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(72) Inventors:

- **TOMINAGA, Rumi**
Yokohama-shi
Kanagawa 230-0045 (JP)
- **WADA, Takuji**
Yokohama-shi
Kanagawa 230-0045 (JP)

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(74) Representative: **Denison, Christopher Marcus et al**

(71) Applicant: **Riken**

Wako-shi, Saitama 351-0198 (JP)

Mewburn Ellis LLP

33 Gutter Lane

London

EC2V 8AS (GB)

(54) **MUTANT PLANT AND METHOD FOR PRODUCTION THEREOF**

(57) According to the present invention, a novel mutated plant is provided based on findings obtained by analyzing the function of a functionally unknown gene having a CPC-like Myb sequence.

The function of any one of the following proteins is suppressed:

(a) a protein comprising the amino acid sequence set forth in SEQ ID NO: 2; and

(b) a protein which comprises an amino acid sequence having a substitution, deletion, addition, or insertion of

one or more amino acids in the amino acid sequence set forth in SEQ ID NO:2, and is capable of acting as a transcription factor for controlling epidermal cell differentiation.

A plant having a characteristic phenotype, for example which can grow to a larger size than the wild-type plant, can be obtained by suppressing the function of the above protein.

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Description

Technical Field

5 **[0001]** The present invention relates to a mutated plant having characteristic morphologies and a method for production thereof.

Background Art

10 **[0002]** Non-Patent Document 1 discloses CAPRICE (hereinafter referred to as a CPC protein), which is a protein having the R3 MYB motif and is usually involved in root hair cell differentiation in *Arabidopsis thaliana*. Non-Patent Document 2 describes that cell-to-cell movement of such CPC protein (from non-root hair cells to root hair cells) takes place such that the CPC protein suppresses the expression of the homeodomain-leucine zipper gene (GLABRA2).

15 **[0003]** Four types of CPC-like Myb sequences such as the TRIPTYCHON gene (TRY gene) have been found in the genome of *Arabidopsis thaliana*. Since trichome clusters are observed in a strain lacking the TRY gene, the TRY protein controls trichome formation. Kirik et al. have already identified the two of the above four types of CPC-like Myb sequences (referred to as ETC 1 and ETC2) and genetically analyzed the relationships between such sequences and CPC or TRY (Non-Patent Documents 3 and 4). The group of David Marks analyzed the relationship between TRY and At1g01380 (ETC1) and they demonstrated that GL3 controls TRY gene expression but not ETC1 expression (Non-Patent Document 5).

20 **[0004]** However, other genes each comprising a CPC-like Myb sequence have not been analyzed thus far. Therefore, the functions of such genes remain unknown.

[0005] Non-Patent Document 1: Wada, T., Tachibana, T., Shimura, Y., and Okada, K. (1997). Epidermal cell differentiation in *Arabidopsis* determined by a Myb homolog, CPC. *Science* 277, 1113-1116.

25 **[0006]** Non-Patent Document 2: Wada, T., Kurata, T., Tominaga, R., Koshino-Kimura, Y., Tachibana, T., Goto, K., Marks, M.D., Shimura, Y., and Okada, K. (2002). Role of a positive regulator of root hair development, CAPRICE, in *Arabidopsis* root epidermal cell differentiation. *Development* 129, 5409-5419.

30 **[0007]** Non-Patent Document 3: Kirik, V., Simon, M., Huelskamp, M., and Schiefelbein, J. (2004b). The ENHANCER OF TRY AND CPC1 gene acts redundantly with TRIPTYCHON and CAPRICE in trichome and root hair cell patterning in *Arabidopsis*. *Dev Biol* 268, 506-513.

[0008] Non-Patent Document 4: Kirik, V., Simon, M., Wester, K., Schiefelbein, J., and Huelskamp, M. (2004a). ENHANCER of TRY and CPC 2 (ETC2) reveals redundancy in the region-specific control of trichome development of *Arabidopsis*. *Plant Mol Biol* 55, 389-398.

35 **[0009]** Non-Patent Document 5: Esch, J. J., Chen, M. A., Hillestad, M., and Marks, M.D. (2004). Comparison of TRY and the closely related At1g01380 gene in controlling *Arabidopsis* trichome patterning. *Plant J* 40, 860-869.

Disclosure of the Invention

40 **[0010]** It is an objective of the present invention to provide a novel mutated plant based on findings obtained by analyzing the function of a functionally unknown gene having a CPC-like Myb sequence and to provide a method for producing the mutated plant.

[0011] As a result of analysis of the function of a functionally unknown gene having a CPC-like Myb sequence, the present inventors have found that a strain lacking such gene has characteristic morphologies compared with the wild-type plant. This has led to the completion of the present invention.

45 **[0012]** Specifically, the present invention encompasses the following.

(1) A mutated plant in which the function of any one of the following proteins (a) to (c) is suppressed:

- 50 (a) a protein comprising the amino acid sequence set forth in SEQ ID NO: 2;
 (b) a protein which comprises an amino acid sequence having a substitution, deletion, addition, or insertion of one or more amino acids in the amino acid sequence set forth in SEQ ID NO: 2, and is capable of acting as a transcription factor for controlling epidermal cell differentiation; and
 (c) a protein which is encoded by a polynucleotide having a nucleotide sequence with a substitution, deletion, addition, or insertion of one or more nucleotides in the nucleotide sequence set forth in SEQ ID NO: 1, and is
 55 capable of acting as a transcription factor for controlling epidermal cell differentiation.

(2) The mutated plant according to (1), wherein the mutated plant lacks a gene encoding any one of the above proteins (a) to (c).

- (3) The mutated plant according to (1), wherein the mutated plant is a dicotyledon.
 (4) The mutated plant according to (1), wherein the mutated plant is a plant of the family *Brassicaceae*.
 (5) A method for producing a mutated plant, comprising suppressing the function of any one of the following proteins (a) to (c) in a target plant:

- (a) a protein comprising the amino acid sequence set forth in SEQ ID NO: 2;
 (b) a protein which comprises an amino acid sequence having a substitution, deletion, addition, or insertion of one or more amino acids in the amino acid sequence set forth in SEQ ID NO: 2, and is capable of acting as a transcription factor for controlling epidermal cell differentiation; and
 (c) a protein which is encoded by a polynucleotide having a nucleotide sequence with a substitution, deletion, addition, or insertion of one or more nucleotides in the nucleotide sequence set forth in SEQ ID NO: 1, and is capable of acting as a transcription factor for controlling epidermal cell differentiation.

- (6) The method for producing a mutated plant according to (5), comprising deleting a gene encoding any one of the above proteins (a) to (c).

- (7) The method for producing a mutated plant according to (5), wherein the target plant is a dicotyledon.

- (8) The method for producing a mutated plant according to (5), wherein the target plant is a plant of the family *Brassicaceae*.

- (9) A method for producing a mutated plant which grows to a larger size or flowers earlier than the wild-type plant, comprising suppressing the function of any one of the following proteins (a) to (c) in a target plant:

- (a) a protein comprising the amino acid sequence set forth in SEQ ID NO: 2;
 (b) a protein which comprises an amino acid sequence having a substitution, deletion, addition, or insertion of one or more amino acids in the amino acid sequence set forth in SEQ ID NO: 2, and is capable of acting as a transcription factor for controlling epidermal cell differentiation; and
 (c) a protein which is encoded by a polynucleotide having a nucleotide sequence with a substitution, deletion, addition, or insertion of one or more nucleotides in the nucleotide sequence set forth in SEQ ID NO: 1, and is capable of acting as a transcription factor for controlling epidermal cell differentiation.

- (10) The method for producing a mutated plant according to (9), comprising deleting a gene encoding any one of the above proteins (a) to (c).

- (11) The method for producing a mutated plant according to (9), wherein the target plant is a dicotyledon.

- (12) The method for producing a mutated plant according to (9), wherein the target plant is a plant of the family *Brassicaceae*.

[0013] This description includes part or all of the contents as disclosed in the description of Japanese Patent Application No. 2006-278988, which is a priority document of the present application.

Brief Description of the Drawings

[0014]

Fig. 1-1 is a phylogenetic tree showing the relationship between the CPL3 gene and rice-derived orthologs.

Fig. 1-2 shows the alignment of the amino acid sequences of LOC_Os01g43180, LOC_Os01g43230, CPC, TRY, ETC1, ETC2, and CPL3.

Fig. 2 shows the alignment of the amino acid sequences of the CPC protein, the ETC1 protein, the ETC2 protein, the TRY protein, and the CPL3 protein, and schematically shows the T-DNA insertion sites of mutants each lacking the relevant gene.

Fig. 3 is a characteristic graph showing the results of determining the root hair number in double mutants and triple mutants with respect to the CPC gene, the TYR gene, the ETC1 gene, the ETC2 gene, and the CPL3 gene.

Fig. 4 is a characteristic graph showing the results of determining the trichome number in double mutants and triple mutants with respect to the CPC gene, the TYR gene, the ETC1 gene, the ETC2 gene, and the CPL3 gene.

Fig. 5 is a characteristic graph showing the results of measuring the raw weight in the cpl3-1 mutant, the 35S::CPL3 transformant, the CPL3::CPL3 transformant, and the wild-type plant, at the same growth stage.

Fig. 6 shows images of the cpl3-1 mutant, the 35S::CPL3 transformant, the CPL3::CPL3 transformant, and the wild-type plant, at the same growth stage.

Fig. 7 is a characteristic graph showing the results of measuring the endoreduplication level in the cpl3-1 mutant, the CPL3::CPL3 transformant, and the wild-type plant.

Fig. 8 is a characteristic graph showing the results of determining the true leaf number in each of mutants and transformants.

Fig. 9 is a characteristic graph showing the results of determining the number of days until bolting of individual mutants and transformants.

Fig. 10 shows images indicating the results of observing the cell phenotypes in leaves and hypocotyls of the *cpl3-1* mutant, the CPL3::CPL3 transformant, and the wild-type plant, with the use of an optical microscope.

Fig. 11 is a characteristic graph showing the results of comparing the root hair numbers among transformants each overexpressing the CPC gene, the ETC1 gene, the ETC2 gene, the TRY gene, or the CPL3 gene under the control of the 35S promoter or its own promoter.

Fig. 12 is a characteristic graph showing the results of comparing the trichome numbers among transformants each overexpressing the CPC gene, the ETC1 gene, the ETC2 gene, the TRY gene, or the CPL3 gene under the control of the 35S promoter or its own promoter.

Fig. 13 shows images indicating the results of examining the gene expression pattern with the use of transformants expressing the GUS protein under the control of the promoter of the CPC gene, the ETC1 gene, the ETC2 gene, the TRY gene, or the CPL3 gene.

Fig. 14 shows the results of examining the expression pattern of each of the CPC gene, the ETC 1 gene, the ETC2 gene, the TRY gene, and the CPL3 gene with the use of transformants highly expressing the GFP fusion protein, and the results of examining the expression pattern of the respective gene via RT-PCR and in *situ* hybridization.

Fig. 15 shows real-time PCR results indicating the CO, FT, SOC1, and CPL3 expression levels in *cpc*, *try*, *etc1*, *etc2*, and *cpl3* mutated plants.

Best Mode for Carrying Out the Invention

[0015] Hereinafter, the present invention will be described in greater detail.

[0016] The mutated plant of the present invention is obtained by deleting the function of a specific protein found in the wild-type plant. The mutated plant of the present invention is characterized in that it grows to a larger size than the wild-type plant. Specifically, the mutated plant of the present invention is characterized in that its weight (measured when cultivated to attain a size comparable to that of the wild-type plant under the same conditions) is significantly greater than that of the wild-type plant. In addition, the mutated plant of the present invention is characterized in that it flowers earlier than the wild-type plant.

[0017] The phrase "deleting the function of a protein" used herein is intended to include deletion of a gene encoding the protein from the genome, inhibition of the expression of a gene encoding the protein, and reduction of the activity of the protein.

[0018] More specifically, examples of a method for deleting a gene encoding particular protein that can be used include, but are not limited to, a method involving homologous recombination and a method using a transposon(s). In addition, upon deletion of the gene, the full-length sequence or a partial sequence of the gene may be deleted.

[0019] Further, examples of a method for inhibiting the expression of a gene encoding a particular protein that can be used include, but are not limited to, a method for deleting a promoter that controls the expression of the gene, a method for substituting a promoter that controls the expression of the gene with an expression-inducing promoter, a method for mutating a promoter that controls the expression of the gene, a method for degrading transcription products of the gene via RNA interference, and a method for inhibiting translation of the gene with the use of antisense RNA.

