



(12) **CORRECTED EUROPEAN PATENT SPECIFICATION**

- (15) Correction information:
Corrected version no 1 (W1 B1)
Corrections, see
Description Paragraph(s) 10, 15, 23, 25, 35,
39, 40, 112, 114, 117, 126, 133,
134, 136, 138, 139, 140, 156
Claims DE 1, 3, 15
Claims EN 15
Claims FR 1, 3, 9, 15
- (51) Int Cl.:
C12N 15/00 ^(2006.01) **C12N 5/10** ^(2006.01)
C12N 15/09 ^(2006.01) **C12P 21/02** ^(2006.01)
C12P 21/08 ^(2006.01) **C12N 15/85** ^(2006.01)
- (86) International application number:
PCT/JP2010/059881
- (87) International publication number:
WO 2010/143698 (16.12.2010 Gazette 2010/50)
- (48) Corrigendum issued on:
06.03.2019 Bulletin 2019/10
- (45) Date of publication and mention
of the grant of the patent:
22.08.2018 Bulletin 2018/34
- (21) Application number: **10786229.4**
- (22) Date of filing: **10.06.2010**

(54) **PROCESS FOR PRODUCTION OF PROTEIN**
VERFAHREN ZUR HERSTELLUNG EINES PROTEINS
PROCÉDÉ DE PRODUCTION D'UNE PROTÉINE

- | | |
|--|--|
| <p>(84) Designated Contracting States:
AL AT BE BG CH CY CZ DE DK EE ES FI FR GB
GR HR HU IE IS IT LI LT LU LV MC MK MT NL NO
PL PT RO SE SI SK SM TR</p> <p>(30) Priority: 11.06.2009 JP 2009140626
11.06.2009 US 186138 P</p> <p>(43) Date of publication of application:
18.04.2012 Bulletin 2012/16</p> <p>(73) Proprietors:
• Inter-University Research Institute Corporation
Research Organization of Information and
Systems
Tachikawa-shi
Tokyo 190-0014 (JP)
• Kyowa Hakko Kirin Co., Ltd.
Tokyo 100-8185 (JP)</p> | <p>(72) Inventors:
• KAWAKAMI, Koichi
Shizuoka 411-8540 (JP)
• YAMAGUCHI, Keina
Gunma 370-0013 (JP)
• OGAWA, Risa
Gunma 370-0013 (JP)
• TSUKAHARA, Masayoshi
Gunma 370-0013 (JP)</p> <p>(74) Representative: J A Kemp
14 South Square
Gray's Inn
London WC1R 5JJ (GB)</p> <p>(56) References cited:
WO-A1-00/65042 JP-A- 2002 262 879

• KAWAKAMI, K.: 'Tol2: a versatile gene transfer
vector in vertebrates' GENOME BIOLOGY vol. 8,
no. 1, 2007, pages S7.1 - S7.10, XP008156743</p> |
|--|--|

Note: Within nine months of the publication of the mention of the grant of the European patent in the European Patent Bulletin, any person may give notice to the European Patent Office of opposition to that patent, in accordance with the Implementing Regulations. Notice of opposition shall not be deemed to have been filed until the opposition fee has been paid. (Art. 99(1) European Patent Convention).

- URASAKI, A. ET AL.: 'Functional dissection of the Tol2 transposable element identified the minimal cis-sequence and a highly repetitive sequence in the subterminal region essential for transposition' GENETICS vol. 174, October 2006, pages 639 - 649, XP008156737
- KOGA, A. ET AL.: 'Germline transgenesis of Zebrafish using the medaka Toll transposon system' DEVELOPMENTAL DYNAMICS vol. 237, 2008, pages 2466 - 2474, XP008156738
- KOGA, A. ET AL.: 'The Toll element of medaka fish is transposed with only terminal regions and can deliver large DNA fragments into chromosomes' JOURNAL OF HUMAN GENETICS vol. 52, 2007, pages 1026 - 1030, XP019545326
- KOGA, A. ET AL.: 'The Toll transposable element of the medaka fish moves in human and mouse cells' JOURNAL OF HUMAN GENETICS vol. 52, 2007, pages 628 - 635, XP019493753
- KODAMA, K. ET AL.: 'The Toll element of the medaka fish, a member of the hAT transposable element family, jumps in Caenorhabditis elegans' HEREDITY vol. 101, 2008, pages 222 - 227, XP008156740
- SCHIFFERLI, K. P. ET AL.: 'Transfection of suspension cultures of CHO cells' FOCUS vol. 21, no. 1, 1999, pages 16 - 17, XP008156741
- KOO, T. Y. ET AL.: 'Beneficial effect of 30Kc6 gene expression on production of recombinant interferon-P in serum-free suspension culture of CHO cells' PROCESS BIOCHEMISTRY vol. 44, February 2009, pages 146 - 153, XP025769083
- KAWAKAMI ET AL: "Tol2: a versatile gene transfer vector in vertebrates", GENOME BIOLOGY (ONL, BIOMED CENTRAL LTD, GB, vol. 8, no. Suppl. 1, 1 January 2007 (2007-01-01), pages S7.1-S7.10, XP008156743, ISSN: 1465-6914, DOI: 10.1186/GB-2007-8-S1-S7
- URASAKI ET AL: "Functional dissection of the Tol2 transposable element identified the minimal cis-sequence and a highly repetitive sequence in the subterminal region essential for transposition", GENE, GENETICS SOCIETY OF AMERICA, AUSTIN, TX, US, vol. 174, no. 2, 1 October 2006 (2006-10-01), pages 639-649, XP008156737, ISSN: 0016-6731, DOI: 10.1534/GENETICS.106.060244

Description

Technical Field

5 **[0001]** This invention relates to a method for producing a protein of interest, comprising introducing a protein expression vector which comprises a gene fragment comprising a DNA encoding a protein of interest and a selectable marker gene and transposon sequences at both terminals of the gene fragment, into a suspension mammalian cell, integrating the gene fragment inserted between a pair of the transposon sequences into a chromosome of the mammalian cell to obtain a mammalian cell capable of expressing the protein of interest; and suspension-culturing the mammalian cell; and a
10 suspension mammalian cell capable of expressing the protein of interest.

Background Art

15 **[0002]** Production of exogeneous proteins by recombinant DNA techniques is used in various industries such as pharmaceutical industry and food industry. In most cases, production of recombinant proteins is carried out by introducing an expression vector comprising a nucleotide sequence encoding a protein of interest into a host, such as *Escherichia coli*, yeast, insect cell, plant cell, and animal cell, selecting a transformant in which the expression vector is integrated into the chromosome, and further culturing the cell line under appropriate culture conditions.

20 **[0003]** However, in order to develop a host which can produce an exogeneous protein efficiently, it is necessary to select a host cell having good productivity for each protein of interest, so that a further technical innovation is desired on the exogeneous protein production techniques for individual host.

25 **[0004]** In the bacteria systems, such as *Escherichia coli*, and yeast systems, different from animal cells, post-translational modifications, such as sugar chain modification, are difficult to attain in many cases and thus cause a problem in producing a protein having its activity.

30 **[0005]** Since the produced protein is subject to a post-translational modification such as phosphorylation and addition of sugar chains in the insect system, this system has a merit that the protein having its original physiological activity can be expressed. However, since the sugar chain structure of the secreted protein is different from that of mammals-derived cells, antigenicity and the like become a problem when the protein is applied to pharmaceutical use.

35 **[0006]** In addition, since a recombinant virus is used in the insect cell system when an exogeneous gene is introduced, there is a problem that its inactivation and containment of the virus are required from the viewpoint of safety.

40 **[0007]** In the animal cell system, post-translational modifications, such as phosphorylation, sugar chain addition, and folding, can be conducted to proteins derived from higher animals including human, in more similar manner to those produced in the living body. Such accurate post-translational modifications are necessary for recreating the physiological activity originally possessed by a protein in its recombinant protein, and a protein production system in which a mammalian cell is used as a host is usually applied to pharmaceutical products and the like that requires such physiological activity.

45 **[0008]** However, a protein expression system in which a mammalian cell is used as the host is generally low in productivity, and also causes a problem of the stability of introduced genes in many cases. Improvement of productivity of a protein using a mammalian culture cell as a host is not only very important in producing medicaments for treatment, diagnostic agents and the like, but also greatly contributes to research and development of them. Thus, it is urgent to develop a gene expression system which easily makes it possible to obtain a cell line of a high productivity using a mammalian culture cell, particularly Chinese hamster ovary cell (CHO cell), as the host.

50 **[0009]** A transposon is a transposable genetic element which can transfer from one locus to other locus on the chromosome. A transposon is a strong tool for the study on molecular biology and genetics and used for a purpose, such as mutagenesis, gene trapping, and preparation of transgenic individuals, in insects or nematode (e.g., *Drosophila melanogaster* or *Caenorhabditis elegans*) and plants. However, development of such a technique has been delayed for vertebral animals including mammalian cells.

55 **[0010]** In recent years, however, transposons which have activities also in vertebral animals have been reported, and some of them were shown to have an activity in mammalian cells, such as cell derived from mouse and human. Typical examples include transposons Tol1 (Patent Reference 1) and Tol2 (Non-patent Reference 1 and Non-patent Reference 13) cloned from a medaka (killifish), Sleeping Beauty reconstructed from a non-autonomous transposon existed in *Onchorhynchus* fish genome (Non-patent Reference 2), an artificial transposon Frog prince (Non-patent Reference 3) which is derived from frog and a transposon piggyBac (Non-patent Reference 4) which is derived from insect.

60 **[0011]** These DNA transposons have been used for mutagenesis, gene trapping, preparation of transgenic individuals, expression of drug-resistant proteins, and the like, as a gene transfer tool for bringing a new phenotype in a genome of a mammalian cell (Non-patent References 5 to 12).

65 **[0012]** In the case of insects, a method in which an exogeneous gene is introduced into silkworm chromosome using the transposon piggyBac derived from a Lepidoptera insect to express the protein encoded by said exogeneous gene was studied, and a protein production method using the above techniques has been disclosed (Patent Reference 2).

[0013] However, since the expressed protein of interest is not sufficient in expression level and is produced in the whole body of silkworm, it causes an economical problem due to the necessity of an advanced purification technique for recovering the expressed exogeneous protein in a highly purified form from the body fluid including a large amount of contaminated proteins.

[0014] In addition, an example in which a protein relating to G418 resistance is expressed in a mammalian cell using the medaka-derived transposon Tol2 (Non-patent Reference 12) is known.

[0015] The minimal cis-sequence and a highly repetitive sequence in the sub-terminal region of the Tol2 transposon that is essential for transposition have been identified (Non-patent Reference 14).

[0016] A technique for selecting cells, into which a gene has been transferred in a stable state, by using a novel drug resistance gene as a stable marker, and a technique for obtaining cells in which a gene is highly expressed are disclosed in Patent Reference 3.

Citation List

Patent Literature

[0017]

Patent Literature 1 WO2008/072540

Patent Literature 2 Japanese Published Unexamined Patent Application No. 2001-532188

Patent Literature 3 Japanese Published Patent Application No. 2002-262879 Non Patent Literature

[0018]

Non Patent Literature 1 Nature 383, 30 (1996)

Non Patent Literature 2 Cell 91, 501-510 (1997)

Non Patent Literature 3 Nucleic Acids Res, 31, 6873-6881 (2003)

Non Patent Literature 4 Insect Mol.Biol.5, 141-151 (1996)

Non Patent Literature 5 Genetics.166, 895-899 (2004)

Non Patent Literature 6 PLoS Genet, 2, e169 (2006)

Non Patent Literature 7 Proc. Natl. Acad. Sci. USA 95, 10769-10773 (1998)

Non Patent Literature 8 Proc. Natl. Acad. Sci. USA 98:6759-6764 (2001)

Non Patent Literature 9 Nature 436,221-22 6 (2005)

Non Patent Literature 10 Nucleic Acids Res., 31, 6873-6881 (2003)

Non Patent Literature 11 Nucleic Acids Res., 35, e87 (2007)

Non Patent Literature 12 Proc Natl. Acad. Sci. USA, 103, 15008-15013 (2006)

Non Patent Literature 13 Genome Biology, 8 suppl I, S7.1-S7.10 (2007)

Non Patent Literature 14 Genetics 174, 639-649 (2006)

Summary of Invention

Technical Problem

[0019] In order to produce and analyze a protein of interest, it is necessary to select a cell line which stably and highly expresses a protein of interest, using a mammalian-derived culture cell, but preparation and culture of the cell that produces the protein of interest require considerable labor and time.

