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(54) **Extracts obtained from cell line cultures from Syringa vulgaris IRB-SV25/B (DMS:16857), their preparation and use**

Extrakten aus Zelllinien von Syringa vulgaris IRB-SV25/B (DMS:16857), ihre Vorbereitung und Verwendung davon

Des extraits obtenus à partir des lignées cellulaires des Syringa vulgaris IRB-SV25/B (DMS:16857), leur préparation et utilisation

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Remarks:

The file contains technical information submitted after the application was filed and not included in this specification

Description

[0001] The present invention refers to phenylpropanoid extracts titrated in verbascoside, the use thereof in human and veterinary medicine, for nutritional and cosmetic purposes. Said extracts are derived from "*in vitro*" cell cultures from *Syringa vulgaris* IRB SV25/B (DSM: 16857) cell line, belonging to the Oleaceae family, obtained by fermentation. Furthermore, the present invention concerns the preparation and use of said extracts, for the production of drugs or nutritional or cosmetic substances, said extracts possessing not only antioxidant activity but also other important pharmacological properties.

[0002] Numerous studies have confirmed that oxidative stress is a phenomenon which progresses over time, causing biological damage leading to cell death, and which is responsible for both the effects connected with lipoprotein peroxidation and death due to degeneration, in addition to death due to apoptosis. Indeed, oxidative stress has been recognised as a mediator of programmed cell death, responsible for the activation of T lymphocytes and the depletion of CD4+ T cells in AIDS (Buttke T.M., Sandstrom P.A. "Oxidative stress as a mediator of apoptosis" Immunology Today, 1994, 15(1): 7-10).

[0003] The antioxidant action of phenylpropanoids, and verbascoside in particular, has been amply described and is well known in the literature (Seidel et al. Phenylpropanoids from *Ballota nigra* L. inhibit *in vitro* LDL peroxidation. *Phytother. Res.*, 2000, 14(2): 93-98; Chion et al. Acteoside protects endothelial cells against free radical-induced oxidative stress. *J. Pharm. Pharmacol.*, 2004, 56(6): 743-8; Lee et al. Protective effect of acteoside on carbon tetrachloride induced hepatotoxicity. *Life Sci.*, 2004, 74(8): 1051-64), but suitable sources for the large scale production thereof are completely absent, due to the very low quantities typically present in plant tissues and the high costs associated with the purification process.

[0004] Isoverbascoside was first identified as an "acteoside isomer," in *Leucosceptrum japonicum* (MIQ.) Kitamura et Murata and also from *Paulownia tomentosa* (Thumb) Steud. (Kawada et al., 2002, 48: 512-515).

[0005] *Syringa vulgaris* cell suspension cultures produce mixture of hydroxyphenylethanol glycosides, namely verbascoside (acteoside) and salidroside, where the main component is verbascoside (Ellis, *Phytochemistry*, 1983, 22(9): 1941-1943). Cell suspension cultures producing verbascoside at more than 1 g/l have been established in three plant species, namely *Leucosceptrum japonicum* f. *barbinerve*, *Syringa josikaea*, and *S. vulgaris*. Of the three species, suspension cultures of *L. japonicum* f. *barbinerve* showed rapid growth and the highest yield of verbascoside (1.89 g/l) (Inagaki et al., *Plant Cell Reports*, 1991, 9(9): 484-487). Stabilized plant cell cultures obtained by clonal selection of *Olea europea*, *Appia citrobata* and *Syringa vulgaris* producing 1.500, 1.800 and 900 µg verbascoside, have been described in patent document WO2004/033673 (Istituto di Ricerche Biotechnologiche, S.R.L., 22.04.2004). Hence, a first subject of the invention is a process for the production of extracts from plant cells belonging to *Syringa vulgaris*, according to claim 1, allowing the attainment of industrial quantities of phenylpropanoids.

[0006] Further subjects of the invention relate to an extract according to claim 13, obtainable according to this process, and uses thereof in medical, nutritional and cosmetics fields.

[0007] Further subjects and the advantages of the present invention will become clearer from the following detailed description of the invention.

[0008] The process of the invention comprises a first stage of cell culture selection based on the metabolite of interest.

[0009] Briefly, this process envisages the collection of plant tissue, the cleaning thereof, for example under running water, cutting into fragments of 2-5 cm and sterilisation on plates, for example by sequential treatment with 70% ethanol for approx. 15 minutes, 2% sodium hypochlorite for approx. 5 minutes and finally 0.05% HgCl₂ for approx. 1 minute. Between treatments, the plant fragments are washed, typically three or more times with sterile distilled water.

[0010] Each fragment of said tissue, chopped-up even further (explants), is placed on a Petri dish containing nutrient medium, solidified by the addition of Agar and supplemented with growth hormones without the addition of any antibiotics. The number of explants performed influences the outcome of the subsequent stages. Generally from 2000 to 5000 uncontaminated explants are sufficient to proceed to the subsequent selection stage. Following a suitable period of time, undifferentiated callus tissue, forms, which is then multiplied following transfer onto a greater surface area with fresh medium.

[0011] Furthermore, the plant cell line derived from the undifferentiated callus tissue is preferably stabilised by means of a certain number of transfers (sub-cultures) onto fresh culture medium. Particularly, it has been observed that in order to obtain a stable cell line, it is important to perform at least ten sub-cultures. Such medium is solid in nature, and may be advantageously constituted by 0.8-1% agar in a standard culture medium, to which has been added plant peptone, allowing a balanced supply of aminoacids and guaranteeing the maintenance of good cell wall integrity. Preferably, plant peptone will be added in quantities ranging between 500 and 4000 mg/l of culture medium.

[0012] A "stable cell line" is defined as a culture having the following characteristics:

- high and constant proliferation rate over time;
- preservation of the same phenotypic characteristics throughout various subcultures (cell colour, aggregate friability,

size etc.);

- constant secondary metabolite levels per unit of mass, over the course of the various subculture steps (secondary metabolite content is assessed by chemical analysis of the extracts);
- constant primary metabolite content (protein, lipids and polysaccharides) per unit mass.

[0013] Subsequently, at the stabilisation stage, the cell line is subjected to "clonal selection". Such selection consists of growing the stabilised cells for an appropriate amount of time (typically, 10-15 days of culture). Subsequently, individual cell aggregates are taken from the solid culture medium and each of said cell aggregates is inoculated into liquid culture medium described above.

[0014] Following fermentation for such time necessary to obtain adequate multiplication of the cell aggregate (hereinafter referred to as "clone"), a time generally comprised of between 10 and 15 days, the content of the metabolite of interest is determined for each clone.

[0015] Such operations may be repeated until a plant cell line clone is selected, in which production of the metabolite of interest is the highest.

[0016] It should be remembered that alternating periods of culture on solid and liquid media is essential for the clonal selection process. Hence, it is essential that the clonal selection process described above does not conclude with the identification of the most active clone, but is constantly repeated so as to keep the selected clone phenotypically homogeneous.

[0017] The selected plant cell line is then multiplied in order to obtain a sufficient quantity of biomass to carry out the production fermentation stage. Said quantity will depend on the specific production requirements, the plant cell line typology used and the type of metabolite it is desired to produce.

[0018] The biomass thus obtained may be passed directly into the final fermenter, or can be subjected to one or more further growth stages in liquid medium, working with intermediate volumes.

[0019] Preferably, the process just illustrated includes the stages of:

- a) cultivating a predetermined plant cell line, stabilised for a sufficient period of time to allow the multiplication of said cell line to give substantially distinct cell clusters, on solid medium;
- b) removing said substantially distinct cell clusters from said solid medium and placing each of them in a separate liquid culture medium;
- c) cultivating each of the said substantially distinct cell clusters in said liquid culture medium for a sufficient period of time to allow the multiplication of said cell clusters, and the analytical determination of the primary and/or secondary metabolites produced thereby;
- d) performing a qualitative and quantitative determination of the primary and/or secondary metabolites produced by each of said cell clusters in said liquid culture medium;
- e) selecting the cell cluster capable of producing the greatest quantity of said metabolite of interest;
- f) repeating the process cycle according to stages a), b), c), d) and e) on said cell cluster, selected according to stage e), a sufficient number of times until the quantity of said metabolite of interest produced by the selected cell cluster, and by the cell clusters deriving from further selection cycles, is essentially constant.

[0020] In addition, the subsequent fermentation may preferably consist of the following stages:

- A) the inoculation of said plant clone into liquid medium and the multiplication thereof for a sufficient period of time to obtain an increase in cellular mass of at least 300% of the weight thereof;
- B) optionally, transfer of the suspension obtained from stage A) into fresh liquid culture medium and the multiplication thereof for a sufficient period of time to obtain an increase in cellular mass of at least 300% of the weight thereof;
- C) optionally, the repetition- of stage B) at least one additional time;
- D) the transfer of the suspension obtained in stages A), B) or C) into fresh liquid culture medium in a fermenter to give a biomass, and conducting the fermentation under such conditions and for a sufficient period of time so as to obtain within said biomass, the production of said at least one metabolite of interest;
- E) the separation of said at least one metabolite of interest from said biomass. The fermentation will normally be performed at a temperature of between 15°C and 35°C, typically around 25°C and for a period of time normally between 7 and 42 days, preferably between 7 and 21 days. It is essential that the biomass be adequately aerated and that at the same time be kept stirred by means of stirring external to the fermenter. Indeed, it has been observed that plant biomass is comprised of cells having cell-walls that are poorly resistant to rupture. A stirrer submerged into the biomass acts mechanically on the cells and can easily cause the lysis thereof. However, it is necessary that the stirring, although delicate, be efficient, above all in the final fermentation stages when the biomass greatly increases in density. For the purposes of the present invention, particularly appropriate methods of stirring are orbital

means of stirring. Such means of stirring preferably operate at 10-200 rpm, more preferably at around 120 rpm.

[0021] It is appropriate that the volume of the container (fermenter) in which the fermentation occurs be considerably greater than the volume of the biomass. Typically, the volume of the reactor will be from 50% to 200% greater than the biomass volume.

[0022] As already mentioned, efficient fermentation requires adequate oxygenation. Oxygenation is normally performed by using sterile air with a flow rate of 0.1-5 l/minute, more preferably 2-3 l/minute, for a volume of 10 litres of biomass. Alternatively, gas mixtures containing from 10% to 100% v/v of oxygen may be used.

[0023] As mentioned previously in relation to stirring, even oxygenation by means of over violent bubbling can cause rupturing of the cell walls. Hence it is necessary to ensure that oxygenation is performed delicately, for example by bubbling through appropriate diffusers. It will be preferable to use means of air or oxygen diffusion with nozzle delivery flow speeds comprised of between 10 m/min and 600 m/min, more preferably between 50 m/min and 350 m/min.

[0024] In addition, the shape of the fermentation chamber has significant importance. Indeed, it is recommended that it has a smooth and uniform surface, i.e. that there are no edges, corners or other parts which can cause the cell walls of the biomass rupture. Additives increasing water oxygen solubility will be added to the biomass. Such additives will preferably be selected from those substances defined as "artificial blood", for example the perfluorinated hydrocarbons (PFC).

[0025] According to the invention, stabilised cell lines derived from *Syringa vulgaris* IRB-SV25/B (DSM16857) have been selected for their ability to produce phenylpropanoids (FP), and particularly verbascoside, in suitable qualitative and quantitative amounts.

[0026] Subsequently, said plant cell lines, selected in the aforesaid manner, have been extracted by means of the following procedure.

[0027] The details of a *Syringa vulgaris* cell line, the extraction conditions and an extract thereof, will now be described.

Morphological characteristics of the cell line

[0028] The *Syringa vulgaris* cell line denominated IRB-SV25/B DSMZ (Deutsche Sammlung von Mikroorganismen und Zellkulturen, Braunschweig, D) accession N°: DSM 16857, accession date: 2004-09-15) has a clear hazelnut colouration and mucid (sticky) appearance.

Extraction process

[0029] The extraction comprises the following stages in sequence:

- a) subjecting the biomass, obtained in accordance with the selection process described previously, to an enzyme deactivation treatment;
- b) homogenising the product obtained following step a);
- c) separating the aqueous phase from the cell residues by means of gravity filtration, pressure filtration or centrifugation through a porous mesh, for example nylon, steel or cotton mesh etc., preferably having a porosity comprised of between 10 µm and 300 µm, more preferably between 50 µm and 150 µm;
- d) performing an extraction of the aqueous phase obtained following step c) using hydrophobic interaction resin, preferably in batch;
- e) recovering the absorbed substances (extract) from the resin by means of elution with ethanol, or a mixture of alcohols having up to 5 carbon atoms (preferably ethanol), in water in percentages varying from 30% up to just under 100% by volume.

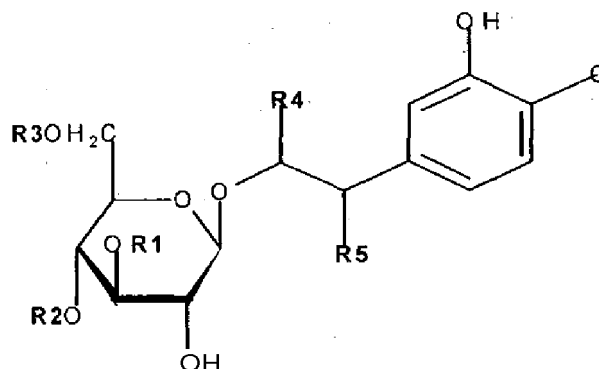
[0030] According to one particular aspect of the invention, between steps a) and b) a biomass filtration step may be performed under the same conditions reported in step c), with the aim of recovering just the biomass itself without any fermenter culture medium.

[0031] In particular, cultures of the cell line IRB-SV25/B in suspension, obtained in accordance with the cited procedures, are harvested with ages comprised of between 7 and 21 days. The enzyme deactivation treatment occurs, for example, by means of thermal treatment. Particularly, said thermal treatment occurs by heating the biomass at a temperature comprised of between 50°C and 150°C, preferably between 60°C and 120°C, for a time of less than 5 minutes, so as to inactivate all enzymes. Preferably, the thermal treatment involves the use of steam. Subsequently, as described previously, the biomass is homogenised using an ultraturrax and then filtered or centrifuged. The substances present in the clear aqueous phase are batch extracted with hydrophobic interaction resin, preferably selected from polystyrene-divinylbenzene or acrylic matrix resins, and the absorbed products are recovered from the resin by means of one or more elution steps with aqueous ethanol, from 30% to 95% by volume.

[0032] In accordance with one particular embodiment, the entire biomass is filtered as described above, and subjected to thermal treatment with steam at 120°C for a period of time varying between 5 and 30 minutes.

[0033] By the term "extract" is meant an extract obtained from cell cultures from *Syringa vulgaris* plant, belonging to the Oleaceae family, deriving from any of the extraction methods described below and comprising varying percentages of phenylpropanoids, such as for example verbascoside and isoverbascoside and a series of verbascoside analogues. Said "extract" further comprises a fraction, devoid of any characteristic chromophores, present in variable quantities depending on the preparation method and consisting mainly, but not exclusively, of oligo- and poly-saccharide, protein and lipid molecules.