[0020] Furthermore, an example of a method for reducing the activity of a particular protein that can be used includes a method which comprises making a substance having a function to specifically bind to the protein and to suppress its activity to be acted on the protein. Examples of such substance that can be used include an antibody and a suppressor capable of inhibiting the function of the protein.

[0021] In the mutated plant of the present invention, the function of the protein encoded by the gene registered as At4g01060 in *Arabidopsis thaliana* is deleted. Specifically, the gene registered as At4g01060 is herein referred to as the CPL3 gene, which encodes a protein having a CPC-like Myb sequence. The nucleotide sequence of the CPL3 gene is depicted in SEQ ID NO: 1. The amino acid sequence of the CPL3 protein encoded by the CPL3 gene is depicted in SEQ ID NO: 2.

[0022] Thus, the mutated plant of the present invention lacks the function of the following protein (1), (2) or (3):

- (1) a protein comprising the amino acid sequence set forth in SEQ ID NO: 2;
- (2) a protein which comprises an amino acid sequence having a substitution, deletion, addition, or insertion of one or more amino acids in the amino acid sequence set forth in SEQ ID NO: 2, and is capable of acting as a transcription factor for controlling epidermal cell differentiation; or
- (3) a protein which is encoded by a polynucleotide having a nucleotide sequence with a substitution, deletion, addition, or insertion of one or more nucleotides in the nucleotide sequence set forth in SEQ ID NO: 1, and is capable

of acting as a transcription factor that controls epidermal cell differentiation.

[0023] The expression "more amino acids" used herein means, for example, 2 to 33 amino acid residues, preferably 2 to 15 amino acid residues, and more preferably 2 to 5 amino acid residues. In addition, examples of a positions into which a substitution, deletion, addition, or insertion of one or more amino acids is introduced include regions of positions 3 to 5 and 21 to 32, and the position 77 and the following regions in the amino acid sequence set forth in SEQ ID NO: 2. Since these regions are non-CPC-like Myb sequence regions, the function of the CPL3 protein would not be significantly affected by introducing a substitution, deletion, addition, or insertion of one or more amino acids into said regions.

[0024] In addition, the expression "more nucleotides" means, as derived from the above expression "more amino acids," for example, 6 to 99 nucleotides, preferably 6 to 45 nucleotides, and more preferably 6 to 15 nucleotides. Also, examples of a position into which a substitution, deletion, addition, or insertion of one or more nucleotides is introduced include, as derived from the above "more amino acids," the regions of the positions 9 to 15 and 63 to 96, and the position 231 and the following regions in the nucleotide sequence set forth in SEQ ID NO: 1.

[0025] Further, the present invention can be applied to any plant. Thus, the present invention can be applied to any plant as long as the wild-type plant contains a protein defined as above. For instance, the present invention is applied particularly preferably to a dicotyledon and most preferably to a plant of the family *Brassicaceae* represented by *Arabidopsis thaliana*.

[0026] Examples of a plant of the family *Brassicaceae* that can be used include plants belonging to the genus *Brassica* such as napa cabbage (*Hakusai*) (*B. campestris*), Japanese mustard spinach (*Komatsuna*) (*B. campestris* var. *peru-viridis*), *Karashina* (*B. juncea*), *Takana* (*B. juncea* var. *integrifolia*), *Hagoromo kanran* (kale) (*B. oleracea* var. *acephala*), *Habotan* (*B. oleracea* var. *acephala*), cauliflower (*B. oleracea* var. *botrytis*), cabbage (*B. oleracea* var. *capitata*), Brussels sprouts (*B. oleracea* var. *gemmifera*), kohlrabi (*B. oleracea* var. *gongylodes*), broccoli (*B. oleracea* var. *italica*), turnip (*B. rapa*), Bok Choy (*B. rapa* var. *chinensis*), turnip green (*B. rapa* var. *hakabura*), Mizuna (*B. rapa* var. *lancinifolia*), and *Brassica rapa* var. *nippo-oleifera* (*B. rapa* var. *nippo-oleifera*). In addition, examples of a plant of the family *Brassicaceae* that can be used include plants belonging to the genus *Nasturtium* such as Cresson (watercress) (*N. officinale*); plants belonging to the genus *Lepidium* such as Garden cress (cress) (*L. sativum*); plants belonging to the genus *Raphanus* such as seeding radish (*R. sativus*) and radish (*R. sativus* var. *radicula*); plants belonging to the genus *Eruca* such as Rocket (*Rucola*) (*E. vesicaria*); plants belonging to the genus *Wasabia/Eutrema* such as Japanese horseradish (*W. japonica*); plants belonging to the genus *Armoracia* such as horseradish (*A. rusticana*); plants belonging to the genus *Matthiola* such as *Matthiola incana* (stock) (*M. incana*); plants belonging to the genus *Capsella* such as shepherd's purse (*C. bursa-pastoris*); plants belonging to the genus *Arabidopsis* such as *Arabidopsis thaliana* (Thale Cress or Mouse-ear Cress); plants belonging to the genus *Orychophragmus* such as *Orychophragmus violaceus* (*Shokatsusai* or *Murasakihanana*); and plants belonging to the genus *Rorippa*.

[0027] Meanwhile, the mutated plant of the present invention is not limited to the above dicotyledons and it may be a monocotyledon represented by rice. For instance, it is possible to identify a gene corresponding to the CPL3 gene in the rice genome by searching for such gene in a database containing the rice genome sequence data with the use of the amino acid sequence set forth in SEQ ID NO: 2 as a search key. Specifically, it was possible to identify two orthologs of the CPL3 gene in the rice genome database. These two ortholog genes each existing on the chromosome 1 have been registered as LOC_Os01g43180 and LOC_Os01g43230. The CPL3 gene has 63% homology at the amino acid level with LOC_Os01g43180 and 59% homology at the amino acid level with LOC_Os01g43230. The amino acid sequence of the protein encoded by LOC_Os01g43180 is depicted in SEQ ID NO: 3. The amino acid sequence of the protein encoded by LOC_Os01g43230 is depicted in SEQ ID NO: 4.

[0028] In addition, regarding LOC_Os01g43180 and LOC_Os01g43230, homologies with the TRY gene, the CPC gene, the ETC1 gene, and the ETC2 gene were calculated. LOC_Os01g43180 has 61% homology at the amino acid level with CPC, 54% homology at the amino acid level with ETC2, 53% homology at the amino acid level with ETC1, and 51% homology at the amino acid level with TRY. LOC_Os01g43230 has 56% homology at the amino acid level with ETC2, 54% homology at the amino acid level with CPC, 49% homology at the amino acid level with ETC1, and 44% homology at the amino acid level with TRY. Fig. 1-1 is a phylogenetic tree showing the above results. Further, fig. 1-2 shows the results of alignment of the amino acid sequences of LOC_Os01g43180, LOC_Os01g43230, CPC, TRY, ETC1, ETC2, and CPL3.

[0029] As described above, LOC_Os01g43180 and LOC_Os01g43230 found in the rice genome have the highest homology with CPL3 among proteins each comprising a CPC-like Myb sequence. Therefore, it can be judged that LOC_Os01g43180 and LOC_Os01g43230 have a high probability of having functions equivalent to those of the CPL3 gene of *Arabidopsis thaliana*.

[0030] The mutated plant of the present invention as defined above is characterized in that it grows to a larger size than the wild-type plant. Thus, in a case in which a target plant for mutation is edible, productivity improvement can be expected as a result of size enlargement according to the present invention. In addition, in a case in which a substance is produced in a target plant for mutation, it is possible to improve the productivity of such useful substance according

to the present invention. Herein, such substance may be a substance originally produced by the wild-type plant or a substance produced via transformation of the wild-type plant.

[0031] Further, the mutated plant of the present invention is characterized in that it flowers earlier than the wild-type plant. Therefore, fruit or seed formation can be promoted and thus the mutated plant of the present invention is advantageous in that it is possible to harvest a crop that can usually be harvested just once a year a plurality of times a year. Moreover, such characteristics of the mutated plant of the present invention enable to save time and cost for crop harvesting.

[0032] The present invention is hereafter described in greater detail with reference to the following examples, although the scope of the present invention is not limited thereto.

(Experimental example 1) Screening for a mutated plant lacking the CPL3 gene

[0033] In this Example, the *Arabidopsis thaliana* ecotype Col-0 (hereinafter referred to as the "wild-type *Arabidopsis thaliana*") was used as the wild-type plant. In this Example, a cpl3-1 mutant in which a gene comprising the nucleotide sequence set forth in SEQ ID NO: 1 had been mutated was screened from the Wisconsin T-DNA group.

[0034] Further, in this Example, a try-29760 mutant in which the TRY gene had been mutated, an etc1-1 mutant in which the ETC1 gene had been mutated, and an etc2-40390 mutant in which the ETC2 gene had been mutated were isolated from the SALK T-DNA group. In addition, a cpc-2 mutant in which the CPC gene had been mutated (disclosed in Kurata, T et al. Cell-to-cell movement of the CAPRICE protein in *Arabidopsis* root epidermal cell differentiation. Development 132, 5387-5398 (2005)) was prepared. Fig. 2 shows T-DNA insertion sites in the obtained mutants.

(Experimental example 2) Transformed plant preparation

[0035] In this Example, the following transformants were prepared: transformants constitutively expressing the CPC gene, the ETC1 gene, the ETC2 gene, the TRY gene, and the CPL3 gene under the control of the 35S promoter; transformants highly expressing GFP fusion proteins, each of which comprises the respective gene linked to the GFP gene; and transformants expressing the GUS protein, each of which comprises the GUS gene linked to the promoter sequence of the respective gene.

[0036] Names, nucleotide sequences, and SEQ ID NOs of promoters used in this Example are listed in table 1.

Table 1

Primer name	Sequence (5' → 3')	SEQ ID NO
TW1165	ATATGGTACCAATAAAAAATAAATCAC	SEQ ID NO: 5
TW1166	TGCTTGTCGACTGTATACACTAA	SEQ ID NO: 6
TW1167	ATATGGTACCTTTAACATAGAAACCGAC	SEQ ID NO: 7
TW1168	ATATGTCGACATCTACGACTTAGCTTC	SEQ ID NO: 8
TW1169	ATATGGTACCACTTCATGTTCTTCCCTT	SEQ ID NO: 9
TW1170	ATATGTCGACAAGCCAATACATATCCA	SEQ ID NO: 10
RT46	GGCCAGTCGACAGAAAACCTCACTCACTATTACATC	SEQ ID NO: 11
RT47	CGAGGATCCACGCTGCGTATTCATCTCAA	SEQ ID NO: 12
RT48	GGCCAGTCGACGCTTGGCTAGCTCATAAACG	SEQ ID NO: 13
RT49	CGATCTAGAACGGTTGGTATTATCCATAACTACT	SEQ ID NO: 14
RT50	GGCCAGTCGACAGCCCTGAAAACAGCTAAGAA	SEQ ID NO: 15
RT51	CGAGGATCCGCGATGGTTATCCATGTCAAAC	SEQ ID NO: 16
RT67	ATATGTCGACAGAAAACCTCACTCACTATTACATC	SEQ ID NO: 17
RT68	ATATCCCGGGACGTAATTGAGATCTTCGATGATTC	SEQ ID NO: 18
RT69	ATATGTCGACGCTAGCTCATAAACGTTGGTACG	SEQ ID NO: 19
RT70	ATATGATATCCAATTTTAGATTTTCTTGGAGATTAAG	SEQ ID NO: 20
RT71	ATATGTCGACAGCCCTGAAAACAGCTAAGAA	SEQ ID NO: 21
RT72	ATATGATATCATTTTTCATGACCCAAAACCTCT	SEQ ID NO: 22
RT73	CGATGGATAACACTGACCGTCGTC	SEQ ID NO: 23
RT88	ATATGGATCCACGGTCAGTGTTATCCATTACTATT	SEQ ID NO: 24
RT89	ATATGTCGACCTCAATATATCAAATTCAAACATTCA	SEQ ID NO: 25

(continued)

	Primer name	Sequence (5' → 3')	SEQ ID NO
	RT90	ATATCCCGGGGAAGGATAGATAGAAAAGCGAG	SEQ ID NO: 26
5	RT91	ATATGGATCCGTTGGACATTTCTTCTCTCTC	SEQ ID NO: 27
	RT92	ATATGTCGACCTAACCGCATGGATTAAAGTTG	SEQ ID NO: 28
	RT122	GCGATCGTAAATCTTTGTGTACTAAG	SEQ ID NO: 29
	RT123	CTCAGGAACAAAAGTGCAGAATTAC	SEQ ID NO: 30
	RT124	GATAATACCAACCGTCTTCGTCTTC	SEQ ID NO: 31
10	RT125	TTCTTGGAGATTAAGAGGAGAAGTAG	SEQ ID NO: 32
	RT126	GATAACCATCGCAGGACTAAGC	SEQ ID NO: 33
	RT127	TACAACGGAATATAATCGAAACAATC	SEQ ID NO: 34
	RT128	CTTCTTGTTTCTCGAGATTATTCTC	SEQ ID NO: 35
15	RT129	AATAGTAATTCAAGGACAGGTACATTTC	SEQ ID NO: 36

[0037] In order to obtain transformants constitutively expressing the respective gene under the control of the 35S promoter, the following constructs were prepared. Specifically, a 0.7-kb PCR fragment containing the ETC1 gene was first amplified using TW1169 and TW1170. Also, a 1.4-kb PCR fragment containing the ETC2 gene was amplified using TW1165 and TW1166. Further, a 0.8-kb PCR fragment containing the CPL3 gene was amplified using TW1167 and TW1168. Furthermore, a 1.4-kb PCR fragment containing the TRY gene was amplified using RT91 and RT92. Next, the obtained PCR fragments were each subcloned into pBluescript SK+ (Stratagene) with the use of Pyrobest DNA polymerase (Takara, Japan). The prepared plasmids were designated as pBS-ETC1, pBS-ETC2, pBS-CPL3, and pBS-TRY.

