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(72) Inventors:  
• **Badwan, Adnan, Dr.  
11185 Amman (JO)**  
• **Murshed, Abedel-qader Mohammed  
11118 Amman (JO)**

(71) Applicant: **The Jordanian Pharmaceutical  
Manufacturing Co.  
11710 Naor (JO)**

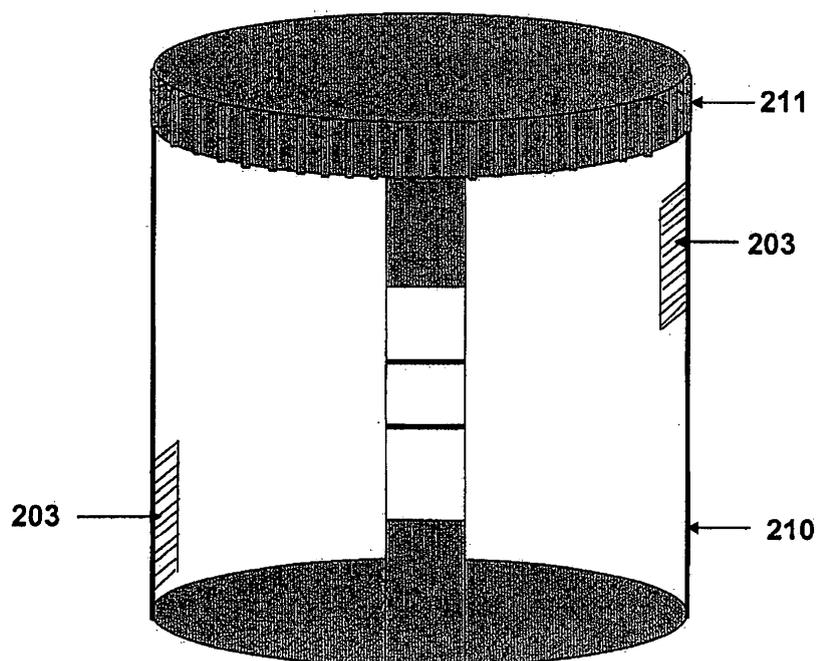
(74) Representative: **Winkler, Andreas Fritz Ernst  
Forrester & Boehmert  
Pettenkoferstrasse 20-22  
80336 München (DE)**

(54) **Antibody detection method involving an oligonucleotide enhanced colloidal gold signal**

(57) The present invention refers to a rapid immunochromatographic test device for antibody detection, comprising at least a first and a second conjugate releasing pad, wherein the first conjugate pad comprises a gold conjugated protein linked first oligonucleotide and a gold conjugated anti-antibody, and the second conjugate pad comprises a gold conjugated protein linked second oli-

gonucleotide, which second oligonucleotide is complementary to the first oligonucleotide. The present invention further refers to a use of such test device for antibody detection in urine or saliva, e.g. HIV antibody. Embodiments of the test device are a test strip and a detection cup. The present invention also refers to a method for manufacturing such test device.

**Figure 2b**



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**Description**

5 [0001] The present invention refers to a rapid immunochromatographic test device in the form of a test strip or a detection cup comprising a first and a second conjugate pad, wherein the first conjugate pad comprises a protein linked oligonucleotide and an anti-antibody, and the second conjugate pad comprises a protein linked complementary oligonucleotide, wherein the protein linked oligonucleotides and the anti-antibody are gold conjugated. The present invention further refers to a use of such test device for antibody detection in urine or saliva. The present invention also refers to a method for manufacturing such test device.

**Background of the Invention**

10 [0002] In recent years, the *in vitro* diagnostics (IVD) industry has made enormous efforts to develop immunochromatographic tests. Such tests have found applications in both clinical and non-clinical fields<sup>1</sup>. A clinical utility of this test format has been shown for more than 150 different analytes, and many of them are target now of commercially available diagnostic products<sup>3</sup>. The wide range of applications for such devices has been reviewed<sup>1,2</sup>.

15 [0003] Rapid immunochromatographic test devices, e.g. in the form of a test strip, are made up of a number of components (see Fig. 1). Such a test strip 101 commonly includes a sample pad 102, a conjugate pad 103, a membrane 104, e.g. a nitrocellulose membrane, and an absorbent pad 105. The membrane 104 is usually attached by means of an adhesive 106 to a supporting backing 107, e.g. made of plastic. In practice, the user dispense a patient sample (usually urine or whole blood) onto the sample pad 102. The sample then flows through the sample pad 102 into the conjugate pad 103, where it mixes with and releases the detector reagent. This mixture then flows across the membrane 104, where it binds with the test and control reagents located in the capture test zone 108 (sample zone) and negative control zone 109, respectively. When the mixture binds to the reagent that forms the test line, a positive result is indicated. The colour intensity of the test line is proportional to the concentration of analyte in the sample. Excess sample that flows beyond the test and control zones 108, 109 is taken up in the absorbent pad 105.

20 [0004] Rapid immunochromatographic test devices for diagnostic purposes are easy to operate and thus do not only contribute to the comfort of professional users, e.g. medical staff, but also allow the operation by non-professionals users, e.g. most patients.

25 [0005] However, despite the wide use of rapid immunochromatographic test devices, their suitability is still limited with regard to certain applications. Urine, for example, contains very low levels of IgG, frequently around 1 mg/l. Therefore, the detection of antibodies, e.g. directed to HIV or HCV, require very sensitive techniques. To date, the tests for antibodies in urine samples are based on ELISA and Western blot techniques, which are labour-intensive, time-consuming and need to be carried out by qualified persons. Efforts are being made to develop simple and/or rapid tests for the detection of antibody to HIV in urine specimens<sup>4</sup>.

30 [0006] Oral fluid specimens consist mostly of saliva, which predominantly contains IgA class antibody, and oral mucosal transudates, which mostly contain IgG, and therefore also have much lower levels of IgG than serum. The levels of IgG normally found in oral fluid specimens (approximately 15 mg/l) are, however, higher than in urine specimens and innovative simple and rapid technology that has been shown to be effective for whole blood, serum and plasma, e.g. lateral flow through a chromatographic membrane, has been developed for use with these specimens<sup>4</sup>.

35 [0007] It is therefore an object of the present invention to provide a method and means for sensitive antibody detection, e.g. for the detection of antibodies directed against HIV antigen, in urine and saliva.