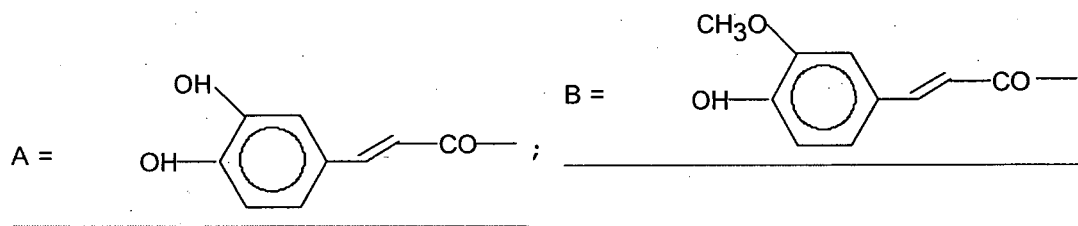
[0034] The extracts contain phenylpropanoids of the following general formula (I)



wherein:

- R1 is a hydrogen atom or a monosaccharide with 5 or 6 carbon atoms and preferably rhamnose;
- R2 is a hydrogen atom or is a caffeoyl (A) or feruloyl (B) group;
- R3 is a hydrogen atom or a monosaccharide with 5 or 6 carbon atoms or a disaccharide with 10 to 12 carbon atoms or is a caffeoyl (A) or feruloyl (B) group;
- R4 is a hydrogen atom or an alkyl group, preferably methyl, or a hydroxyl group;
- R5 is a hydrogen atom or an alkyl group, preferably methyl, or a hydroxyl group, either free or condensed with the alcohol group at position C2 of the monosaccharide group bound to it, in order to form a 1,4 dioxane ring;
- R6 is a hydrogen atom or an alkyl group, preferably methyl;

when R3 is a caffeoyl (A) or feruloyl (B) group,



then R2 is always a hydrogen atom and *vice versa*.

[0035] The structures of verbascoside and isoverbascoside have been confirmed using mass spectrometry (MS) and nuclear magnetic resonance (NMR).

[0036] Verbascoside is a phenylpropanoid known also as acteoside, Kusagin or Orobanchin (C. Jimenez et al. Nat. Prod Report 1994, 591-606) and is defined as b-(3,4-dihydroxyphenyl)-ethyl-O-a-L-ramnopyranosyl (1"-3')-b-D-(4'-O-caffeoyl)-glucopyranoside. It is found along with isoverbascoside or b-(3,4-dihydroxyphenyl)-ethyl-O-a-L-ramnopyranosyl (1"-3')-b-D-(6'-O-caffeoyl)-glucopyranoside.

[0037] Hence, one of the aspects of the present invention consists in the fact that, by performing the extraction of the biomass obtained from the cell lines described above, extracts containing phenylpropanoids having general formula (I) are obtained.

[0038] Extraction occurs by bringing the clear solution obtained from treatment of the biomass into contact with an appropriate quantity of resin, separating the resin and eluting said resin with a suitable eluent.

[0039] Attainment of the extracts varies as a function of numerous parameters. The use of fermentation supernatant (which contains the part of the fraction devoid of chromophores and which is released from the cells during culture) during the filtration stage leads to a slight enrichment in phenylpropanoids due to the dilution effect. A second factor is the quantity of resin used in relation to the quantity of material to be extracted, with less resin promoting the selective attachment of phenylpropanoids and hence consequent enrichment of the eluate. Furthermore, different enrichments may be obtained depending on the EtOH/H₂O ratios used to recover the compounds absorbed onto the resin, since lesser percentages of ethanol in the eluent predominantly extract the fraction devoid of chromophore, while subsequent elution with a mixture containing a higher percentage of ethanol leads to the preferential extraction of phenylpropanoids.

[0040] Thus it is possible to obtain extracts containing from 20% to 90% phenylpropanoids by weight, preferably between 30% and 60%, with the remaining percentage fraction, comprised of between 80 and 10%, preferably between 70% and 40%, being comprised of a chromophore-free fraction, comprising mainly, but not exclusively, oligo- and polysaccharide, protein and lipid type molecules.

[0041] It has also been surprisingly discovered that the thermal treatment performed on the biomass causes the isomerisation of verbascoside to isoverbascoside, and that the extent of said isomerisation depends on the treatment temperature and time. Temperatures comprised of between 50°C and 150°C, preferably between 60°C and 120°C, for a period of time sufficient to cause isomerisation (measurable by HPLC analysis as reported previously), preferably between 5 and 30 minutes, are generally used to obtain the transformation of a percentage ranging between 40% and 60% of the verbascoside initially present into isoverbascoside. Hence the content of such substances in the filtered clear solution will also be modified. The thermal treatment may be performed by heating externally or internally for example by the insufflation of steam into the biomass.

[0042] Non-thermally treated biomass will generally contain a percentage of between 5% and 20% isoverbascoside with respect to the total mass of phenylpropanoids. Following thermal treatment, the percentage of isoverbascoside will generally be comprised of between 45% and 65% of the total mass of phenylpropanoids.

[0043] The above described isomerisation may occur in all those phenylpropanoids of general formula I wherein R₃ is a hydrogen atom and R₂ is anything other than hydrogen.

[0044] Hence, a process for the isomerisation of phenylpropanoids of general formula I, wherein R₃ is a hydrogen atom and R₂ is anything other than hydrogen, comprising a thermal treatment stage of said phenylpropanoids as described above, constitutes the subject of the present invention.

EXAMPLE 1

[0045] Calluses grown on solid medium (GAMBORG B5 in 1% agar supplemented with 20 g/l sucrose, 2 g/l plant peptone, 1 mg/l kinetin, 1mg/l naphthalenacetic acid, 0.2 mg/l indolacetic acid at pH 6.5) and subjected to subculture for at least three months, are inoculated into 20 x 300 ml flasks, each containing 50 ml of liquid medium (GAMBORG B5 supplemented with 20 g/l sucrose, 2 g/l plant peptone, 1 mg/l kinetin, 1mg/l naphthalenacetic acid, 0.2 mg/l indolacetic acid at pH 6.5). Following fermentation for 7 days, the cultures are used to likewise inoculate 1000 ml flasks, each containing 250 ml of liquid medium which are then unloaded after a period 14 days. The biomass is combined and filtered through nylon mesh with a porosity of 100 µm. The cells are suspended in an equal volume of H₂O and briefly treated with steam (3-4 minutes). The suspension is then homogenised using an ultraturrax and centrifuged in order to remove the solid residue. The latter is then taken up in 5 vol. of H₂O then homogenised and centrifuged. The supernatants are combined and titrated by HPLC.

[0046] The analysis shows a total phenylpropanoid content, expressed as verbascoside, equal to 12.3 g. To the aqueous solution is added 2 kg of Diaion HP 2MG resin, and the slurry left stirring overnight. After removal of the aqueous phase by filtration, the resin is washed with a first portion of 10 litres of hydroalcoholic mixture EtOH:H₂O (20:80), then eluted with three 10 litre batches of EtOH:H₂O mixture (80:20). The combined eluates are concentrated to a small volume in an evaporator under reduced pressure. The lyophilised residual aqueous phase yields 10.3 g of a yellow powder. Typically, the extract thus obtained (**Extract 1**) has a titre equal to approx. 90% in phenylpropanoids expressed as verbascoside, and approx. 10%, in other components, constituted mainly, but not exclusively, by oligo- and polysaccharides, protein and lipid type molecules.

EXAMPLE 2

[0047] 40 x 1000 ml flasks, each containing 250 ml of culture obtained from calluses grown on solid medium (GAMBORG B5 in 1% agar supplemented with 20 g/l sucrose, 2 g/l plant peptone, 1 mg/l kinetin, 1mg/l naphthalenacetic acid, 0.2 mg/l indolacetic acid at pH 6.5) following 15 days of fermentation are used to similarly inoculate 3 litre flasks, each containing 1 litre of liquid medium (GAMBORG B5 supplemented with 20 g/l sucrose, 2 g/l plant peptone, 1 mg/l kinetin, 1mg/l naphthalenacetic acid, 0.2 mg/l indolacetic acid at pH 6.5). After a further 15 days of fermentation, these are in turn used to inoculate 10 30 litre fermenters, each containing 20 litres of liquid medium. After a further 15 days, the

fermenters are in turn used as the inoculum (equal to 200 litres) for a fermenter with a capacity of 1000 litres, containing 600 litres of medium. Fermentation is monitored by means of samples taken sequentially over time and, is terminated upon reaching an age of 20 days. Final production is equal to 3.1 g/l, for a total of 2,480g of phenylpropanoids expressed as verbascoside (600 litres + 200 litres of inoculum). Steam at 120°C is insufflated into the reactor for 20 minutes, then the biomass is unloaded therefrom, homogenised and centrifuged or filtered under the same conditions adopted for Example 1. To the clear aqueous phase is then added 150 kg of Diaion HP 2MG resin and this is then left stirring overnight. At the end of said period, the resin is allowed to settle into a column with a diameter of 50 cm and height of 150 cm. The spent aqueous phase is left to slowly flow out from the bottom of the column, which is then washed with copious amounts of water (500 litres), then firstly eluted with 200 litres of hydroalcoholic mixture EtOH:H₂O (20:80), then with 500 litres of EtOH:H₂O mixture (80:20), at a flow rate of 300 ml/min. The eluate is concentrated under reduced pressure to give an aqueous residue which is then lyophilised. 4,960g of lyophilisate are obtained. Typically, the extract thus obtained (**Extract 2**) is constituted by approx. 50% by weight of phenylpropanoids expressed as verbascoside, and approx. 50% of other components, constituted mainly, but not exclusively, by oligo- and polysaccharides, protein and lipid type molecules.

EXAMPLE 3

[0048] >From HPLC analysis of the crude product obtained in example 2, verbascoside and its analogue isoverbascoside are the main components belonging to the class of caffeic derivatives, and are typically accompanied by at least six products belonging to the same class.

[0049] With the aim of purifying the compounds of interest, in order to assess their activities, chromatographic purification using a 7 x 150 cm column, packed with Sephadex- LH20 resin and equilibrated in 8% EtOH in water was used. The column was loaded with 90g of extract obtained from example 2 and eluted isocratically using the same solvent. All the fractions were analysed by HPLC using the method described below and those containing the same products combined, concentrated and lyophilised. In particular, two groups of fractions have been identified containing the pure molecules, verbascoside (27.1 g) and isoverbascoside (8.8 g).

[0050] High pressure liquid chromatography (HPLC) analysis is performed using a Phenomenex 4.6 x 150 mm C18 (2) column. Phase A - water/0.01 N phosphoric acid; Phase B - acetonitrile/0.01 N phosphoric acid; flow rate - 0.8 ml/min.

Gradient:

Time	%B
0	0
10	10
15	20
20	25
25	35
30	45
35	55
40	0

[0051] The retention times of the 2 major phenylpropanoids present in the extracts are reported in table 1.

Table 1 - Retention times of the phenylpropanoids verbascoside and isoverbascoside

Phenylpropanoid:	verbascode	isoverbascoside
RT*	20.3-20.6	21.3-21.6
*Retention time in minutes (interval)		

[0052] Figure 1, enclosed herein, shows the chromatogram of an extract measured at 330 nm.

[0053] Table 2 (below) shows the percentage concentration of each component of Extract 1 and Extract 2 compared to the composition of purified verbascoside and isoverbascoside.

Table 2

Code	Concentration %		
	Verbascoside	FP	FS
Extract 1	82±3	8±2	10±2,5
Extract 2	35±5	17±3	48±4
Purified verbascoside	99	1	0
Purified isoverbascoside	1	99	0
FP = fraction containing isoverbascoside and less abundant phenylpropanoids FS = chromophore-free fraction			

[0054] In particular, it has been surprisingly observed that when an extract in accordance with the present invention is used, i.e. an extract also comprising the chromophore-free fraction, then the pharmacological activities are greater in comparison to the use of pure phenylpropanoids.

[0055] In other cases, verbascoside and isoverbascoside have been found to exhibit pharmacological activities comparable to the inventive extracts. Particularly, the verbascoside and isoverbascoside have a platelet aggregation, anti-inflammatory and anti-lipoxygenase-, anti-5 α reductase, anti-tyrosinase, antifungal and metal-chelating (Fe²⁺, Fe³⁺, Cu²⁺ e Ni²⁺) activity.

[0056] In the following are reported some examples of the pharmacological action of the inventive extracts, verbascoside and isoverbascoside.

EXAMPLE 4

ANTIOXIDANT ACTIVITIES OF THE *SYRINGA VULGARIS* EXTRACTS BY MEANS OF BIOCHEMICAL ASSAYS BASED ON CHEMILUMINESCENCE MEASUREMENTS.

[0057] *Syringa vulgaris* Extracts 1 and 2, prepared as described in examples 1 and 2, have been used to assess *in vitro* antioxidant action with specific oxygen radical generating systems such as the superoxide anion (O₂⁻), the hydroxyl radical (HO[•]), the peroxynitrite radical (ONOO⁻) and the inhibition of lipid peroxidation. The superoxide anion has been measured by a chemiluminescent assay based on the hypoxanthine(HPX)/xanthine oxidase (XOD) reaction using the Lumimax Superoxide Anion Detection Kit (Stratagene, La Jolla, CA, USA) and following the manufacturer's instructions. Hydroxyl radical scavenger activity has been measured by means of the Fenton reaction for the production of hydroxyl radicals and chemiluminescent dosage (using a Victor³ chemiluminometer, Wallac, Finland) with luminol. Peroxynitrite ions have been determined by means of dosage with a spontaneous generator of peroxynitrites (3-morpholinosisidnonimine hydrochloride; SIN-1) and a stable reagent for the nitroxyl ions (2-(4-carboxyphenyl)-4,4,5,5-tetramethylimidazoline-1-oxyl-3-oxide potassium salt PTIO spin trap) then dosed by means of electron spin resonance (ESR) using a Nippon Denshi JES-FR30. Lipid peroxidation has been generated using Fe²⁺ induced-oxygen reactive species, and detected using thiobarbituric acid in accordance with Buege, JA & Aust, SD: Microsomal lipid peroxidation (Methods Enzymol. 1978, 52:302-310).

Table 3: The effects of *Syringa vulgaris* extracts on various assays of antioxidant activity (expressed as IC₅₀)

	Superoxide radical	Hydroxyl radical	Peroxynitrite radical	Lipid peroxidation
Extract 1	0.20 μ g/ml	0.80 μ g/ml	1.7 μ g/ml	6.0 μ g/ml
Extract 2	0.15 μ g/ml	0.61 μ g/ml	1.2 μ g/ml	5.8 μ g/ml
Verbascoside	3.1 μ g/ml	2.2 μ g/ml	1.6 μ g/ml	7.4 μ g/ml
Rutin	10 μ g/ml	3.9 μ g/ml	10 μ g/ml	Not determined
Aloin	7.6 μ g/ml	2.9 μ g/ml	12.3 μ g/ml	Not determined

[0058] Extract 1 and Extract 2 listed in Table 3 are derived from *Syringa vulgaris* cell cultures. In said extracts, verbascoside is present at 82% and 35% by weight respectively. The results are expressed as the concentration inhibiting 50% of activity (IC₅₀). The values reported are the means of 6 independent assays.

[0059] In the assays, *Syringa vulgaris* extracts 1 and 2 have shown themselves to be extremely effective scavengers of oxygen free radicals-, independently of their degree of purity. For the superoxide, hydroxyl and peroxynitrite radicals, both Extract 1, and Extract 2 possess greater capacity with respect to the classic flavonoids, such as for example rutin and aloin. In the superoxide and peroxynitrite generation assay, the IC₅₀ for rutin and aloin is 10 µg/ml and 7.6 µg/ml respectively, concentrations approximately 6 to 30 fold higher with respect to *Syringa vulgaris* Extract 1 and Extract 2 with verbascoside. The IC₅₀ for rutin and aloin for the inhibition of the production of hydroxyl radicals is 3.9 µg/ml and 2.9 µg/ml respectively, from 5 to 7 times higher with respect to the value determined for Extracts 1 -and 2. Furthermore,- the extracts have also shown themselves to be extremely effective in reducing iron ion-induced lipid peroxidation.