[0038] Next, fragments obtained by treating pBS-ETC1, pBS-ETC2, and pBS-CPL3 with *Acc65I* and *SaI* were separately inserted into the *Acc65I* and *SaI* sites of a pCHF3 binary vector (Jarvis, P. et al., An Arabidopsis mutant defective in the plastid general protein import apparatus. *Science* 282, 100-103 (1998)) such that 35S::ETC1, 35S::ETC2, and 35S::CPL3 constructs were prepared. In addition, 35S::TRY construct was prepared in the manner described above, except that pBS-TRY was treated with *Bam*HI and *SaI*.

[0039] The following constructs were prepared in order to obtain transformants highly expressing a GFP fusion protein, each of which comprises the respective gene linked to the GFP gene. Specifically, a 2.3-kb PCR fragment containing the ETC1 gene was amplified using RT67 and RT68. The resultant was subjected to restriction enzyme treatment with the use of *SaI* and *SmaI*. Also, a 4.0-kb PCR fragment containing the ETC2 gene was amplified using RT69 and RT70. The resultant was digested with *SaI* and *EcoRV*. Further, a 3.0-kb PCR fragment was amplified using RT71 and RT72. The resultant was subjected to restriction enzyme treatment with the use of *SaI* and *EcoRV*. Furthermore, a 4.0-kb PCR fragment was amplified using RT89 and RT90. The resultant was subjected to restriction enzyme treatment with the use of *SaI* and *SmaI*. The obtained fragments were separately inserted into the *SaI* and *EcoRV* sites of pBS-2xGFP (Kurata, T. et al., Cell-to-cell movement of the CAPRICE protein in Arabidopsis root epidermal cell differentiation. *Development* 132, 5387-5398 (2005)). Accordingly, pBS-ETC1:2xGFP, pBS-ETC2:2xGFP, pBS-CPL3:2xGFP, and pBS-TRY:2xGFP were prepared.

[0040] Next, a fragment obtained by treating pBS-ETC1:2xGFP with *SaI* and *XbaI* was inserted into the *SaI* and *XbaI* sites of a pJHA212K binary vector (Yoo, S. Y. et al., The 35S promoter used in a selectable marker gene of a plant transformation vector affects the expression of the transgene. *Planta* 221, 523-530 (2005)). In addition, a fragment obtained by treating pBS-ETC2:2xGFP with *SaI* and *SacI* was inserted into the *SaI* and *SmaI* sites of a pJHA212K binary vector. Further, fragments obtained by treating pBS-CPL3:2xGFP and pBS-TRY:2xGFP with *SaI* and *SacI* were separately inserted into the *SaI* and *SacI* sites of a pJHA212K binary vector.

[0041] The following constructs were prepared in order to obtain transformants expressing the GUS protein, each of which comprises the GUS gene linked to the promoter sequence of the respective gene. Specifically, a 1.9-kb PCR fragment containing the ETC1 gene promoter region was amplified using RT46 and RT47. Also, a 3.0-kb PCR fragment containing the ETC2 gene promoter region was amplified using RT48 and RT49. Further, a 2.4-kb PCR fragment containing the CPL3 gene promoter region was amplified using RT50 and RT51. Furthermore, a 3.0-kb PCR fragment containing the TRY gene promoter region was amplified using RT88 and RT89. The obtained amplification fragments were each treated with *NotI* and *AccI* and then subcloned into pBS. The prepared plasmids were designated as pBS-ETC1, pBS-ETC2, pBS-CPL3, and pBS-TRY, respectively.

[0042] Next, fragments obtained by treating pBS-ETC1 and pBS-CPL3 with *SaI* and *Bam*HI were separately inserted into the *SaI* and *Bam*HI sites of a pBI101 binary vector (Clontech Laboratories, Inc., CA, USA). The obtained plasmids were designated as ETC1promoter::GUS and CPL3promoter::GUS. In addition, fragments obtained by treating pBS-

ETC2 and pBS-TRY with *Sall* and *XbaI* were separately inserted into the *Sall* and *XbaI* sites of a pBI101 binary vector. The obtained plasmids were designated as ETC2promoter::GUS and TRYpromoter::GUS.

[0043] Transformation was carried out in accordance with the method disclosed in Kurata, T. et al., The YORE-YORE gene regulates multiple aspects of epidermal cell differentiation in Arabidopsis. Plant J 36, 55-66 (2003) with the use of the resultant 35S::ETC1, 35S::ETC2, 35S::CPL3, ETC1::ETC1:2xGFP, ETC2::ETC2:2xGFP, CPL3::CPL3:2xGFP, TRY::TRY:2xGFP, ETC1 promoter::GUS, CPL3 promoter::GUS, ETC2 promoter::GUS, and TRY promoter::GUS.

[Example 1] The phenotype of a mutant lacking the CPL3 gene

[0044] Double mutants and triple mutants with respect to the CPC gene, the TRY gene, the ETC1 gene, the ETC2 gene, and the CPL3 gene were identified by identifying homozygotes of the *etc1-1* mutant, the *etc2-40390* mutant, and the *cpl3-1* mutant isolated in Experimental example 1 in the F2 generation, by PCR. Fig. 3 shows the results of determining the root hair numbers in mutants, the obtained double mutants and triple mutants with respect to the respective genes.

[0045] As seen from fig. 3, the root hair number slightly decreased in the *cpl3-1* mutant. In addition, the root hair number significantly decreased or substantially no root hair was observed in the double mutant with respect to the CPC gene and the TRY gene, the double mutant with respect to the CPC gene and the ETC1 gene, and the triple mutant with respect to the TRY gene, the CPL3 gene, and the CPC gene.

[0046] Fig. 4 shows the results of determining the trichome number in mutants, the obtained double mutants and triple mutants, with respect to the respective gene. As seen from fig. 4, the trichome number increased in the *cpl3-1* mutant compared with the wild-type plant, as in the case of the *cpc-2* mutant. Also, the trichome number further increased in the triple mutant with respect to the TRY gene, the CPC gene, and the CPL3 gene. Further, in the quadruple mutant with respect to the TRY gene, the CPC gene, the ETC1 gene, and the CPL3 gene, a further increased number of trichomes were formed and the entire primary surfaces of leaves were covered with trichomes.

[0047] In addition, the *cpl3-1* mutant isolated in Experimental example 1 and the transformants prepared in Experimental example 2 were observed in terms of growth.

[0048] Fig. 5 shows the results of measuring the raw weight in the respective plant at the same growth stage. Fig. 6 shows images of the respective plant at such stage. As seen from figs. 5 and 6, the mutant lacking the CPL3 gene grew to a larger size than the wild-type plant. On the other hand, the growth of transformant overexpressing the CPL3 gene was equivalent to 30% of the growth of the wild-type plant. In addition, the growth of each of the mutants lacking the CPC gene, the ETC 1 gene, the ETC2 gene, or the TRY gene was merely comparable to the growth of the wild-type plant (the results are not shown).

[0049] Further, fig. 7 shows the results of polyploidy verification for the respective plant. Incidentally, the respective plant 2 weeks after seeding was subjected to measurement for polyploidy with the use of a Ploidy Analyzer PA Flow Cytometer (Partec) according to the manufacturer's instructions. As seen from fig. 7, the 16C peak significantly increased in the mutant lacking the CPL3 gene. The results suggest that the mutant lacking the CPL3 gene exhibits a phenotype related to an improvement in cell size due to promoted endoreduplication.

[0050] Further, fig. 8 shows the results of determining the true leaf number in the respective plant. Fig. 9 shows results of determining the number of days until bolting of the respective plant. As seen from figs. 8 and 9, the mutant lacking the CPL3 gene flowered earlier than the wild-type plant and the other mutants and transformants.

[0051] In addition, leaf and hypocotyl cell phenotypes of the respective plants were observed with an optical microscope. Fig. 10 shows the results. As seen from fig. 10, the following features were exhibited by the mutant lacking the CPL3 gene compared with the wild-type plant and transformants overexpressing the CPL3 gene: enlargement in leaf epidermal cells and elongation in hypocotyls. In addition, table 2 lists the results of measuring the leaf size and determining epithelial cell number for the respective plant.

Table 2

Genotype	Leaf size (mm ²)	Epithelial cells (number / mm ²)	Epithelial cells / leaf
Col-0	16.8±0.3	181±19	3035±311
<i>cpl3</i>	19.0±1.3	158±26	3014±535
<i>CPL3::CPL3</i>	9.5±0.9	311±14	2960±306

[0052] Upon epithelial cell number determination, third leaves were collected 2 weeks after seeding and washed with a washing liquid containing the following components at the following ratio: chloral hydrate (weight) : glycerol (volume) : water (volume) = 8:2:1. Then, visualization was carried out with a Zeiss Axioplan 2 (Carl Zeiss, Germany). In table 2, each value represents the average value of measurement results for five leaves.

[0053] Based on the results shown in fig. 10 and table 2, the mutant lacking the CPL3 gene grew to a larger size than

the wild-type plant, although the cell number of the mutant did not increase beyond that of the wild-type plant. Thus, it has been revealed that growth at the individual cell level is promoted in the mutant lacking the CPL3 gene to a greater extent than those of the wild-type plant.

[Example 2]

[0054] In this Example, transformants overexpressing the CPC gene, the ETC1 gene, the ETC2 gene, the TRY gene, or the CPL3 gene under the control of the 35S promoter or its own promoter were compared with one another in terms of root hair number with the use of the transformants prepared in Experimental example 2. Fig. 11 shows the results. As shown in fig. 11, every transformant constitutively expressing the relevant gene under the 35S promoter exhibited a higher root hair number than the wild-type plant. However, the transformant expressing the CPL3 gene under the control of the 35S promoter exhibited a moderately higher root hair number than the other transformants. Further, the transformant overexpressing the ETC1 gene under the control of the ETC1 promoter exhibited a higher root hair number than the other transformants overexpressing the relevant gene under the control of its own promoter, and the wild-type plant.

[0055] In addition, fig. 12 shows the results of determining the trichome number in the respective transformant in the manner described above. As shown in fig. 12, all transformants, excluding the transformant overexpressing the ETC2 gene under the control of the ETC2 promoter, completely lacked trichomes. Also, the transformant overexpressing the ETC2 gene under the control of the ETC2 promoter had a sharply lower trichome number than the wild-type plant.

[Example 3]

[0056] In this Example, the gene expression pattern was examined with the use of the transformants expressing the GUS protein under the control of any promoter of the above genes, which had been prepared in Experimental example 2. Fig. 13 shows the results. In addition, the expression patterns of gene products of the individual genes were examined with the use of the transformants highly expressing the GFP fusion protein (each of which comprises the respective gene), prepared in Experimental example 2. Fig. 14 shows the results. Fig. 14 also shows the results of examining the expression pattern of the respective gene by RT-PCR and *in situ* hybridization.

[0057] As shown in figs. 13 and 14, gene products of the CPC gene, TRY gene, and ETC1 gene accumulated specifically in non-root hair cells (hairless cells) of young-leaf trichomes and of root epidermis. In addition, gene products of the ETC2 gene and CPL3 gene accumulated specifically in leaf epidermal cells comprising adaxial-side stomatal guard cells, and not in roots and trichomes. In particular, the CPL3 gene was expressed in a limited manner in stomatal guard cells of leaves, cotyledons, hypocotyls, and petioles, compared to the ETC3 gene.