**Summary of the Invention**

40 [0008] The object of the present invention is solved by a rapid immunochromatographic test device for antibody detection, comprising at least a first and a second conjugate releasing pad, wherein the first conjugate pad comprises a gold conjugated protein linked first oligonucleotide and a gold conjugated anti-antibody, and the second conjugate pad comprises a gold conjugated protein linked second oligonucleotide, which second oligonucleotide is complementary to the first oligonucleotide. An anti-antibody is an antibody that is specifically directed against another antibody.

45 [0009] In one embodiment, the test device is in the form of a test strip.

[0010] In an alternative embodiments, the test device is in the form of a detection cup.

50 [0011] In one embodiment, the first and the second conjugate pads are separated from each other. Such separation may be realized by a divider, preferably a plastic divider, sandwiched between the two pads, or by placing the two pads spaced apart from each other at different locations. The purpose of separating the two pads is to prevent untimely mixing of the conjugates comprised in the first and second pad.

55 [0012] In one embodiment, the conjugate pads further comprise optionally modified water-soluble chitosan.

[0013] The object of the present invention is further solved by a use of a test device according to the present invention for the detection of an antibody in a sample of urine or saliva of a subject, preferably human.

**[0014]** In a preferred embodiment, the antibody to be detected is selected from antibodies directed against HIV, HCV, and *H. pylori* antigen.

**[0015]** The object of the present invention is further solved by a method for preparing a test device according to the present invention, comprising the following steps:

- 5
- (a) providing a first conjugate solution comprising a gold conjugated protein linked first oligonucleotide and a gold conjugated anti-antibody, and applying said solution to a first conjugate pad;
- 10
- (b) providing a second conjugate solution comprising a gold conjugated protein linked second oligonucleotide, and applying said solution to a second conjugate pad;
- (c) providing a membrane comprising an antigen recognized by the antibody to be detected, which antigen is immobilized within a sample zone, and a non-specific antibody immobilized within a control zone. Immobilization can be realized by direct or indirect attachment to the membrane and has the effect that the sample zone antigen is not released.
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**[0016]** In one embodiment, the method is for preparing a test device in the form of a test strip and additionally comprises the following steps:

- 20
- (d) placing the membrane onto a backing material;
- (e) placing the first conjugate pad onto the membrane;
- (f) placing a divider, preferably a plastic divider, onto the first conjugate pad;
- 25
- (g) placing the second conjugate pad onto the divider.

**[0017]** In an alternative embodiment, the method is for preparing a test device in the form of a detection cup and additionally comprises the following step:

- 30
- (d') placing the membrane and the first and second conjugate pads into a container, wherein the first and second conjugate pads are separated from each other. Such separation may be realized by placing the two pads spaced apart from each other at different locations in the container interior.

**[0018]** In one embodiment of the method, optionally modified water-soluble chitosan is added during the preparation of the first and second conjugate solutions prior to conjugation of colloid gold with the anti-antibody and/or the protein linked oligonucleotides, i.e. the protein linked first oligonucleotide and the protein linked second oligonucleotide.

**[0019]** In a preferred embodiment of the method, the colloidal gold is prepared by reduction of a 1% aqueous solution of tetrachloroauric acid using trisodium citrate aqueous solution to produce spheroidal gold particles.

**[0020]** Thus, the present invention provides a rapid immunochromatographic detection system for antibody detection comprising a test strip and two conjugate releasing pads with different compositions. The first pad comprises anti-antibody, e.g. mouse anti-human IgG, specifically recognizing the antibody to be detected, and a protein linked oligonucleotide ("sense" or first oligonucleotide). The proteins, i.e. the anti-antibody and the protein linked to the oligonucleotide (e.g. bovine serum albumin, BSA), are conjugated with colloidal gold. The second conjugate pad comprises a further protein linked oligonucleotide ("antisense" or second oligonucleotide) also conjugated with colloidal gold. The both oligonucleotides, i.e. the sense and antisense oligonucleotide, are complementary to each other. When the sample comes into contact with the anti-antibody of the first pad, the antibody to be detected is captured and thereby a complex is formed comprising the antibody, the conjugated anti-antibody and the conjugated protein linked sense oligonucleotide. This complex is carried to the sample zone where an antigen is immobilized which is recognized by the antibody to be detected. The complex will be captured then by the antigen within the sample zone. The conjugated protein linked antisense oligonucleotide which is released from the second conjugate pad also moves to the sample zone where the complementary oligonucleotides, i.e. the sense and the antisense oligonucleotides, bind to each other. Thus, the complex of antibody, anti-antibody and the sense oligonucleotide, which complex is bound within the sample zone serves as a target for the antisense oligonucleotide. Due to sense/antisense oligonucleotide and antibody/anti-antibody/antigen interaction, a multi-complex is formed. The binding between the two conjugates will enhance the signal colour intensity and thus enhance the sensitivity of detection.

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**[0021]** In addition to making use of the oligonucleotide/complementary oligonucleotide interaction, further sensitivity enhancement is achieved by using a large sample volume of urine or saliva and by using water-soluble chitosan (or

modified water-soluble chitosan).

## Detailed Description of the Invention

### 5 Brief Description of the Figures

#### [0022]

10 Figure 1a shows top and side views of a typical rapid-flow immunochromatographic test device in the form of a test strip 101 including a sample pad 102, a conjugate pad 103, a membrane 104, an absorbent pad 105, an adhesive 106, a supporting backing 107, a test or sample zone 108, and a control zone 109.

15 Figure 1b shows top and side views of a rapid-flow immunochromatographic test device according to the present invention in the form of a test strip 101 including a sample pad 102, a first conjugate pad 103.1, a second conjugate pad 103.2, a membrane 104, an absorbent pad 105, an adhesive 106, a supporting backing 107, a test or sample zone 108, a control zone 109, and a conjugates divider 110 separating the conjugate pads 103.1 and 103.2.

Figure 2a shows an assembly comprising a test strip 201, an absorbent sample pad 202, and an absorbent pad 205.

20 Figure 2b shows an immunochromatographic detection cup comprising a sample-collecting container 210, a cap 211, a first conjugate pad 203 and a second conjugate pad 203', and the assembly shown in Fig. 2a.