[0020] In addition, though it is known that a protein of interest is expressed in a mammalian cell using a transposon sequence, preparation of a cell which can highly express a protein of interest and thus can be used as a protein production system by using a transposon sequence; preparation method of a mammalian cell which can highly produce a protein of interest by using a transposon sequence; and a production method of a protein using the cell are not known.

[0021] As described in the above, the expression of a protein of interest in a large amount by establishing a protein production system which can highly produce a protein of interest using a mammalian culture cell efficiently and within a short period has been required. Thus, the objects of the invention are to provide a cell capable of highly expressing a protein of interest which can be efficiently established, and a method for producing the protein of interest using the cell.

Solution to Problems

[0022] To solve the above-mentioned problems, the present inventors have conducted intensive studies and found

as a result that a mammalian cell capable of highly expressing a protein of interest can be efficiently prepared by introducing a protein expression vector which comprises a gene fragment comprising a DNA encoding the protein of interest and a selectable marker gene and transposon sequences at both terminals of the gene fragment, into a suspension mammalian cell; and integrating the gene fragment inserted between a pair (two) of the transposon sequences into a chromosome of the mammalian cell. In addition, it was found that the protein of interest can be produced efficiently by using the cell, and thereby the invention was accomplished.

Detailed Description of the Invention

[0023] Specifically, the invention is as follows:

1. A method for producing a protein of interest, comprising introducing a protein expression vector which comprises a gene fragment comprising a DNA encoding a protein of interest and a selectable marker gene and, both terminals of the gene fragment, a pair of transposon sequences which are the Tol1 nucleotide sequences shown in SEQ ID NO:14 and SEQ ID NO:15 or the Tol2 nucleotide sequences shown in SEQ ID NO:2 and SEQ ID NO:3 into a suspension CHO cell capable of surviving and proliferating in a serum-free medium; introducing an expression vector (b) which comprises a DNA encoding a transposase which recognizes the transposon sequences and has activity of transferring a gene fragment inserted between the transposon sequences into a chromosome into the CHO cell; integrating the gene fragment inserted between the transposon sequences into a chromosome of the CHO cell to obtain a said CHO cell capable of expressing the protein of interest; and suspension-culturing the CHO cell;

2. A method described in the aforementioned item 1 for producing a protein of interest, comprising:

(A) simultaneously introducing the expression vectors (a) and (b) into the CHO cell,

(B) expressing transiently the transposase from the expression vector introduced in the step (A) to integrate the gene fragment inserted between the transposon sequences into a chromosome of the CHO cell to obtain a suspension CHO cell capable of expressing the protein of interest, and

(C) suspension-culturing the suspension CHO cell capable of expressing the protein of interest obtained in the step (B) to produce the protein of interest;

3. A method for obtaining a suspension CHO cell capable of expressing a protein of interest, comprising introducing a protein expression vector which comprises a gene fragment comprising a DNA encoding a protein of interest and a selectable marker gene and, at both terminals of the gene fragment, a pair of transposon sequences which are the Tol1 nucleotide sequences shown in SEQ ID NO:14 and SEQ ID NO:15 or the Tol2 nucleotide sequences shown in SEQ ID NO:2 and SEQ ID NO:3 into a suspension CHO cell capable of surviving and proliferating in a serum-free medium; introducing an expression vector (b) which comprises a DNA encoding a transposase which recognizes the transposon sequences and has activity of transferring a gene fragment inserted between the transposon sequences into a chromosome into the CHO cell; and integrating the gene fragment inserted between a pair of the transposon sequences, into a chromosome of the CHO cell;

4. The method described in any one of the aforementioned items 1 to 3, wherein the CHO cell is at least one selected from CHO-K1, CHO-K1SV, DUKXB11, CHO/DG44, Pro-3 and CHO-S;

5. The method described in any one of the aforementioned items 1 to 4, wherein the selectable marker gene is a cycloheximide resistance gene;

6. The method described in the aforementioned item 5, wherein the cycloheximide resistance gene is a gene encoding a mutant of human ribosomal protein L36a;

7. The method described in the aforementioned item 6, wherein the mutant is a mutant in which proline at position 54 of the human ribosomal protein L36a is substituted with other amino acid;

8. The method described in the aforementioned item 7, wherein the other amino acid is glutamine;

9. A suspension CHO cell capable of surviving and proliferating in a serum-free medium and of producing a protein of interest, which cell comprises an expression vector (a) comprising a gene fragment comprising a DNA encoding a protein of interest and a selectable marker gene and, at both terminals of the gene fragment, a pair of transposon sequences which are the Tol1 nucleotide sequences shown in SEQ ID NO:14 and SEQ ID NO:15 or the Tol2 nucleotide sequences shown in SEQ ID NO:2 and SEQ ID NO:3 and an expression vector (b) comprising a DNA encoding a transposase (a transferase) which recognizes the transposon sequences and has activity of transferring the gene fragment inserted between the transposon sequences into a chromosome to integrate the gene fragment inserted between the transposon sequences into the chromosome of the CHO cell;

10. The cell described in the aforementioned item 9, wherein the CHO cell is at least one selected from CHO-K1, CHO-K1SV, DUKXB11, CHO/DG44, Pro-3 and CHO-S;

11. The cell described in the aforementioned item 9 or 10, wherein the selectable marker gene is a cycloheximide resistance gene;

12. The cell described in the aforementioned item 11, wherein the cycloheximide resistance gene is a gene encoding a mutant of human ribosomal protein L36a;

13. The cell described in the aforementioned item 12, wherein the mutant is a mutant in which proline at position 54 of the human ribosomal protein L36a is substituted with other amino acid;

14. The cell described in the aforementioned item 13, wherein the other amino acid is glutamine; and

15. Use of a protein expression vector (a) comprising a gene fragment comprising a DNA encoding a protein of interest and a selectable marker gene and, at both terminals of the gene fragment, a pair of transposon sequences which are the Tol1 nucleotide sequences shown in SEQ ID NO:14 and SEQ ID NO:15 or the Tol2 nucleotide sequences shown in SEQ ID NO:2 and SEQ ID NO:3 and an expression vector (b) comprising a DNA encoding a transposase which recognizes the transposon sequences and has activity of transferring a gene fragment inserted between the transposon sequences into a chromosome, to integrate the gene fragment inserted between the transposon sequences into a chromosome of a suspension CHO cell capable of surviving and proliferating in a serum-free medium.

Advantageous Effects of Invention

[0024] According to the protein production method of the invention, a protein of interest can be efficiently produced by the use of a mammalian cell. In addition, the cell of the invention can be used as a protein production cell for producing a recombinant protein with a high efficiency.

Brief Description of the Drawings

[0025]

[Fig. 1] Fig. 1 shows a schematic illustration of a transposon vector for expressing an anti-human influenza M2 antibody. Tol2-L represents a left end Tol2 transposon (SEQ ID NO:2), Tol2-R represents a right end Tol2 transposon (SEQ ID NO:3), CMV represents a CMV promoter, poly A represents a polyadenylation site, Hc represents a human antibody H chain cDNA, Lc represents a human antibody L chain cDNA, and CHX-r represents a cycloheximide resistance gene.

[Fig. 2] Fig. 2 shows a schematic illustration of an anti-human influenza M2 antibody expression vector. CMV represents a CMV promoter, poly A represents a polyadenylation site, Hc represents a human antibody H chain cDNA, Lc represents a human antibody L chain cDNA and CHX-r represents a cycloheximide resistance gene.

[Fig. 3] Fig. 3 shows a schematic illustration of a Tol2 transposase expression vector. CAGGS represents a CAGGS promoter, poly A represents a polyadenylation site, and TPase cDNA represents a Tol2 transposase cDNA.

[Fig. 4A] Fig. 4A shows a result of examining expression level of an anti-human influenza M2 antibody in a suspension CHO-K1 cell when a Tol2 transposon vector for expressing an anti-human influenza M2 antibody was used. The ordinate shows the amount of antibody production ($\mu\text{g/ml}$), and the abscissa shows the number of transgenic clones of the suspension CHO-K1 cell.

[Fig. 4B] Fig. 4B shows a result of examining expression level of an anti-human influenza M2 antibody in an adhesive CHO-K1 cell when a Tol2 transposon vector for expressing an anti-human influenza M2 antibody was used. The ordinate shows the amount of antibody production ($\mu\text{g/ml}$), and the abscissa shows the number of transgenic clones of the adhesive CHO-K1 cell.

[Fig. 5] Fig. 5 shows a schematic illustration of a Tol1 transposon vector for expressing an anti-human influenza M2 antibody. Tol1-L represents a left end Tol1 transposon (SEQ ID NO:14), Tol1-R represents a right end Tol1 transposon (SEQ ID NO:15), CMV represents a CMV promoter, poly A represents a polyadenylation site, Hc represents a human antibody H chain cDNA, Lc represents a human antibody L chain cDNA, and CHX-r represents a cycloheximide resistance gene.

[Fig. 6] Fig. 6 shows a schematic illustration of a Tol1 transposase expression vector. CAGGS represents a CAGGS promoter, poly A represents a polyadenylation site, and TPase cDNA represents a Tol1 transposase cDNA.

[Fig. 7] Fig. 7 shows a result of examining expression level of an anti-human influenza M2 antibody in a suspension CHO-K1 cell when a Tol1 transposon vector for expressing an anti-human influenza M2 antibody was used. The ordinate shows the amount of antibody production ($\mu\text{g/ml}$), and the abscissa shows the number of transgenic clones of the suspension CHO-K1 cell.

[0026] This invention relates to a method for producing a protein of interest, comprising introducing a protein expression vector comprising a gene fragment comprising a DNA encoding a protein of interest and a selectable marker gene and

transposon sequences at both terminals of the gene fragment, into a suspension mammalian cell; integrating the gene fragment inserted between a pair (two) of the transposon sequences, into a chromosome of the mammalian cell to obtain a mammalian cell capable of expressing said protein of interest; and suspension-culturing the mammalian cell.

[0027] Examples of the method for producing a protein of interest of the present invention include a method, comprising the following steps (A) to (C):

(A) a step of simultaneously introducing the following expression vectors (a) and (b) into a suspension mammalian cell:

- (a) an expression vector which comprises a gene fragment comprising a DNA encoding a protein of interest and transposon sequences at both terminals of the gene fragment,
- (b) an expression vector which comprises a DNA encoding a transposase which recognizes the transposon sequences and has activity of transferring a gene fragment inserted between a pair of the transposon sequences into a chromosome,

(B) a step of expressing transiently the transposase transiently from the expression vector introduced in the step (A) to integrate the gene fragment inserted between a pair of the transposon sequences into a chromosome of the mammalian cell to obtain a suspension mammalian cell capable of expressing the protein of interest, and
(C) a step of suspension-culturing the suspension mammalian cell capable of expressing the protein of interest obtained in the step (B) to produce the protein of interest.

[0028] In addition, the present invention relates to a suspension mammalian cell capable of producing a protein of interest, into which a protein expression vector comprising a gene fragment comprising a DNA encoding a protein of interest and a selectable marker gene and transposon sequences at both terminals of the gene fragment is introduced, to integrate the gene fragment inserted between a pair of the transposon sequences into a chromosome.

[0029] Furthermore, the present invention relates to a suspension mammalian cell capable of producing a protein of interest, into which an expression vector (a) comprising a gene fragment comprising a DNA encoding a protein of interest and a selectable marker gene and transposon sequences at both terminals of the gene fragment, and an expression vector (b) comprising a DNA encoding a transposase (a transferase) which recognizes the transposon sequences and has activity of transferring the gene fragment inserted between a pair of the transposon sequences into a chromosome to integrate the gene fragment inserted between a pair of the transposon sequences into the chromosome.

[0030] The term "transposon" in the present specification is a transposable genetic element and means a gene unit which moves on a chromosome or from a chromosome to other chromosome (transposition) while keeping a certain structure.

[0031] The transposon comprises a gene unit of a repeating transposon sequences (also called inverted repeat sequence (IR sequence) or terminal inverted repeat sequence (TIR sequence)) which positions in the same direction or the reverse direction at both terminals of the gene unit and a nucleotide sequence encoding a transposase which recognizes the transposon sequence to transfer a gene existing between the transposon sequences.

[0032] The transposase translated from the transposon can transfer a DNA by recognizing transposon sequences of both terminals of the transposon, cutting out the DNA fragment inserted between a pair of the transposon sequences and inserting the fragment into the site to be transferred.

[0033] The term "transposon sequence" in the present specification means the nucleotide sequence of a transposon recognized by a transposase and has the same meaning as the IR sequence or TIR sequence. A DNA comprising the nucleotide sequence may comprise an imperfect repeating moiety as long as it can be transferred (inserted into other position in the genome) by the activity of a transposase, and comprise a transposon sequence specific to the transposase.