[Example 4]

[0058] In this Example, *cpc*, *try*, *etc1*, *etc2*, and *cpl3* knockout mutants identified as in the case of Example 1 and a CPL3-overexpressing plant prepared as in the case of Experimental example 2 were examined in terms of time of flowering and leaf number.

[0059] Table 3 shows the results of examining the above mutant plants in terms of time of flowering and leaf number at flowering. Incidentally, the data is expressed as the mean \pm SD from at least 10 plants for a single experiment.

Table 3

Phenotype	Time of flowering (day)	Leaf number
Col-0	37.6 \pm 0.5	17.4 \pm 0.6
<i>cpl3</i>	28.9 \pm 0.5	8.2 \pm 0.3
<i>cpc</i>	39.6 \pm 0.5	19.2 \pm 0.6
<i>try</i>	36.0 \pm 0.8	14.3 \pm 0.6
<i>etc1</i>	40.6 \pm 0.8	19.3 \pm 0.8
<i>etc2</i>	39.0 \pm 0.5	14.1 \pm 1.9
35S:CPL3	41.1 \pm 1.1	28.5 \pm 1.7

[0060] As shown in table 3, the *cpl3* knockout-mutated plant flowered earlier than the wild-type plant (28.9 \pm 0.5 days versus 37.6 \pm 0.5 days). In addition, the leaf number thereof was lower than that of the wild-type plant (8.2 \pm 0.3 versus 17.4 \pm 0.6). On the other hand, there was no significant difference between the wild-type plant and the *cpc*, *try*, *etc1*, or *etc2* mutated plant (table 3). The 35S:CPL3 transgenic plant flowered slightly later than the wild-type plant (41.1 \pm 1.1 versus 37.6 \pm 0.5). In addition, the leaf number thereof was higher than that of the wild-type plant (28.5 \pm 1.7 versus

17.4 ± 0.6).

[0061] Further, in order to clarify the effects of the CPL3 gene upon flowering, plant lineages with altered CPL3 expression were examined by real-time PCR in terms of the expression of the following genes known to play a central role in the control of floral transition (flowering): FLOWERING LOCUS T (FT), SUPPRESSOR OF OVEREXPRESSION OF CO 1 (SOC1), and CONSTANS (CO) (Bagnall, *Annals of Botany* 71, 75, 1993; Lee et al., *Genes & development* 14, 2366-2376, 2000; Koornneef et al., *Mol Gen Genet* 229, 57-66, 1991).

[0062] Real-time PCR was conducted as follows. Total RNA was extracted using an RNeasy plant mini kit (Qiagen). On-column DNase I digestion was carried out during RNA purification in accordance with protocols described in the RNeasy mini Handbook. The first-strand cDNA was synthesized from total RNA (1 µg) in a reaction liquid mixture (20 µL) with the use of a Prime Script RT reagent kit (Takara). Real-time PCR was performed via the Chromo4 Real-Time PCR Detection System (Bio-Rad, Hercules, CA, USA) with the use of an SYBR Premix Ex Taq (Takara). PCR amplification included a denaturation step at 95°C for 30 seconds, followed by reaction at 95°C for 5 seconds and 60°C for 30 seconds (45 cycles for CPL3 and 40 cycles for CO, FT, SOC1, and ACT2). The relative mRNA levels were calculated with iQ5 software (Bio-Rad) and normalized to the ACT2 mRNA concentration.

[0063] The FT and SOC1 expression levels of the cpl3 knockout mutant were higher than those of the wild-type plant under long day (LD) conditions during leaf growth. The FT expression levels of 21-day-old leaves of the cpl3 knockout mutant were approximately 7.5-fold higher than those of the wild-type plant. The SOC1 expression levels thereof were 3.2-fold higher than the same (figs. 15A and 15B). There was no significant difference between the cpl3 knockout mutant and the wild-type plant in terms of CO expression (fig. 15C). These results indicate that CPL3 is involved in flowering regulation through suppression of FT and SOC 1 expression.

Industrial Applicability

[0064] According to the present invention, it is possible to provide a novel mutated plant that grows to a larger size or flowers earlier than the wild-type plant and a method for production thereof. The mutated plant of the present invention achieves a larger size than that of the wild-type plant. Thus, for instance, the productivity of a substance extracted from a plant can be improved. Also, the mutated plant of the present invention flowers earlier than the wild-type plant and thus fruit or seed formation can be improved. For instance, it is possible to harvest a crop that can generally be harvested just once a year a plurality of times a year. Further, such characteristics of the mutated plant of the present invention result in time and cost saving for crop harvesting.

[0065] All publications, patents, and patent applications cited herein are incorporated herein by reference in their entirety.

SEQUENCE LISTING

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Claims

1. A mutated plant in which the function of any one of the following proteins (1) to (3) is suppressed:

- (1) a protein comprising the amino acid sequence set forth in SEQ ID NO: 2;
 (2) a protein which comprises an amino acid sequence having a substitution, deletion, addition, or insertion of one or more amino acids in the amino acid sequence set forth in SEQ ID NO: 2, and is capable of acting as a transcription factor for controlling epidermal cell differentiation; and
 (3) a protein which is encoded by a polynucleotide having a nucleotide sequence with a substitution, deletion, addition, or insertion of one or more nucleotides in the nucleotide sequence set forth in SEQ ID NO: 1, and is capable of acting as a transcription factor for controlling epidermal cell differentiation.

2. The mutated plant according to claim 1, wherein the mutated plant lacks a gene encoding any one of the proteins (1) to (3).

3. The mutated plant according to claim 1, wherein the mutated plant is a dicotyledon.

4. The mutated plant according to claim 1, wherein the mutated plant is a plant of the family *Brassicaceae*.

5. A method for producing a mutated plant, comprising suppressing the function of any one of the following proteins (1) to (3) in a target plant:

- (1) a protein comprising the amino acid sequence set forth in SEQ ID NO: 2;
 (2) a protein which comprises an amino acid sequence having a substitution, deletion, addition, or insertion of one or more amino acids in the amino acid sequence set forth in SEQ ID NO: 2, and is capable of acting as a transcription factor for controlling epidermal cell differentiation; and
 (3) a protein which is encoded by a polynucleotide having a nucleotide sequence with a substitution, deletion, addition, or insertion of one or more nucleotides in the nucleotide sequence set forth in SEQ ID NO: 1, and is capable of acting as a transcription factor for controlling epidermal cell differentiation.

6. The method for producing a mutated plant according to claim 5, comprising deleting a gene encoding any one of the proteins (1) to (3).

7. The method for producing a mutated plant according to claim 5, wherein the target plant is a dicotyledon.

8. The method for producing a mutated plant according to claim 5, wherein the target plant is a plant of the family *Brassicaceae*.

9. A method for producing a mutated plant which grows to a larger size or flowers earlier than the wild-type plant, comprising suppressing the function of any one of the following proteins (1) to (3) in a target plant:

- (1) a protein comprising the amino acid sequence set forth in SEQ ID NO: 2;
 (2) a protein which comprises an amino acid sequence having a substitution, deletion, addition, or insertion of one or more amino acids in the amino acid sequence set forth in SEQ ID NO: 2, and is capable of acting as a transcription factor for controlling epidermal cell differentiation; and
 (3) a protein which is encoded by a polynucleotide having a nucleotide sequence with a substitution, deletion, addition, or insertion of one or more nucleotides in the nucleotide sequence set forth in SEQ ID NO: 1, and is capable of acting as a transcription factor for controlling epidermal cell differentiation.

10. The method for producing a mutated plant according to claim 9, comprising deleting a gene encoding any one of the proteins (1) to (3).

11. The method for producing a mutated plant according to claim 9, wherein the target plant is a dicotyledon.

12. The method for producing a mutated plant according to claim 9, wherein the target plant is a plant of the family *Brassicaceae*.

Fig. 1-1

[GENETYX-MAC: Phylogenetic tree]

Date: 2006.07.13

Method: NJ

Bootstrap trial: 10

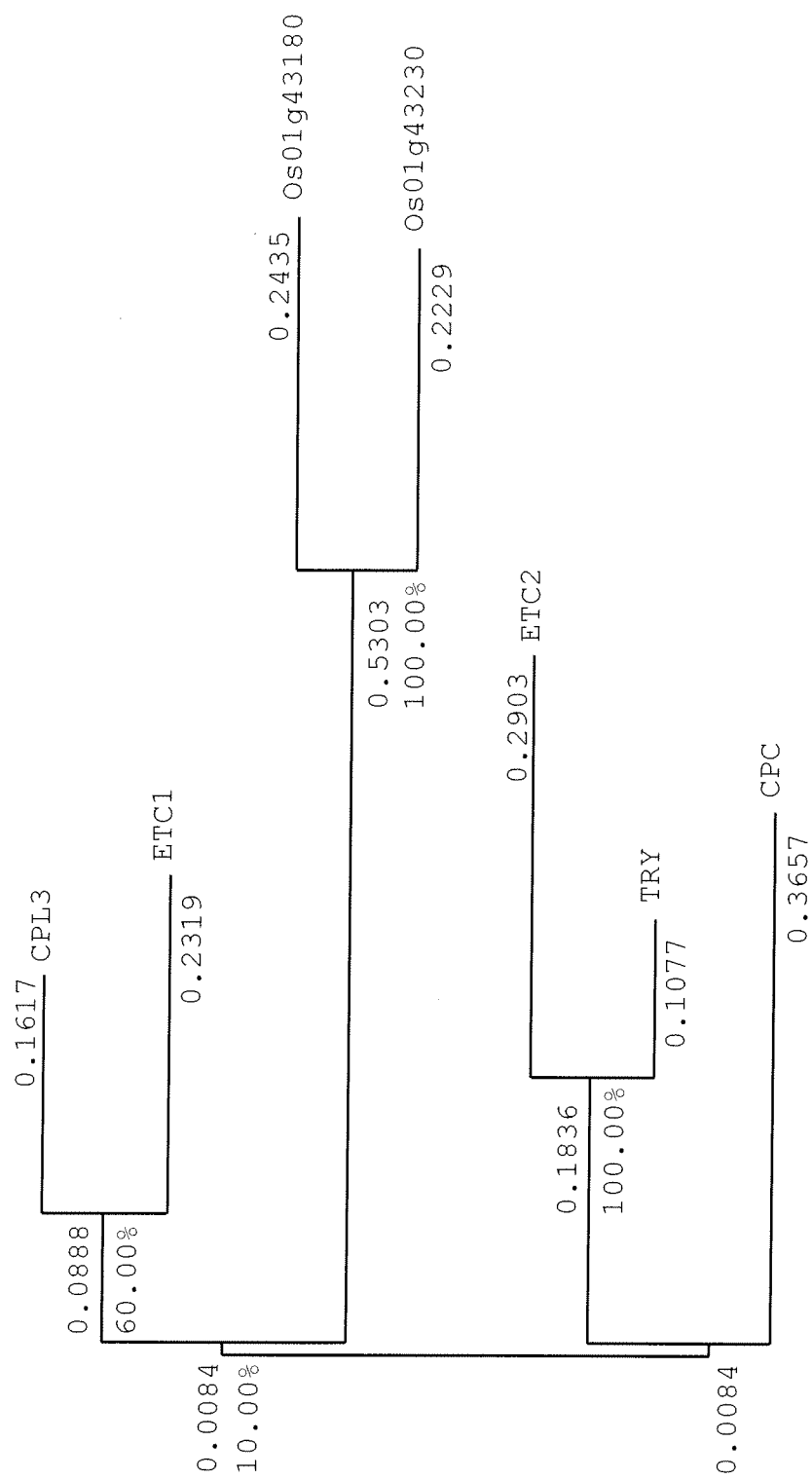
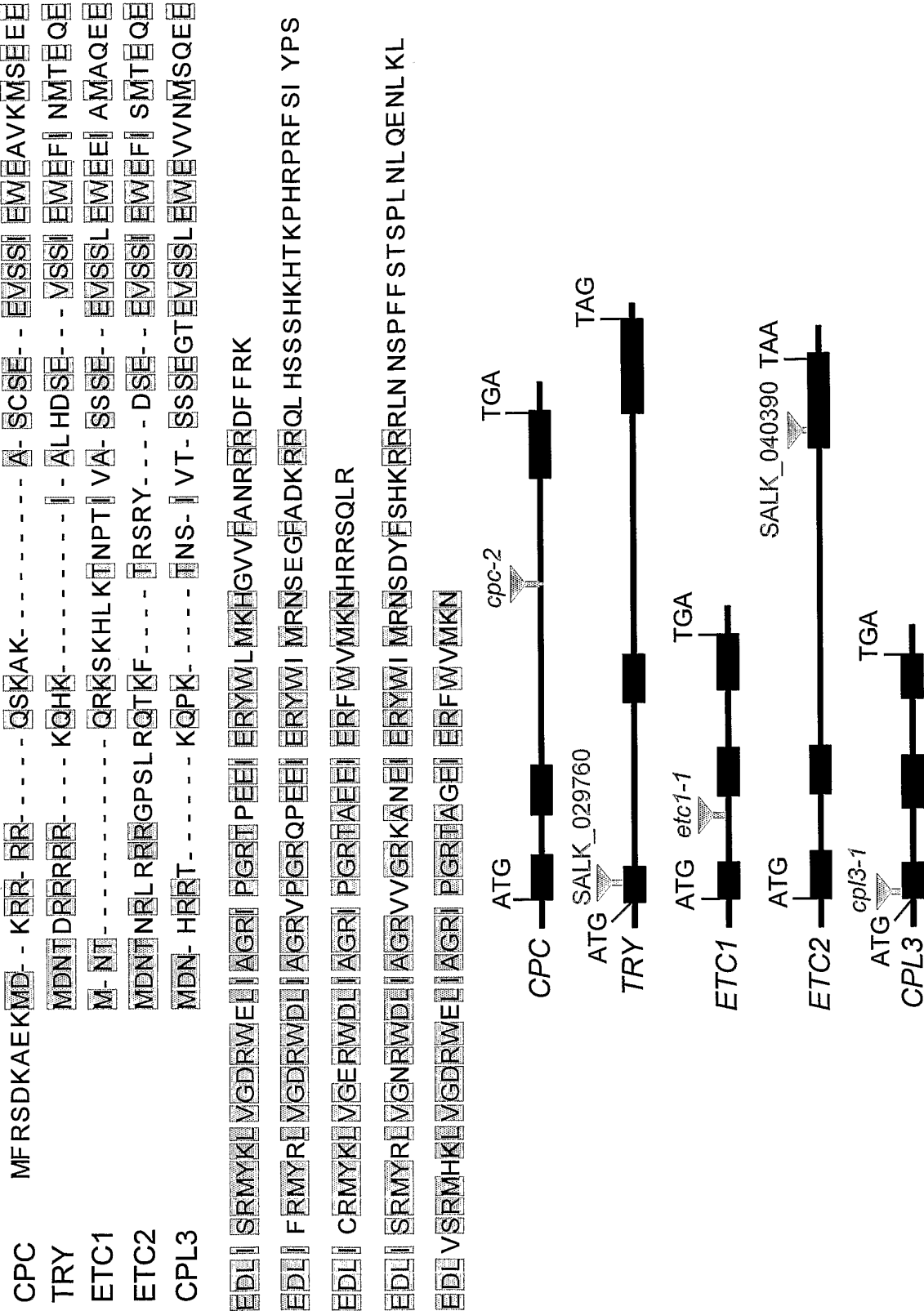