### EXAMPLES

#### 25 EXAMPLE 1: Preparation of protein linked oligonucleotide

30 **[0023]** 5 mg of bovine serum albumin (BSA) each was linked to an oligonucleotide (about 20 nucleotides having an amino group at the 5' terminus) and to a complementary oligonucleotide (about 20 nucleotides having an amino group at the 5' terminus) according to the method of Duncan *et al.* (1983)<sup>5</sup> which can be illustrated as a procedure comprising the following steps:

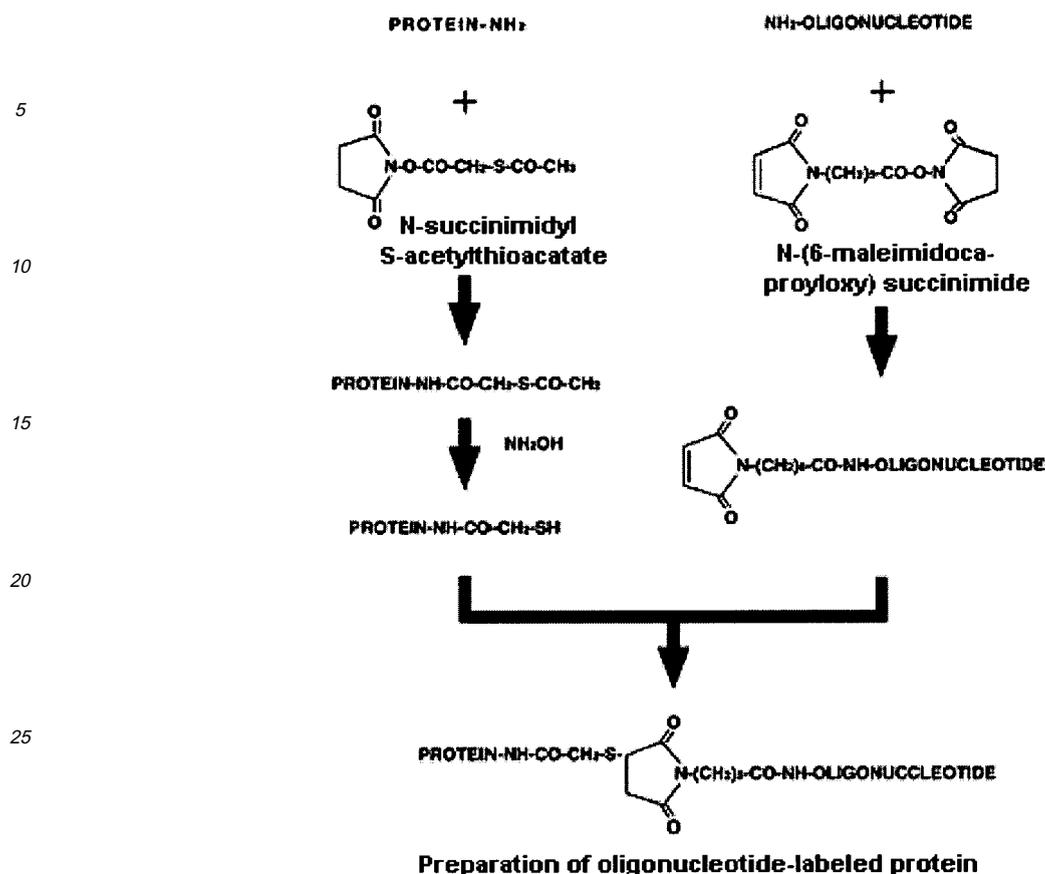
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35 **EXAMPLE 2:** Preparation of conjugate releasing pads comprising oligonucleotide linked protein

[0024] The oligonucleotide and complementary oligonucleotide linked BSA prepared as described in Example 1 are further processed according to a procedure comprising the following steps:

- 40 (a) prepare oligonucleotide linked BSA solution (solution 1);
- (b) prepare complementary oligonucleotide linked BSA solution (solution 2);
- (c) prepare 1% aqueous solution of tetrachloroauric acid at room temperature;
- (d) prepare 4% trisodium citrate aqueous solution at room temperature;
- (e) prepare 0.05 M potassium carbonate aqueous solution at room temperature;
- 45 (f) prepare 400 ml of phosphate stabilizing buffer, pH 7.4, containing BSA, Tween 20, sucrose, polyvinylpyrrolidone and preservative (like sodium azide) at room temperature;
- (g) prepare colloidal gold solution by reduction of 1.7 ml boiling tetrachloroauric acid solution (after dilution in 100 ml) using 1 ml trisodium citrate solution and equilibrate to room temperature;
- (h) dilute the colloidal gold solution 1:1 using distilled water and adjust the pH to 7.4 using potassium carbonate solution at room temperature;
- 50 (i) prepare 200 ml of phosphate conjugation buffer, pH 7.4, at room temperature;
- (j) partition the 200 ml conjugation buffer by dividing it between two flasks (100 ml each), i.e. a first and a second flask;
- (k) add 0.5 mg of an aqueous anti-antibody solution to the conjugation buffer in the first flask while stirring at room temperature;
- 55 (l) add 0.5 mg of the oligonucleotide linked BSA aqueous solution (solution 1) to the first flask at room temperature;
- (m) add 1.0 mg of the complementary oligonucleotide linked BSA aqueous solution (solution 2) to the conjugation buffer in the second flask while stirring at room temperature;
- (n) add 100 ml colloidal gold solution into each flask while stirring at room temperature;
- (o) add 200 ml of stabilizing buffer to each flask and concentrate each conjugate solution by cooled (temperature

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- around 15°C) high speed centrifugation (10,000 rpm for one hour);  
(p) discard the supernatant and re-suspend the concentrated conjugates at room temperature;  
(q) adjust the concentration for each of the two conjugates to  $O.D._{520} = 2.0$ ;  
(r) add 0.1 ml of Tween 20 to the first conjugate solution and soak a glass fibre sheet conjugate pad with the conjugate  
5 solution, then heat dry at temperature around 50°C;  
(s) soak another glass fibre sheet conjugate pad with the second conjugate, then heat dry at temperature around 50°C.

### EXAMPLE 3: Preparation of a test device in the form of a test strip

10 **[0025]** In case of a test strip, the first conjugate releasing pad 103.1 is prepared by soaking with oligonucleotide linked BSA and anti-antibody conjugate, while the other pad 103.2 is prepared by soaking with complementary oligonucleotide linked BSA conjugate (see Example 2).