[0034] As the transposon sequence to be used in the invention, a nucleotide sequence derived from a pair of natural or artificial DNA-type transposons, which can be recognized by a transposase and be transposed in mammalian cells, is used.

[0035] The nucleotide sequence derived from a DNA-type transposon is a pair of nucleotide sequences derived from the medaka fish-derived Tol1 transposon or Tol2 transposon.

[0036] Medaka fish-derived Tol2 and Tol1 transposon nucleotide sequences are shown in SEQ ID NO:6 and SEQ ID NO:13, respectively.

[0037] Examples of a nucleotide sequence derived from a pair of Tol2 transposons include the nucleotide sequence at positions 1 to 2229 and the nucleotide sequence at positions 4148 to 4682 in the Tol2 transposon nucleotide sequence shown in SEQ ID NO:6 of Sequence Listing.

[0038] As the nucleotide sequence derived from a pair of Tol2 transposons, the nucleotide sequence at positions 1 to 200 (SEQ ID NO:2) (hereinafter referred to as "Tol2-L sequence") and the nucleotide sequence at positions 2285 to 2788 (SEQ ID NO:3) (hereinafter referred to as "Tol2-R sequence") in the Tol2 transposon nucleotide sequence shown in SEQ ID NO:1 of Sequence Listing are used.

[0039] Examples of a nucleotide sequence derived from a pair of Tol1 transposons include the nucleotide sequence comprising a nucleotide sequence at positions 1 to 157 and the nucleotide sequence at positions the 1748 to 1855 in the Tol1 transposon nucleotide sequence shown in SEQ ID NO:13 of Sequence Listing.

[0040] As the nucleotide sequence derived from a pair of Tol1 transposons, the nucleotide sequence at positions 1 to 200 (SEQ ID NO:14) (hereinafter referred to as "Tol1-L sequence") and the nucleotide sequence at positions 1351 to 1855 (SEQ ID NO:15) (hereinafter referred to as "Tol1-R sequence") in the Tol2 transposon nucleotide sequence shown in SEQ ID NO:1 of Sequence Listing are used.

[0041] Examples of the transposon sequence include transposon sequences of which transfer reactions are controlled by using a partial sequence of a transposon sequence derived from the above-mentioned transposon, by adjusting the length of the nucleotide sequence and by modifying the nucleotide sequence due to addition, deletion or substitution.

[0042] Regarding the control of the transfer reaction of a transposon, the transfer reaction can be accelerated or suppressed by accelerating or suppressing recognition of the transposon sequence by a transposase, respectively.

[0043] The term "transposase" in the present specification means an enzyme which recognizes nucleotide sequences having transposon sequences and transfers a DNA existing between the nucleotide sequences into a chromosome or from the chromosome to other chromosome.

[0044] Examples of the transposase include the Tol1 and Tol2 which are derived from medaka fish, the Sleeping Beauty reconstructed from a non-autonomous transposon existed in an *Onchorhynchus* fish genome, the artificial transposon Frog prince which is derived from frog and the transposon PiggyBac which is derived from insect.

[0045] As the transposase, a native enzyme may be used, and any transposase in which a part of its amino acids are substituted, deleted, inserted and/or added may be used as long as the same transfer activity as the transposase is maintained. By controlling the enzyme activity of the transposase, the transfer reaction of the DNA existing between the transposon sequences can be controlled.

[0046] In order to analyze whether or not it possesses a transfer activity similar to that of transposase, it can be measured by the 2-components analyzing system disclosed in Japanese Published Unexamined Patent Application No.235575/2003.

[0047] Illustratively, whether or not a non-automatic Tol2 element can be transferred and inserted into a mammalian cell chromosome by the activity of a transposase can be analyzed by separately using a plasmid comprising a Tol2 transposase-deleted Tol2 transposon (Tol2-derived non-autonomous transposon) and a plasmid comprising Tol2 transposase.

[0048] The term "non-autonomous transposon" in the present specification means a transposon which is lost a transposase existed inside the transposon and cannot therefore perform its autonomous transfer. The non-autonomous transposon can transfer the DNA inserted between transposon sequences of the non-autonomous transposon into the host cell chromosome, by allowing a transposase protein, an mRNA encoding the transposase protein or a DNA encoding the transposase protein to simultaneously present in the cell.

[0049] The transposase gene means a gene encoding a transposase. In order to improve its expression efficiency in a mammalian cell, a sequence which adjusts a space between the Kozak's consensus sequence (Kozak M., Nucleic Acids Res., 12, 857 - 872 (1984)) or a ribosome binding sequence, Shine-Dalgarno sequence and the initiation codon, to an appropriate distance (e.g., from 6 to 18 bases) may be connected to an upstream site of the translation initiation codon ATG of the gene.

[0050] According to the method of the invention, in order to integrate a gene fragment comprising a DNA encoding the protein of interest and a selectable marker gene in an expression vector into the chromosome of a host cell, an expression vector which comprises the gene fragment comprising a DNA encoding the protein of interest and a selectable marker gene and transposon sequences at both terminals of the gene fragment is introduced into the host cell, and a transposase is allowed to act upon the transposon sequences comprised in the expression vector which is introduced into the cell.

[0051] In order to allow a transposase to act upon the transposon sequences comprised in the expression vector which is introduced into the cell, the transposase may be injected into the cell, or an expression vector comprising a DNA encoding the transposase may be introduced into the host cell together with an expression vector comprising a DNA encoding the protein of interest and a selectable marker gene. In addition, by introducing an RNA encoding a transposase gene into the host cell, the transposase may be expressed in the cell.

[0052] The expression vector is not particularly limited. Any expression vector can be used by optionally selecting from the expression vectors known to those skilled in the art, depending on a host cell into which an expression vector comprising a transposase gene is introduced; the use; and the like.

[0053] In order that a protein constituted from two or more polypeptides is produced by the method of the invention, the DNA can be integrated into the chromosome of the cell by integrating a DNA encoding the two or more polypeptides into the same or different expression vectors and then introducing the expression vectors into a host cell.

[0054] The transposase may be inserted into an expression vector to express together with the protein of interest or may be inserted into a vector different from the expression vector. The transposase may be allowed to act transiently

or may be allowed to act continuously, but it is preferably to allow the transposase to act transiently in order to prepare a cell for stable production.

[0055] As the method for allowing the transposase to act transiently, examples include a method comprising preparing an expression vector which comprises a DNA encoding the transposase and an expression vector comprising a DNA encoding a protein of interest and then introducing both of the expression plasmids simultaneously into a host cell.

[0056] The term "expression vector" in the present specification means an expression vector to be used for introducing a mammalian cell in order to express a protein of interest. The expression vector used in the invention has a structure in which at least a pair of transposon sequences is present at both sides of an expression cassette.

[0057] The term "expression cassette" in the present specification means a nucleotide sequence which has a gene expression controlling region necessary for expressing a protein of interest and a sequence encoding the protein of interest. Examples of the gene expression controlling region include an enhancer, a promoter, and a terminator. the expression cassette may contain a selectable marker gene.

[0058] Any promoter can be used, so long as it can function in an animal cell. Examples include a promoter of IE (immediate early) gene of cytomegalovirus (CMV), SV40 early promoter, a promoter of retrovirus, a metallothionein promoter, a heat shock promoter, SR α promoter, moloney murine leukemia virus, an enhancer and the like. Also, the enhancer of the IE gene of human CMV can be used together with the promoter.

[0059] The "selectable marker gene" means an arbitral other marker gene which can be used for distinguishing a cell to which a plasmid vector is introduced from a cell lacking of the vector.

[0060] Examples of the selectable marker gene include a drug resistance gene (a neomycin resistance gene, a DHFR gene, a puromycin resistance gene, a blasticidin resistance gene, a hygromycin resistance gene, and a cycloheximide resistance gene (Japanese Published Unexamined Patent Application No.262879/2002)), fluorescence and bio-luminescence marker genes (such as green fluorescent protein GFP) and the like.

[0061] In the invention, preferable selectable marker is a drug resistance gene and particularly preferable selectable marker is a cycloheximide resistance gene. In addition, by carrying out a gene modification of the selectable marker gene, drug resistance performance and luminescence performance of the selectable marker protein can also be modified.

[0062] Cycloheximide (hereinafter sometimes referred to as CHX) is a protein synthesis inhibitor, and as examples of the use of the CHX resistance gene as a selectable marker gene, the cases of yeast (Kondo K. J. Bacteriol., 177, 24, 7171 - 7177 (1995)) and animal cells (Japanese Published Unexamined Patent Application No.262879/2002) are known.

[0063] In the case of the animal cells, it has been found that the resistance to cycloheximide is provided by a transformant which expresses a protein encoded by the nucleotide sequence shown in SEQ ID NO:7 of Sequence Listing in which proline at position 54 in human ribosomal protein subunit L36a encoded by the nucleotide sequence shown in SEQ ID NO:5 of Sequence Listing is substituted with glutamine.

[0064] The method for introducing the above-mentioned protein expression vector comprising a transposon sequence, a transposase expressing plasmid vector and RNA is not particularly limited. Examples include calcium phosphate transfection, electroporation, a liposome method, a gene gun method, lipofection and the like.

[0065] Examples of the method for directly introducing a transposase in the form of a protein include by microinjection or endocytosis for supplying into a cell. The gene transfer can be carried out by the method described in Shin Idenshi Kogaku Handbook (New Genetic Engineering Handbook), edited by Masami Muramatsu and Tadashi Yamamoto, published by Yodo-sha, ISBN 9784897063737.

[0066] The host cell is a suspension mammalian cell. The mammalian cell is a Chinese hamster ovarian cell CHO cell (Journal of Experimental Medicine, 108, 945 (1958); Proc. Natl. Acad. Sci. USA., 601275 (1968); Genetics, 55, 513 (1968); Chromosoma, 41, 129 (1973); Methods in Cell Science, 18, 115 (1996); Radiation Research, 148, 260 (1997); Proc. Natl. Acad. Sci. USA., 77, 4216 (1980); Proc. Natl. Acad. Sci., 60, 1275 (1968); Cell, 6, 121 (1975); Molecular Cell Genetics, Appendix I,II (pp. 883-900)). Examples of the CHO cell include CHO/DG44, CHO-K1 (ATCC CCL-61), DUKXB11 (ATCC CCL-9096), Pro-5 (ATCC CCL-1781), CHO-S (Life Technologies, Cat #11619), Pro-3 and substrain of CHO cell.

[0067] In addition, the above-mentioned host cell can also be used in the protein production method of the invention by modifying it so as to be suitable for the protein production, by modification of chromosomal DNA, introduction of an exogenous gene, and the like.

[0068] Further, in order to control the sugar chain structure bound to a protein of interest to be produced, Lec13 which acquired lectin resistance [Somatic Cell and Molecular Genetics, 12, 55 (1986)] and CHO cell from which α 1,6-fucosyl-transferase gene is deleted (WO2005/35586, WO2002/31140) can also be used as the host cell.

[0069] The protein of interest may be any protein so long as it can be expressed by the method of the invention. Specifically, examples include a human serum protein, a peptide hormone, a growth factor, a cytokine, a blood coagulation factor, a fibrinolysis system protein, an antibody and partial fragments of various proteins, and the like.

[0070] Preferable examples of the protein of interest include a monoclonal antibody such as a chimeric antibody, a humanized antibody and a human antibody; Fc fusion protein; and albumin-bound protein; and a fragment thereof.

[0071] An effector activity of a monoclonal antibody obtained by the method of the present invention can be controlled

by various methods. For example, known methods are a method for controlling an amount of fucose (hereinafter, referred to also as "core fucose") which is bound N-acetylglucosamine (GlcNAc) through α -1,6 bond in a reducing end of a complex type N-linked sugar chain which is bound to asparagine (Asn) at position 297 of an Fc region of an antibody (WO2005/035586, WO2002/31140, and WO00/61739), a method for controlling an effector activity of a monoclonal antibody by modifying amino acid group(s) of an Fc region of the antibody, and the like. The effector activity of the monoclonal antibody produced by the method of the present invention can be controlled by using any of the methods.

[0072] The "effector activity" means an antibody-dependent activity which is induced via an Fc region of an antibody. As the effector activity, an antibody-dependent cellular cytotoxicity (ADCC activity), a complement-dependent cytotoxicity (CDC activity), an antibody-dependent phagocytosis (ADP activity) by phagocytic cells such as macrophages or dendritic cells, and the like are known.

[0073] In addition, by controlling a content of core fucose of a complex type N-linked sugar chain of Fc region of a monoclonal antibody, an effector activity of the antibody can be increased or decreased.