[0060] The antioxidant activity of pure verbascoside is surprisingly less effective in comparison to *Syringa vulgaris* Extracts 1 and 2 in activity assays against superoxide and hydroxyl radicals and, even though only slightly, in the inhibition of lipid peroxidase. This tends to indicate higher antioxidant activity in the extracts with respect to pure verbascoside.

EXAMPLE 5

ANTI- INFLAMMATORY ACTIVITY OF SYRINGA VULGARIS EXTRACTS, VERBASCOSIDE AND ISOVERBASCOSIDE ON WHOLE BLOOD AND WHITE BLOOD CELLS.

[0061] Anti-inflammatory activity has been assessed by measuring luminol dependent chemiluminescence generated by free radicals released from whole blood and from white blood cells. The assay has been performed using the following protocol: 10µl of fresh human blood is mixed with 0.980 ml of Hanks saline solution containing 5x10⁻⁵ M luminol and the test sample (at various concentrations). The chemiluminescent response is recorded continuously for 30 minutes at a temperature of 37°C. The results are expressed as IC₅₀ (mg/ml), i.e. as the minimum sample concentration inhibiting chemiluminescence by 50% with respect to the control. To isolate white blood cells (WBC) from whole blood one proceeds in the following manner: 1 ml of fresh blood is layered onto a Hipaque gradient (p=1.119) and centrifuged at 250 x g for 60 minutes at room temperature. Following centrifugation, the pellet containing the white blood cells is washed twice, using a large volume of cold Hanks saline solution. Finally, the washed WBCs are resuspended in 0.1 ml di HBSS containing foetal calf serum. 10⁶ WBC are added to a chemiluminometer cuvette. 10µl of PMA (final concentration equal to 10nM) are added to induce the formation of free radicals. The chemiluminescent response is recorded continuously for 30 minutes at a temperature of 37°C. The results are expressed as IC₅₀ (mg/ml), i.e. as the minimum sample concentration inhibiting chemiluminescence by 50% with respect to the control. The above assays have been performed on *Syringa vulgaris* Extracts 1 and 2, verbascoside and isoverbascoside.

[0062] Table 4 reports the values obtained for the above assays.

Table 4: anti-inflammatory activity of *Syringa vulgaris* extracts, verbascoside and isoverbascoside on whole blood and white blood cells.

	Whole blood chemiluminescence (IC ₅₀ mg/ml)	White blood cell chemiluminescence (IC ₅₀ mg/ml)
Extract 1	1.25	0.2
Extract 2	0.18	0.14
Verbascoside	1,29	0,23
Isoverbascoside	1,30	0,25

The inhibition of the production of free radicals released by the white blood cells is correlated with the anti-inflammatory activity of the substance assayed. *Syringa vulgaris* Extracts 1 and 2, verbascoside and isoverbascoside show high inhibitory capacity in relation to the release of free radicals from whole blood and stimulated white blood cells. Extract 2 has significantly higher activity with respect to Extract 1, verbascoside and isoverbascoside.

EXAMPLE 6

THE PLATELET ANTI-AGGREGATING AND DISAGGREGATING ACTIVITIES OF SYRINGA VULGARIS EXTRACT 1, VERBASCOSIDE AND ISOVERBASCOSIDE

[0063] Platelet enriched rabbit plasma has been used to assess the platelet anti-aggregating and disaggregating properties of *Syringa vulgaris* Extract 1, verbascoside and isoverbascoside. Platelet rich plasma has been obtained from rabbit venous blood by mixing it with a 3.8% sodium citrate solution (9:1 v/v). The solution has then been centrifuged at

460 x g for 20 minutes. The supernatant has been collected and immediately used for the aggregation assays. Platelet aggregation has been recorded using a Chronolog "Ionised calcium platelet aggregometer" (Chrono-log Co., USA). All measurements have been performed at 37°C with continuous stirring. One millilitre of the platelet suspension (in the presence or absence of the test substances) has been added to the cuvettes and incubated for five minutes to equilibrate the cells to 37°C. Then 10 µl of a solution of ADP (final concentration - 10 µM) has been added and the luminous transmittance continuously recorded. The results are expressed as the D₀/D₀⁺ ratio, where D₀⁺ and D₀ are the luminous transmittance values, in the absence and presence of extract respectively.

[0064] Platelet disaggregation (the disappearance of preformed aggregates) has been determined using the same luminous transmittance curve 10 minutes following the addition of ADP. The results are expressed as the DR/DR_k ratio, where DR_k and DR, are the luminous transmittance in the absence and presence of extract. The test substances, diluted in physiological solution (isotonic saline) (pH 7.4) are pre-incubated with the platelet rich plasma (1:10 v/v) for 5-40 minutes. The aggregation measurements have then been performed. Physiological solution carrier has been added to the control samples (1:10 v/v). The values are reported in Table 5.

Table 5. The effect of Extract 1, verbascoside and isoverbascoside on ADP induced platelet aggregation-disaggregation

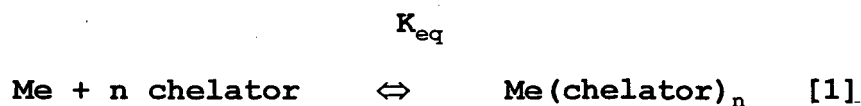
Extract 1 (mg/ml)	Aggregation (% with respect to control)	Disaggregation (% with respect to control)
0,001	100 ± 5*	85 ± 15
0,01	93 ± 11	113 ± 13
0,1	88 ± 10	138 ± 18
1,0	26 ± 5#	75 ± 25
Verbascoside (mg/mL) 1,0	24 ± 6#	73 ± 21
Isoverbascoside (mg/mL) 1,0	25 ± 6#	75 ± 21
"- Mean ± SD (n=6); # - p < 0.001 vs. control		

[0065] Extract 1, verbascoside and isoverbascoside cause a significant reduction in platelet aggregation at a concentration of 1 mg/ml, whilst no significant effects on platelet disaggregation are evident.

EXAMPLE 7

THE Fe²⁺, Fe³⁺, Cu²⁺ AND Ni²⁺ METAL CHELATING ACTION OF SYRINGA VULGARIS EXTRACTS, VERBASCOSIDE AND ISOVERBASCOSIDE

[0066] The chelating activities of *Syringa vulgaris* extracts, verbascoside and isoverbascoside in relation to Fe²⁺, Fe³⁺, Cu²⁺ and Ni²⁺ metal ions has been determined by means of spectrophotometric measurements using a Shimadzu 1770 UV spectrophotometer. The activities of chelating agents depends on the association constant (equilibrium) of their reactions with the metals according to the following reaction [1]:



wherein K_{eq} is the equilibrium constant equal to the ratio of concentrations at equilibrium of a complex and a product of equilibrium concentrations of metal ions and a chelator:

$$K_{eq} = \frac{[\text{Me (chelator)}_n]}{[\text{Me}] [\text{chelator}]^n} \quad [2]$$

The method used for determining chelating activity is well documented in the literature (Korkina LG, Afarras'er IB. Antioxidant and chelating properties of flavonoids. Adv. Pharmacol.,1997, 38:151-63). The results are expressed as **Keq (M⁻¹)** calculated for a pure phenylpropanoid. This way the chelating activity against ferrous (Fe II), ferric (Fe III), copper (Cu II) and nickel ions has been calculated.

Table 6: Chelation equilibrium constants (Keq, M⁻¹) for the *Syringa vulgaris* extracts, verbascoside and isoverbascoside.

Samples	Fe ²⁺ (Keq, M ⁻¹)	Fe ³⁺ (Keq, M ⁻¹)	Cu ²⁺ (Keq, M ⁻¹)	Ni ²⁺ (Keq, M ⁻¹)
Extract 1	0.47x10 ⁶ ± 0.14x10 ⁶	0.14x10 ⁶ ± 0.04x10 ⁶	0.12x10 ⁶ ± 0.03x10 ⁶	1553 ± 499
Extract 2	1. 90x10 ⁶ ± 0.14x10 ⁶	0.13x10 ⁶ ± 0.04x10 ⁶	0.11x10 ⁶ ± 0.03x10 ⁶	1704 ± 387
Verbascoside	0.45x10 ⁶ ±0.11x10 ⁶	0.12x10 ⁶ ±0.05x10 ⁶	0.12x10 ⁶ ±0.04x10 ⁶	1427±319
Isoverbascoside	0.46x10 ⁶ ±0.12x10 ⁶	0.13x10 ⁶ ±0.04x10 ⁶	0.13x10 ⁶ ±0.06x10 ⁶	1398±362
Rutin	0.14x10 ⁶ ± 0.04x10 ⁶	0.16x10 ⁶ ± 0.04x10 ⁶	0.11x10 ⁶ ± 0.04x10 ⁶	635±169

The chelating activities of *Syringa vulgaris* Extracts 1 and 2, verbascoside and isoverbascoside in relation to Fe²⁺ and Ni²⁺ metal ions, are higher with respect to that of rutin.

EXAMPLE 8

ANTI 5 ALPHA REDUCTASE AND ANTI-LIPOXYGENASE ACTIVITIES OF THE SYRINGA VULGARIS EXTRACTS, VERBASCOSIDE AND ISOVERBASCOSIDE, USEFUL FOR THE TRATMENT OF JUVENILE ACNE AND THE PREVENTION OF HAIR LOSS.

[0067] Juvenile acne shows a multifactorial aetiology; one factor which plays a fundamental role is the following: during puberty and adolescence, there is increased production of the hormone testosterone. In the skin, this hormone is converted by the enzyme 5 alpha reductase into the hormone dihydrotestosterone, the latter causes enlargement of the sebaceous glands with a concomitant increase in sebum secretion. High levels of the enzyme 5 alpha reductase is also one of the major causes of hair loss (Thiboutot D., Harris G., Iles V., Cimis G., Gilliland K., Hagari S. Activity of the type 1,5 alpha reductase exhibits regional differences in isolated sebaceous glands and whole skin. J. Invest. Dermatol. 1995; 105:209-14).

[0068] Data reported in the bibliography, show that extracts derived from the plant *Serenoa repens*, have anti-hair loss activity in that they possess inhibitory activities in relation to two enzymes: 5-alpha reductase and lipoxigenase (*Serenoa repens* (Bartram) J.K. Small. Fitoterapia 1997; LXVIII (2):99-113).

[0069] Determination of the 5 alpha reductase inhibitory activities by *Syringa vulgaris* extracts, verbascoside and isoverbascoside in comparison to *Serenoa repens* extracts. Said determination has been performed using enzymes (5 alpha reductase, type I and type II) produced by genetically modified yeast. Enzyme activities have been determined in the presence of *Syringa vulgaris* and *Serenoa repens* extracts, verbascoside and isoverbascoside.

[0070] Type I 5 alpha reductase activity has been assayed by incubating the enzyme in 0.1M Tris-citrate buffer (pH 7.0), containing 5µM [¹⁴C] radio-labelled testosterone (Amersham, Sweden) and 5 mM NADPH (Sigma) in a final volume of 5 ml. The solution thus obtained has been incubated for 1 hour at 37°C.

[0071] Determination of the type II 5 alpha reductase activity has been performed in an analogous manner to that just described, with the exception of a different pH value for the 0.1 M Tris-citrate buffer (pH 5.0).

[0072] In both assays, the steroids have been extracted using 5 ml of methylene chloride. The extracts have then been dried under a stream of nitrogen and the dried residues obtained have been dissolved in 20 µl of a chloroform-methanol mixture (2:1 v/v). An aliquot of said. mixture has been deposited on the surface of a silica gel TLC plate. The solvent mixture used to develop the TLC has been the same used to solubilise the dried residues. At the end of the TLC run, following UV detection, the R_f values of the metabolites have been compared with those of standard steroids, chromatographed simultaneously with the other samples. For quantification of the results, the spots have been scraped from the TLC plate and extracted with 2 ml of ethyl acetate. The extracts obtained have been analysed, using scintillation fluid, with the aid of a Wallach a-counter. The enzyme activity has been calculated from the percentage conversion (C%) of the substrate into final product:

$$C\% = \left[\frac{\text{counted product}}{\text{counted substrate} + \text{product}} \right] \times 100.$$

The results are reported in the following Table 7. Determination of the inhibition of lipoxygenase activity The inhibition of lipoxygenase activity by *Syringa vulgaris* extracts, verbascoside and isoverbascoside has been demonstrated using a method based on the measurement of chemiluminescence (Laakso S., Lilius EM, and Turunen P. Determination of cis-,cis-methylene interrupted polyunsaturated fatty acids in aqueous solutions by lipoxygenase chemiluminescence. J.Biochem. Biophys. Methods 1984; 9(1):61-68).

[0073] The results obtained are summarised in table 7.

Table 7: Inhibition of the enzymes 5 alpha reductase and lipoxygenase (expressed as IC₅₀) by *Syringa vulgaris* extracts, verbascoside and isoverbascoside, and *Serenoa repens* plant extracts.

Samples assayed	Type I 5 alpha reductase inhibition (IC ₅₀)	Type II 5 alpha reductase inhibition (IC ₅₀)	Lipoxygenase inhibition (IC ₅₀)
Extract 1	12 µg/ml	19 µg/ml	11 µg/ml
Extract 2	9.2 µg/ml	12.3 µg/ml	10.5 µg/ml
Verbascoside	13 µg/ml	21 µg/ml	12 µg/ml
Isoverbascoside	12 µg/ml	20 µg/ml	11,5 µg/ml
Serenoa repens extract	12 µg/ml	44 µg/ml	18.5 µg/ml

Syringa vulgaris Extracts 1 and 2, verbascoside and isoverbascoside have significant inhibitory capacity in relation to the enzymes 5 alpha reductase (type I and type II) and lipoxygenase. The inhibitory activity of the *Syringa vulgaris* extracts, verbascoside and isoverbascoside is greater than that shown by *Serenoa repens* plant extracts, particularly in relation to type II 5-alpha reductase, which is the principal agent responsible for the onset of acne and androgenetic alopecia (male pattern baldness).

EXAMPLE 9

ANTI-TYROSINASE ACTIVITY OF THE SYRINGA VULGARIS EXTRACTS, VERBASCOSIDE AND ISOVERBASCOSIDE TO BE USED FOR SKIN BLEACHING

[0074] The enzyme tyrosinase catalyses the reactions leading to melanin biosynthesis from tyrosine. L-tyrosine is oxidised to 3,4 dihydroxyphenylalanine (L-DOPA), through the action of the enzyme tyrosinase, and again by means of a reaction catalysed by the same enzyme, 3,4 dihydroxyphenylalanine is converted to DOPAquinone. Substances inhibiting tyrosinase activity produce a clearing/bleaching effect on the skin, since they block melanin synthesis. The method used to determine the inhibition of tyrosinase activity in the oxidation reaction of L-tyrosine to L-DOPA is that described in Kim et al. (4,4'-Dihydroxybiphenyl as a new potent tyrosinase inhibitor. Biol. Pharm. Bull. 28(2), 323-327, 2005). The method applied for determination of the inhibition of tyrosinase activity in the conversion of L-DOPA to DOPAquinone is that described in Masamoto et al. (Mushroom tyrosinase inhibitory activity of esculetin isolated from seeds of Euphorbia lathyris L. Biosci. Biotechnol. Biochem., 67(3), 631-634, 2003). The results obtained from the inhibition assay of tyrosinase activity by *Syringa vulgaris* total extracts, verbascoside and isoverbascoside in comparison to resorcinol and kojic acid, are summarised in tables 8 and 9.