Fig. 1-2

[GENETYX-MAC: Multiple alignment]
Date : 2006.09.27

CPC	1	-MFRSDKAEKMDK	-RRRR-	QS	K-A	KA-S	-CS-	25
CPL3	1	MDNH-BRT	-K-	QP	KTNS			20
ETC1	1	MDNT		QR	C	SKHLKTNPTIV	SSS	21
ETC2	1	MDNTNRLRLRRG	PSLRT	CF	TR			22
TRY	1	MDNTDRRRRK		QH	KIALH			18
Os01g43180	1	MESGGG		QL	GN	NSK	-TSDGRETKATSSLLLLITAIHFVAL	39
Os01g43230	1	MDSSGG		D	GN	NSK	-TSDGCETK	21
CPC	26	EEVSSIEWEAMK	MS	EEEE	DL	SR	MYKLVGD	75
CPL3	21	EEVSSIEWEAMK	MS	EEEE	DL	SR	MYKLVGD	72
ETC1	22	EEVSSIEWEAMK	MS	EEEE	DL	SR	MYKLVGD	71
ETC2	23	EEVSSIEWEAMK	MS	EEEE	DL	SR	MYKLVGD	77
TRY	19	EEVSSIEWEAMK	MS	EEEE	DL	SR	MYKLVGD	70
Os01g43180	40	CPKGENEDARKVNTAQN	FVHF	EEEE	DL	SR	MYKLVGD	99
Os01g43230	22	EVNNTAQN	EVHF	EEEE	DL	SR	MYKLVGD	68
CPC	76	MLMK-HGVVFANRRD	FFRK					94
CPL3	73	MMKN						77
ETC1	72	MMKNH						83
ETC2	78	MMKNSDYF	SHKRRRLMNSPFFST	SP	LN	QEN	LKL	112
TRY	71	MMKNSEG	FADKRRQLHSSSHKHTK	PHR	P	RS	IYPS	106
Os01g43180	100	MAIX-HQAT						107
Os01g43230	69	TEGEIWCLET	FRM					83