**[0026]** In more detail, the test device is prepared according to a procedure comprising the following steps:

15 (a) prepare a phosphate sample buffer containing goat serum, ethylenediamine tetraacetic acid (EDTA), non-fat dry milk, preservative (like sodium azide) and Tween 20;

(b) soak a sample pad with the phosphate sample buffer and heat dry at a temperature around 50°C;

20 (c) prepare a colloidal gold solution (see Example 2);

(d) conjugate colloidal gold with oligonucleotide linked BSA and anti-antibody to prepare the first conjugate (see Example 2), add Tween 20 to this first conjugate solution, soak a conjugate pad with this first conjugate solution and heat dry at a temperature around 50°C;

25 (e) conjugate colloidal gold with complementary oligonucleotide linked BSA to prepare the second conjugate, soak a conjugate pad with this second conjugate solution (see Example 2) and heat dry at a temperature around 50°C;

30 (f) print sample zone antigen (i.e. the antigen recognized by the antibody to be detected, e.g. envelop antigens of HCV) and control zone antibody (i.e. non-specific antibody) onto a nitrocellulose membrane and heat dry at a temperature around 50°C;

(g) prepare a membrane blocking solution containing Tween 20 and non fat dry milk;

35 (h) block the nitrocellulose membranes using the blocking solution and heat dry at a temperature around 50°C;

(i) laminate in the following order components of the test strip onto a backing material:

(i.i) laminate the nitrocellulose membrane nearly in the middle of the test strip;

40 (i.ii) laminate the absorbent pad at the end of the test strip (overlaps from the nitrocellulose membrane side);

(i.iii) laminate the first conjugate pad on the other side of the nitrocellulose membrane;

(i.iv) laminate the plastic divider onto the first conjugate (overlaps from the nitrocellulose membrane side);

(i.v) laminate the second conjugate pad onto the divider (overlaps from the nitrocellulose membrane side);

45 (i.vi) laminate the sample pad onto the other end of the sheet, the sample pad shall overlap with the two conjugate pads;

(j) pack the test strip into an aluminium pouch with a silica gel desiccant.

50 **[0027]** In steps (d) or (e), a sheet can be used instead of a single conjugate pad which is cut into several pads after drying. Similarly, in step (i), a sheet can be used which is cut into strips after having laminated all components.

**[0028]** The antigens used may be synthetic or recombinant.

**[0029]** Control and sample zone may be realized by lines. More than one sample zone and/or more than one control zone are also contemplated.

55 **[0030]** A urine wick can be used in case of urinary testing, while a blood filter is used in case of blood testing.

### EXAMPLE 4: Urinary HCV IgG test strip

**[0031]** The first conjugate releasing pad 103.1 comprises gold conjugate 1, i.e. conjugated anti-antibody, and conju-

gated BSA linked oligonucleotide 1. The second conjugate releasing pad 103.2 comprises gold conjugate 2, i.e. conjugated complementary oligonucleotide 2. The sample zone 208 comprises a mixture of synthetic and recombinant NS3 and envelop antigens of HCV which antigens are immobilized onto the nitrocellulose membrane 104. The control zone 109 comprises anti-mouse IgG. The sample and control zones 108, 109 turn into purple colour in case that HCV IgG is present in the sample; only the control line 109 turns into purple colour in case of HCV IgG free sample.

#### EXAMPLE 5: Immunochromatographic detection cup

**[0032]** Another embodiment of the rapid immunochromatographic detection system of the present invention is shown in Fig. 2a, b. This immunochromatographic detection cup comprises a sample-collecting container 210, actually the "cup", a cap 211 for closing the container, and a detection test strip 201 inserted into the container 210. The test strip 201 may be placed inside the transparent container wall, and the test result can be read on the outer surface. The test strip 201 comprises a nitrocellulose membrane 204 and is linked via an absorbent pad 202 on the test strip 201 to an absorbent sample pad 202 placed on the bottom of the container 210. The absorbent sample pad 202 is designed to cover the inner surface of the container bottom. At the opposite end, the test strip 201 is linked via an absorbent pad 205 on the test strip to an absorbent pad 205 which is fitted into the cap 211 of the container and is designed to be thick enough to absorb more than 15 ml of sample.

**[0033]** A first gold conjugate releasing pad 203 is fixed at the inner surface of the container wall which pad 203 comprises gold conjugated anti-antibody and gold conjugated BSA linked oligonucleotide. A second gold conjugate releasing pad 203' is also fixed to the inner surface of the container wall which pad 203' comprises gold conjugated albumin linked complementary oligonucleotide. Importantly, both conjugate pads are fixed at different locations. The gold conjugates will begin releasing from pad 203 and 203' during the urine sample is streaming into the container 210. Release into the sample will increase the possibility of interaction between the conjugates and the antibodies to be detected.

**[0034]** The sample zone 208 comprises an antigen which is specifically recognized by the antibody to be detected. The antigens used may be synthetic or recombinant. Non-specific antibody is immobilized as a control zone 209 onto the nitrocellulose membrane 204. In the embodiment shown in Fig. 2a, b the control zone 209 is realized by a control line. More than two sample zones 208 and/or more than one control zone 209 are also contemplated.

#### EXAMPLE 6: Preparation of a test device in the form of a detection cup

**[0035]** In case of a urinary or saliva immunochromatographic detection cup, two pads 203 and 203' are soaked each with one of the solutions containing gold conjugate, the first solution containing oligonucleotide linked BSA and anti-antibody, the second solution containing complementary oligonucleotide linked BSA. The two different conjugate releasing pads 203 and 203' are fixed to the inner surface of the container 210 at different locations (see Fig. 2b).