Table 8: Percentage tyrosinase enzyme activity inhibition in the oxidation of L-tyrosine to L-DOPA, by *Syringa vulgaris* extracts, verbascoside and isoverbascoside in comparison to resorcinol and kojic acid.

Compound	concentration	Test L-tyrosine-DOPA inhibitory activity (%)
Resorcinol	250 µg/ml	56.9
Kojic acid	250 µg/ml	94.1
Extract 1	250 µg/ml	29

(continued)

Compound	concentration	Test L-tyrosine-DOPA inhibitory activity (%)
Extract 2	250 µg/ml	31.3
Verbascoside	250 µg/ml	19,6
Isoverbascoside	250 µg/ml	10.9

As may be observed from the data summarised in table 8, *Syringa vulgaris* Extracts 1 and 2, verbascoside and isoverbascoside display inhibitory activity towards the enzyme tyrosinase.

Table 9: Percentage tyrosinase enzyme activity inhibition in the conversion of L-DOPA to L-DOPAquinone, by *Syringa vulgaris* extracts, verbascoside and isoverbascoside in comparison to resorcinol and kojic acid.

Compound	concentration	Test-LDOPA-DOPAquinone inhibitory activity (%)
Resorcinol	100 µg/ml	0
Kojic acid	100 µg/ml	35
Extract 1	100 µg/ml	40.4
Extract 2	100 µg/ml	35.4
Verbascoside	100 µg/ml	20,6
Isoverbascoside	100 µg/ml	10

Table 9 highlights the high inhibitory capacity of Extracts 1 and 2, verbascoside and isoverbascoside towards the enzyme tyrosinase. The enzyme inhibitory activities of the *Syringa vulgaris* extracts 1 and 2 are higher with respect to those of the reference molecules resorcinol and kojic acid.

EXAMPLE 10

OXIDATIVE STRESS ON HUMAN PULMONARY ENDOTHELIAL CELL (A549) CULTURES

[0075] In the following example, reference is made to certain substances, herein reported using the corresponding abbreviations:

BSO buthionine [S,R] sulfoximine
 GSH reduced glutathione
 DCF Dichlorofluorescein
 DCF-DA Dichlorofluorescein diacetate
 FCS foetal calf serum
 MTT 3-(4,5-dimethylthiazol)-2,5-diphenyl tetrazol bromide
 ROS Reactive Oxygen Species

[0076] 10x10⁵ A549 cells (ATCC-CCL185) have been seeded into wells with 3 ml of DMEM/ medium (HAM'S F12K) with 10% FCS and incubated at 37°C in a 5% CO₂, water vapour saturated atmosphere. After 24 hours the culture medium is replaced by medium supplemented with 5.55 mM galactose, which is left in contact with the cells for 20 hours. The medium is once again replaced with medium containing galactose in the presence or absence of the test compound. After 5 hours, quantitative measurement of the ROSs is performed by washing the cells with phosphate buffer and then incubating them with 30µM DCF-DA in buffer solution. At the end of a 15 minute incubation, excess DCF-DA is removed from the cells by washing, prior to measuring the fluorescence emitted.

Parameters

Fluorimetry.

[0077] Antioxidant activity measurements are performed using a (Hitachi F3010) spectrophotometer with excitation at 488 nm and measuring emission at 525 nm, and values are expressed in pmoles, by extrapolation from the calibration curve constructed using known quantities of DCF, in relation to total cellular protein. Values are thus expressed in %ages with respect to the ROS content of the cells not subjected to oxidative stress, to which are assigned a value of 100.

Sample preparation

[0078] *Syringa vulgaris* Extracts 1 and 2 and ascorbic acid have been dissolved in phosphate buffer.

Results

[0079] Extracts 1 and 2 significantly reduce ROS concentrations in a dose-dependent manner and are more potent than ascorbic acid, as demonstrated in Table 10.

Table 10. The antioxidant effect of *Syringa vulgaris* Extracts 1 and 2

Treatment	Concentration	% ROS in picomoles
Oxidative stress		100.02 ± 3.11
Extract 1	20mM	67.74 ± 3.81*
Extract 1	30mM	57.21 ± 2.77*
Extract 2	20mM	65.83 ± 5.68*
Extract 2	30mM	57.23 ± 2.31*
Ascorbic acid	1mM	62.91 ± 3.82*
Verbascoside	20mM	85 ± 7*
- Mean ± SD (n=6); *p>0.05 vs. BSO. Student- Newman- Keuls test		

[0080] In the different percentage proportions of verbascoside, Extracts 1 and 2 show overlapping biological activities, markedly superior to ascorbic acid without displaying any peroxidising activity, frequently encountered with low concentrations (<100 µM) of ascorbic acid. Furthermore, the biological activity is unexpectedly greater than pure verbascoside.

EXAMPLE 11

THE PROTECTIVE EFFECT OF SYRINGA VULGARIS EXTRACTS AGAINST OXIDATIVE STRESS IN PC12 CELLS

[0081] This assay has the aim of determining the protective capacity, of *Syringa vulgaris* extracts, in relation to oxidative stress induced cell death in rat pheochromocytoma cell cultures (PC12 cell line).

[0082] The assay has been performed as described in the literature (Echevery C. et al. Cytoprotection by neutral fraction of tannat red wine against oxidative stress-induced cell death. J. Agric. Food Chem., 2004, 52(24): 7395-9. Ochiai T. et al. Crocin prevents the death of rat pheo-chromocytoma (PC12) cells by its antioxidant effects stronger than those of alpha-tocopherol. Neurosci. Lett., 2004, 362(1):61-4).

[0083] The results obtained are summarised in table 11, and are expressed as the ratio between the numbers of viable cells after 72 hours in the samples with respect to control.

TABLE 11: Increase in PC12 viability with respect to control.

Sample	Concentration	Ratio of viable treated cells over control
Control	0 ug/ml	1
Extract 1	100 µg/ml	10
Extract 1	500 µg/ml	30

(continued)

Sample	Concentration	Ratio of viable treated cells over control
Extract 1	1000 µg/ml	35
Extract 2	100 µg/ml	15
Extract 2	500 µg/ml	25
Extract 2	1000 µg/ml	37
Isoverbascoside	100 µg/ml	4
Isoverbascoside	500 µg/ml	10
Isoverbascoside	1000 µg/ml	10

The results obtained indicate that *Syringa vulgaris* Extracts 1 and 2 display higher protective activity and survival in relation to oxidative stress in PC12 cells, since the quantity of viable cells after a 72 hour assay is up to 37 fold higher than the viable cells in the control. This activity is much more evident with Extracts 1 and 2 in comparison to pure isoverbascoside.

EXAMPLE 12

ANTIPROLIFERATIVE ACTIVITY OF SYRINGA VULGARIS EXTRACTS

[0084] The antiproliferative activity of the *Syringa vulgaris* extracts has been assayed, using an *in vitro* model, envisaging the use of a stabilised human melanoma cell line (A375). The above cell lines, provided by the American Type Culture Collection, Rockville, MD (USA), have been maintained in culture at 37°C and 5% CO₂, in a defined nutrient medium (Dulbecco's Modified Eagle's Medium (DMEM)) supplemented with 10% foetal calf serum (FCS). The *Syringa vulgaris* extracts have been diluted in culture medium, so as to obtain the minimum possible dilution at which no toxic effects are observed. Each extract diluted in medium has been sterilised by filtration (using 0.2 µm filters). The assay consists of two experimental stages: A cytotoxicity test and an antiproliferative test. Such assays have been performed as described in the literature (Niles, RM et al. Resveratrol is a potent inducer of apoptosis in human melanoma cells. Cancer Lett., 2003, 190(2): 157-63; Wong D. et al. Cytotoxic effects of mansone E and F isolated from Ulmus pumula. Biol. Pharm. Bull., 2004, 27(7): 1025-30) .

The data obtained from the above assay are summarised in tables 12 and 13.

Table 12: cytotoxic and antiproliferative activities of *Syringa vulgaris* Extract 1 on A375 melanoma cells

	h	conc ug/mL Estratto1						IC50
		1	10	50	100		1000	
citotoc	24	0	0	0	0	0	0	
	48	0	0	0	0	0	0	
	72	0	0	0	0	0	80%	
antiprolif	24	0	-10%	0	0	-25%	-75%	529
	48	0	0	0	0	-65%	-90%	366
	72	-8%	-15%	-10%	-15%	-70%	-90%	200

Table 13: cytotoxic and antiproliferative activities of

	h	conc ug/mL Estratto 2						IC50
		1	10	50	100	500	1000	ug/mL
citotox	24	0	0	0	0	0	0	
	48	0	0	0	0	0	0	
	72	0	0	0	0	0	30%	
antiprolif	24	0	0	0	0%	-30%	-70%	515
	48	0	0	0	0%	-50%	-90%	493
	72	0	-5%	-15%	-25%	-70%	-95%	220

Syringa vulgaris Extract 2 on A375 melanoma cells

[0085] The results obtained indicate that *Syringa vulgaris* Extracts 1 and 2 display a significant anti-proliferative effect towards A375 melanoma cells.

EXAMPLE 13

ANTIMYCOTIC ACTIVITY OF SYRINGA VULGARIS EXTRACTS, VERBASCOSIDE AND ISOVERBASCOSIDE TOWARDS STRAINS OF MALASSEZIA FURFUR AND TRICOPHYTON RUBRUM

[0086] *Tricophyton rubrum* and *Malassezia furfur* are fungal microorganisms responsible for giving rise to a number of skin diseases. Particularly, *Malassezia furfur* is the etiological agent responsible for seborrhoeic dermatitis and Pityriasis versicolor (Eichstedt's disease), while *Tricophyton rubrum* is the agent responsible for onychomycosis, athletes foot, tinea barbae (beard ringworm) and tinea corporis (body ringworm).

[0087] With the aim of identifying any potential antifungal activity in the *Syringa vulgaris* extracts, microbiological assays have been prepared, generally known as multiple dilution assays. Such assays have been performed using the following fungal strains:

Tricophyton rubrum ATCC 28188 and *Malassezia furfur* ST 8036.

Syringa vulgaris Extracts 1 and 2, verbascoside and isoverbascoside have been assayed at concentrations of 0.625 mg/ml, 1.25 mg/ml, 2.5 mg/ml and 5mg/ml. The fungal strains used for testing have been inoculated into culture-broth (in Tryptone soy broth medium, for *Malassezia furfur* the culture medium has been supplemented with 2 ml/100ml of olive oil) containing *Syringa vulgaris* extracts 1 and 2, verbascoside and isoverbascoside at the concentrations described above. In parallel with these tests, culture broths containing the microbial strain alone have been prepared (controls). The culture-broths have been incubated at 30°C for a period of 28 days. Samples of the various culture-broths have been taken at 0, 7, 14, 21 and 28 days, with the aim of counting the number of CFU/ml in relation to the various fungal strains, by plating the cultures on solid medium. A certain number of serial dilutions have been prepared for each sample in order to determine the fungal load of each culture-broth. Each dilution has been assayed in triplicate on solid medium. Then number of CFUs reported in the table is the mean of the counts performed on the triplicate assays. The results obtained are reported in the following table.

Table 14: the effect of *Syringa vulgaris* Extracts 1 and 2 on the growth of *Tricophyton rubrum* ATCC 28188

Samples	TO (CFU/ml)	T7d (CFU/ml)	T14d (CFU/ml)	T21d (CFU/ml)	T28d (CFU/ml)
Control	9.8 x 10 ⁴	7.1 X 10 ⁵	9.2 X 10 ⁵	1.8 X 10 ⁵	1.5 X 10 ⁵
Extract 1 (5mg/ml)	8.5 X 10 ⁴	<10	<10	<10	<10
Extract 1 (2.5mg/ml)	8.3 X 10 ⁴	<10	<10	<10	<10
Extract 1 (1.25mg/ml)	7.7 X 10 ⁴	<10	<10	<10	<10
Extract 1 (0.625mg/ml)	7.2 X 10 ⁴	1.3 X 10 ³	3.3 X 10 ⁵	3.1 X 10 ⁵	3.3 X 10 ⁵
Extract 2 (5mg/ml)	6.8 X 10 ⁴	<10	<10	<10	<10

(continued)

Samples	TO (CFU/ml)	T7d (CFU/ml)	T14d (CFU/ml)	T21d (CFU/ml)	T28d (CFU/ml)
Extract 2 (2.5mg/ml)	8×10^4	5.8×10^2	5.2×10^4	3.5×10^4	2.1×10^4
Extract 2 (1.25mg/ml)	9×10^4	2.4×10^3	2×10^5	5.5×10^4	3.5×10^4
Extract 2 (0.625mg/ml)	6.1×10^4	6.6×10^3	2.7×10^5	2.3×10^5	1.2×10^5

Table 15: the effect of *Syringa vulgaris* Extracts 1 and 2 on the growth of *Malassezia furfur* ST 8036

Samples	T0 (CFU/ml)	T7d (CFU/ml)	T14d(CFU/ml)	T21d (CFU/ml)	T28d (CFU/ml)
Control	3.9×10^3	1.1×10^5	2.7×10^5	3.8×10^4	3.6×10^4
Extract 1 (5mg/ml)	2.7×10^3	<10	<10	<10	<10
Extract 1 (2.5mg/ml)	2.8×10^3	<10	<10	<10	<10
Estratto 1 (1,25mg/ml)	3.2×10^3	<10	<10	<10	<10
Extract 1 (0.625mg/ml)	2.9×10^3	<10	<10	<10	<10
Extract 2 (5mg/ml)	3×10^3	<10	<10	<10	<10
Extract 2 (2.5mg/ml)	2.6×10^3	<10	<10	<10	<10
Extract 2 (1.25mg/ml)	2.2×10^3	<10	<10	<10	<10
Extract 2 (0.625mg/ml)	3.9×10^3	1.7×10^4	6.1×10^4	3.5×10^4	3.1×10^4

>From the results obtained it is clear that both Extract 1 and Extract 2 display significant inhibitory activity towards the growth of the two dermatophytes assayed.

In particular, Extract 1 shows fungicidal activity towards *Trycophyton rubrum* at a concentration as low as 1.25 mg/ml, while Extract 2 displays analogous activity at a concentration of 5 mg/ml.

Extract 1 displays fungicidal activity towards *Malassezia furfur* at a concentration as low as 0.625 mg/ml, while Extract 2 displays such activity at a concentration of 1.25 mg/ml.

Verbascoside and isoverbascoside exhibited an activity comparable to Extract 1.

Preferably, the verbascoside and isoverbascoside of the invention have a purity of about 99%.