[0074] As a method for lowering a content of fucose which is bound to a complex type N-linked sugar chain bound to Fc of the antibody, an antibody to which fucose is not bound can be obtained by the expression of an antibody using a CHO cell which is deficient in a gene encoding α 1,6-fucosyltransferase. The antibody to which fucose is not bound has a high ADCC activity.

[0075] On the other hand, as a method for increasing a content of fucose which is bound to a complex type N-linked sugar chain bound to Fc of an antibody, an antibody to which fucose is bound can be obtained by the expression of an antibody using a host cell into which a gene encoding α 1,6-fucosyltransferase is introduced. The antibody to which fucose is bound has a lower ADCC activity than the antibody to which fucose is not bound.

[0076] Further, by modifying amino acid residue(s) in an Fc region of an antibody, the ADCC activity or CDC activity can be increased or decreased. For example, the CDC activity of an antibody can be increased by using the amino acid sequence of the Fc region described in US2007/0148165.

[0077] Further, the ADCC activity or CDC activity of an antibody can be increased or decreased by modifying the amino acid as described in US Patent Nos. 6,737,056, or 7,297,775 or 7,317,091.

[0078] The term "suspension mammalian cell" in the present invention means a cell which does not adhere to a cell culture anchorage coated for facilitating adhesion of culture cells, such as microbeads, a culture container for tissue culture (also referred to as a tissue culture or adhesion culture container and the like) and the like, and can survive and grow by suspending in the culture liquid.

[0079] When the cell does not adhere to the cell culture anchorage, it may survive and grow under a state of a single cell in the culture liquid or survive and grow under a state of a cell mass formed by the agglutination of two or more cells.

[0080] In addition, as the suspension mammalian cell to be used in the present invention, a cell which can survive and grow in a serum-free medium that does not contain fetal calf serum (hereinafter referred to as FCS) and the like, while suspending in the culture liquid without adhering to the cell culture anchorage, is preferable, and a mammalian cell which can survive and grow while suspending in a protein-free medium that does not contain protein is more preferable.

[0081] As the culture container for tissue culture, it may be any culture container such as a flask, a Petri dish and the like, so long as coating for adhesion culture is applied thereto. Specifically, for example, whether or not it is a suspension mammalian cell can be confirmed by the use of commercially available tissue culture flask (manufactured by Greiner), adhesion culture flask (manufactured by Sumitomo Bakelite) and the like.

[0082] As the suspension mammalian cell to be used in the present invention, it may be either a CHO cell prepared by further adapting a CHO cell originally having a suspension property to suspension culture or a suspension CHO cell prepared by adapting an adhesive CHO cell to suspension culture conditions.

[0083] Examples of the cell originally having a suspension property include CHO-S cell (manufactured by Invitrogen) and the like.

[0084] The aforementioned "suspension mammalian cell prepared by adapting an adhesive mammalian cell to suspension culture conditions" can be prepared by the method described in Mol. Biotechnol., 2000, 15(3), 249 - 57 or by the method shown in the following, and can be prepared by establishing a cell which shows proliferation property and surviving property similar to those before the suspension culture adaptation or superior to those before adapting to suspension culture (J. Biotechnol., 2007, 130(3), 282 - 90).

[0085] The term "similar to those before the suspension culture adaptation" means that survival ratio, proliferation rate (doubling time) and the like of the cell adapted to the suspension culture are substantially the same as those of the cell before adapting suspension culture.

[0086] Examples of the method for adapting an adhesive mammalian cell to suspension culture conditions according to the present invention include the following method. The serum content of a serum-containing medium is reduced to 1/10 and sub-culturing is repeated at relatively high concentration of cell. When the mammalian cell comes to be able to survive and proliferate, the serum content is further reduced and the sub-culturing is repeated. By this method, a suspension mammalian cell which can survive and proliferate under serum-free conditions can be prepared.

[0087] In addition, a suspension mammalian cell can also be prepared by a method comprising culturing with the

addition of an appropriate nonionic surfactant such as Pluronic-F68 or the like in the culture liquid.

[0088] In the present invention, as a property possessed by the suspension mammalian cell, when 2×10^5 cells/ml of the cell is suspension-cultured, the cell concentration after culturing for 3 or 4 days is preferably 5×10^5 cells/ml or more, more preferably 8×10^5 cells/ml or more, particularly preferably 1×10^6 cells/ml or more, most preferably 1.5×10^6 cells/ml or more.

[0089] In addition, doubling time of the suspension mammalian cell of the present invention is preferably 48 hours or less, more preferably 24 hours or less, particularly preferably 18 hours or less, most preferably 11 hours or less.

[0090] Examples of the medium for suspension culturing include commercially available media, such as CD-CHO medium (manufactured by Invitrogen), EX-CELL 325-PF medium (manufactured by SAFC Biosciences), SFM4CHO medium (manufactured by HyClone) and the like. In addition, it can also be obtained by mixing saccharides, amino and the like acids which are necessary for the culturing of mammalian cells.

[0091] The suspension mammalian cell can be cultured using a culture container which can be used for suspension culturing under a culture condition capable of suspension culturing. Examples of the culture container include a 96 well plate for cell culture (manufactured by Corning), a T-flask (manufactured by Becton Dickinson), an Erlenmeyer flask (manufactured by Corning) and the like.

[0092] Regarding the culture conditions, for example, it can be statically cultured in an atmosphere of 5% CO_2 at a culture temperature of 37°C . A shaking culture equipment, such as culturing equipment for suspension culture exclusive use, Wave Bioreactor (manufactured by GE Healthcare Bioscience), can also be used.

[0093] Regarding the suspension culture conditions of a suspension mammalian cell using the Wave Bioreactor equipment, the cell can be cultured by the method described on the GE Healthcare Bioscience homepage <http://www.gelifesciences.co.jp/tech-support/manual/pdf/cellcult/wave-03-16.pdf>.

[0094] In addition to the shaking culture, culturing by a rotation agitation equipment such as a bioreactor, can also be used. Culturing using a bioreactor can be carried out by the method described in Cytotechnology, (2006) 52: 199 - 207, and the like.

[0095] In the present invention, when a cell line other than the suspension mammalian cells is used, any cell line can be used so long as it is a mammalian cell line adapted to the suspension culture by the above-mentioned method and is a cell line which can be used in the protein producing method of the present invention.

[0096] Purification of the protein of interest produced by the suspension mammalian cell is carried out by separating the protein of interest from impurities other than the protein of interest in a culture liquid or cell homogenate containing the protein of interest. Examples of the separation method include centrifugation, dialysis, ammonium sulfate precipitation, column chromatography, a filter and the like. The separation can be carried out based on the difference in physicochemical properties of the protein of interest and impurities and based on the difference in their affinity for the column carrier.

[0097] The method for purifying the protein of interest can be carried out, for example, by the method described in Protein Experimentation Note (the first volume) - Extraction, Separation and Expression of Recombinant Protein (translation of a textbook written in Japanese) (edited by Masato Okada and Kaori Miyazaki, published by Yodo-sha, ISBN 9784897069180).

[0098] The present invention has been described in the foregoing by showing preferred embodiments thereof for the sake of easy understanding. Hereinafter, the present invention is further described specifically based on examples, but the above-mentioned explanations and the following examples are provided merely for the purpose of exemplifications and not provided for the purpose of limiting the invention. Accordingly, the scope of the invention is not limited to the embodiments and examples which are specifically described herein, but is limited by the claims alone.

[0099] Various experimental techniques relating to genetic recombination described hereinafter, such as the cloning and the like were carried out in accordance with the genetic engineering techniques described in Molecular Cloning 2nd edition edited by J. Sambrook, E. F. Fritschy and T. Maniatis, Current Protocols in Molecular Biology edited by Frederick M. Ausubel et al, published by Current Protocols, and the like.

EXAMPLES

[Example 1]

Preparation of transposon vector for expressing anti-human influenza M2 antibody

[0100] A plasmid which contains a gene expression cassette for mammalian cells comprising an arbitrary human antibody gene and a drug resistance marker gene inserted between a pair of Tol2 transposon sequences was used as a plasmid vector for protein expression.

[0101] Each DNA of the used genes was chemically and artificially synthesized based on a known nucleotide sequence or obtained by preparing primers for its both terminal sequences and then carrying out PCR using an appropriate DNA source as a template. In order to carry out the gene manipulation later, a restriction site for a restriction enzyme was

added to the terminal of the primer.

[0102] Among the nucleotide sequence of the non-autonomous Tol2 transposone disclosed by Japanese Published Unexamined Patent Application No.235575/2003 (SEQ ID NO:1), the nucleotide sequence at position 1 to 200 (Tol2-L sequence) (SEQ ID NO:2) and the nucleotide sequence at positions 2285 to 2788 (Tol2-R sequence) (SEQ ID NO:3) were used as the transposon sequences.

[0103] Each synthetic DNA fragments comprising a pair of transposon sequences (manufactured by TAKARA BIO INC.) was prepared by the following method. A DNA fragment comprising a nucleotide sequence in which a recognition sequence of a restriction enzyme *NruI* was attached to both of the 5'-terminal and 3'-terminal of the Tol2-R sequence was prepared. Then, a DNA fragment comprising a nucleotide sequence in which a recognition sequence of a restriction enzyme *FseI* was attached to the 5'-terminal of the Tol2-L sequence and a restriction enzyme *AscI* was attached to the 3'-terminal thereof was prepared.

[0104] Next, the thus prepared DNA fragments comprising Tol2-R sequence and Tol2-L sequence were inserted into an expression vector N5LG1-M2-Z3 vector (WO2006/061723) comprising a nucleotide sequence encoding an amino acid sequence of anti-human influenza M2 antibody Z3G1.

[0105] The N5LG1-M2-Z3 vector (WO2006/061723) into which a nucleotide sequence (SEQ ID NO:8) encoding the H chain of the anti-human influenza M2 antibody Z3G1 (ATCC Deposit No. PTA-5968: deposited March 13, 2004, American Type Culture Collection, Manassas, VA, USA) and a nucleotide sequence (SEQ ID NO:10 and SEQ ID NO:11) encoding the L chain (SEQ ID NO:9) of the same were inserted under the control of the CMV enhancer/promoter control was used as an antibody gene expression cassette.

[0106] The DNA fragment comprising the Tol2-R sequence was inserted into the restriction enzyme *NruI* site of the N5LG1-M2-Z3 vector, at the 5'-terminal side of a gene fragment comprising the antibody gene expression cassette and a resistance marker gene. Then, the DNA fragment comprising the Tol2-L sequence was inserted into the restriction enzyme *FseI* and *AscI* sites at the 3'-terminal side.

[0107] In addition, a transposon vector for expressing an anti-human influenza M2 antibody was constructed (Fig. 1) by inserting a cycloheximide resistance gene expression cassette connected with a nucleotide sequence (SEQ ID NO:5) encoding a resistance gene for cycloheximide (a gene in which proline at position 54 of the human ribosomal protein L36a was substituted with glutamine) into the *FseI* recognition site of the N5LG1-M2-Z3 vector connected with the Tol2 transposon sequence, under the control of the CMV enhancer/promoter.

[0108] On the other hand, a vector containing no transposon sequences was named anti-human influenza M2 antibody expression vector and used as the control vector (Fig. 2).

[Example 2]

Preparation of transposase expression vector

[0109] The transposase was expressed using an expression vector independent of the expression vector of the antibody of interest. That is, a gene which is encoding a medaka fish-derived Tol2 transposase (SEQ ID NO:4) was inserted into a downstream of the CAGGS promoter of a pCAGGS vector (Gene, 108, 193 - 200, 1991) and used as the expression vector (Fig. 3).

[Example 3]

(1) Preparation of suspension CHO cell

[0110] An adhesive CHO cell which had been cultured using an α -MEM medium (manufactured by Invitrogen) containing 10% serum (FCS) was peeled off and recovered by a trypsin treatment and shaking-cultured at 37°C in a 5% CO₂ incubator using fresh α -MEM medium containing 10% FCS. Several days thereafter, growth of these cells was confirmed and then shaking culture was carried out by seeding them into a α -MEM medium containing 5% FCS at a concentration of 2×10^5 cells/ml.

[0111] Further several days thereafter, the inoculation was similarly carried out using the α -MEM medium containing 5% FCS. Finally, a cell adapted to the suspension culture was prepared by repeating the sub-culture and shaking culture using serum-free α -MEM medium and confirming that the cells have the same growing ability of the case of their culturing in the presence of serum.