**[0036]** In more detail, a detection cup is prepared according to a procedure comprising the following steps:

(a) prepare a phosphate sample buffer containing goat serum, ethylenediamine tetraacetic acid (EDTA), non-fat dry milk, preservative (like sodium azide) and Tween 20;

(b) soak a sample pad with the phosphate sample buffer and heat dry at a temperature around 50°C;

(c) prepare a colloidal gold solution (see Example 2);

(d) conjugate colloidal gold with oligonucleotide linked BSA and anti-antibody to prepare the first conjugate (see Example 2), add Tween 20 to this first conjugate solution, soak a conjugate pad with this first conjugate solution and heat dry at a temperature around 50°C;

(e) conjugation colloidal gold with complementary oligonucleotide linked BSA to prepare the second conjugate (see Example 2), soak a conjugated pas with this second conjugate solution and heat dry at a temperature around 50°C;

(f) print sample zone antigen (i.e. an antigen recognized by the antibody in the sample to be detected, e.g. HIV envelop protein gp160) and control zone antibody (e.g. non-specific antibody) onto a nitrocellulose membrane and heat dry at a temperature around 50°C;

(g) prepare a membrane blocking solution containing Tween 20 and non fat dry milk;

(h) block the nitrocellulose membranes using the blocking solution and heat dry at a temperature around 50°C;

(i) laminate in the following order the components of a test strip to be inserted in the container onto a backing material sheet:

(i.i) laminate the nitrocellulose membrane nearly in the middle of the test strip;

(i.ii) laminate the absorbent pad at the end of the test strip (overlaps from the nitrocellulose membrane side);

(j) assemble the test strip into a plastic housing, i.e. the container;

(k) attach the first and the second conjugate pad inside the container;

(l) pack the detection cup into an aluminium pouch with a silica gel desiccant.

**[0037]** In steps (d) or (e), a sheet can be used instead of a single conjugate pad which is cut into several pads after drying. Similarly, in step (i), a sheet can be used which is cut into strips after having laminated all components.

**[0038]** The antigens used may be synthetic or recombinant. Control and sample zone may be realized by lines. More than one sample zone and/or more than one control zone are also contemplated.

**[0039]** A urine wick can be used in case of urinary testing, while a blood filter is used in case of blood testing.

#### EXAMPLE 7: Urinary HIV IgG detection cup

**[0040]** The first conjugate releasing pad 203 comprises gold conjugate 1, i.e. conjugated anti-antibody, and oligonucleotide 1. The second conjugate releasing pad 203' comprises gold conjugate 2, i.e. conjugated complementary oligonucleotide 2. The sample zone 208 comprises recombinant envelope protein gp160 immobilized onto the nitrocellulose membrane 204. The control zone 209 comprises mouse anti-mouse IgG. Sample and control zones 208, 209 turn into purple colour in case that of HIV IgG is present in the sample; only the control zone 209 turns into purple colour in case of HIV IgG free sample.

#### EXAMPLE 8: Amplification of gold conjugate colour intensity

**[0041]** The colloidal gold conjugate colour intensity was amplified using water-soluble chitosan (or modified water-soluble chitosan) as a colour intensity modification agent. The action of water-soluble chitosan (or modified water-soluble chitosan) in connection with the antibodies or antigens is on the colour intensity of the colloidal gold. Chitosan is added during the preparation of colloidal gold, but prior to the conjugation of colloidal gold with the proteins. Water-soluble chitosan (or modified water-soluble chitosan) affect the colour intensity of colloidal gold and so increases the ability of the human eye to identify the colour, and, thus, enables to detect very low concentrations of the analyte. Signal amplification lies in the range of up to 10folds. Colloidal gold could be prepared by the reduction of 1% aqueous solution of tetrachloroauric acid (HAuCl<sub>4</sub>) using trisodium citrate aqueous solution to produce spheroidal gold particles. After colloidal gold preparation, water-soluble chitosan (or modified water-soluble chitosan) aqueous solution was added with a suitable volume and concentration to convert colour from purple to violet pending on the volume and concentration of the added modification solution.

#### **References**

##### **[0042]**

(1) J Chandler, N Robinson, and K Whiting, "Handling False Signals in Gold-Based Rapid Tests", IVD Technology 7, no. 2 (2001): 34-45; <http://www.devicelink.com/ivdt/archive/01/03/002.html>.

(2) J Chandler, T Gurmin, and N Robinson, "The Place of Gold in Rapid Tests", IVD Technology 6, no. 2 (2000): 37-49; <http://www.devicelink.com/ivdt/archive/00/03/004.html>

(3) TC Tisone et al., "Image Analysis for Rapid-Flow Diagnostics", IVD Technology 5, no. 5 (1999): 52-58; <http://www.devicelink.com/ivdt/archive/99/09/010.html>.

(4) World Health Organization, HIV assays: operational characteristics (Phase I). Report 13: urine specimens, oral fluid (saliva) specimens. [Material originally distributed as WHOBCT/02.08]

(5) Duncan, R.J.S., Weston, P.D., Wrigglesworth, R., 1983. A new reagent which may be used to introduce sulfhydryl groups into proteins, and its use in the preparation of conjugates for immunoassay. Anal. Biochem. 132, 68.

Claims

- 5
1. A rapid immunochromatographic test device for antibody detection, comprising at least a first and a second conjugate releasing pad, wherein the first conjugate pad comprises a gold conjugated protein linked first oligonucleotide and a gold conjugated anti-antibody, and the second conjugate pad comprises a gold conjugated protein linked second oligonucleotide, which second oligonucleotide is complementary to the first oligonucleotide.
  2. The test device according to claim 1 in the form of a test strip.
  - 10 3. The test device according to claim 1 in the form of a detection cup.
  4. The test device according to any of the preceding claims, wherein the first and the second conjugate pads are separated from each other.
  - 15 5. The test device according to any of the preceding claims, wherein the conjugate pads further comprise optionally modified water-soluble chitosan.
  6. A use of a test device according to any of claims 1 to 5 for the detection of an antibody in a sample of urine or saliva of a subject, preferably human.
  - 20 7. The use according to claim 6 wherein the antibody to be detected is selected from antibodies directed against HIV, HCV, and anti-*H. pylori* antigen.
  - 25 8. A method for preparing a test device according to any of claims 1 to 5, comprising the following steps:
    - (a) providing a first conjugate solution comprising a gold conjugated protein linked first oligonucleotide and a gold conjugated anti-antibody, and applying said solution to a first conjugate pad;
    - (b) providing a second conjugate solution comprising a gold conjugated protein linked second oligonucleotide, and applying said solution to a second conjugate pad;
    - 30 (c) providing a membrane comprising an antigen recognized by the antibody to be detected, which antigen is immobilized within a sample zone, and a non-specific antibody immobilized within a control zone.
  - 35 9. The method according to claim 8 for preparing a test device in the form of a test strip, additionally comprising the following steps:
    - (d) placing the membrane onto a backing material;
    - (e) placing the first conjugate pad onto the membrane;
    - (f) placing a divider, preferably a plastic divider, onto the first conjugate pad;
    - 40 (g) placing the second conjugate pad onto the divider.
  - 45 10. The method according to claim 8 for preparing a test device in the form of a detection cup, additionally comprising the following step:
    - (d') placing the membrane and the first and second conjugate pads into a container, wherein the first and second conjugate pads are spaced apart from each other at different locations.
  - 50 11. The method according to any of claims 8 to 10, wherein optionally modified water-soluble chitosan is added during the preparation of the first and second conjugate solutions prior to conjugate colloid gold with the antibody and/or protein linked oligonucleotide.
  - 55 12. The method according to claim 11, wherein the colloidal gold is prepared by reduction of a 1% aqueous solution of tetrachloroauric acid using trisodium citrate aqueous solution to produce spheroidal gold particles.