With regard to their potent effect on oxidative stress, as described in examples 4, 10 and 11, the *Syringa vulgaris* extracts forming the subject of the invention are valid tools in the prevention and treatment of disease states associated with damage due to free radicals such as:

- anoxia, trauma and inflammatory based diseases of the nervous system such as cerebral stroke, cerebral and spinal trauma, haemorrhagic shock, epilepsy, cerebral ischemia and ischemic spinal injury, peripheral neuropathy and cephalalgia (B. Halliwell et al, 1992); degenerative type pathologies of the nervous system also associated with ageing or autoimmune based or having inflammatory components such as amyotrophic lateral sclerosis, multiple sclerosis, Parkinson's disease, Alzheimer's disease, senile dementia and Huntington's disease (Olanow C.W. "A Radical Hypothesis for Neurodegeneration", TINS, 1993, 16(11):439-444; Dib M. Amyotrophic lateral sclerosis: progress and prospects for treatment. Drugs 2003; 63(3): 289-310);
- reperfusion damage and cardiovascular diseases including angina, percutaneous transluminal coronary angioplasty (PTCA), myocardial infarction and vasculopathies (Bagchi D, Sen CK, Ray SD, Das DK, Bagchi M, Preuss HG, Vinson JA. Molecular mechanisms of cardioprotection by a novel grape seed proanthocyanidin extract. Mutat. Res. 2003 Feb-Mar; 523-524: 87-97);
- atherosclerosis and muscular damage (Mertens A, Verhamme P, Bielicki JK, Phillips MC, Quarck R, Verreth W, Stengel D, Ninio E, Navab M, Mackness B, Mackness M, Holvoet P. Increased low-density lipoprotein oxidation and impaired high-density lipoprotein antioxidant defence are associated with increased macrophage homing and atherosclerosis in dyslipidemic obese mice: LCAT gene transfer decreases atherosclerosis. Circulation 2003 Apr 1; 107(12): 1640-6.);
- dimetabolic disorders such as diabetes mellitus and the neurological, vascular and ophthalmic complications thereof (Yoshida K., Hirokawa J., Tagami S., Kawakami Y., Urata Y., Kondo T. "Weakened cellular scavenging activity against oxidative stress in diabetes mellitus: regulation of glutathione synthesis and efflux" Diabetologia, 1995 Feb.,

38(2): 201-210);

- toxic disorders such as alcoholism and the neurological complications thereof besides cirrhosis of the liver (Clot P., Tabone M., Arico S., Albano E. "Monitoring oxidative damage in patients with liver cirrhosis and different daily alcohol intake" *Glut.*, 1994 Nov., 35(11):1637-43);
- peripheral neuropathies including toxic neuropathies (Leonetti C, Biroccio A, Gabellini C, Scarsella M, Maresca V, Flori E, Bove L, Pace A, Stoppacciaro A, Zupi G, Cognetti F, Picardo M. Alpha-tocopherol protects against cisplatin-induced toxicity without interfering with antitumour efficacy. *Int. J. Cancer* 2003 Mar 20; 104(2): 243-50);
- skin and mucosal disorders such as photolysis, actinic damage, skin ageing, atopy, diseases associated with keratinocyte hyperproliferation for example psoriasis, xeroderma, skin tumours (Mittal A, Elmetts CA, Katiyar SK CD11b+ cells are the major source of oxidative stress in UV radiation-irradiated skin: possible role in photo-ageing and photo-carcinogenesis. *Photochem. Photobiol.* 2003 Mar; 77(3): 259-64; Pinnell SR. Cutaneous photo-damage, oxidative stress, and topical antioxidant protection. *J. Am. Acad. Dermatol.* 2003 Jan; 48(1): 1-19; Tsukahara H, Shibata R, Ohshima Y, Todoroki Y, Sato S, Ohta N, Hiraoka M, Yoshida A, Nishima S, Mayumi M. Oxidative stress and altered antioxidant defences in children with acute exacerbation of atopic dermatitis. *Life Sci.* 2003 Apr 18; 72(22): 2509-16);
- dental disorders including diseases of bacterial origin, gingivitis; viral infections such as HIV (Greenspan and Aruoma, 1994 ref. cit.); Herpes Zoster, Herpes Simplex and Cytomegalovirus;
- disorders of the respiratory apparatus such as cystic fibrosis, asthma, rhinitis (including allergic), "respiratory distress" and neonatal pulmonary hypertension (Fortoul TI, Valverde M, Lopez Mdel C, Lopez I, Sanchez I, Avila-Costa MR, Avila-Casado Mdel C, Mussali-Galante P, Soria E, Rojas E. Nasal cytology and genotoxic damage in nasal epithelium and leukocytes: asthmatics versus non-asthmatics.. *Int. Arch. Allergy Immunol.* 2003 Mar; 130(3): 232-5);
- articular disorders including rheumatoid arthritis (Lee SH, Chang DK, Goel A, Boland CR, Bugbee W, Boyle DL, Firestein GS. Microsatellite instability and suppressed DNA repair enzyme expression in rheumatoid arthritis. *J. Immunol.* 2003 Feb 15; 170(4): 2214-20);
- tumour pathologies; ophthalmic disorders for example cataract, retinal degeneration (Organisciak DT, Darrow RM, Barsalou L, Kutty RK, Wiggert Susceptibility to retinal light damage in transgenic rats with rhodopsin mutations. *Invest. Ophthalmol. Vis. Sci.* 2003 Feb; 44(2): 486-92);
- hearing apparatus damage (Shi X, Nuttall AL. Up-regulated iNOS and oxidative damage to the cochlear stria vascularis due to noise stress. *Brain Res.* 2003 Mar 28; 967(1-2): 1-10);
- diminished immune response (Halliwell B et al., 1992 ref. cit.; Toyokuni S. et al. "Persistent oxidative stress in cancer" *FEBS Lett.*, 1995 Jan. 16, 358(1): 1-3; Ames B.N., Shigenaga M.K.) gastric lesions (Yajima N., Hiraishi H., Harada T., "Protection of Cultured Rat gastric cells Against Oxidant Stress by Iron Chelation" *Digestive Diseases and Sciences* 1995, 40(4): 879-886).

[0088] Furthermore, in relation to their potent effectiveness against platelet aggregation as described in example 6, the Extracts of the invention, verbascoside and isoverbascoside are valid tools in the prevention and treatment of disease states associated with the formation of platelet clots and atheromatous plaques, besides finding use for the preservation and stabilisation of purified platelets. Due to the marked anti-inflammatory activities described in example 5, antifungal activities (described in example 13) and antibacterial activities (Didry N. Isolation and antibacterial activity of phenylpropanoid derivatives from *Ballota nigra*. *J. Ethnopharmacol.* 1999, 67(2): 197-202), the *Syringa vulgaris* extracts of the invention, verbascoside and isoverbascoside are valid tools in the prevention and treatment of pathological states associated with lesions of the epidermis and skin caused by burns, cuts or lesions and mechanical trauma, diabetic sores and bedsores. Furthermore, due to the aforementioned biological activities, the extracts, verbascoside and isoverbascoside may be used as promoters of the tissue repair processes in cases of mucosal and endothelial lesions and ulcers, including those of the gastrointestinal tract and vaginal mucosa.

[0089] As a result of the high chelating activity shown in example 7, the extracts, verbascoside and isoverbascoside may be used for the treatment of numerous allergy-related pathologies, both systemic and cutaneous (for example: allergic skin reaction to Ni²⁺ in topical cosmetic and pharmaceutical products). Said activity may be exploited in order to protect the body from harm caused by the presence of excess heavy metals, such as iron and copper, for example following thalassemia. A combination of radical scavenging, anti-oxidant and chelating activities, may form the basis for the use of *Syringa vulgaris* extracts, verbascoside and isoverbascoside against disease processes caused by free radicals generated by the presence of excess heavy metals.

[0090] The *Syringa vulgaris* extracts, verbascoside and isoverbascoside possess high inhibitory capacity in relation to the activities of the enzymes 5 alpha reductase, both type I and type II, and lipoxygenase, as shown in example 8. This inhibitory capacity may be exploited for the treatment and prevention of hair loss in cases of androgenetic alopecia (male pattern baldness) and alopecia areata (patchy hair loss). Furthermore, this activity may be exploited for the treatment of juvenile acne and in subjects affected by prostate diseases dependent on the activation of 5 alpha reductase.

[0091] As reported in example 9, *Syringa vulgaris* extracts, verbascoside and isoverbascoside demonstrate high

inhibitory activity- in relation to the enzyme tyrosinase. This activity may be exploited in order to bring about bleaching effects on the skin.

[0092] As shown in example 13, total extracts, verbascoside and isoverbascoside display fungicidal activity in relation to two dermatophytic strains: *Tricophyton rubrum* and *Malassezia furfur*. This characteristic permits the use of such products in the topical treatment of dermatitis, particularly seborrhoeic dermatitis, onychomycosis, Pityriasis versicolor (Eichstedt's disease), athletes foot, tinea barbae (beard ringworm) and tinea corporis (body ringworm).

[0093] The doses, times and routes of administration of the treatment will be selected on the basis of the type, stage, severity and location of manifestation of the pathology. For all the pathologies mentioned, systemic and oral administration are indicated, but additionally also topical and transdermal, and in any case so as to make the active ingredient maximally available. For the oral formulations, administration in the form of tablets, lozenges and capsules, but also powders and suspensions are preferred: for topical treatment, gels, creams, ointments and solutions compatible with dermal and mucosal use are preferred, in addition to eye washes for administration into the conjunctival sac. The injectable forms are formulated using solvents for pharmaceutical use, compatible with intravenous, intramuscular and subcutaneous administration.

[0094] The therapeutic dose varies, depending on the patients age, weight, sex and type of pathology, between 0.1 mg and 2 g per day and preferably between 5 and 150 mg per day, taken in a single dose or in 2-4 doses or in slow release form, depending on the patients therapeutic need, and for periods of time varying between 1 and 120 days.

[0095] At appropriate concentrations, the same extracts, verbascoside and isoverbascoside may be formulated in the form of supplements, to be taken orally for prevention, or as aids- in the treatment of changes resulting from disreactive states in humans or in veterinary medicine. Total extracts, verbascoside and isoverbascoside active at the appropriate concentrations, and in appropriate formulations, may be likewise used in cosmetics and trichological treatments.

[0096] Thanks to the absorbance spectrum of the phenylpropanoids, which ranges from 240 nm to 350nm (UVB absorption range), such substances may be used for the preparation of sun filters.

[0097] Some examples of pharmaceutical, cosmetic and supplement preparations are reported in the following, for the purposes of non-limiting illustration:

Example 1: Capsules for oral use

[0098]

Active ingredient: Extract titrated in verbascoside as 500 mg

Excipients: Carbowax 6000, gelatin, distilled water, E172, Titanium dioxide

Example 2:

[0099]

Active ingredient: Extract titrated in verbascoside as 10 mg

Excipients: pH 7 buffered injectable solution, sufficient for 2ml

Example 3: 0.2% emulsion for topical application

[0100]

Active ingredient: Extract titrated in verbascoside as 200 mg

Excipients: Synthetic glycerides, anionic emulsifying wax, anhydrous lanolin, polysorbate 80, dimethylpolyoxane 100, dimethylpolyoxane 500, methyl-p-hydroxybenzoate, propyl-p- hydroxybenzoate, lemon essence, sufficient water to make up to 100g

Example of an oral preparation:

Extract: 10 mg

[0101] Magnesium, Selenium, Folic acid, Mannitol, sodium saccharin, citric acid, lemon flavouring.

[0102] The *Syringa vulgaris* extracts, verbascoside and isoverbascoside may be formulated for use in the veterinary sector and as dietary supplements for zootechnical use, such as, as a non-limiting example, for the raising of fowl and fish. From the above description, it is clear that the extracts for use in the preparation of drugs, cosmetics or nutritional substances advantageously allow the provision of compounds capable of having unexpectedly greater antioxidant ac-

tivity, in some cases, with respect to any individual phenylpropanoid.

[0103] In addition, said extracts, verbascoside and isoverbascoside may be advantageously used as wound-healing agents.

Claims

1. A process for the preparation of a plant cell line extract from plants, wherein said plant cell line is the *Syringa vulgaris* IRB-SV25/B (DSM:16857) cell line, comprising the following stages in sequence:

a) subjecting the biomass obtained following culture of said *Syringa vulgaris* cell line to an enzyme deactivating treatment by thermal treatment, wherein the thermal treatment takes place for a period of time of less than 5 minutes or comprised between 5 and 30 minutes, and at a temperature comprised between 50°C and 150°C in such a manner as to isomerise from 40% to 60% of the verbascoside initially present in the biomass into isoverbascoside;

b) homogenising the product obtained following step a);

c) separating the aqueous phase from the cell residues by means of gravity filtration, pressure filtration or centrifugation;

d) performing an extraction of the aqueous phase obtained following step c) with hydrophobic interaction resin;

e) recovering the absorbed substances (extract) from the resin by means of elution with a mixture of alcohols having up to 5 carbon atoms, in water in percentages varying from 30% up to just under 100% by volume.

2. The process according to claim 1 additionally comprising, between steps a) and b), a preceding biomass filtration step performed under the same conditions reported in step c), with the aim of recovering just the biomass, without the fermenter culture medium.

3. The process according to claims 1 or 2, wherein said filtration stage c) and said prior filtration occur through nylon, steel or cotton mesh or the like having porosity comprised between 10 µm and 300 µm.

4. The process according to anyone of claims 1 to 3, wherein the thermal treatment takes place with the use of steam.

5. The process according to any of the claims 1 to 4, wherein the extraction step d) occurs by chromatography on a column or in batch.

6. The process according to any of the claims 1 to 5, wherein the recovery step e) of the substances absorbed onto the resin occurs with the use of a mixture of ethanol in water.

7. The process according to any of the claims 1 to 6, wherein step a) is performed when the cells have an age comprised between 7 and 21 days.

8. The process according to any of the claims 1 to 7, wherein step d) is performed using a resin selected from polystyrene-divinylbenzene or acrylic matrix resins.

9. The process according to any of the claims 1 to 8, wherein the *Syringa vulgaris* IRB-SV25/B (DSM: 16857) cell lines are first stabilised and selected in accordance with a procedure comprising:

a) cultivating a *Syringa vulgaris* IRB-SV25/B (DSM: 16857) cell line, stabilised for a sufficient period of time to allow the multiplication of said cell line to give substantially distinct cell clusters, on solid medium;

b) removing said substantially distinct cell clusters from said solid medium and placing each of them in a separate liquid culture medium;

c) cultivating each of the said substantially distinct cell clusters in said liquid culture medium for a sufficient period of time to allow the multiplication of said cell clusters, and the analytical determination of the primary and/or secondary metabolites produced thereby;

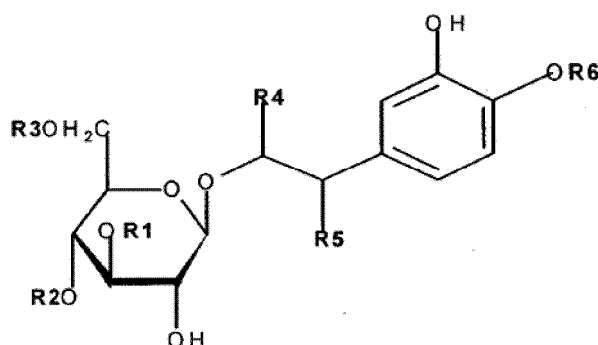
d) performing a qualitative and quantitative determination of the primary and/or secondary metabolites produced by each of said cell clusters in said liquid culture medium;

e) selecting the cell cluster capable of producing the greatest quantity of said metabolite of interest;

f) repeating the process cycle according to stages a), b), c), d) and e) on said cell cluster, selected according to stage e), a sufficient number of times until the quantity of said metabolite of interest produced by the selected

cell cluster, and by the cell clusters deriving from further selection cycles, is essentially constant.