(2) Preparation of antibody-producing CHO cell

[0112] The transposon vector for expressing the anti-human influenza M2 antibody prepared in Example 1 and Example 2 (hereinafter referred to as transposon vector) and Tol2 transposase expression vector pCAGGS-T2TP (Fig. 3, Kawaka-

mi K. & Noda T., Genetics, 166, 895 - 899 (2004)) were used as the expression vectors. In addition, the anti-human influenza M2 antibody expression vector having no transposon sequences was used as the control.

[0113] By introducing the aforementioned expression vectors into the suspension culture-adapted CHO-K1 cell (American Type Culture Collection Cat. No. CCL-61) or HEK293 cell (FreeStyle 293F cell, manufactured by Invitrogen), the frequencies of obtaining cycloheximide-resistant clones were compared.

[0114] Each cells (4×10^6 cells) was suspended in 400 μ l of PBS, and the transposon vector for expressing the anti-human influenza M2 antibody (10 μ g) and Tol2 transposase expression vector (25 μ g) were co-transfected directly in the form of circular DNA by electroporation. In this connection, in order to express the Tol2 transposase transiently, the Tol2 transposase expression vector was directly introduced in the form of circular DNA for the purpose of preventing from integrateing into the host chromosome.

[0115] In addition, as the control, the anti-human influenza M2 antibody expression vector (10 μ g) was linearized by a restriction enzyme and then introduced into each cells, in accordance with the standard gene transfer method by electroporation.

[0116] The electroporation was carried out using a cuvette of 4 mm in gap width (manufactured by Bio-Rad), using an electroporator (Gene Pulser Xcell System (manufactured by Bio-Rad)) under conditions of 300 V in voltage, 500 μ F in electrostatic capacity and room temperature.

[0117] After the transfection by electroporation, each cell was seeded into three 96-well plates and cultured in a CO₂ incubator for 3 days using the EX-CELL 325-PF medium manufactured by SAFC Biosciences for the CHO cell, and the FreeStyle-293 medium (manufactured by Invitrogen) for the HEK293 cell.

[0118] Next, from the day of medium exchange on the 4th day of the transfection, 3 μ g/ml of cycloheximide was added to the medium so that the cells were cultured in the presence of cycloheximide, followed by culturing for 3 weeks while carrying out the medium exchange in every week.

[0119] After culturing for 3 weeks, the number of wells in which cycloheximide-resistant colonies were found was counted. The results are shown in Table 1 and Table 2.

[Table 1]

[0120]

Table 1 Comparison of the numbers of cycloheximide-resistant cells (CHO cell)

	Transposon vector	Conventional vector
Test 1	155 / 288	0 / 288
Test 2	100 / 288	0 / 288
Test 3	94 / 288	0 / 288

[Table 2]

[0121]

Table 2 Comparison of the numbers of cycloheximide-resistant cells (HEK293 cell)

	Transposon vector	Conventional vector
Test 1	0 / 288	0 / 288
Test 2	0 / 288	0 / 288
Test 3	0 / 288	0 / 288

[0122] As shown in Table 1, each the anti-human influenza M2 antibody expression transposon vector or anti-human influenza M2 antibody expression vector was introduced into the suspension CHO-K1 cell. As a result, cycloheximide-resistant transformants were not obtained from the cell introduced with anti-human influenza M2 antibody expression vector like the case of other cell lines, but cycloheximide-resistant transformants were obtained from the cell introduced with transposon vector for expressing anti-human influenza M2 antibody with a high frequency.

[0123] On the other hand, as shown in Table 2, cycloheximide-resistant transformants were not obtained when either of the transposon vector for expressing anti-human influenza M2 antibody and anti-human influenza M2 antibody expression vector was introduced into the HEK293 cell.

[0124] Based on these results, it was found that the intended protein-encoded gene and cycloheximide resistance

gene which were inserted between a pair of transposon sequences are efficiently introduced into the chromosome of the host cell, namely a suspension mammalian cell.

(3) Examination on the antibody production by suspension CHO cell and adhesive CHO cell

[0125] In order to examine antibody production efficiency by a suspension CHO cell or an adhesive CHO cell, the amounts of antibodies produced by respective cell lines were examined. As the suspension CHO cell, the suspension CHO-K1 cell adapted to suspension culture was used. In addition, as the adhesive CHO cell, the adhesive CHO-K1 cell before adaptation to suspension culture was used.

[0126] The anti-human influenza M2 antibody expression transposon vector (10 µg) and Tol2 transposase expression vector (25 µg) were introduced into the suspension CHO-K1 cell and adhesive CHO-K1 cell by means of electroporation, respectively. Thereafter, the suspension CHO-K1 cell and the adhesive CHO-K1 cell were seeded into three 96-well plates for each cell.

[0127] A medium for suspension cells (EX-CELL 325-PF, manufactured by SAFC Biosciences) was used for the suspension CHO-K1 cell, and the α-MEM medium containing 10% serum was used for the adhesive CHO-K1 cell. Each cell was cultured in a CO₂ incubator for 3 days. From the day of medium exchange on the 4th day of the transfection, 3 µg/ml of cycloheximide was added to the medium so that the cells were cultured in the presence of cycloheximide and the cells were further cultured for 3 weeks. In this case, the medium exchange was carried out every week.

[0128] For the suspension CHO-K1 cell, 1 x 10⁶ of the cells were seeded into a 6-well plate and shaking-cultured in a CO₂ incubator for 3 days, and the amount of the anti-human influenza M2 antibody protein was measured by HPLC using the culture supernatant.

[0129] For the adhesive CHO-K1 cell, medium exchange was carried out when the cell reached confluent on a 6-well plate (2 x 10⁶ cells), and 3 days after static culture, the amount of the antibody protein was measured by HPLC using the culture supernatant.

[0130] The antibody concentration in the culture supernatant was measured in accordance with the method described in Yeast Res., 7 (2007), 1307 - 1316. The results are shown in Fig. 4A and Fig. 4B.

[0131] As shown in Fig. 4A, a large number of cells showing a markedly high antibody expression level were obtained when the CHO-K1 cell adapted to suspension culture was used. On the other hand, as shown in Fig. 4B, only the cells showing an expression level of the HPLC detection limit (5 µg/ml) or less were obtained when the adhesive CHO-K1 cell was used.

[0132] Based on these results, it was found that, for the expression of a protein of interest using a transposon vector, the protein of interest can be expressed at a high level when a suspension mammalian cell is used.

[0133] In addition, it was found from the results of Examples 1 to 3 that the method of the invention can be used as a novel method for producing a protein of interest, by efficiently preparing a production cell which can highly express an exogenous gene using a suspension mammalian cell adapted to suspension culture.

[Example 4]

Preparation of Tol1 transposon vector for expressing anti-human influenza M2 antibody

[0134] In the same manner as in Example 1, a plasmid which contains a gene expression cassette for mammalian cells, comprising an arbitrary human antibody gene and a drug resistance marker gene inserted between a pair of Tol1 transposon sequences, was used as a protein expression plasmid vector.

[0135] Each DNA of the used genes was chemically synthesized artificially based on the known sequence information or obtained by preparing primers of its both terminal sequences and carrying out PCR using an appropriate DNA source as the template. For the gene manipulation to be carried out later, a site cleaved by a restriction enzyme was added to the end of the primer.

[0136] Among the non-autonomous Tol1 transposon nucleotide sequence shown in SEQ ID NO:13 of Sequence Listing (WO2008/072540), the nucleotide sequence at positions 1 to 200 (Tol1-L sequence) (SEQ ID NO:14) and the nucleotide sequence at positions 1351 to 1855 (Tol1-R sequence) (SEQ ID NO:15) were used as the transposon sequences.

[0137] Each of the synthetic DNA fragments comprising each a pair of transposon sequences was prepared by the following method. A DNA fragment comprising a nucleotide sequence in which a recognition sequence of a restriction enzyme *Nru*I was connected to both of the 5'-terminal and 3'-terminal of the Tol1-R sequence. Then, a DNA fragment comprising a nucleotide sequence in which a recognition sequence of a restriction enzyme *Fse*I was connected to the 5'-terminal of the Tol1-L sequence and a restriction enzyme *Asc*I was connected to the 3'-terminal thereof.

[0138] Next, the thus prepared DNA fragments comprising Tol1-R sequence and Tol1-L sequence were inserted into the expression vector N5LG1-M2-Z3 vector. The DNA fragment comprising the Tol1-R sequence was inserted into the

restriction enzyme *Nru*I site of the N5LG1-M2-Z3 vector, existing on the 5'-terminal side of a gene fragment comprising the antibody gene expression cassette and a resistance marker gene, and the DNA fragment comprising the Tol1-L sequence was inserted into the restriction enzyme *Fse*I and *Asc*I sites existing on the 3'-terminal side.

[0139] In addition, Tol1 transposon vector for expressing an anti-human influenza M2 antibody was constructed (Fig. 5) by inserting a cycloheximide resistance gene expression cassette connected with a resistance gene for cycloheximide (a gene in which proline at position 54 in the human ribosomal protein L36a was mutated to glutamine) into the *Fse*I recognition site of the N5LG1-M2-Z3 vector connected with the Tol1 transposon sequence, under the control of the CMV enhancer/promoter.

[Example 5]

Preparation of Tol1 transposase expression vector

[0140] The transposase was expressed using an expression vector independent from the expression vector of the antibody of interest. That is, a Tol1 transposase gene expression cassette connected with a DNA fragment encoding a medaka fish-derived Tol1 transposase, containing the nucleotide sequence shown in SEQ ID NO:16 of Sequence Listing, was inserted into pBluescriptII SK (+) (manufactured by Stratagene) under the CMV enhancer/promoter control and used as the expression vector pTol1ase (Fig. 6).

[Example 6]

(1) Preparation of antibody-producing CHO cell

[0141] The Tol1 transposon vector for expressing the anti-human influenza M2 antibody (hereinafter referred to as Tol1 transposon vector) and Tol1 transposase expression vector pTol1ase of Example 4 and Example 5 were used as the expression vectors. In addition, the CHO-K1 cell prepared by adapting to suspension culture in the same manner as in Example 3(1) was used as the cell.

[0142] The aforementioned expression vectors were introduced into the CHO-K1 cell adapted to suspension culture, and the frequency of obtaining clones resistant to cycloheximide was measured. The CHO-K1 cell adapted to suspension culture (4×10^6 cells) were suspended in 400 μ l of PBS, and the Tol1 transposon vector for expressing the anti-human influenza M2 antibody (10 μ g) and Tol1 transposase expression vector (50 μ g) were co-transfected directly in the form of circular DNA by electroporation. In order to effect transient expression of the Tol1 transposase, the Tol1 transposase expression vector was directly introduced in the form of circular DNA for the purpose of preventing from integrating into the host chromosome.

[0143] The electroporation was carried out using a cuvette of 4 mm in gap width (manufactured by Bio-Rad), using an electroporator (Gene Pulser Xcell System (manufactured by Bio-Rad)) under conditions of 300 V in voltage, 500 μ F in electrostatic capacity and room temperature.

[0144] After the transfection by electroporation, each cell was seeded into two 96-well plates and cultured in a CO₂ incubator for 3 days using the EX-CELL 325-PF medium (manufactured by SAFC Biosciences) for the CHO cell. Next, from the day of medium exchange on the 4th day of the transfection, 3 μ g/ml of cycloheximide was added to the medium so that the cells were cultured in the presence of cycloheximide, followed by culturing for 3 weeks while carrying out the medium exchange every week.

[0145] After the culturing for 3 weeks, the number of wells in which cycloheximide-resistant colonies were found was counted. The results are shown in Table 3. Each of the tests 1 to 3 in Table 3 shows a result of carrying out the gene transfer three times.

[Table 3]

Tol1 transposon vector	
Tests 1	133 / 192
Tests 2	67 / 192
Tests 3	122 / 192

[0146] As shown in Table 3, when the Tol1 transposon vector for expressing the anti-human influenza M2 antibody was introduced into the suspension CHO-K1 cell, cycloheximide-resistant transformants were obtained at a high frequency similarly to Example 3 in which the Tol2 transposon vector for expressing the anti-human influenza M2 antibody was introduced.

[0147] It was found based on these results that the antibody gene and cycloheximide resistance gene inserted between a pair of transposon sequences are efficiently transduced into the chromosome of the host cell, namely the suspension mammalian cell, in the case of using the Tol1 transposon, too.

(2) Examination on antibody production by suspension CHO-K1 cell

[0148] Antibody production efficiency of the suspension CHO-K1 cell was examined using the suspension CHO-K1 cell. The transposon vector for expressing the anti-human influenza M2 antibody (10 μ g) and Tol1 transposase expression vector (50 μ g) were introduced by electroporation into the suspension CHO-K1 cell adapted to suspension culture.