Figure 1a

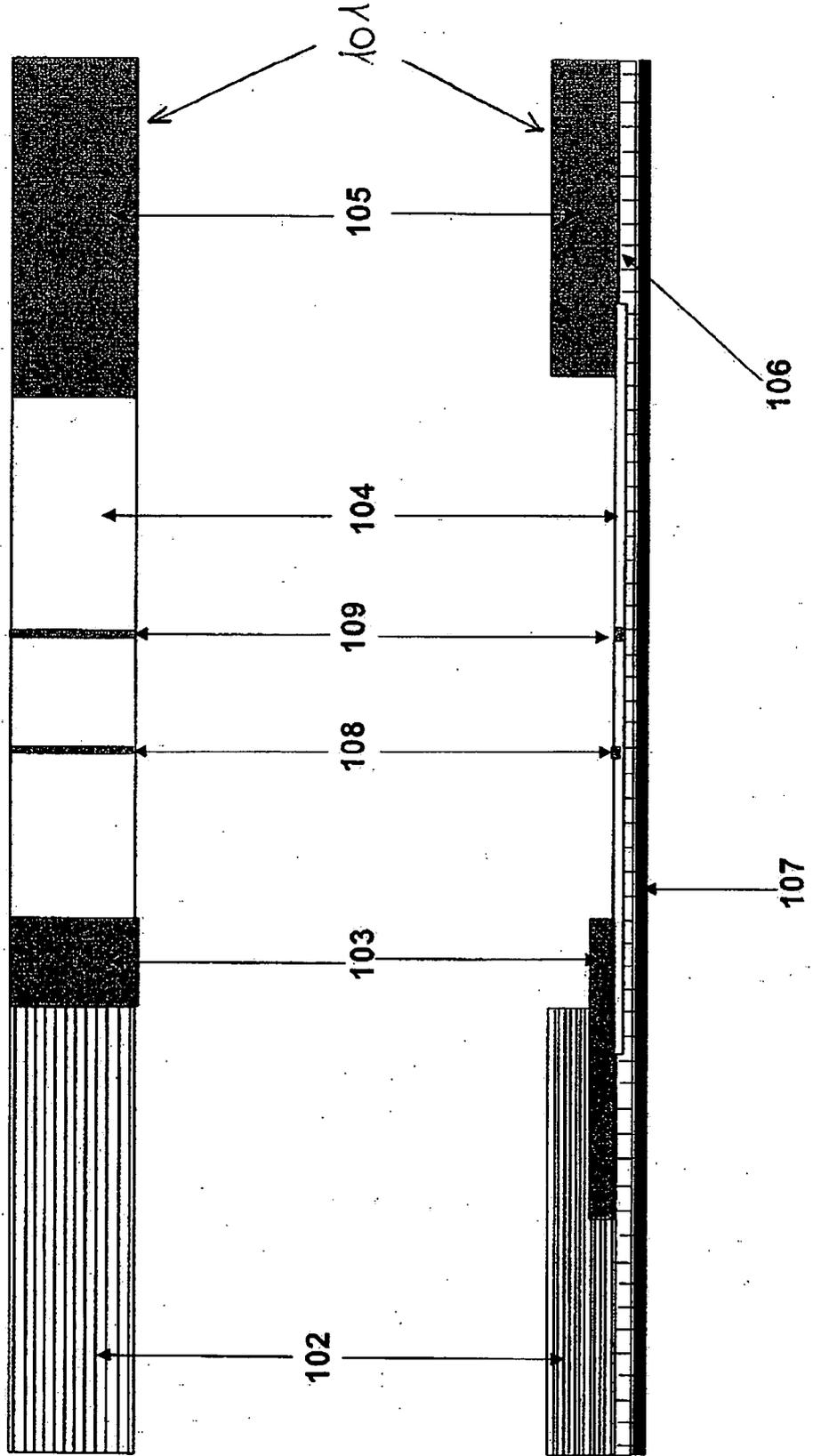


Figure 1b

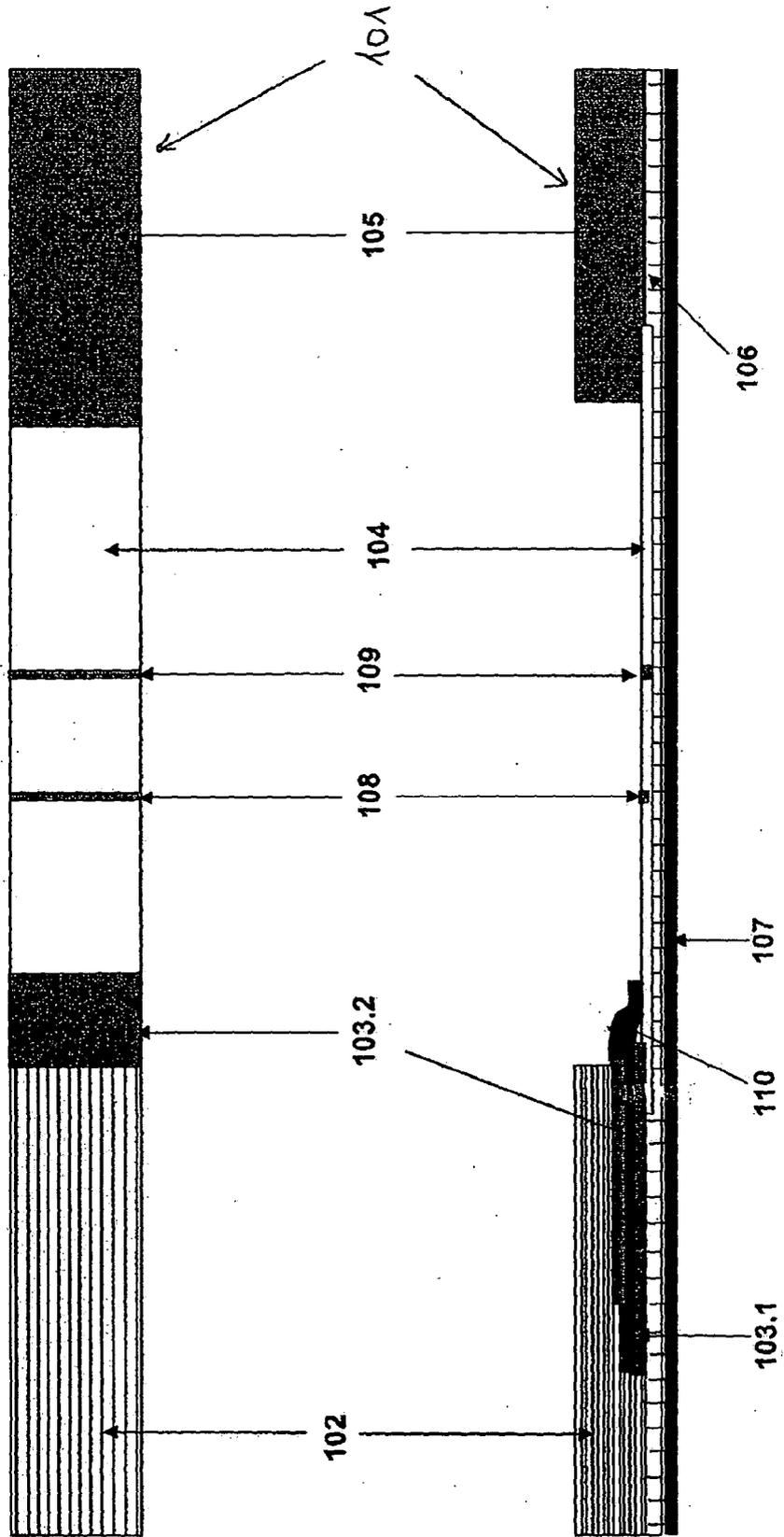