Fig. 2



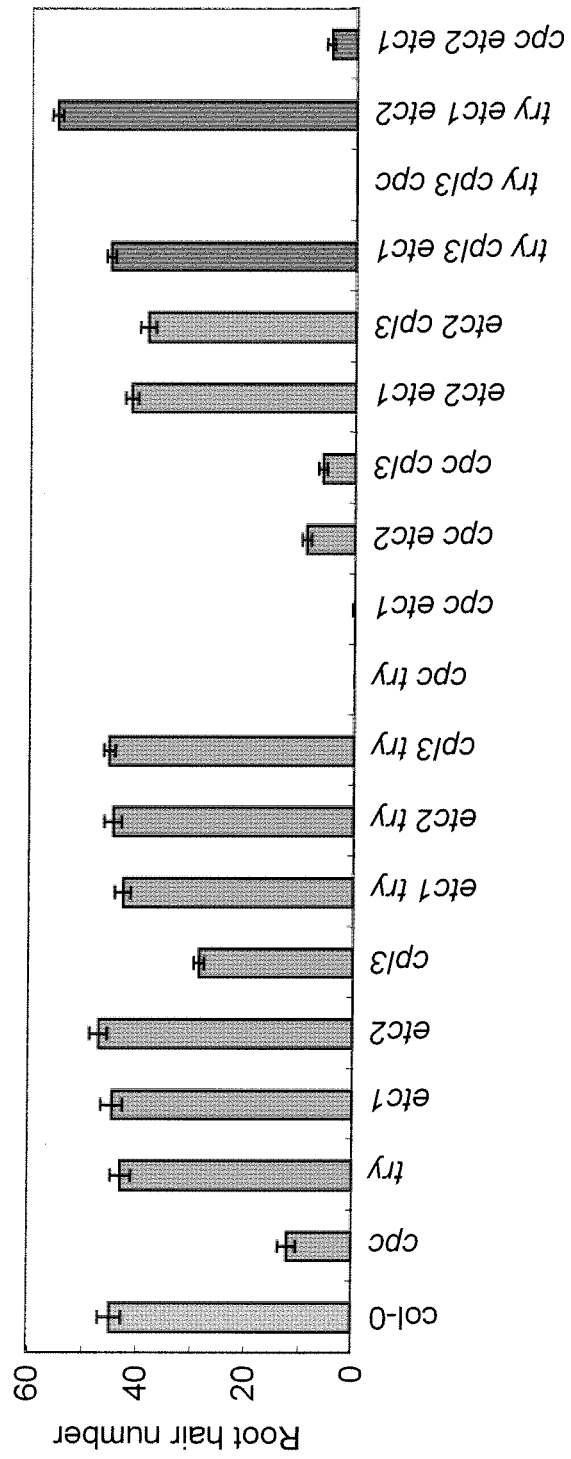


Fig. 3

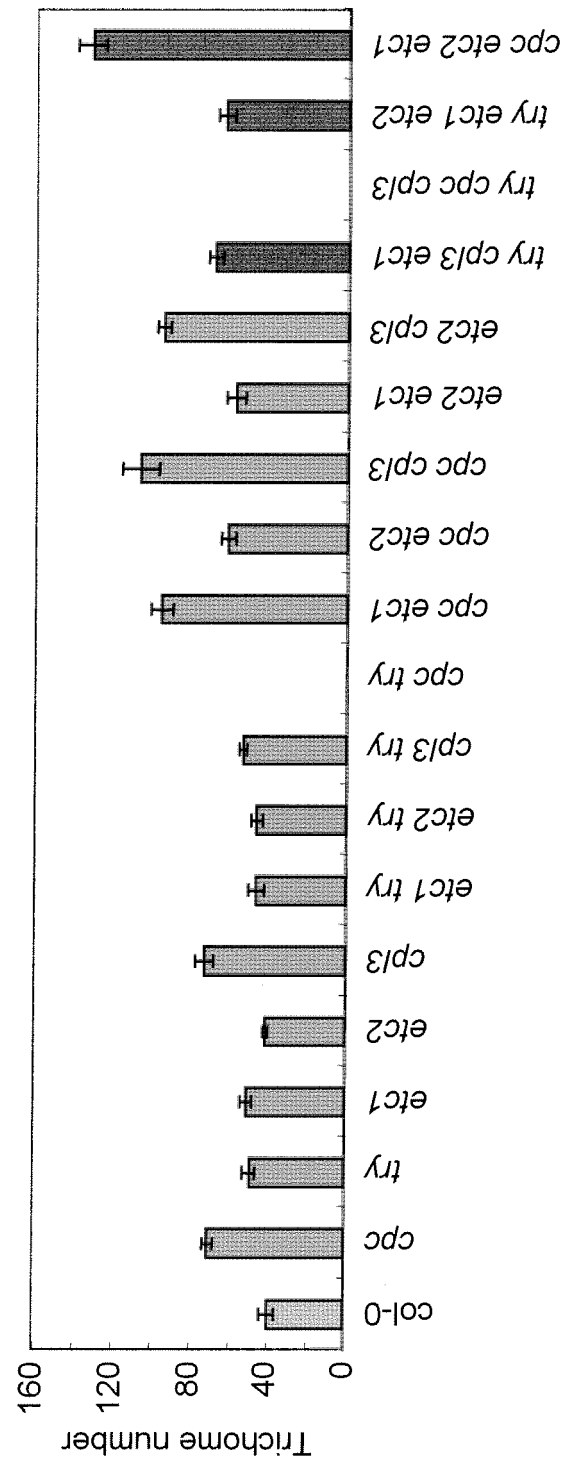


Fig. 4

Fig. 5

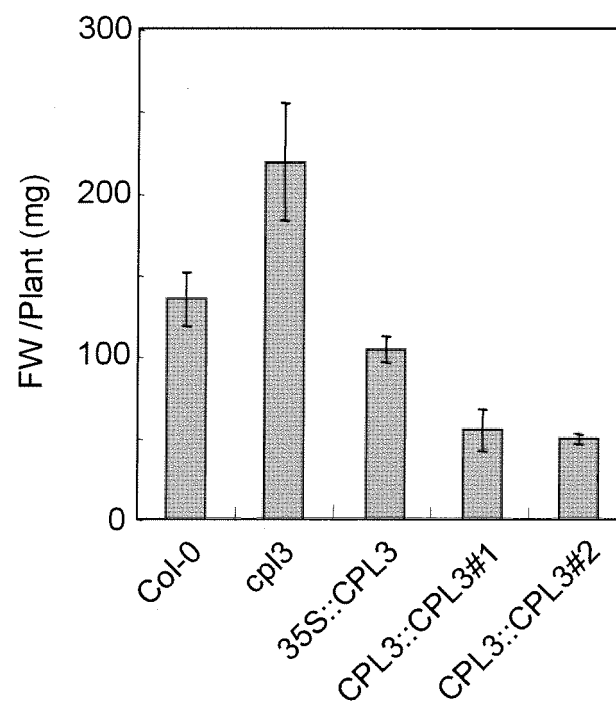


Fig. 6

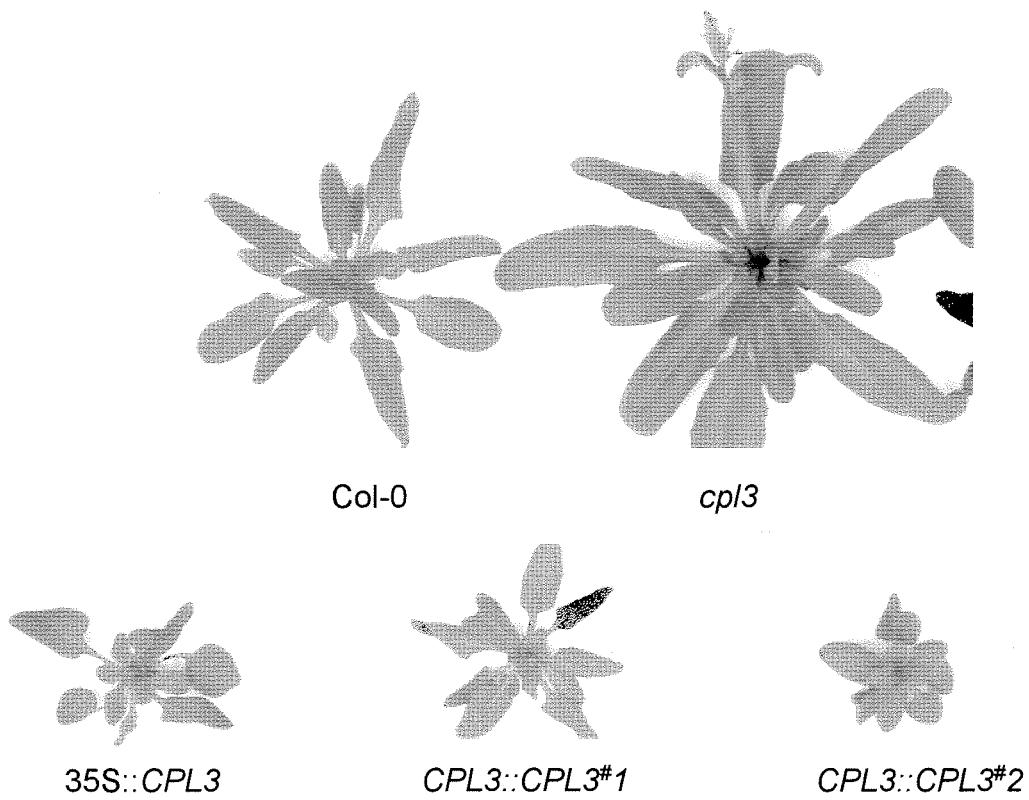


Fig. 7

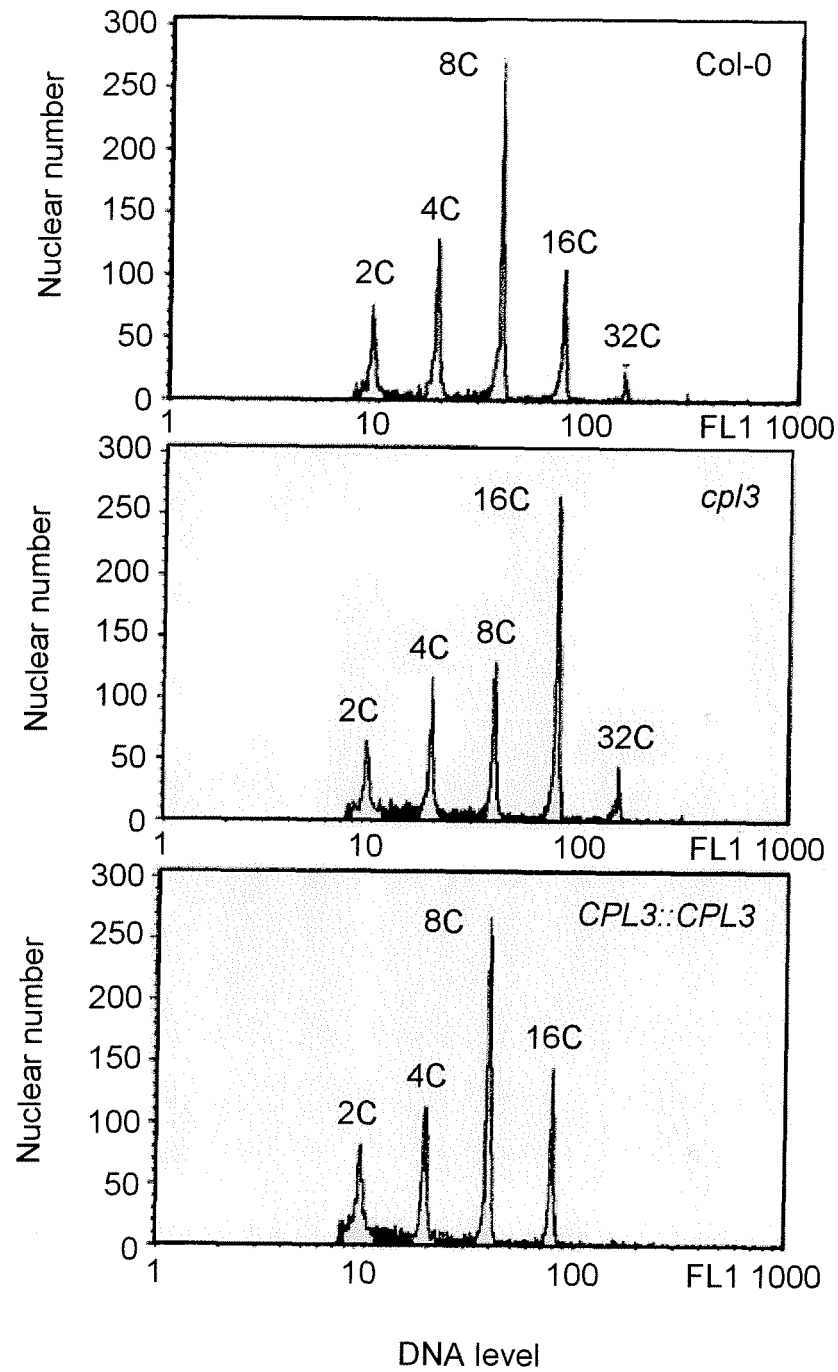


Fig. 8

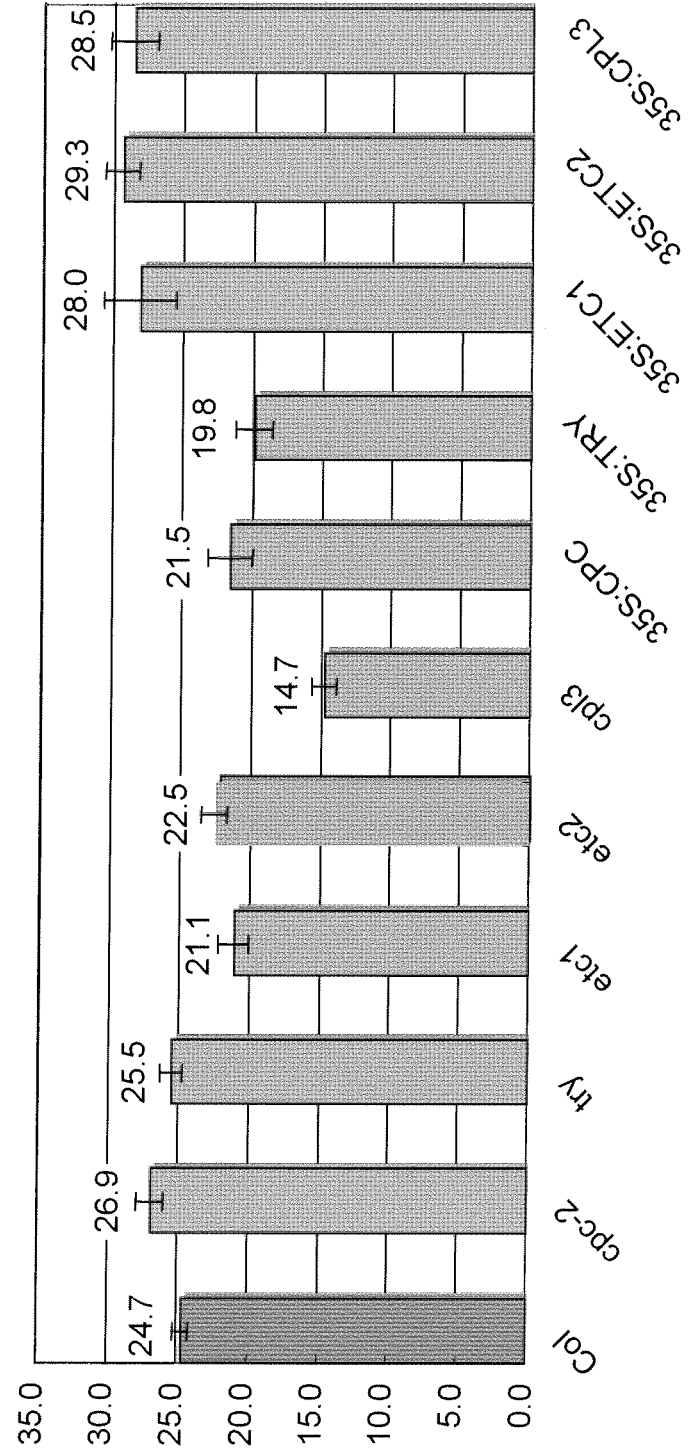


Fig. 9

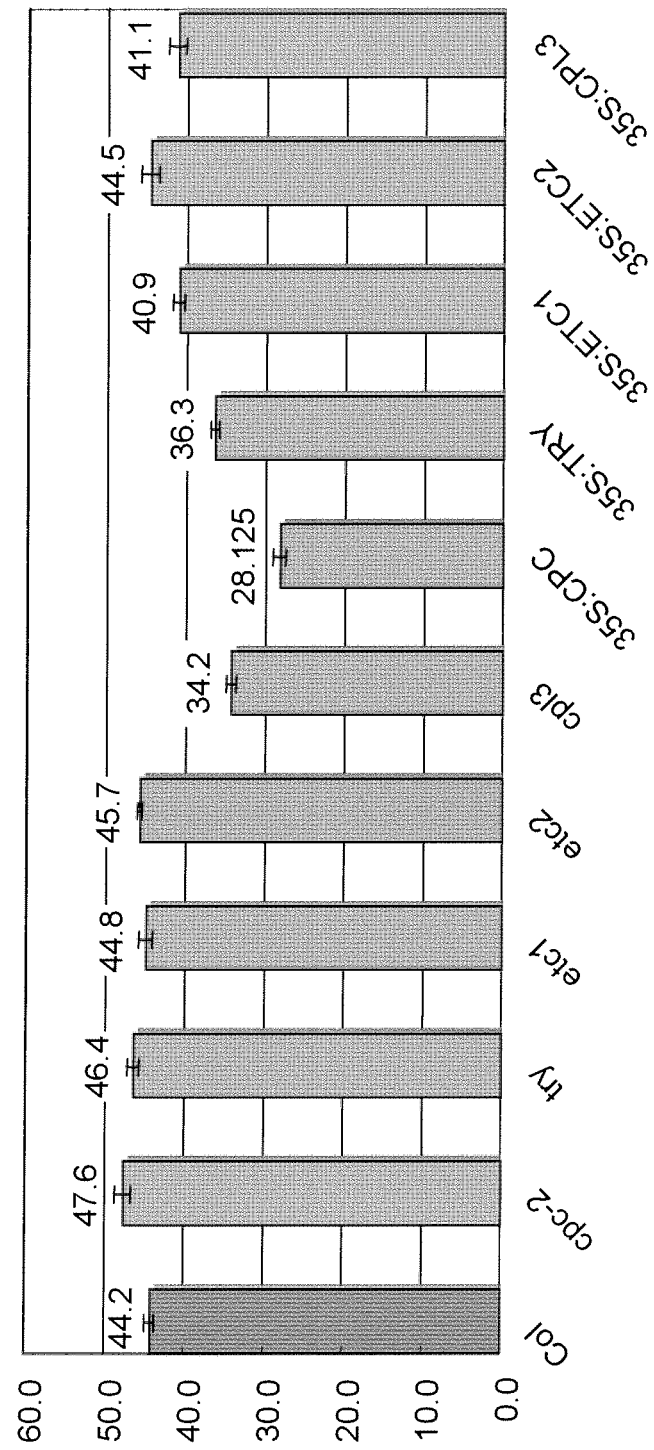


Fig. 10

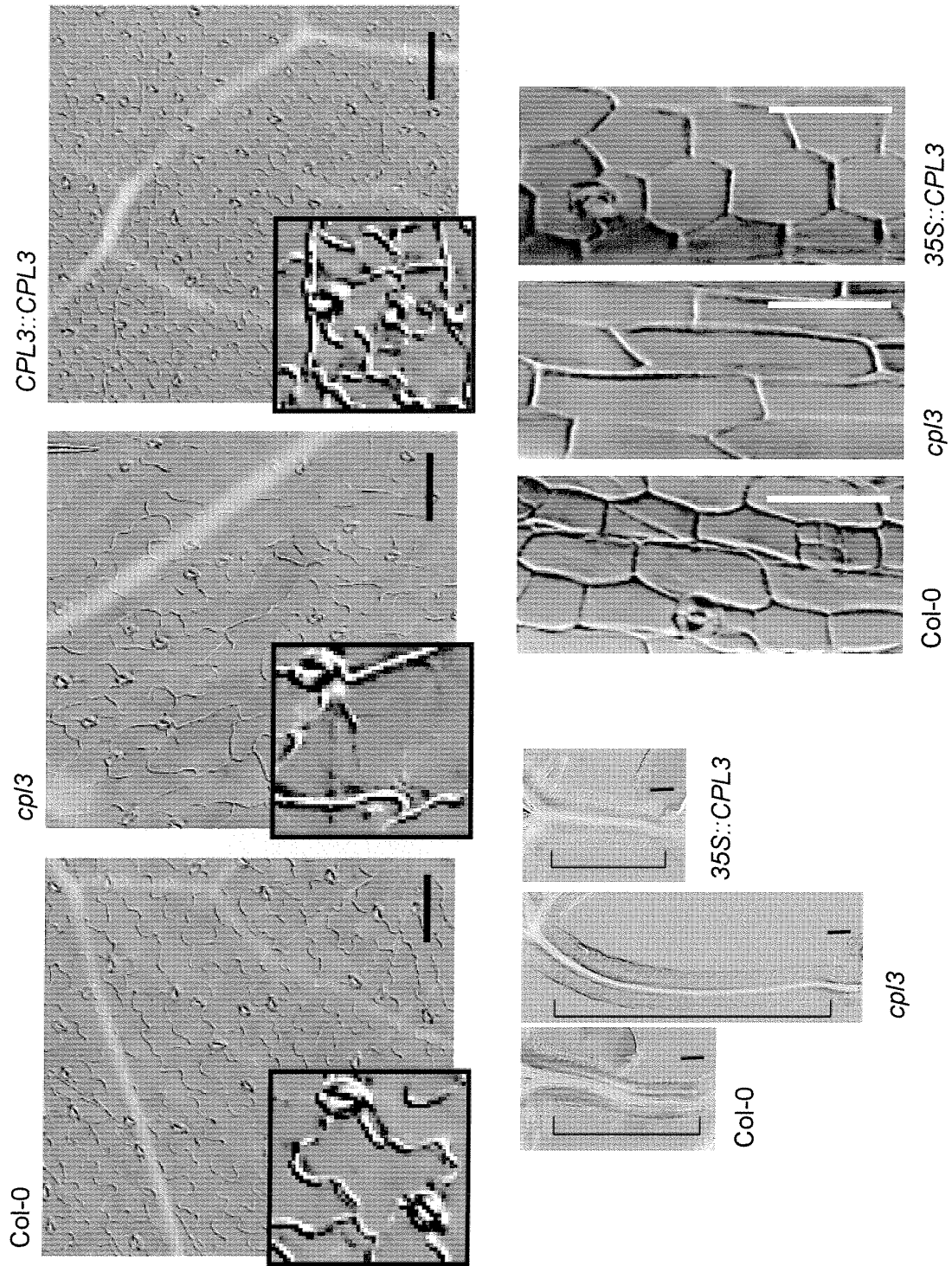


Fig. 11

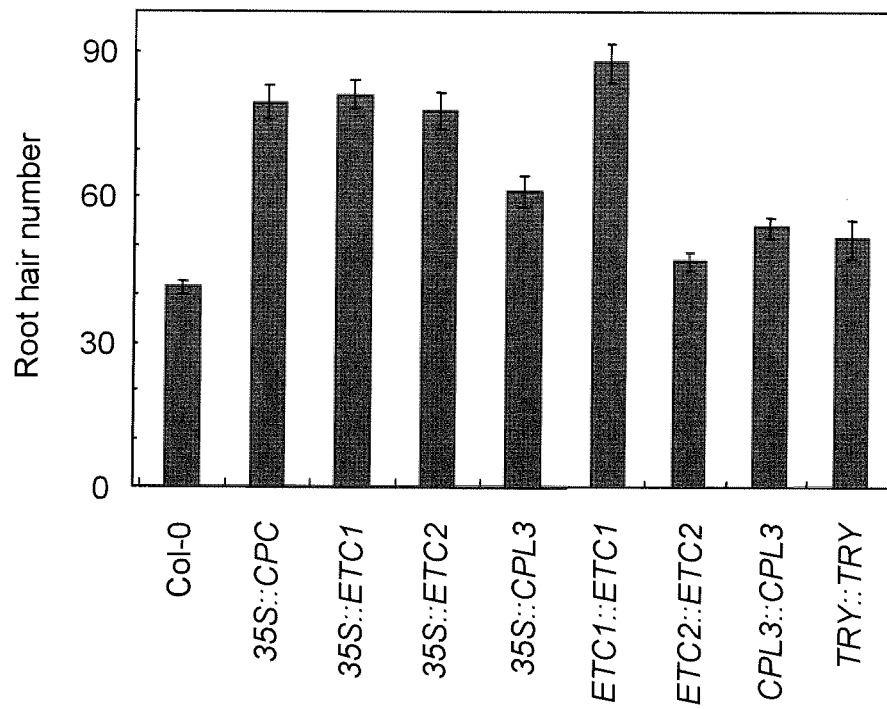


Fig. 12

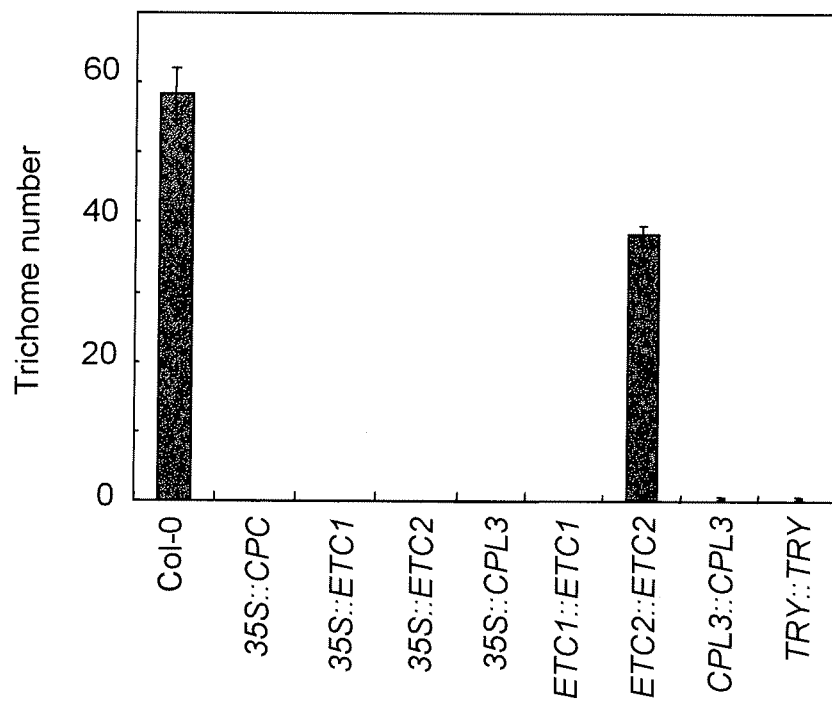


Fig. 13

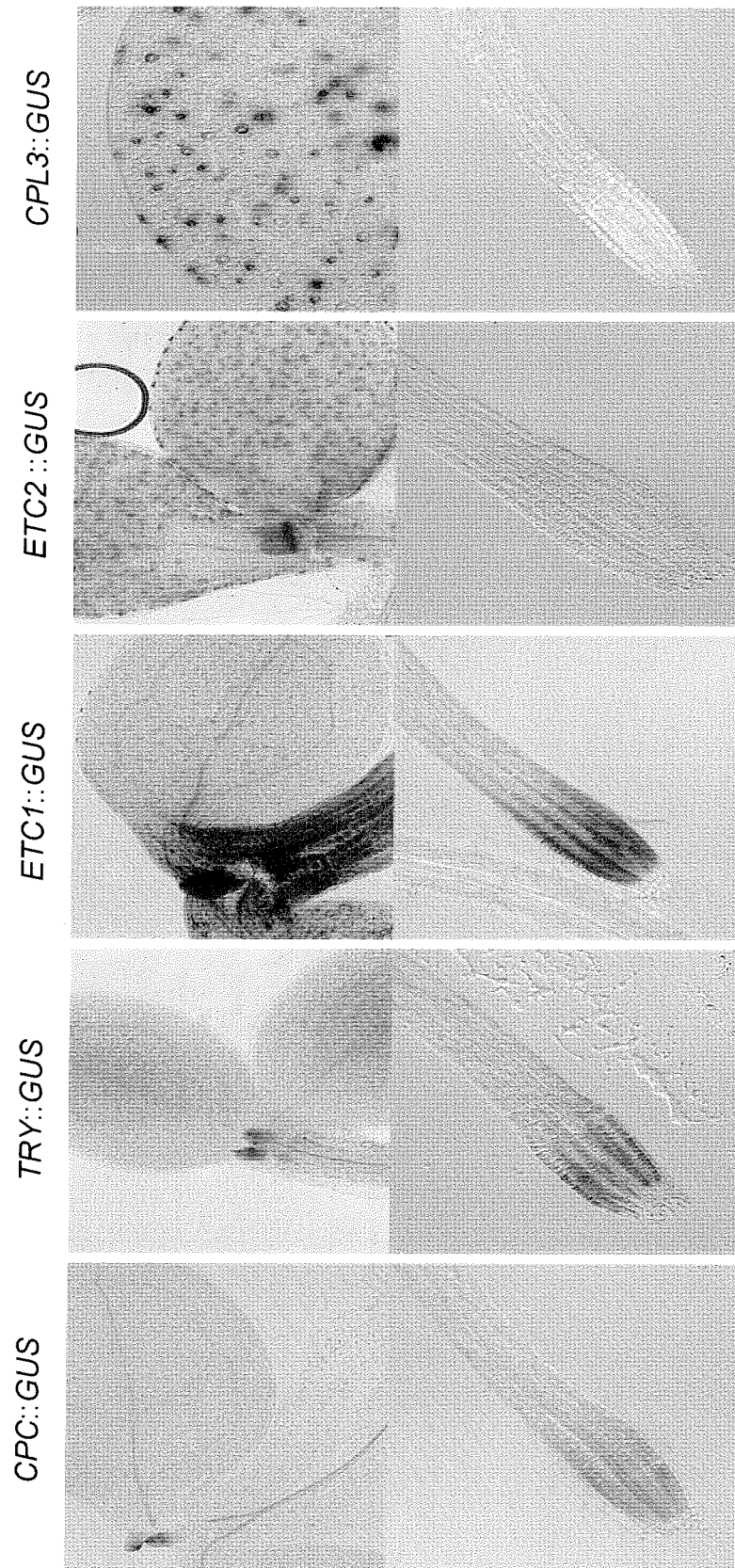


Fig. 14

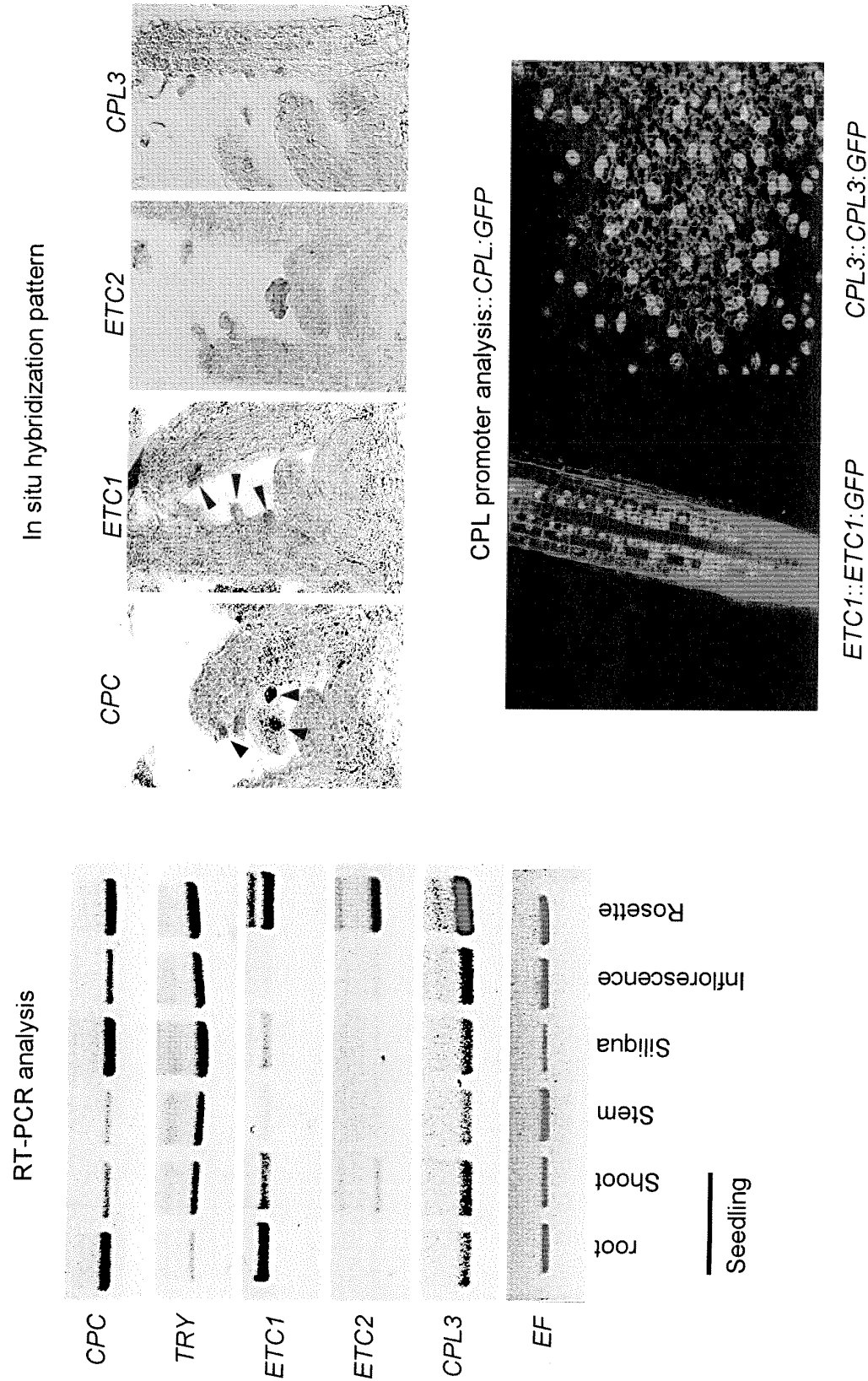
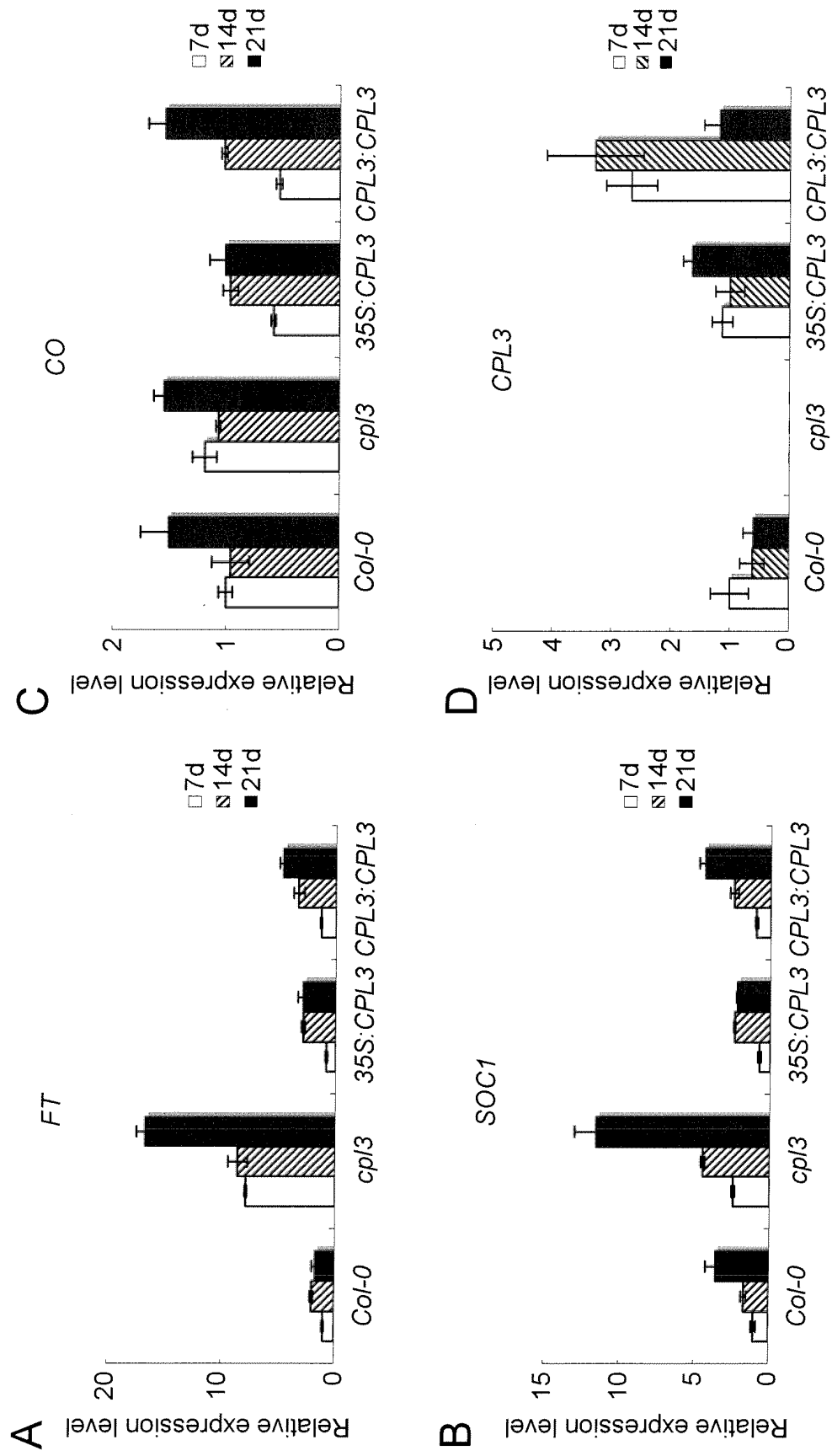


Fig. 15



INTERNATIONAL SEARCH REPORT

International application No.

PCT/JP2007/070396

A. CLASSIFICATION OF SUBJECT MATTER

C12N15/29(2006.01)i, A01H1/00(2006.01)i, A01H5/00(2006.01)i, C07K14/415(2006.01)n

According to International Patent Classification (IPC) or to both national classification and IPC

B. FIELDS SEARCHED

Minimum documentation searched (classification system followed by classification symbols)

C12N15/00-15/90, A01H1/00, A01H5/00, C07K14/415, C12N5/00-5/10

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched

Electronic data base consulted during the international search (name of data base and, where practicable, search terms used)

WPIDS (STN), BIOSIS (STN), MEDLINE (STN), AGRICOLA (STN), BIOENG (STN), CA (STN), CABA (STN), CONFSCI (STN), CROPU (STN), SCISEARCH (STN), JSTPlus (JDreamII),

C. DOCUMENTS CONSIDERED TO BE RELEVANT

Category*	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
X/A	Mineko IWATA, et al., "Shiroinunazuna no Hyohi Saibo Bunka ni Kan'yo suru MYB Idenshi no Kaiseki", Dai 45 Kai The Japanese Society of Plant Physiologists Nenkai Yoshishu, 20 March, 2004 (20.03.04), page 211. (Abstract 3aE06(437))	1-8/9-12
X/A	US 2004/0006797 A1 (SHI L et al.), 08 January, 2004 (08.01.04), Seq ID: 3, 15; example 1; table 1; Claims 11, 13 (Family: none)	1-8/9-12
X/A	KIRIK V et al., The ENHANCER OF TRY AND CPC1 gene acts redundantly with TRIPTYCHON and CAPRICE in trichome and root hair cell patterning in Arabidopsis., Dev. Biol., 2004, vol. 268, 506-513.	1-8/9-12