[0149] Thereafter, the cells were seeded into respective two 96-well plates and cultured for 3 days in a CO₂ incubator using the suspension culture medium EX-CELL 325-PF. From the medium exchange on the 4th days after the electroporation, the cells were cultured for 3 weeks in the presence of 3 μ g/ml of cycloheximide. In this case, the medium exchange was carried out every week.

[0150] For the suspension CHO-K1 cell, 1 x 10⁶ of the cells were seeded into a 6-well plate and shaking-cultured in a CO₂ incubator for 3 days, and amount of the anti-human influenza M2 antibody protein was measured by HPLC using the culture supernatant.

[0151] The antibody concentration in culture supernatant was measured in accordance with the method described in Yeast Res., 7 (2007), 1307 - 1316. The results are shown in Fig. 7.

[0152] As shown in Fig. 7, a large number of cells showing a markedly high antibody expression level were obtained in the case of the use of the Tol1 transposon, too. From this result, it was found that similar to the case of the use of the Tol2 transposon-derived nucleotide sequence, a suspension mammalian cell capable of highly expressing the protein of interest can also be obtained when a Tol1 transposon-derived nucleotide sequence is used as the transposon sequence.

[0153] This application is based on Japanese application No. 2009-140626, filed on June 11, 2009, and U.S. provisional application No. 61/186,138, filed on June 11, 2009.

Industrial Applicability

[0154] By the method for producing the protein of the present invention, a protein of interest can be efficiently produced using a suspension mammalian cell. The cell of the present invention can be used as a protein producing cell for producing a recombinant protein.

Sequence Listing Free Text

[0155]

SEQ ID NO:1 - Description of Artificial sequence: Nucleotide sequence of non-autonomous Tol2 transposon

SEQ ID NO:2 - Description of Artificial sequence: Tol2-L sequence

SEQ ID NO:3 - Description of Artificial sequence: Tol2-R sequence

SEQ ID NO:7 - Description of Artificial sequence: Nucleotide sequence of cycloheximide resistance gene

SEQ ID NO:8 - Description of Artificial sequence: Amino acid sequence of protein encoded by cycloheximide resistance gene

SEQ ID NO:9 - Description of Artificial sequence: Nucleotide sequence encoding H chain of M2Z3 antibody

SEQ ID NO:10 - Description of Artificial sequence: Nucleotide sequence encoding H chain of M2Z3 antibody

SEQ ID NO:11 - Description of Artificial sequence: Nucleotide sequence encoding L chain of M2Z3 antibody

SEQ ID NO:12 - Description of Artificial sequence: Amino acid sequence encoding L chain of M2Z3 antibody

SEQ ID NO:13 - Description of Artificial sequence: Nucleotide sequence of non-autonomous Tol1 transposon

SEQ ID NO:14 - Description of Artificial sequence: Tol1-L sequence

SEQ ID NO:15 - Description of Artificial sequence: Tol1-R sequence

SEQUENCE LISTING

[0156]

<110> Kyowa Hakko Kirin Co., Ltd.

Inter-University Research Institute Corporation Research Organization of Information and Systems

<120> Protein production method

EP 2 441 831 B9

<130> WO69874

<150> JP2009-140626

<151> 2009-06-11

<150> US61/186138

<151> 2009-06-11

<160> 17

<170> PatentIn version 3.3

<210> 1

<211> 2788

<212> DNA

<213> Artificial

<220>

<223> Description of artificial sequence; nonautologous Tol2 transposon

<400> 1

```

cagaggtgta aagtacttga gtaattttac ttgattactg tacttaagta ttatTTTTTgg      60
ggatTTTTTtac tttacttgag tacaattaaa aatcaatact tttactTTTta cttaattaca      120
TTTTTTTtaga aaaaaaagta ctttttactc cttacaattt tatttacagt caaaaagtac      180
ttatTTTTTtg gagatcactt cattctattt tcccttgcta ttaccaaacc aattgaattg      240
cgctgatgcc cagtTTtaatt taaatgttat ttattctgcc tatgaaaatc gttttcacat      300
tatatgaaat tggtcagaca tgttcattgg tcctttggaa gtgacgtcat gtcacatcta      360
ttaccacaat gcacagcacc ttgacctgga aattagggaa attataacag tcaatcagtg      420
gaagaaaatg gaggaagtat gtgattcatc agcagctgcg agcagcacag tccaaaatca      480
gccacaggat caagagcacc cgtggccgta tcttcgcgaa ttcttttctt taagtgggtg      540
aaataaagat tcattcaaga tgaaatgtgt cctctgtctc ccgcttaata aagaaatatc      600
ggccttcaaa agttcgccat caaacctaag gaagcatatt gaggtaagta cattaagtat      660
tttgTTTTTtac tgatagTTTT tttttttttt tttttttttt tttttgggtg tgcattTTTT      720
gacgttgatg gcgcgccttt tatatgtgta gtaggcctat tttcactaat gcatgcgatt      780
gacaatataa ggctcacgta ataaaatgct aaaatgcatt tgtaattggg aacgttaggt      840
ccacgggaaa tttggcgcct attgcagctt tgaataatca ttatcattcc gtgctctcat      900
tgtgtttgaa ttcattgcaa acacaagaaa accaagcgag aaatTTTTTt ccaaacatgt      960
tgtattgtca aaacggtaac actttacaat gaggttgatt agttcatgta ttaactaaca     1020
ttaaataacc atgagcaata catttggttac tgtatctgtt aatctttgtt aacgttaggt     1080

```

EP 2 441 831 B9

	aatagaaata cagatgttca ttgtttgttc atgttagttc acagtgcatt aactaatggt	1140
	aacaagatat aaagtattag taaatgttga aattaacatg tatacgtgca gttcattatt	1200
5	agttcatggt aactaatgta gttaactaac gaaccttatt gtaaaagtgt taccatcaaa	1260
	actaatgtaa tgaaatcaat tcaccctgtc atgtcagcct tacagtcctg tgtttttgtc	1320
	aatataatca gaaataaaaat taatgtttga ttgtcactaa atgctactgt atttctaaaa	1380
10	tcaacaagta tttaacatta taaagtgtgc aattggctgc aaatgtcagt tttattaaag	1440
	ggtagttca cccaaaaatg aaaataatgt cattaatgac tcgccctcat gtcgttccaa	1500
	gcccgtaga cctccgttca tcttcagaac acagtttaag atattttaga tttagtccga	1560
15	gagctttctg tgcctccatt gagaatgtat gtacggata ctgtccatgt ccagaaaggt	1620
	aataaaaaca tcaaagtagt ccatgtgaca tcagtgggtt agttagaatt ttttgaagca	1680
20	tcgaatacat tttgggtccaa aaataacaaa acctacgact ttattcggca ttgtattctc	1740
	ttccgggtct gttgtcaatc cgcgttcacg acttcgcagt gacgctacaa tgctgaataa	1800
	agtcgtaggt tttgttattt ttggaccaa atgtattttc gatgcttcaa ataattctac	1860
25	ctaaccact gatgtcacat ggactacttt gatgttttta ttacctttct ggacatggac	1920
	agtataccgt acatacatTT tcagtggagg gacagaaagc tctcggacta aatctaaaat	1980
	atcttaaaact gtgttccgaa gatgaacgga ggtgttacgg gcttgaacg acatgaggg	2040
30	gagtcattaa tgacatcttt tcatttttgg gtgaactaac cctttaatgc tgtaatcaga	2100
	gagtgtatgt gtaattgtta catTTattgc atacaatata aatatttatt tgttgttttt	2160
35	acagagaatg caccctaaatt acctcaaaaa ctactctaaa ttgacagcac agaagagaaa	2220
	gatcgggaca gatctcatat gctcgagggc ccatctggcc tgtgtttcag acaccaggga	2280
	gtctctgctc acgtttcctg ctatttgcag cctctctatc aagactaata cacctcttcc	2340
40	cgcacgggct gcctgtgaga ggcttttcag cactgcagga ttgcttttca gcccctaaag	2400
	agctaggctt gacactaaca attttgagaa tcagcttcta ctgaagttaa atctgaggtt	2460
45	ttacaacttt gagtagcgtg tactggcatt agattgtctg tcttatagtt tgataattaa	2520
	atacaaacag ttctaaagca ggataaaacc ttgtatgcat ttcatTTaat gttttttgag	2580
	attaaaagct taaacaagaa tctctagttt tctttcttgc ttttactttt acttccttaa	2640
50	tactcaagta caattttaat ggagtacttt tttactttta ctcaagtaag attctagcca	2700
	gatactttta ctttttaattg agtaaaattt tccctaagta cttgtacttt cacttgagta	2760
55	aaatttttga gtacttttta cacctctg	2788

<210> 2

<211> 200

<212> DNA

EP 2 441 831 B9

<213> Artificial

<220>

<223> Description of artificial sequence; Tol2-L transposon sequence

5

<400> 2

10	cagaggtgta aagtacttga gtaattttac ttgattactg tacttaagta ttatTTTTTgg	60
	ggattttttac tttacttgag tacaattaaa aatcaatact tttactttta cttaattaca	120
	ttttttttaga aaaaaaagta cttttttactc cttacaattt tatttacagt caaaaagtac	180
15	ttatTTTTTtg gagatcactt	200

<210> 3

<211> 504

<212> DNA

20 <213> Artificial

<220>

<223> Description of artificial sequence; Tol2-R transposon sequence

25 <400> 3

30	ctgctcacgt ttcctgctat ttgcagcctc tctatcaaga ctaatacacc tcttcccgca	60
	tcggtgcct gtgagaggct tttcagcact gcaggattgc ttttcagccc caaaagagct	120
	aggcttgaca ctaacaattt tgagaatcag cttctactga agttaaatct gaggttttac	180
	aactttgagt agcgtgtact ggcattagat tgtctgtctt atagtttgat aattaaatac	240
35	aaacagttct aaagcaggat aaaaccttgt atgcatttca tttaatgttt tttgagatta	300
	aaagcttaaa caagaatctc tagttttctt tcttgctttt acttttactt ccttaatact	360
	caagtacaat tttaatggag tactttttta cttttactca agtaagattc tagccagata	420
40	cttttacttt taattgagta aaattttccc taagtacttg tactttcact tgagtaaaat	480
	ttttgagtac tttttacacc tctg	504

45 <210> 4

<211> 2156

<212> DNA

<213> Oryzias latipes

50 <220>

<221> CDS

<222> (85)..(2034)

<400> 4

55

EP 2 441 831 B9

	acgtcatgtc acatctatta ccacaatgca cagcaccttg acctggaaat tagggaaatt	60
	ataacagtca atcagtggaa gaaa atg gag gaa gta tgt gat tca tca gca	111
5	Met Glu Glu Val Cys Asp Ser Ser Ala	
	1 5	
	gct gcg agc agc aca gtc caa aat cag cca cag gat caa gag cac ccg	159
	Ala Ala Ser Ser Thr Val Gln Asn Gln Pro Gln Asp Gln Glu His Pro	
10	10 15 20 25	
	tgg ccg tat ctt cgc gaa ttc ttt tct tta agt ggt gta aat aaa gat	207
	Trp Pro Tyr Leu Arg Glu Phe Phe Ser Leu Ser Gly Val Asn Lys Asp	
	30 35 40	
15		
20		
25		
30		
35		
40		
45		
50		
55		