Figure 2a

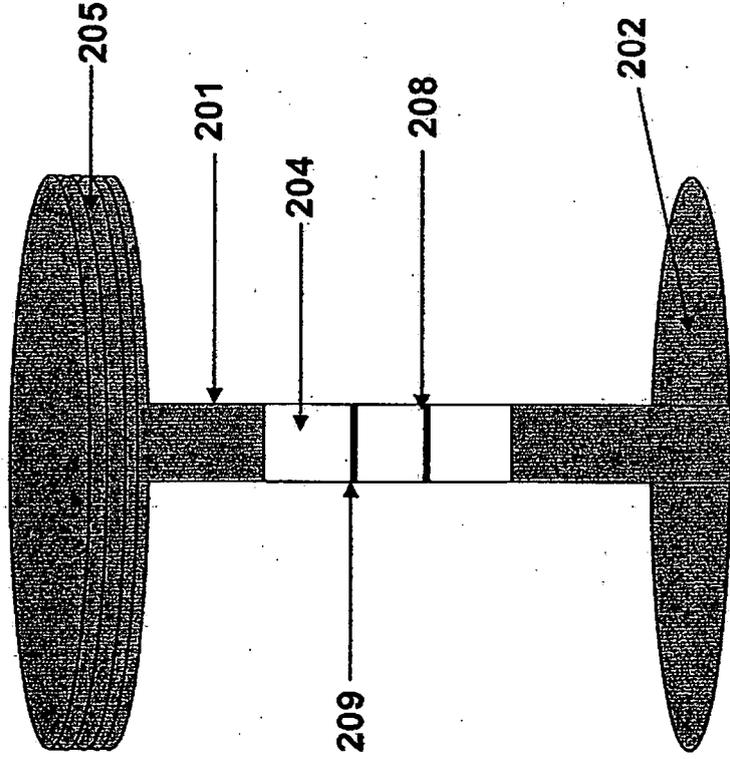
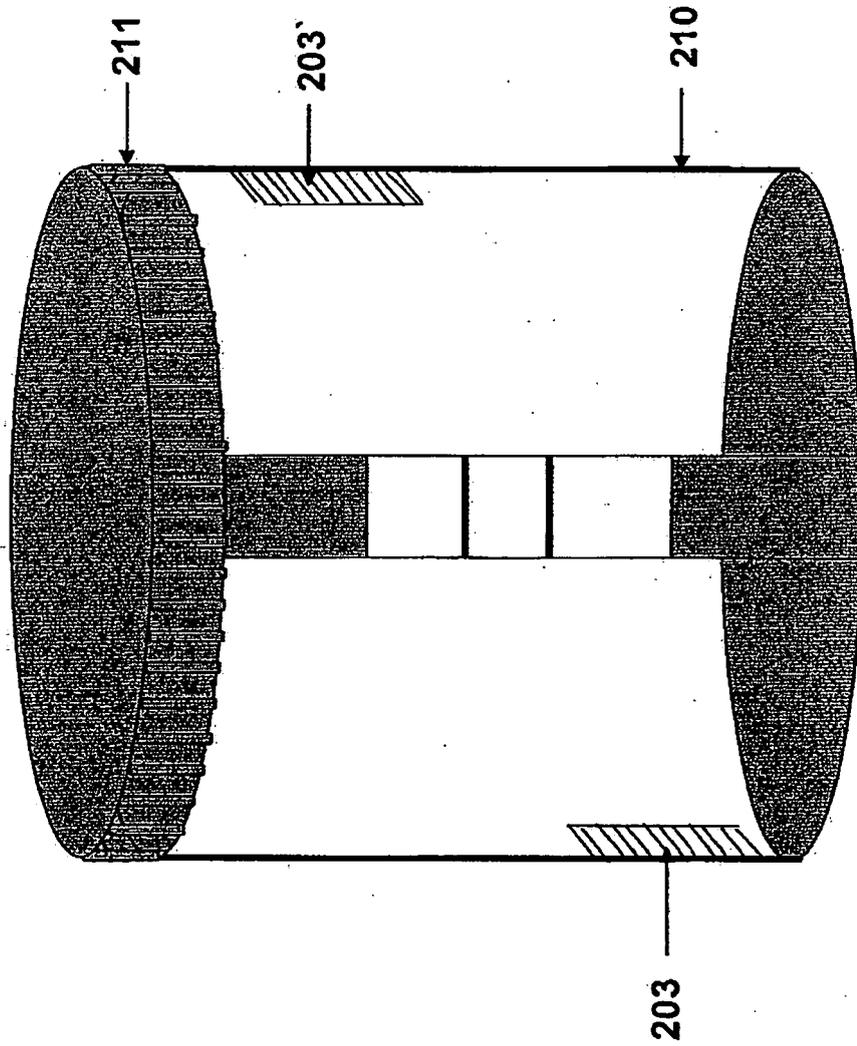