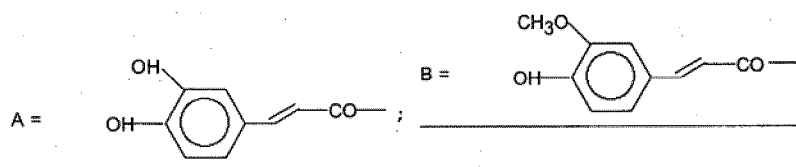
10. The process according to claim 9, wherein following the stabilisation and selection stage and prior to extraction, the *Syringa vulgaris* IRB-SV25/B (DSM: 16857) cell line is subjected to fermentation.
11. The process according to anyone of claims 1 to 10, wherein the thermal treatment stage occurs at a temperature of between 60°C and 120°C.
12. A *Syringa vulgaris* IRB-SV25/B (DSM: 16857) cell line extract obtainable according to the process any of the claims 1 to 11.
13. The extract according to claim 12, comprising from 20% to 90% by weight phenylpropanoids and from 80 to 10% of a chromophore-free fraction, mainly, but not exclusively comprising, oligo- and polysaccharide, protein and lipid molecules.
14. The extract according to claim 12, comprising between 30% and 60% by weight phenylpropanoids and between 70% and 40% of said chromophore-free fraction.
15. The extract according to any of the claims 12 to 14, wherein the phenylpropanoids comprise between 5% and 20% isoverbascoside with respect to the total mass of phenylpropanoids.
16. The extract according to any of the claims 12 to 14, when ancillary to claim 7, wherein the phenylpropanoids comprise between 45% and 65% isoverbascoside with respect to the total mass of phenylpropanoids.
17. The extract according to any of the claims 12 to 14, wherein the phenylpropanoids are represented by the following general formula (I)



wherein:

- R1 is a hydrogen atom or a monosaccharide with 5 or 6 carbon atoms;
- R2 is a hydrogen atom or is a caffeoyl (A) or feruloyl (B) group;
- R3 is a hydrogen atom or a monosaccharide with 5 or 6 carbon atoms or a disaccharide with 10 to 12 carbon atoms or is a caffeoyl (A) or feruloyl (B) group;
- R4 is a hydrogen atom or an alkyl group, or a hydroxyl group;
- R5 is a hydrogen atom or an alkyl group, or a hydroxyl group, either free or condensed with the alcohol group at position C2 of the monosaccharide group bound to it, in order to form a 1,4 dioxane ring;
- R6 is a hydrogen atom or an alkyl group,

when R3 is a caffeoyl (A) or feruloyl (B) group,



then R2 is always a hydrogen atom and *vice versa*.

10 18. The extract according to claim 17, wherein R1 is a ramnhose.

19. The extract according to claim 17 or 18, wherein R4 is a methyl.

20. The extract according to anyone of claims 17-19, wherein R5 is a methyl.

15 21. The extract according to anyone of claims 17-20, wherein R6 is a methyl.

22. The extract according to any of the claims 12 to 21 for medicinal and nutritional use.

20 23. The extract according to any of the claims 12 to 21 for use as an anti-oxidant, antiplatelet aggregating factor.

24. The extract according to any of the claims 12 to 21 for use as a platelet disaggregating factor.

25 25. Use of the extract as specified according to any of the claims 12 to 21 for the preparation of a medication having metal ion chelating activity for topical and systemic use.

26. The use of the extract as specified according to any of the claims 12 to 21 for the preparation of a medication having anti-alpha-reductase and antilipoxygenase activities for the treatment of juvenile acne and for the prevention of hair loss and in the treatment of diseases of the prostate.

30 27. The use of the extract as specified according to any of the claims 12 to 21 for the preparation of a medication having anti-tyrosinase activity for skin bleaching treatments.

35 28. The use of the extract as specified according to any of the claims 12 to 21 for the preparation of a medication having anti-proliferative activity towards cells.

29. The use of the extract as specified according to any of the claims 12 to 21 for the preparation of a medication having antimicotic activity.

40 30. The use of the extract according to any of the claims 12 to 21 for the preparation of a medication for the treatment of anoxic, trauma and inflammatory based disorders of the nervous system selected from the group of cerebral stroke, cerebral and spinal trauma, haemorrhagic shock, epilepsy, cerebral ischemia and ischemic spinal injury, peripheral neuropathy and cephalgia.

45 31. The use of the extract according to any of the claims 12 to 21 for the preparation of a medication for the treatment of degenerative disorders of the nervous system selected from the group of disorders linked to ageing or based on autoimmune and inflammatory components selected from the group of amyotrophic lateral sclerosis, multiple sclerosis, Parkinson's disease, Huntington's disease, Alzheimer's disease, dementia or behavioural disorders selected from the group of schizophrenia and depression.

50 32. The use of the extract according to any of the claims 12 to 21 for the preparation of a medication for the treatment of damage due to reperfusion and cardiovascular diseases including angina, percutaneous transluminal coronary angioplasty (PTCA), myocardial infarction and vasculopathies, atherosclerosis and muscular damage.

55 33. Use of the extract according to any of the claims 12 to 21 for the preparation of a medication for the treatment or prevention of platelet aggregation in thrombotic diseases, and for the treatment and storage of purified platelets.

34. The use of the extract according to any of the claims 12 to 21 for the preparation of a medication for the treatment

of dismetabolic disorders selected from the group of diabetes mellitus and the neurological, vascular and ophthalmic complications thereof.

- 5 35. The use of the extract according to any of the claims 12 to 21 for the preparation of a medication for the treatment of toxic disorders selected from the group of alcoholism and the neurological complications thereof and liver cirrhosis.
36. The use of the extract according to any of the claims 12 to 21 for the preparation of a medication for the treatment of peripheral neuropathic disorders including toxic neuropathies.
- 10 37. The use of the extract according to any of the claims 12 to 21 for the preparation of -a medication for the treatment of skin and mucosal diseases comprising photolysis, actinic damage, atopy, diseases associated with keratinocyte hyperproliferation selected from the group of psoriasis, xeroderma, skin tumours, seborrheic dermatitis, atopic dermatitis; skin lesions selected from the group of burns, abrasions, cuts, scars, surgical wounds and bedsores, as well as ulcers and lesions of the mucosa including the oral, gastric and intestinal mucosa and the vaginal mucosa.
- 15 38. The use of the extract according to any of the- claims 12 to 21 for the preparation of a medication for the treatment of viral infections selected from HIV, Herpes zoster, Herpes simplex and cytomegalovirus and bacterial infections including dental disorders.
- 20 39. The use of the extract according to any of the claims 12 to 21 for the preparation of a medication for the treatment of disorders of the respiratory apparatus selected from the group of cystic fibrosis, asthma, rhinitis including due to allergies, "respiratory distress" and neonatal pulmonary hypertension.
- 25 40. The use of the extract according to any of the claims 12 to 21 for the preparation of a medication for the treatment of articular disorders including rheumatoid arthritis and psoriatic arthritis.
41. The use of the extract -according to any of the claims 12 to 21 for the preparation of a medication for the treatment of tumors and disorders associated with a diminished immune response.
- 30 42. The use of the extract according to any of the claims 12 to 21 for the preparation of a medication for the treatment of sensory apparatus disorders selected from the group of cataract, retinal degeneration and hearing apparatus damage.
- 35 43. A formulation comprising the extract according to any of the claims 12 to 21 as active ingredient in association with pharmaceutically, nutritionally or cosmetically acceptable excipients for human and veterinary use.
44. The formulation according to claim 43 for systemic, parenteral, oral, rectal, inhalatory, topical, transdermal, intravenous, intramuscular or subcutaneous administration.
- 40 45. The formulation according to claim 43, wherein the form for oral administration is represented by tablets, lozenges, capsules, powders, solutions, suspensions or a nebulised form.
46. The formulations according to claim 43, wherein the form for administration by inhalation is represented by powders, solutions and suspensions.
- 45 47. The formulations according to claim 43, wherein the form for topical administration is an emulsion, gel or compatible solutions for dermal and mucosal use and eye washes.
48. The use of the extract according to any of the claims 12 to 21 for the preparation of a medication for veterinary care or a product for zootechnical feeds.
- 50 49. The use of the extract according to any of the claims 12 to 21, wherein the doses of extract per treatment cycle are comprised of between 0.1 and 100 mg/kg for 1-100 days.
- 55 50. Use of a *Syringa vulgaris* IRB-SV25/B (DMS:16857) cell line extract comprising between 20% and 90% by weight, over the total dry weight of said extract, of phenylpropanoids and between 80% and 10% of a chromophore-free fraction as a cosmetic product.

51. Use of a *Syringa vulgaris* IRB-SV25/B (DMS:16857) line extract comprising between 20% and 90% by weight, over the total dry weight of said extract, of phenylpropanoids and between 80% and 10% of a chromophore-free fraction as a nutritional product.

52. The use of the extract according to any of the claims 12 to 21 for the preparation of sun filters.

53. The use of the extract according to any of the claims 12 to 21 for the cosmetic non-therapeutical treatment of skin ageing.

54. The *Syringa vulgaris* plant cell line denominated IRB-SV25/B and having DSMZ accession number: DSM 16857.

Patentansprüche

1. Verfahren zum Herstellen eines Pflanzenzelllinienextrakts aus Pflanzen, wobei die Pflanzenzelllinie die *Syringa-vulgaris*-IRB-SV25/B-(DSM:16857)-Zelllinie ist, das die folgenden Stufen in Sequenz umfasst:

a) Unterziehen der Biomasse, die nach der Kultur der *Syringa-vulgaris*-Zelllinie erhalten wurde, einer Enzymdeaktivierungsbehandlung durch Wärmebehandlung, wobei die Wärmebehandlung für einen Zeitraum von weniger als 5 Minuten oder zwischen 5 und 30 Minuten und bei einer Temperatur zwischen 50 °C und 150 °C erfolgt, um 40 % bis 60 % des ursprünglich in der Biomasse vorhandenen Verbascosids zu Isoverbascosid zu isomerisieren;

b) Homogenisieren des nach Schritt a) erhaltenen Produkts;

c) Trennen der wässrigen Phase von den Zellresten mithilfe von Schwerkraftfiltration, Druckfiltration von Zentrifugation;

d) Durchführen einer Extraktion der nach Schritt c) erhaltenen wässrigen Phase mit hydrophobem Interaktionsharz;

e) Gewinnen der absorbierten Substanzen (Extrakt) aus dem Harz mittels Elution mit einer Mischung von Alkoholen mit bis zu 5 Kohlenstoffatomen in Wasser in Prozentsätzen, die von 30 Vol.-% bis zu knapp unter 100 Vol.-% variieren.

2. Verfahren nach Anspruch 1, das des Weiteren zwischen Schritt a) und b) einen vorangehenden Biomassefiltrationsschritt umfasst, der unter den gleichen Bedingungen wie in Schritt c) durchgeführt wird, mit dem Ziel, nur die Biomasse ohne das Fermenterkulturbedium zu gewinnen.

3. Verfahren nach Anspruch 1 oder 2, wobei die Filtrationsstufe c) und die zuverige Filtration durch ein Nylon-, Stahl- oder Baumwollnetz oder dergleichen mit einer Porosität zwischen 10 pm und 300 pm erfolgt.

4. Verfahren nach einem der Ansprüche 1 bis 3, wobei die Wärmebehandlung unter Verwendung von Dampf erfolgt.

5. Verfahren nach einem der Ansprüche 1 bis 4, wobei der Extraktionsschritt d) durch Chromatographie auf einer Säule oder chargenweise erfolgt.

6. Verfahren nach einem der Ansprüche 1 bis 5, wobei der Gewinnungsschritt e) der Substanzen, die auf das Harz absorbiert wurden, unter Verwendung einer Mischung von Ethanol in Wasser erfolgt.

7. Verfahren nach einem der Ansprüche 1 bis 6, wobei Schritt a) durchgeführt wird, wenn die Zellen ein Alter zwischen 7 und 21 Tagen aufweisen.

8. Verfahren nach einem der Ansprüche 1 bis 7, wobei Schritt d) unter Verwendung eines Harzes durchgeführt wird, das aus Polystyrol-Divinylbenzol- oder Acrylmatrixharzen ausgewählt ist.

9. Verfahren nach einem der Ansprüche 1 bis 8, wobei die *Syringa-vulgaris*-IRB-SV25/B-(DSM:16857)-Zelllinien zunächst stabilisiert und gemäß einer Verfahrensweise ausgewählt werden, umfassend:

a) Kultivieren einer *Syringa-vulgaris*-IRB-SV25/B-(DSM:16857)-Zelllinie, die für einen ausreichenden Zeitraum stabilisiert wurde, um die Vervielfachung der Zelllinie zu ermöglichen, so dass im Wesentlichen einzelne Zellcluster erhalten werden, auf festem Medium;

b) Entfernen der im Wesentlichen einzelnen Zellcluster aus dem festen Medium und Platzieren jedes dieser in einem 15 separaten Flüssigkulturmedium;
 c) Kultivieren jedes der im Wesentlichen einzelnen Zellcluster im Flüssigkulturmedium für einen ausreichenden Zeitraum, um die Vervielfachung der Zellcluster zu ermöglichen, und analytisches Bestimmen der dadurch hergestellten primären und/oder sekundären Metaboliten;
 d) Durchführen einer qualitativen und quantitativen Bestimmung der durch jeden der Zellcluster im Flüssigkulturmedium hergestellten primären und/oder sekundären Metaboliten;
 e) Auswählen des Zellclusters, der in der Lage ist, die größte Menge des Metaboliten von Interesse herzustellen;
 f) Wiederholen des Prozesszyklus gemäß den Stufen a), b), c), d) und e) am Zellcluster, der gemäß Stufe e) ausgewählt wurde, ausreichend oft, bis die Menge des Metaboliten von Interesse, der von dem ausgewählten Zellcluster und von den Zellclustern hergestellt wird, die sich aus weiteren Auswahlzyklen ableiten, im Wesentlichen konstant ist.

10. Verfahren nach Anspruch 9, wobei die *Syringa-vulgaris*-IRB-SV25/B-(DSM:16857)-Zelllinie nach der Stabilisierungs- und Auswahlstufe und vor der Extraktion einer Fermentation unterzogen wird.

11. Verfahren nach einem der Ansprüche 1 bis 10, wobei die Wärmebehandlungsstufe bei einer Temperatur zwischen 60 °C und 120 °C erfolgt.

12. *Syringa-vulgaris*-IRB-SV25/B-(DSM: 16857)-Zelllinienextrakt, das mit dem Verfahren nach einem der Ansprüche 1 bis 11 erhältlich ist.

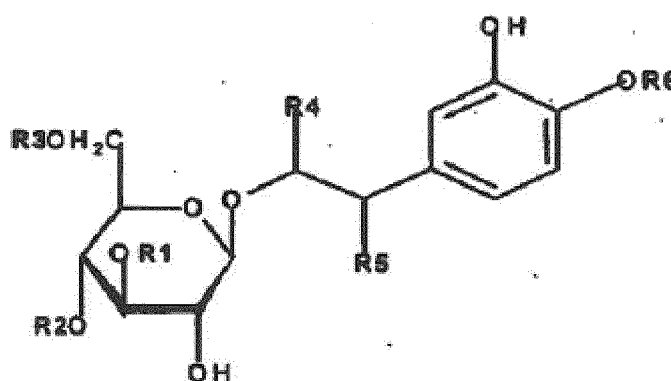
13. Extrakt nach Anspruch 12, das 20 Gew.-% bis 90 Gew.-% Phenylpropanoide und 80 Gew.-% bis 10 Gew.-% einer chromophorfreien Fraktion umfasst, vorwiegend umfassend Oligo- und Polysaccharide, Protein- und Lipidmoleküle, ohne diese jedoch ausschließlich zu umfassen.

