protéine qui comprend une séquence d'aminoacides incluant une délétion, une substitution, une insertion ou une addition de 1-22 aminoacides dans la séquence d'aminoacides montrée dans SEQ ID NO : 11 dans le listage de séquences, et qui a une activité de production d'une bactérie ayant une résistance accrue aux L-aminoacides et/ou leurs analogues ;

15 les activités des protéines étant augmentées par transformation de ladite bactérie avec un ADN codant une protéine telle que définie en (E) ou (F), ou par modification de la séquence promotrice dudit ADN sur le chromosome de la bactérie.

2. Bactérie selon la revendication 1 où la transformation est accomplie avec un vecteur à copies multiples.

- 20 3. Procédé pour produire un L-aminoacide qui comprend la culture de la bactérie selon l'une quelconque des revendications 1 ou 2 dans un milieu de culture et la collecte à partir du milieu de culture du L-aminoacide destiné à être produit et accumulé.

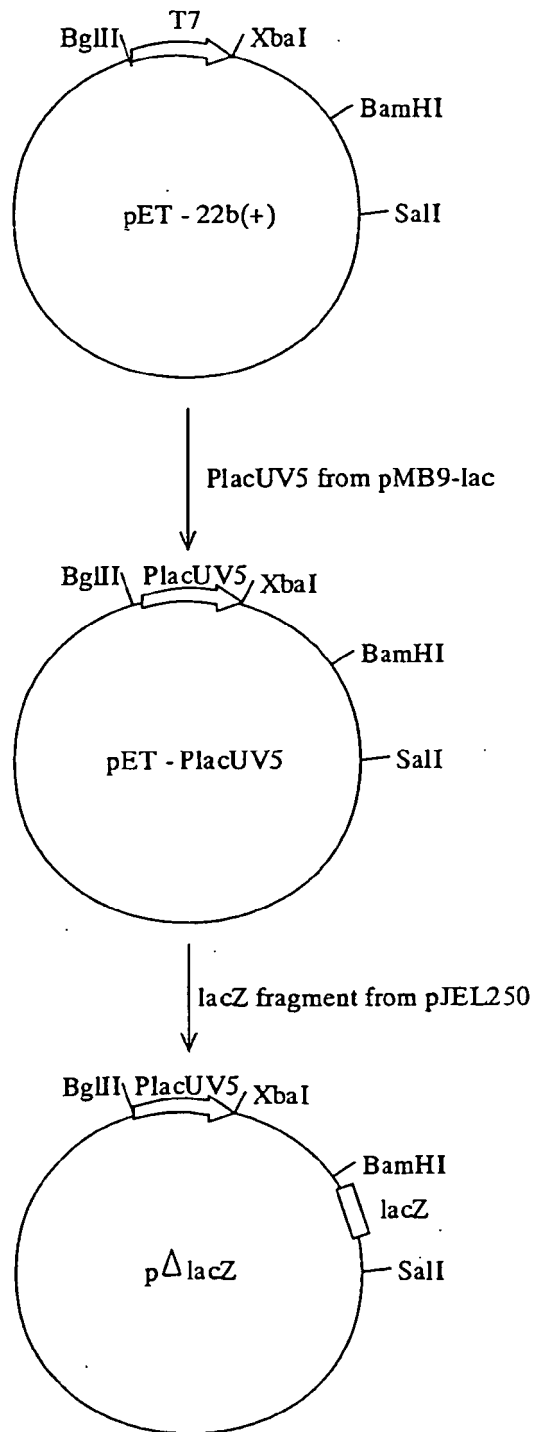
- 25 4. Procédé selon la revendication 3 où le L-aminoacide est la L-thréonine.

5. Procédé selon la revendication 4 où la bactérie a été modifiée de telle manière que la bactérie a une expression accrue de l'opération thréonine.

- 30 6. Procédé selon la revendication 3 où le L-aminoacide est la L-valine.

7. Procédé selon la revendication 6 où la bactérie a été modifiée de telle manière que la bactérie a une expression accrue de l'opération *ilv*.

Fig. 1



REFERENCES CITED IN THE DESCRIPTION

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