Figure 2b





DOCUMENTS CONSIDERED TO BE RELEVANT			
Category	Citation of document with indication, where appropriate, of relevant passages	Relevant to claim	CLASSIFICATION OF THE APPLICATION (IPC)
X	<p>ANONYMOUS: "DoubleCheckGoldTM HIV 1&amp;2" INTERNET ARTICLE, [Online] September 2006 (2006-09), pages 1-3, XP002423790 Retrieved from the Internet: URL:http://orgenics.com/files/pdf/70632100E.pdf&gt; [retrieved on 2007-03-08] * the whole document * in particular par. "Principle of the Test".</p>	1-4,6-10	<p>INV. G01N33/558 G01N33/543 G01N33/58 C12Q1/68</p>
A	<p>WO 02/46472 A (NANOSPHERE INC; MIRKIN C A; LETSINGER R L; MUCI) 13 June 2002 (2002-06-13) * the whole document * in particular fig. 25b.</p>	1-4,6-10	<p>TECHNICAL FIELDS SEARCHED (IPC)</p> <p>G01N C12Q C07K</p>
A	<p>OKU Y ET AL: "Development of oligonucleotide lateral-flow immunoassay for multi-parameter detection" JOURNAL OF IMMUNOLOGICAL METHODS, vol. 258, no. 1-2, 1 December 2001 (2001-12-01), pages 73-84, XP004311918 * the whole document * in particular fig. 1.</p>	1-4,6-10	
A	<p>SIMS P W ET AL: "IMMUNOPOLYMERASE CHAIN REACTION USING REAL-TIME POLYMERASE CHAIN REACTION FOR DETECTION" ANALYTICAL BIOCHEMISTRY, vol. 281, no. 2, 1 June 2000 (2000-06-01), pages 230-232, XP001147707 * the whole document * in particular fig. 1.</p>	1-4,6-10	
The present search report has been drawn up for all claims			
Place of search The Hague		Date of completion of the search 12 March 2007	Examiner Weber, Peter
<p>CATEGORY OF CITED DOCUMENTS</p> <p>X : particularly relevant if taken alone Y : particularly relevant if combined with another document of the same category A : technological background O : non-written disclosure P : intermediate document</p>		<p>T : theory or principle underlying the invention E : earlier patent document, but published on, or after the filing date D : document cited in the application L : document cited for other reasons &amp; : member of the same patent family, corresponding document</p>	

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EPO FORM 1503 03.82 (P04C001)



DOCUMENTS CONSIDERED TO BE RELEVANT			
Category	Citation of document with indication, where appropriate, of relevant passages	Relevant to claim	CLASSIFICATION OF THE APPLICATION (IPC)
A	SUGUNAN ET AL: "Heavy-metal ion sensors using chitosan-capped gold nanoparticles" SCIENCE AND TECHNOLOGY OF ADVANCED MATERIALS, vol. 6, no. 3-4, April 2005 (2005-04), pages 335-340, XP005039763 * the whole document * in particular abstract; fig. 3a. -----	5,11,12	
			TECHNICAL FIELDS SEARCHED (IPC)
The present search report has been drawn up for all claims			
Place of search The Hague		Date of completion of the search 12 March 2007	Examiner Weber, Peter
CATEGORY OF CITED DOCUMENTS X : particularly relevant if taken alone Y : particularly relevant if combined with another document of the same category A : technological background O : non-written disclosure P : intermediate document		T : theory or principle underlying the invention E : earlier patent document, but published on, or after the filing date D : document cited in the application L : document cited for other reasons ..... & : member of the same patent family, corresponding document	

2  
EPO FORM 1503 03.02 (P04C01)

**ANNEX TO THE EUROPEAN SEARCH REPORT  
ON EUROPEAN PATENT APPLICATION NO.**

EP 06 02 5523

This annex lists the patent family members relating to the patent documents cited in the above-mentioned European search report. The members are as contained in the European Patent Office EDP file on The European Patent Office is in no way liable for these particulars which are merely given for the purpose of information.

12-03-2007

Patent document cited in search report		Publication date	Patent family member(s)	Publication date
WO 0246472	A	13-06-2002	AU 3059302 A	18-06-2002
			EP 1356109 A2	29-10-2003
-----				

EPO FORM P0459

For more details about this annex : see Official Journal of the European Patent Office, No. 12/82

**REFERENCES CITED IN THE DESCRIPTION**

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**Non-patent literature cited in the description**

- **J CHANDLER ; N ROBINSON ; K WHITING.** Handling False Signals in Gold-Based Rapid Tests. *IVD Technology*, 2001, vol. 7 (2), 34-45, <http://www.devicelink.com/ivdt/archive/01/03/002.html>. [0042]
- **J CHANDLER ; T GURMIN ; N ROBINSON.** The Place of Gold in Rapid Tests. *IVD Technology*, 2000, vol. 6 (2), 37-49, <http://www.devicelink.com/ivdt/archive/00/03/004.html> [0042]
- **TC TISONE et al.** Image Analysis for Rapid-Flow Diagnostics. *IVD Technology*, 1999, vol. 5 (5), 52-58, <http://www.devicelink.com/ivdt/archive/99/09/010.html>. [0042]
- World Health Organization, HIV assays: operational characteristics (Phase I). *Report 13: urine specimens, oral fluid (saliva) specimens* [0042]
- **DUNCAN, R.J.S. ; WESTON, P.D. ; WRIGGLESWORTH, R.** A new reagent which may be used to introduce sulfhydryl groups into proteins, and its use in the preparation of conjugates for immunoassay. *Anal. Biochem.*, 1983, vol. 132, 68 [0042]