14. Extrakt nach Anspruch 12, das zwischen 30 Gew.-% und 60 Gew.-% Phenylpropanoide und zwischen 70 Gew.-% und 40 Gew.-% der chromophorfreien Fraktion umfasst.

15. Extrakt nach einem der Ansprüche 12 bis 14, wobei die Phenylpropanoide bezogen auf die Gesamtmasse von Phenylpropanoiden zwischen 5 % und 20 % Isoverbascosid umfassen.

16. Extrakt nach einem der Ansprüche 12 bis 14, wenn von Anspruch 7 abhängig, wobei die Phenylpropanoide bezogen auf die Gesamtmasse von Phenylpropanoiden zwischen 45 % und 65 % Isoverbascosid umfassen.

17. Extrakt nach einem der Ansprüche 12 bis 14, wobei die Phenylpropanoide durch die folgende allgemeine Formel (I) dargestellt sind:



wobei:

- R1 ein Wasserstoffatom oder ein Monosaccharid mit 5 oder 6 Kohlenstoffatomen ist;
- R2 ein Wasserstoffatom oder eine Caffeoyl-(A)- oder Feruloyl-(B)-Gruppe ist;
- R3 ein Wasserstoffatom oder ein Monosaccharid mit 5 oder 6 Kohlenstoffatomen oder ein Disaccharid mit 10 bis 12 Kohlenstoffatomen oder eine Caffeoyl-(A)- oder Feruloyl-(B)-Gruppe ist;

- R4 ein Wasserstoffatom oder eine Alkylgruppe oder eine Hydroxylgruppe ist;
- R5 ein Wasserstoffatom oder eine Alkylgruppe oder eine Hydroxylgruppe ist, die frei oder mit der Alkoholgruppe an Position C2 der Monosaccharidgruppe kondensiert ist, die daran gebunden ist, um einen 1,4-Dioxanring zu bilden;
- R6 ein Wasserstoffatom oder eine Alkylgruppe ist,

wenn R3 eine Caffeoyle-(A)- oder Feruloyl-(B)-Gruppe ist,



ist R2 sodann immer ein Wasserstoffatom und umgekehrt.

18. Extrakt nach Anspruch 17, wobei R1 eine Ramnhose ist.
19. Extrakt nach Anspruch 17 oder 18, wobei R4 ein Methyl ist.
20. Extrakt nach einem der Ansprüche 17 bis 19, wobei R5 ein Methyl ist.
21. Extrakt nach einem der Ansprüche 17 bis 20, wobei R6 ein Methyl ist.
22. Extrakt nach einem der Ansprüche 12 bis 21 zur medizinischen und ernährungsbezogenen Verwendung.
23. Extrakt nach einem der Ansprüche 12 bis 21 zur Verwendung als antioxidativer Anti-Blutplättchenaggregation-Faktor.
24. Extrakt nach einem der Ansprüche 12 bis 21 zur Verwendung als Blutplättchenzerfallsfaktor.
25. Verwendung des Extrakts nach einem der Ansprüche 12 bis 21 zur Herstellung eines Medikaments mit Metallionenchelatisierungsaktivität zur topischen und systemischen Verwendung.
26. Verwendung des Extrakts nach einem der Ansprüche 12 bis 21 zur Herstellung eines Medikaments mit Anti-Alpha-Reduktase- und Antilipoxygenaseaktivitäten zur Behandlung von juveniler Akne und zur Prophylaxe von Haarverlust und bei der Behandlung von Erkrankungen der Prostata.
27. Verwendung des Extrakts nach einem der Ansprüche 12 bis 21 zur Herstellung eines Medikaments mit Anti-Tyrosinase-Aktivität für Hautbleichungsbehandlungen.
28. Verwendung des Extrakts nach einem der Ansprüche 12 bis 21 zur Herstellung eines Medikaments mit antiproliferativer Aktivität in Bezug auf Zellen.
29. Verwendung des Extrakts nach einem der Ansprüche 12 bis 21 zur Herstellung eines Medikaments mit antimykotischer Aktivität.
30. Verwendung des Extrakts nach einem der Ansprüche 12 bis 21 zur Herstellung eines Medikaments zur Behandlung von anoxisch, traumata- und entzündungsbasierten Störungen des Nervensystems, die aus der Gruppe von Hirnschlag, Hirn- und Spinaltrauma, hämorrhagischem Schock, Epilepsie, zerebraler Ischämie und ischämischer Spinalverletzung, peripherer Neuropathie und Cephalalgie ausgewählt sind.
31. Verwendung des Extrakts nach einem der Ansprüche 12 bis 21 zur Herstellung eines Medikaments zur Behandlung von degenerativen Störungen des Nervensystems, die aus der Gruppe von Störungen ausgewählt sind, die mit dem Altern verknüpft sind oder auf Autoimmun- und Entzündungskomponenten basieren, ausgewählt aus der Gruppe von amyotropher Lateralsklerose, multipler Sklerose, Morbus Parkinson, Morbus Huntington, Morbus Alzheimer, Demenz oder Verhaltensstörungen, ausgewählt aus der Gruppe von Schizophrenie und Depression.
32. Verwendung des Extrakts nach einem der Ansprüche 12 bis 21 zur Herstellung eines Medikaments zur Behandlung eines Reperfusionsschadens und von kardiovaskulären Krankheiten, einschließlich Angina, perkutaner translumi-

naler koronarer Angioplastie (PTCA), Myokardinfarkt und Vaskulopathien, Atherosklerose und Muskelschaden.

33. Verwendung des Extrakts nach einem der Ansprüche 12 bis 21 zur Herstellung eines Medikaments zur Behandlung oder Prophylaxe von Blutplättchenaggregation bei Thromboseerkrankungen und zur Behandlung und Lagerung gereinigter Blutplättchen.
34. Verwendung des Extrakts nach einem der Ansprüche 12 bis 21 zur Herstellung eines Medikaments zur Behandlung von dismetabolischen Störungen, die aus der Gruppe von Diabetes mellitus und den neurologischen, vaskulären und ophthalmischen Komplikationen davon ausgewählt sind.
35. Verwendung des Extrakts nach einem der Ansprüche 12 bis 21 zur Herstellung eines Medikaments zur Behandlung von toxischen Störungen, die aus der Gruppe von Alkoholismus und den neurologischen Komplikationen davon und Leberzirrhose ausgewählt sind.
36. Verwendung des Extrakts nach einem der Ansprüche 12 bis 21 zur Herstellung eines Medikaments zur Behandlung von peripheren neuropathischen Störungen, einschließlich toxischer Neuropathien.
37. Verwendung des Extrakts nach einem der Ansprüche 12 bis 21 zur Herstellung eines Medikaments zur Behandlung von Haut- und Schleimhautkrankheiten, umfassend Photolyse, aktinische Schädigung, Atopie, Krankheiten, die mit Keratinozytenhyperproliferation assoziiert sind, ausgewählt aus der Gruppe von Psoriasis, Xerodermie, Hauttumoren, seborrhoischer Dermatitis, atopischer Dermatitis; Hautläsionen, ausgewählt aus der Gruppe von Verbrennungen, Abschürfungen, Schnitten, Narben, chirurgischen Wunden und wundgelegenen Stellen, sowie Ulzera und Läsionen der Schleimhaut, einschließlich der Mund-, Magen- und Darmschleimhaut und der Vaginalschleimhaut.
38. Verwendung des Extrakts nach einem der Ansprüche 12 bis 21 zur Herstellung eines Medikaments zur Behandlung von Virusinfektionen, die aus HIV, Herpes zoster, Herpes simplex und Cytomegalievirus ausgewählt sind, und bakteriellen Infektionen, einschließlich Dentalstörungen.
39. Verwendung des Extrakts nach einem der Ansprüche 12 bis 21 zur Herstellung eines Medikaments zur Behandlung von Störungen des Atmungsapparats, die aus der Gruppe von zystischer Fibrose, Asthma, Rhinitis, einschließlich allergiebedingt, "Atemnot" und neonatalem Lungenhochdruck ausgewählt sind.
40. Verwendung des Extrakts nach einem der Ansprüche 12 bis 21 zur Herstellung eines Medikaments zur Behandlung von Gelenksstörungen, einschließlich rheumatoider Arthritis und psoriatischer Arthritis.
41. Verwendung des Extrakts nach einem der Ansprüche 12 bis 21 zur Herstellung eines Medikaments zur Behandlung von Tumoren und Störungen, die mit einer verringerten Immunantwort assoziiert sind.
42. Verwendung des Extrakts nach einem der Ansprüche 12 bis 21 zur Herstellung eines Medikaments zur Behandlung von Störungen des sensorischen Apparats, die aus der Gruppe von Katarakt, Netzhautdegeneration und Schädigung des Hörapparats ausgewählt sind.
43. Formulierung, die das Extrakt nach einem der Ansprüche 12 bis 21 als Wirkstoff in Assoziation mit pharmazeutisch, ernährungsbezogen oder kosmetisch annehmbaren Exzipienten zur Verwendung bei Mensch und Tier umfasst.
44. Formulierung nach Anspruch 43 zur systemischen, parenteralen, oralen, rektalen, inhalatorischen, topischen, transdermalen, intravenösen, intramuskulären oder subkutanen Verabreichung.
45. Formulierung nach Anspruch 43, wobei die Form für eine orale Verabreichung durch Tabletten, Lutschtabletten, Kapseln, Pulver, Lösungen, Suspensionen oder eine zerstäubte Form dargestellt ist.
46. Formulierungen nach Anspruch 43, wobei die Form für eine Verabreichung durch Inhalation durch Pulver, Lösungen und Suspensionen dargestellt ist.
47. Formulierungen nach Anspruch 43, wobei die Form für eine topische Verabreichung eine Emulsion, ein Gel oder kompatible Lösungen zur dermalen und mukosalen Verwendung und für Augenspülungen ist.
48. Verwendung des Extrakts nach einem der Ansprüche 12 bis 21 zur Herstellung eines Medikaments zur Tierversor-

gung oder eines Produkts für Tierzuchtfutter.

49. Verwendung des Extrakts nach einem der Ansprüche 12 bis 21, wobei die Extrakt Dosen pro Behandlungszyklus zwischen 0,1 und 100 mg/kg für 1 bis 100 Tage umfasst sind.

50. Verwendung eines *Syringa-vulgaris*-IRB-SV251B-(DSM:16857)-Zelllinienextrakts, das in Bezug auf das Gesamttrockengewicht des Extrakts zwischen 20 Gew.-% und 90 Gew.-% Phenylpropanoide und zwischen 80 % und 10 % einer chromophorfreien Fraktion umfasst, als Kosmetikprodukt umfasst.

51. Verwendung eines *Syringa-vulgaris*-IRB-SV251B-(DSM:16857)-Zelllinienextrakts, das in Bezug auf das Gesamttrockengewicht des Extrakts zwischen 20 Gew.-% und 90 Gew.-% Phenylpropanoide und zwischen 80 % und 10 % einer chromophorfreien Fraktion umfasst, als Ernährungsprodukt umfasst.

52. Verwendung des Extrakts nach einem der Ansprüche 12 bis 21 zur Herstellung von Sonnenfiltern.

53. Verwendung des Extrakts nach einem der Ansprüche 12 bis 21 zur kosmetischen nicht-therapeutischen Behandlung von Hautalterung.

54. *Syringa-vulgaris*-Pflanzenzelllinie mit der Bezeichnung IRB-SV25/B und mit der DSMZ-Hinterlegungsnummer DSM 16857.

Revendications

1. Procédé de préparation d'un extrait de lignée cellulaire végétale à partir de plantes, dans lequel ladite lignée cellulaire végétale est la lignée cellulaire *Syringa vulgaris* IRB-SV25/B (DSM : 16857), comprenant les étapes suivantes en séquence :

a) soumettre la biomasse obtenue après culture de ladite lignée cellulaire de *Syringa vulgaris* à un traitement de désactivation enzymatique par traitement thermique, dans lequel le traitement thermique a lieu pendant une période de temps inférieure à 5 minutes ou comprise entre 5 et 30 minutes, et à une température comprise entre 50°C et 150°C de manière à isomériser de 40% à 60% du verbascoside initialement présent dans la biomasse en isoverbascoside ;

b) homogénéiser le produit obtenu à l'issue de l'étape a) ;

c) séparer la phase aqueuse des résidus cellulaires par filtration par gravité, filtration sous pression ou centrifugation ;

d) réaliser une extraction de la phase aqueuse obtenue à l'issue de l'étape c) avec une résine d'interaction hydrophobe ;

e) récupérer les substances absorbées (extrait) de la résine par élution avec un mélange d'alcools ayant jusqu'à 5 atomes de carbone, dans de l'eau en des pourcentages variant de 30% jusqu'à un peu moins de 100% en volume.

2. Procédé selon la revendication 1, comprenant en outre, entre les étapes a) et b), une étape de filtration de la biomasse préalable effectuée dans les mêmes conditions que celles indiquées à l'étape c), dans le but de récupérer juste la biomasse, sans le milieu de culture du fermenteur.

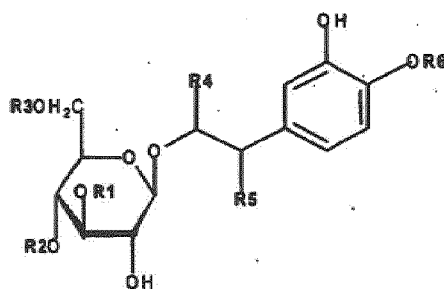
3. Procédé selon l'une quelconque des revendications 1 ou 2, **caractérisé en ce que** ladite étape de filtration c) et ladite filtration préalable se produisent à travers une toile de nylon, d'acier ou de coton ou analogue ayant une porosité comprise entre 10 pm et 300 pm.

4. Procédé selon l'une quelconque des revendications 1 à 3, **caractérisé en ce que** le traitement thermique s'effectue à l'aide de vapeur d'eau.

5. Procédé selon l'une quelconque des revendications 1 à 4, **caractérisé en ce que** l'étape d'extraction d) se produit par chromatographie sur une colonne ou par lot.

6. Procédé selon l'une quelconque des revendications 1 à 5, dans lequel l'étape de récupération e) des substances absorbées sur la résine est réalisée en utilisant un mélange d'éthanol dans l'eau.