Further documents are listed in the continuation of Box C.



See patent family annex.

* Special categories of cited documents:

"A" document defining the general state of the art which is not considered to be of particular relevance

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"O" document referring to an oral disclosure, use, exhibition or other means

"P" document published prior to the international filing date but later than the priority date claimed

"T"

later document published after the international filing date or priority date and not in conflict with the application but cited to understand the principle or theory underlying the invention

"X"

document of particular relevance; the claimed invention cannot be considered novel or cannot be considered to involve an inventive step when the document is taken alone

"Y"

document of particular relevance; the claimed invention cannot be considered to involve an inventive step when the document is combined with one or more other such documents, such combination being obvious to a person skilled in the art

"&"

document member of the same patent family

Date of the actual completion of the international search
05 November, 2007 (05.11.07)

Date of mailing of the international search report
13 November, 2007 (13.11.07)

Name and mailing address of the ISA/
Japanese Patent Office

Authorized officer

Facsimile No.

Telephone No.

INTERNATIONAL SEARCH REPORT

International application No.

PCT/JP2007/070396

C (Continuation). DOCUMENTS CONSIDERED TO BE RELEVANT		
Category*	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
X/A	SIMON M and SCHIEFELBEIN J, Role of the CAPRICE family of myb genes in root epidermal development., 16th International conference on Arabidopsis research, 2005, TAIR accession: 501716944, http://www.arabidopsis.org/servlets/TairObject?type=publication&id=501716944	1-8/9-12
X/A	WO 2005/047516 A2 (MENDEL BIOTECHNOLOGY, INC.), 26 May, 2005 (26.05.05), Pages 49 to 54, 261 to 262, 265 to 268; Claims 16 to 22 & EP 1682668 A2	1-8/9-12
X/A	US 2004/0045049 A1 (ZHANG J et al.), 04 March, 2004 (04.03.04), Seq ID: 467, 468, 1808, 1809; Pages 40, 51, 144; Claims 20, 47, 52 & US 2007/0061911 A1	1-8/9-12