EP 2 441 831 B9

	tca ttc aag atg aaa tgt gtc ctc tgt ctc ccg ctt aat aaa gaa ata	255
	Ser Phe Lys Met Lys Cys Val Leu Cys Leu Pro Leu Asn Lys Glu Ile	
	45 50 55	
5	tcg gcc ttc aaa agt tcg cca tca aac cta agg aag cat att gag aga	303
	Ser Ala Phe Lys Ser Ser Pro Ser Asn Leu Arg Lys His Ile Glu Arg	
	60 65 70	
10	atg cac cca aat tac ctc aaa aac tac tct aaa ttg aca gca cag aag	351
	Met His Pro Asn Tyr Leu Lys Asn Tyr Ser Lys Leu Thr Ala Gln Lys	
	75 80 85	
15	aga aag atc ggg acc tcc acc cat gct tcc agc agt aag caa ctg aaa	399
	Arg Lys Ile Gly Thr Ser Thr His Ala Ser Ser Ser Lys Gln Leu Lys	
	90 95 100 105	
20	gtt gac tca gtt ttc cca gtc aaa cat gtg tct cca gtc act gtg aac	447
	Val Asp Ser Val Phe Pro Val Lys His Val Ser Pro Val Thr Val Asn	
	110 115 120	
25	aaa gct ata tta agg tac atc att caa gga ctt cat cct ttc agc act	495
	Lys Ala Ile Leu Arg Tyr Ile Ile Gln Gly Leu His Pro Phe Ser Thr	
	125 130 135	
30	gtt gat ctg cca tca ttt aaa gag ctg att agt aca ctg cag cct ggc	543
	Val Asp Leu Pro Ser Phe Lys Glu Leu Ile Ser Thr Leu Gln Pro Gly	
	140 145 150	
35	att tct gtc att aca agg cct act tta cgc tcc aag ata gct gaa gct	591
	Ile Ser Val Ile Thr Arg Pro Thr Leu Arg Ser Lys Ile Ala Glu Ala	
	155 160 165	
40	gct ctg atc atg aaa cag aaa gtg act gct gcc atg agt gaa gtt gaa	639
	Ala Leu Ile Met Lys Gln Lys Val Thr Ala Ala Met Ser Glu Val Glu	
	170 175 180 185	
45	tgg att gca acc aca acg gat tgt tgg act gca cgt aga aag tca ttc	687
	Trp Ile Ala Thr Thr Thr Asp Cys Trp Thr Ala Arg Arg Lys Ser Phe	
	190 195 200	
50	att ggt gta act gct cac tgg atc aac cct gga agt ctt gaa aga cat	735
	Ile Gly Val Thr Ala His Trp Ile Asn Pro Gly Ser Leu Glu Arg His	
	205 210 215	
55	tcc gct gca ctt gcc tgc aaa aga tta atg ggc tct cat act ttt gag	783
	Ser Ala Ala Leu Ala Cys Lys Arg Leu Met Gly Ser His Thr Phe Glu	
	220 225 230	
60	gta ctg gcc agt gcc atg aat gat atc cac tca gag tat gaa ata cgt	831
	Val Leu Ala Ser Ala Met Asn Asp Ile His Ser Glu Tyr Glu Ile Arg	
	235 240 245	
65	gac aag gtt gtt tgc aca acc aca gac agt ggt tcc aac ttt atg aag	879
	Asp Lys Val Val Cys Thr Thr Thr Asp Ser Gly Ser Asn Phe Met Lys	
	250 255 260 265	
70	gct ttc aga gtt ttt ggt gtg gaa aac aat gat atc gag act gag gca	927
	Ala Phe Arg Val Phe Gly Val Glu Asn Asn Asp Ile Glu Thr Glu Ala	
	270 275 280	
75	aga agg tgt gaa agt gat gac act gat tct gaa ggc tgt ggt gag gga	975
	Arg Arg Cys Glu Ser Asp Asp Thr Ser Glu Gly Cys Gly Glu Gly	
	285 290 295	
80	agt gat ggt gtg gaa ttc caa gat gcc tca cga gtc ctg gac caa gac	1023

EP 2 441 831 B9

	Ser	Asp	Gly	Val	Glu	Phe	Gln	Asp	Ala	Ser	Arg	Val	Leu	Asp	Gln	Asp	
			300					305					310				
5	gat	ggc	ttc	gaa	ttc	cag	cta	cca	aaa	cat	caa	aag	tgt	gcc	tgt	cac	1071
	Asp	Gly	Phe	Glu	Phe	Gln	Leu	Pro	Lys	His	Gln	Lys	Cys	Ala	Cys	His	
		315					320					325					
10	tta	ctt	aac	cta	gtc	tca	agc	ggt	gat	gcc	caa	aaa	gct	ctc	tca	aat	1119
	Leu	Leu	Asn	Leu	Val	Ser	Ser	Val	Asp	Ala	Gln	Lys	Ala	Leu	Ser	Asn	
	330					335				340						345	
15	gaa	cac	tac	aag	aaa	ctc	tac	aga	tct	gtc	ttt	ggc	aaa	tgc	caa	gct	1167
	Glu	His	Tyr	Lys	Lys	Leu	Tyr	Arg	Ser	Val	Phe	Gly	Lys	Cys	Gln	Ala	
					350					355					360		
20	tta	tgg	aat	aaa	agc	agc	cga	tcg	gct	cta	gca	gct	gaa	gct	ggt	gaa	1215
	Leu	Trp	Asn	Lys	Ser	Ser	Arg	Ser	Ala	Leu	Ala	Ala	Glu	Ala	Val	Glu	
				365					370					375			
25	tca	gaa	agc	cgg	ctt	cag	ctt	tta	agg	cca	aac	caa	acg	cgg	tgg	aat	1263
	Ser	Glu	Ser	Arg	Leu	Gln	Leu	Leu	Arg	Pro	Asn	Gln	Thr	Arg	Trp	Asn	
				380				385						390			
30	tca	act	ttt	atg	gct	ggt	gac	aga	att	ctt	caa	att	tgc	aaa	gaa	gca	1311
	Ser	Thr	Phe	Met	Ala	Val	Asp	Arg	Ile	Leu	Gln	Ile	Cys	Lys	Glu	Ala	
		395					400						405				
35	gga	gaa	ggc	gca	ctt	cgg	aat	ata	tgc	acc	tct	ctt	gag	ggt	cca	atg	1359
	Gly	Glu	Gly	Ala	Leu	Arg	Asn	Ile	Cys	Thr	Ser	Leu	Glu	Val	Pro	Met	
	410					415					420					425	
40	ttt	aat	cca	gca	gaa	atg	ctg	ttc	ttg	aca	gag	tgg	gcc	aac	aca	atg	1407
	Phe	Asn	Pro	Ala	Glu	Met	Leu	Phe	Leu	Thr	Glu	Trp	Ala	Asn	Thr	Met	
				430						435					440		
45	cgt	cca	gtt	gca	aaa	gta	ctc	gac	atc	ttg	caa	gcg	gaa	acg	aat	aca	1455
	Arg	Pro	Val	Ala	Lys	Val	Leu	Asp	Ile	Leu	Gln	Ala	Glu	Thr	Asn	Thr	
				445					450					455			
50	cag	ctg	ggg	tgg	ctg	ctg	cct	agt	gtc	cat	cag	tta	agc	ttg	aaa	ctt	1503
	Gln	Leu	Gly	Trp	Leu	Leu	Pro	Ser	Val	His	Gln	Leu	Ser	Leu	Lys	Leu	
			460					465						470			
55	cag	cga	ctc	cac	cat	tct	ctc	agg	tac	tgt	gac	cca	ctt	gtg	gat	gcc	1551
	Gln	Arg	Leu	His	His	Ser	Leu	Arg	Tyr	Cys	Asp	Pro	Leu	Val	Asp	Ala	
		475					480						485				
60	cta	caa	caa	gga	atc	caa	aca	cga	ttc	aag	cat	atg	ttt	gaa	gat	cct	1599
	Leu	Gln	Gln	Gly	Ile	Gln	Thr	Arg	Phe	Lys	His	Met	Phe	Glu	Asp	Pro	
	490					495					500					505	
65	gag	atc	ata	gca	gct	gcc	atc	ctt	ctc	cct	aaa	ttt	cgg	acc	tct	tgg	1647
	Glu	Ile	Ile	Ala	Ala	Ala	Ile	Leu	Leu	Pro	Lys	Phe	Arg	Thr	Ser	Trp	
					510					515					520		
70	aca	aat	gat	gaa	acc	atc	ata	aaa	cga	ggc	atg	gac	tac	atc	aga	gtg	1695
	Thr	Asn	Asp	Glu	Thr	Ile	Ile	Lys	Arg	Gly	Met	Asp	Tyr	Ile	Arg	Val	
				525					530					535			
75	cat	ctg	gag	cct	ttg	gac	cac	aag	aag	gaa	ttg	gcc	aac	agt	tca	tct	1743
	His	Leu	Glu	Pro	Leu	Asp	His	Lys	Lys	Glu	Leu	Ala	Asn	Ser	Ser	Ser	
			540					545					550				
80	gat	gat	gaa	gat	ttt	ttc	gct	tct	ttg	aaa	ccg	aca	aca	cat	gaa	gcc	1791
	Asp	Asp	Glu	Asp	Phe	Phe	Ala	Ser	Leu	Lys	Pro	Thr	Thr	His	Glu	Ala	

EP 2 441 831 B9

	555	560	565	
5	agc aaa gag ttg gat gga tat ctg gcc tgt gtt tca gac acc agg gag Ser Lys Glu Leu Asp Gly Tyr Leu Ala Cys Val Ser Asp Thr Arg Glu 570 575 580 585	1839		
10	tct ctg ctc acg ttt cct gct att tgc agc ctc tct atc aag act aat Ser Leu Leu Thr Phe Pro Ala Ile Cys Ser Leu Ser Ile Lys Thr Asn 590 595 600	1887		
15	aca cct ctt ccc gca tcg gct gcc tgt gag agg ctt ttc agc act gca Thr Pro Leu Pro Ala Ser Ala Ala Cys Glu Arg Leu Phe Ser Thr Ala 605 610 615	1935		
20	gga ttg ctt ttc agc ccc aaa aga gct agg ctt gac act aac aat ttt Gly Leu Leu Phe Ser Pro Lys Arg Ala Arg Leu Asp Thr Asn Asn Phe 620 625 630	1983		
25	gag aat cag ctt cta ctg aag tta aat ctg agg ttt tac aac ttt gag Glu Asn Gln Leu Leu Leu Lys Leu Asn Leu Arg Phe Tyr Asn Phe Glu 635 640 645	2031		
30	tag cgtgtactgg cattagattg tctgtcttat agtttgataa ttaaatacaa	2084		
35	acagttctaa agcaggataa aaccttgtat gcatttcatt taatgttttt tgagattaaa	2144		
40	agcttaaaca ag	2156		
45	<210> 5			
50	<211> 649			
55	<212> PRT			
	<213> Oryzias latipes			
	<400> 5			

EP 2 441 831 B9

	Met	Glu	Glu	Val	Cys	Asp	Ser	Ser	Ala	Ala	Ala	Ser	Ser	Thr	Val	Gln
	1				5					10					15	
5	Asn	Gln	Pro	Gln	Asp	Gln	Glu	His	Pro	Trp	Pro	Tyr	Leu	Arg	Glu	Phe
				20					25					30		
10	Phe	Ser	Leu	Ser	Gly	Val	Asn	Lys	Asp	Ser	Phe	Lys	Met	Lys	Cys	Val
			35					40					45			
15	Leu	Cys	Leu	Pro	Leu	Asn	Lys	Glu	Ile	Ser	Ala	Phe	Lys	Ser	Ser	Pro
		50					55					60				
20	Ser	Asn	Leu	Arg	Lys	His	Ile	Glu	Arg	Met	His	Pro	Asn	Tyr	Leu	Lys
	65					70					75					80
25	Asn	Tyr	Ser	Lys	Leu	Thr	Ala	Gln	Lys	Arg	Lys	Ile	Gly	Thr	Ser	Thr
					85					90					95	
30	His	Ala	Ser	Ser	Ser	Lys	Gln	Leu	Lys	Val	Asp	Ser	Val	Phe	Pro	Val
				100					105					110		
35	Lys	His	Val	Ser	Pro	Val	Thr	Val	Asn	Lys	Ala	Ile	Leu	Arg	Tyr	Ile
40																
45																
50																
55																

EP 2 441 831 B9

	115					120					125					
5	Ile	Gln	Gly	Leu	His	Pro	Phe	Ser	Thr	Val	Asp	Leu	Pro	Ser	Phe	Lys
	130						135					140				
	Glu	Leu	Ile	Ser	Thr	Leu	Gln	Pro	Gly	Ile	Ser	Val	Ile	Thr	Arg	Pro
	145					150					155					160
10	Thr	Leu	Arg	Ser	Lys	Ile	Ala	Glu	Ala	Ala	Leu	Ile	Met	Lys	Gln	Lys
					165					170					175	
	Val	Thr	Ala	Ala	Met	Ser	Glu	Val	Glu	Trp	Ile	Ala	Thr	Thr	Thr	Asp
15				180					185					190		
	Cys	Trp	Thr	Ala	Arg	Arg	Lys	Ser	Phe	Ile	Gly	Val	Thr	Ala	His	Trp
			195					200					205			
20	Ile	Asn	Pro	Gly	Ser	Leu	Glu	Arg	His	Ser	Ala	Ala	Leu	Ala	Cys	Lys
	210						215					220				
	Arg	Leu	Met	Gly	Ser	His	Thr	Phe	Glu	Val	Leu	Ala	Ser	Ala	Met	Asn
25	225					230					235					240
	Asp	Ile	His	Ser	Glu	Tyr	Glu	Ile	Arg	Asp	Lys	Val	Val	Cys	Thr	Thr
					245					250					255	
30	Thr	Asp	Ser	Gly	Ser	Asn	Phe	Met	Lys	Ala	Phe	Arg	Val	Phe	Gly	Val
				260					265					270		
	Glu	Asn	Asn	Asp	Ile	Glu	Thr	Glu	Ala	Arg	Arg	Cys	Glu	Ser	Asp	Asp
35			275					280					285			
	Thr	Asp	Ser	Glu	Gly	Cys	Gly	Glu	Gly	Ser	Asp	Gly	Val	Glu	Phe	Gln
	290						295					300				
40	Asp	Ala	Ser	Arg	Val	Leu	Asp	Gln	Asp	Asp	Gly	Phe	Glu	Phe	Gln	Leu
	305					310					315					320
	Pro	Lys	His	Gln	Lys	Cys	Ala	Cys	His	Leu	Leu	Asn	Leu	Val	Ser	Ser
					325					330					335	
	Val	Asp	Ala	Gln	Lys	Ala	Leu	Ser	Asn	Glu	His	Tyr	Lys	Lys	Leu	Tyr
50				340					345					350		
	Arg	Ser	Val	Phe	Gly	Lys	Cys	Gln	Ala	Leu	Trp	Asn	Lys	Ser	Ser	Arg
			355					360					365			
	Ser	Ala	Leu	Ala	Ala	Glu	Ala	Val	Glu	Ser	Glu	Ser	Arg	Leu	Gln	Leu
55							375					380				