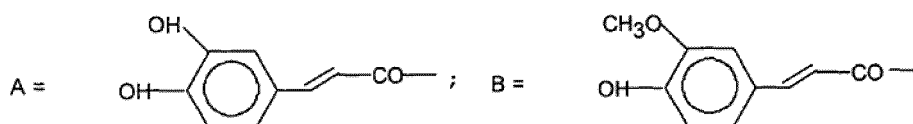
7. Procédé selon l'une quelconque des revendications 1 à 6, dans lequel l'étape a) est réalisée lorsque les cellules ont un âge compris entre 7 et 21 jours.
8. Procédé selon l'une quelconque des revendications 1 à 7, dans lequel l'étape d) est réalisée en utilisant une résine choisie parmi une résine de polystyrène-divinylbenzène et une résine à matrice acrylique.
9. Procédé selon l'une quelconque des revendications 1 à 8, **caractérisé en ce que** les lignées cellulaires *Syringa vulgaris* IRB-SV25/B (DSM : 16857) sont d'abord stabilisées et sélectionnées selon un mode opératoire comprenant :
 - a) la culture d'une lignée cellulaire de *Syringa vulgaris* IRB-SV25/B (DSM : 16857), stabilisée pendant une période de temps suffisante pour permettre la multiplication de ladite lignée cellulaire pour former des amas de cellules sensiblement distincts, sur un milieu solide ;
 - b) enlever lesdits amas de cellules sensiblement distincts dudit milieu solide et placer chacun d'eux dans un milieu de culture liquide séparé ;
 - c) cultiver chacun desdits amas de cellules sensiblement distincts dans ledit milieu de culture liquide pendant une période de temps suffisante pour permettre la multiplication desdits amas de cellules, et déterminer de manière analytique les métabolites primaires et/ou secondaires ainsi produits ;
 - d) effectuer une détermination qualitative et quantitative des métabolites primaires et/ou secondaires produits par chacun desdits amas de cellules dans ledit milieu de culture liquide ;
 - e) sélectionner l'amas cellulaire capable de produire la plus grande quantité dudit métabolite d'intérêt ;
 - f) répéter le cycle du procédé selon les étapes, a), b), c), d) et e) sur ledit amas de cellules, sélectionné selon l'étape e), un nombre de fois nécessaire jusqu'à ce que la quantité dudit métabolite d'intérêt produite par ledit amas de cellules, et par les amas de cellules dérivant d'autres cycles de sélection, est essentiellement constant.
10. Procédé selon la revendication 9, **caractérisé en ce que**, après l'étape de stabilisation et de sélection et avant l'extraction, la lignée cellulaire *Syringa vulgaris* IRB-SV25/B (DSM : 16857) est soumise à une fermentation.
11. Procédé selon l'une quelconque des revendications 1 à 10, **caractérisé en ce que** l'étape de traitement thermique est réalisé à une température comprise entre 60°C et 120°C.
12. Extrait de lignée cellulaire *Syringa vulgaris* IRB-SV25/B (DSM : 16857) pouvant être obtenu selon le procédé selon l'une quelconque des revendications 1 à 11.
13. Extrait selon la revendication 12, comprenant de 20% à 90% en poids de phénylpropanoïdes et de 80 à 10% d'une fraction exempte de chromophores, comprenant principalement, mais non exclusivement, des molécules d'oligo- et de polysaccharides, de protéines et de lipides.
14. Extrait selon la revendication 12, comprenant entre 30% et 60% en poids de phénylpropanoïdes et entre 70% et 40% de ladite fraction exempte de chromophores.
15. Extrait selon l'une quelconque des revendications 12 à 14, dans lequel les phénylpropanoïdes comprennent entre 5% et 20% d'isoverbascoside par rapport à la masse totale des phénylpropanoïdes.
16. Extrait selon l'une quelconque des revendications 12 à 14, lorsqu'elle est dépendante de la revendication 7, dans lequel les phénylpropanoïdes comprennent entre 45% et 65% d'isoverbascoside par rapport à la masse totale de phénylpropanoïdes.
17. Extrait selon l'une quelconque des revendications 12 à 14, **caractérisé en ce que** les phénylpropanoïdes sont représentés par la formule générale (I) suivante:



dans laquelle :

- R1 est un atome d'hydrogène ou un monosaccharide ayant 5 ou 6 atomes de carbone ;
- R2 est un atome d'hydrogène ou est un groupe caffeoyl (A) ou feruloyle (B) ;
- R3 est un atome d'hydrogène ou un monosaccharide ayant 5 ou 6 atomes de carbone ou un disaccharide ayant 10 à 12 atomes de carbone ou est un groupe caffeoyl (A) ou feruloyle (B) ;
- R4 est un atome d'hydrogène ou un groupe alkyle, ou un groupe hydroxyle ;
- R5 représente un atome d'hydrogène ou un groupe alkyle ou un groupe hydroxyle, soit libre soit condensé avec le groupe alcool en position C2 du groupe monosaccharide lié à celui-ci, afin de former un cycle 1,4 dioxane ;
- R6 est un atome d'hydrogène ou un groupe alkyle,

lorsque R3 est un groupe caffeoyl (A) ou feruloyle (B)



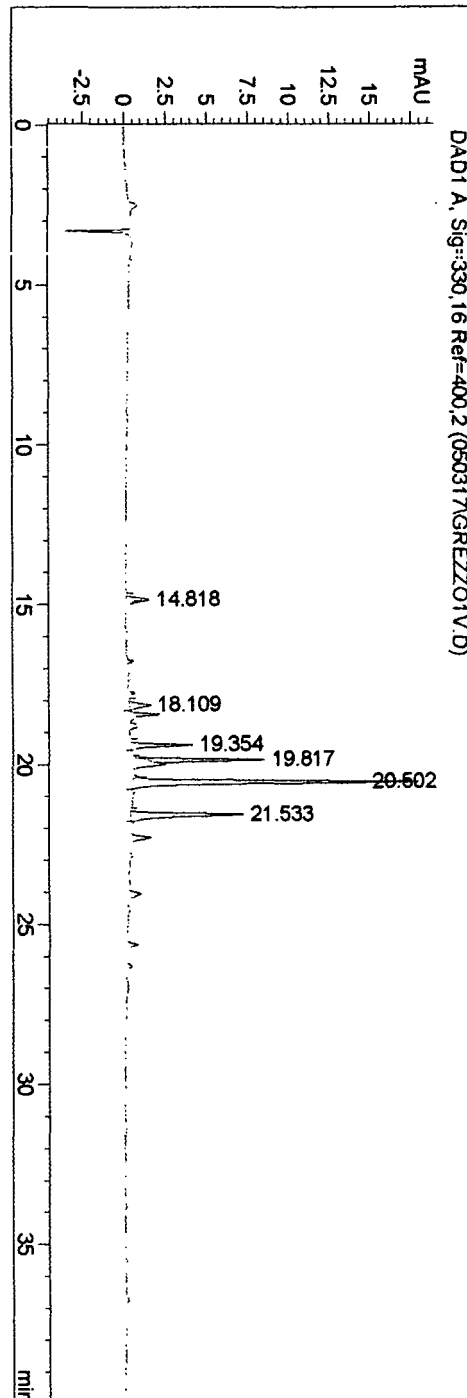
alors R2 est toujours un atome d'hydrogène et *vice-versa*.

18. Extrait selon la revendication 17, dans lequel R1 est un ramnhose.
19. Extrait selon la revendication 17 ou 18, dans lequel R4 est un méthyle.
20. Extrait selon l'une quelconque des revendications 17-19, dans lequel R5 est un méthyle.
21. Extrait selon l'une quelconque des revendications 17-20, dans lequel R6 est un méthyle.
22. Extrait selon l'une quelconque des revendications 12 à 21 pour une utilisation médicale et nutritionnelle.
23. Extrait selon l'une quelconque des revendications 12 à 21, pour une utilisation en tant qu'antioxydant, facteur antiagrégant plaquettaire.
24. Extrait selon l'une quelconque des revendications 12 à 21, pour une utilisation en tant que facteur de désagrégation plaquettaire.
25. Utilisation de l'extrait tel que défini selon l'une quelconque des revendications 12 à 21 pour la préparation d'un médicament ayant une activité chélatante d'ions métalliques pour une utilisation topique et systémique.
26. Utilisation de l'extrait selon l'une quelconque des revendications 12 à 21 pour la préparation d'un médicament ayant des activités anti-alpha-réductase et anti-lipoxygénase pour le traitement de l'acné juvénile et pour la prévention de la chute des cheveux et dans le traitement des maladies de la prostate.
27. Utilisation de l'extrait selon l'une quelconque des revendications 12 à 21 pour la préparation d'un médicament ayant une activité anti-tyrosinase pour des traitements de blanchiment de la peau.

28. Utilisation de l'extrait selon l'une quelconque des revendications 12 à 21 pour la préparation d'un médicament ayant une activité anti-proliférative vis-à-vis des cellules.
- 5 29. Utilisation de l'extrait selon l'une quelconque des revendications 12 à 21 pour la préparation d'un médicament ayant une activité anti-micotique.
- 10 30. Utilisation de l'extrait selon l'une quelconque des revendications 12 à 21 pour la préparation d'un médicament destiné au traitement des troubles de types anoxiques, traumatiques et inflammatoires du système nerveux choisis dans le groupe des attaques cérébrales, des traumatismes cérébraux et spinaux, d'un choc hémorragique, de l'épilepsie, de l'ischémie cérébrale et d'une lésion spinale ischémique, de la neuropathie périphérique et de la céphalalgie.
- 15 31. Utilisation de l'extrait selon l'une quelconque des revendications 12 à 21 pour la préparation d'un médicament destiné au traitement de troubles dégénératifs du système nerveux choisi dans le groupe de troubles liés au vieillissement ou à base de composants auto-immunes et inflammatoires choisis parmi le groupe comprenant la sclérose latérale amyotrophique, la sclérose en plaques, la maladie de Parkinson, la maladie de Huntington, la maladie d'Alzheimer, la démence ou les troubles du comportement choisis dans le groupe de la schizophrénie et la dépression.
- 20 32. Utilisation de l'extrait selon l'une quelconque des revendications 12 à 21 pour la préparation d'un médicament pour le traitement des dommages dus à la reperfusion et aux maladies cardio-vasculaires comprenant l'angine, l'angioplastie coronarienne transluminale percutanée (PTCA), l'infarctus du myocarde et les vasculopathies, l'athérosclérose et les dommages musculaires.
- 25 33. Utilisation de l'extrait selon l'une quelconque des revendications 12 à 21 pour la préparation d'un médicament pour le traitement ou la prévention de l'agrégation plaquettaire dans des maladies thrombotiques et pour le traitement et le stockage de plaquettes purifiées.
- 30 34. Utilisation de l'extrait selon l'une quelconque des revendications 12 à 21 pour la préparation d'un médicament destiné au traitement de troubles métaboliques choisis dans le groupe du diabète sucré et ses complications neurologiques, vasculaires et ophtalmiques.
- 35 35. Utilisation de l'extrait selon l'une quelconque des revendications 12 à 21 pour la préparation d'un médicament destiné au traitement de troubles toxiques choisis dans le groupe de l'alcoolisme et de ses complications neurologiques et de la cirrhose du foie.
- 40 36. Utilisation de l'extrait selon l'une quelconque des revendications 12 à 21 pour la préparation d'un médicament pour le traitement de troubles neuropathiques périphériques comprenant des neuropathies toxiques.
- 45 37. Utilisation de l'extrait selon l'une quelconque des revendications 12 à 21 pour la préparation d'un médicament pour le traitement des maladies de la peau et des muqueuses comprenant la photolyse, les dommages actiniques, l'atopie, les maladies associées à l'hyperprolifération des kératinocytes sélectionnées dans le groupe du psoriasis, la xérodermie, les tumeurs cutanées, la dermatite séborrhéique, la dermatite atopique ; des lésions cutanées choisies dans le groupe des brûlures, des abrasions, des coupures, des cicatrices, des plaies chirurgicales et des escarres, ainsi que des ulcères et des lésions de la muqueuse incluant la muqueuse orale, gastrique et intestinale et la muqueuse vaginale.
- 50 38. Utilisation de l'extrait selon l'une quelconque des revendications 12 à 21 pour la préparation d'un médicament pour le traitement d'infections virales choisies parmi le VIH, l'herpès zoster, l'herpès simplex et le cytomégalovirus et des infections bactériennes y compris des troubles dentaires.
- 55 39. Utilisation de l'extrait selon l'une quelconque des revendications 12 à 21 pour la préparation d'un médicament destiné au traitement de troubles de l'appareil respiratoire choisi dans le groupe de la fibrose kystique, de l'asthme, de la rhinite, y compris en raison d'allergies, de "détresse respiratoire » et l'hypertension pulmonaire néonatale.
40. Utilisation de l'extrait selon l'une quelconque des revendications 12 à 21 pour la préparation d'un médicament destiné au traitement de troubles articulaires comprenant la polyarthrite rhumatoïde et l'arthrite psoriasique.

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41. Utilisation de l'extrait selon l'une quelconque des revendications 12 à 21 pour la préparation d'un médicament destiné au traitement de tumeurs et de troubles associés à une réponse immunitaire diminuée.
- 5 42. Utilisation de l'extrait selon l'une quelconque des revendications 12 à 21 pour la préparation d'un médicament destiné au traitement de troubles de l'appareil sensoriel choisis dans le groupe de la cataracte, la dégénérescence rétinienne et une lésion de l'appareil auditif.
- 10 43. Formulation comprenant l'extrait selon l'une quelconque des revendications 12 à 21 en tant qu'ingrédient actif en association avec des excipients pharmaceutiquement, nutritionnellement ou cosmétiquement acceptables à usage humain et vétérinaire.
44. Formulation selon la revendication 43 pour l'administration systémique, parentérale, orale, rectale, inhalatoire, topique, transdermique, intraveineuse, intramusculaire ou sous-cutanée.
- 15 45. Formulation selon la revendication 43, dans laquelle la forme pour l'administration orale est représentée par des comprimés, des pastilles, des gélules, des poudres, des solutions, des suspensions ou une forme nébulisée.
- 20 46. Formulation selon la revendication 43, **caractérisées en ce que** la forme d'administration par inhalation est représentée par des poudres, des solutions et des suspensions.
47. Formulation selon la revendication 43, dans lesquelles la forme pour l'administration topique est une émulsion, un gel ou des solutions compatibles pour l'utilisation dermique et au contact de muqueuses et les lavages oculaires.
- 25 48. Utilisation de l'extrait selon l'une quelconque des revendications 12 à 21 pour la préparation d'un médicament pour soins vétérinaires ou d'un produit pour aliments zootechniques.
49. Utilisation de l'extrait selon l'une quelconque des revendications 12 à 21, dans laquelle les doses d'extrait par cycle de traitement sont comprises entre 0,1 et 100 mg/kg pendant 1-100 jours.
- 30 50. Utilisation d'un extrait de la lignée cellulaire *Syringa vulgaris* IRB-SV25/B (DMS : 16857) comprenant entre 20% et 90% en poids, par rapport au poids sec total dudit extrait, de phénylpropanoïdes, et entre 80% et 10% d'une fraction d'extrait de la lignée cellulaire exempte de chromophores, en tant que produit cosmétique.
- 35 51. Utilisation d'un extrait de la lignée de *Syringa vulgaris* IRB-SV25/B (DMS : 16857) comprenant entre 20% et 90% en poids, par rapport au poids sec total dudit extrait, de phénylpropanoïdes, et entre 80% et 10% de chromophores, en tant que produit nutritionnel.
52. Utilisation de l'extrait selon l'une quelconque des revendications 12 à 21, pour la préparation de filtres solaires.
- 40 53. Utilisation de l'extrait selon l'une quelconque des revendications 12 à 21 pour le traitement cosmétique non thérapeutique du vieillissement de la peau.
- 45 54. Lignée cellulaire végétale de *Syringa vulgaris* dénommée IRB-SV25/B et ayant un numéro d'accès DSMZ : DSM 16857.



REFERENCES CITED IN THE DESCRIPTION

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