X/A	WO 2004/076638 A2 (MENDEL BIOTECHNOLOGY, INC.), 10 September, 2004 (10.09.04), Seq ID: 147, 148; Pages 40 to 43, 80; Claims 20 to 22 & US 2005/0086718 A1 & US 2005/0155117 A1 & US 2006/0242738 A1 & EP 1601758 A2	1-8/9-12
X/A	US 2004/0019927 A1 (SHERMAN B K et al.), 29 January, 2004 (29.01.04), Seq ID: 147, 148; Fig. 3; tables 4, 6; Pages 48, 356 to 357; Claims 1, 15 & US 2006/0162006 A1	1-8/9-12
X/A	US 2003/0229915 A1 (KEDDIE J et al.), 11 December, 2003 (11.12.03), Seq ID 161, 162, Ex. VI (page 14, left column); Claims 7, 8, 10 & US 7223904 B2	1-8/9-12
A	KIRIK V et al., ENHANCER of TRY and CPC2 (ETC2) reveals redundancy in the region-specific control of trichome development in Arabidopsis., Plant Mol. Biol., 2004, vol. 55, p. 389-398.	1-12
P,X	Rumi TOMINAGA, et al., "Shiroinunazuna no Hyohi Saibo Bunka ni Kan'yo suru CPC LIKE MYB Idenshi no Kaiseki", Dai 48 Kai The Japanese Society of Plant Physiologists Nenkai Yoshishu, 15 March, 2007 (15.03.07), page 157. (Abstract 2aB08(263))	1-12

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INTERNATIONAL SEARCH REPORT

International application No.

PCT/JP2007/070396

C (Continuation). DOCUMENTS CONSIDERED TO BE RELEVANT

Category*	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
P, X	TOMINAGA R et al., The CAPRICE-like Myb3 (CPL3) is involved in the determination of epidermal cell fate in Arabidopsis., 18th International conference on Arabidopsis research, 2007.06, TAIR accession: 501722033, http://www.arabidopsis.org/servlets/TairObject?type=publication&id=501722033	1-12

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INTERNATIONAL SEARCH REPORT

International application No.

PCT/JP2007/070396

Continuation of B. FIELDS SEARCHED

Electronic data base consulted during the international search
(name of data base and, where practicable, search terms used)

GenBank/EMBL/DDBJ/Geneseq, UniProt/PDB/Geneseq

REFERENCES CITED IN THE DESCRIPTION

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Patent documents cited in the description

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