EP 2 441 831 B9

	Leu	Arg	Pro	Asn	Gln	Thr	Arg	Trp	Asn	Ser	Thr	Phe	Met	Ala	Val	Asp	
	385					390					395					400	
5	Arg	Ile	Leu	Gln	Ile	Cys	Lys	Glu	Ala	Gly	Glu	Gly	Ala	Leu	Arg	Asn	
				405						410					415		
	Ile	Cys	Thr	Ser	Leu	Glu	Val	Pro	Met	Phe	Asn	Pro	Ala	Glu	Met	Leu	
10				420					425					430			
	Phe	Leu	Thr	Glu	Trp	Ala	Asn	Thr	Met	Arg	Pro	Val	Ala	Lys	Val	Leu	
			435					440					445				
15	Asp	Ile	Leu	Gln	Ala	Glu	Thr	Asn	Thr	Gln	Leu	Gly	Trp	Leu	Leu	Pro	
	450						455					460					
	Ser	Val	His	Gln	Leu	Ser	Leu	Lys	Leu	Gln	Arg	Leu	His	His	Ser	Leu	
20	465					470					475					480	
	Arg	Tyr	Cys	Asp	Pro	Leu	Val	Asp	Ala	Leu	Gln	Gln	Gly	Ile	Gln	Thr	
				485						490					495		
25	Arg	Phe	Lys	His	Met	Phe	Glu	Asp	Pro	Glu	Ile	Ile	Ala	Ala	Ala	Ile	
				500					505					510			
	Leu	Leu	Pro	Lys	Phe	Arg	Thr	Ser	Trp	Thr	Asn	Asp	Glu	Thr	Ile	Ile	
30			515					520					525				
	Lys	Arg	Gly	Met	Asp	Tyr	Ile	Arg	Val	His	Leu	Glu	Pro	Leu	Asp	His	
35		530					535					540					
	Lys	Lys	Glu	Leu	Ala	Asn	Ser	Ser	Ser	Asp	Asp	Glu	Asp	Phe	Phe	Ala	
	545					550					555					560	
40	Ser	Leu	Lys	Pro	Thr	Thr	His	Glu	Ala	Ser	Lys	Glu	Leu	Asp	Gly	Tyr	
				565						570					575		
	Leu	Ala	Cys	Val	Ser	Asp	Thr	Arg	Glu	Ser	Leu	Leu	Thr	Phe	Pro	Ala	
45				580					585					590			
	Ile	Cys	Ser	Leu	Ser	Ile	Lys	Thr	Asn	Thr	Pro	Leu	Pro	Ala	Ser	Ala	
50			595					600					605				
	Ala	Cys	Glu	Arg	Leu	Phe	Ser	Thr	Ala	Gly	Leu	Leu	Phe	Ser	Pro	Lys	
		610					615					620					
55	Arg	Ala	Arg	Leu	Asp	Thr	Asn	Asn	Phe	Glu	Asn	Gln	Leu	Leu	Leu	Lys	
	625					630					635					640	

EP 2 441 831 B9

Leu Asn Leu Arg Phe Tyr Asn Phe Glu
645

5 <210> 6
<211> 4682
<212> DNA
<213> Oryzias latipes

10 <400> 6

15

20

25

30

35

40

45

50

55

EP 2 441 831 B9

	cagaggtgta aagtacttga gtaattttac ttgattactg tacttaagta ttatttttgg	60
	ggattttttac tttacttgag tacaattaaa aatcaatact tttactttta cttaattaca	120
5	tttttttaga aaaaaaagta cttttttactc cttacaattt tatttacagt caaaaagtac	180
	ttatttttttg gagatcactt cattctattt tcccttgcta ttaccaaacc aattgaattg	240
	cgctgatgcc cagtttaatt taaatgttat ttattctgcc tatgaaaatc gttttcacat	300
10	tatatgaaat tggtcagaca tgttcattgg tcctttggaa gtgacgtcat gtcacatcta	360
	ttaccacaat gcacagcacc ttgacctgga aattagggaa attataacag tcaatcagtg	420
	gaagaaaatg gaggaagtat gtgattcatc agcagctgcg agcagcacag tccaaaatca	480
15	gccacaggat caagagcacc cgtggccgta tcttcgcgaa ttcttttctt taagtgggtg	540
	aaataaagat tcattcaaga tgaaatgtgt cctctgtctc ccgcttaata aagaaatatc	600
20	ggccttcaaa agttcgccat caaacctaag gaagcatatt gaggtaagta cattaagtat	660
	tttgttttac tgatagtttt tttttttttt tttttttttt tttttgggtg tgcatgtttt	720
	gacgttgatg gcgcgccttt tatatgtgta gtaggcctat tttcactaat gcatgcgatt	780
25	gacaatataa ggctcacgta ataaaatgct aaaatgcatt tgtaattggg aacgttaggt	840
	ccacgggaaa tttggcgcct attgcagctt tgaataatca ttatcattcc gtgctctcat	900
30	tgtgtttgaa ttcatgcaaa acacaagaaa accaagcgag aaattttttt ccaaacatgt	960
	tgtattgtca aaacggtaac actttacaat gaggttgatt agttcatgta ttaactaaca	1020
	ttaaataacc atgagcaata catttggttac tgtatctgtt aatctttgtt aacgttaggt	1080
35	aatagaaata cagatgttca ttgtttgttc atgttagttc acagtgcatt aactaatgtt	1140
	aacaagatat aaagtattag taaatgttga aattaacatg tatacgtgca gttcattatt	1200
	agttcatgtt aactaatgta gttaactaac gaaccttatt gtaaaagtgt taccatcaaa	1260
40	actaatgtaa tgaaatcaat tcaccctgtc atgtcagcct tacagtcctg tgtttttgtc	1320
	aatataatca gaaataaaat taatgtttga ttgtcactaa atgctactgt atttctaaaa	1380
45	tcaacaagta tttaacatta taaagtgtgc aattggctgc aaatgtcagt tttattaaag	1440
	ggttagttca cccaaaaatg aaaataatgt cattaatgac tcgccctcat gtcgttccaa	1500
	gcccgtaaga cctccgttca tcttcagaac acagtttaag atattttaga tttagtccga	1560
50	gagctttctg tgcctccatt gagaatgtat gtacgggtata ctgtccatgt ccagaaaggt	1620

55

EP 2 441 831 B9

	aataaaaaaca	tcaaagtagt	ccatgtgaca	tcagtgggtt	agttagaatt	ttttgaagca	1680
	tcgaatacat	tttgggccaa	aaataacaaa	acctacgact	ttattcggca	ttgtattctc	1740
5	ttccgggtct	gttgtcaatc	cgcgttcacg	acttcgcagt	gacgctacaa	tgctgaataa	1800
	agtcgtaggt	tttgttattt	ttggacccaa	atgtattttc	gatgcttcaa	ataattctac	1860
	ctaaccctact	gatgtcacat	ggactacttt	gatgttttta	ttacctttct	ggacatggac	1920
10	agtataccgt	acatacat	tcagtggagg	gacagaaagc	tctcggacta	aatctaaaat	1980
	atcttaaaact	gtgttccgaa	gatgaacgga	ggtgttacgg	gcttggaacg	acatgagggg	2040
	gagtcattaa	tgacatcttt	tcatttttgg	gtgaactaac	cctttaatgc	tgtaatcaga	2100
15	gagtgtatgt	gtaattgtta	cattttattgc	atacaatata	aatatttatt	tggtgttttt	2160
	acagagaatg	cacccaaatt	acctcaaaaa	ctactctaaa	ttgacagcac	agaagagaaa	2220
	gatcgggacc	tccacccatg	cttcacgacg	taagcaactg	aaagttgact	cagttttccc	2280
20	agtcaaacat	gtgtctccag	tcactgtgaa	caaagctata	ttaaggtaca	tcattcaagg	2340
	acttcatcct	ttcagcactg	ttgatctgcc	atcattttaa	gagctgatta	gtacactgca	2400
	gcctggcatt	tctgtcatta	caaggcctac	tttacgctcc	aagatagctg	aagctgctct	2460
25	gatcatgaaa	cagaaagtga	ctgctgccat	gagtgaagtt	gaatggattg	caaccacaac	2520
	ggattgttgg	actgcacgta	gaaagtcatt	cattggtgta	actgctcact	ggatcaaccc	2580
	tggaagtctt	gaaagacatt	ccgctgcact	tgcttgcaaa	agattaatgg	gctctcatac	2640
30	ttttgaggta	ctggccagtg	ccatgaatga	tatccactca	gagtatgaaa	tacgtgacaa	2700
	ggttgtttgc	acaaccacag	acagtgggtc	caactttatg	aaggctttca	gagtttttgg	2760
	tgtggaaaac	aatgatatcg	agactgaggc	aagaaggtgt	gaaagtgatg	acactgattc	2820
35	tgaaggctgt	ggtgagggaa	gtgatggtgt	ggaattccaa	gatgcctcac	gagtcctgga	2880
	ccaagacgat	ggcttcgaat	tccagctacc	aaaacatcaa	aagtgtgcct	gtcacttact	2940
	taacctagtc	tcaagcgttg	atgccccaaa	agctctctca	aatgaacact	acaagaaact	3000
40	ctacagatct	gtctttggca	aatgccaaagc	tttatggaat	aaaagcagcc	gatcggctct	3060
	agcagctgaa	gctgttgaat	cagaaagccg	gcttcagctt	ttaaggccaa	accaaacgcg	3120
	gtggaattca	acttttatgg	ctgttgacag	aattcttcaa	atttgcaaag	aagcaggaga	3180
45	aggcgcactt	cggaatatat	gcacctctct	tgaggttcca	atgtaagtgt	ttttcccctc	3240
	tatcgatgta	aacaaatgtg	ggttgttttt	gtttaatact	ctttgattat	gctgatttct	3300
	cctgtagggt	taatccagca	gaaatgctgt	tcttgacaga	gtgggccaac	acaatgcgtc	3360
50	cagttgcaaa	agtactcgac	atcttgcaag	cggaaacgaa	tacacagctg	gggtggctgc	3420
	tgcttagtgt	ccatcagtta	agcttgaaac	ttcagcgact	ccaccattct	ctcaggtact	3480
55	gtgaccctact	tgtggatgcc	ctacaacaag	gaatccaaac	acgattcaag	catatgtttg	3540
	aagatcctga	gatcatagca	gctgccatcc	ttctccctaa	atttcggacc	tcttggacaa	3600

EP 2 441 831 B9

	atgatgaaac catcataaaa cgaggtaa	aat gaatgcaagc aacatacact tgacgaattc	3660
	taatctgggc aacctttgag ccataccaaa	attattcttt tatttattta tttttgcact	3720
5	tttttaggaat gttatatccc atctttggct	gtgatctcaa tatgaatatt gatgtaaagt	3780
	attcttgcag caggttgtag ttatccctca	gtgtttcttg aaaccaaact catatgtatc	3840
	atatgtgggt tggaaatgca gttagatttt	atgctaaaat aagggatttg catgatttta	3900
10	gatgtagatg actgcacgta aatgtagtta	atgacaaaat ccataaaatt tgttcccagt	3960
	cagaagcccc tcaaccaa	ac ttttctttgt gtctgtctcac tgtgcttgta	ggcatggact 4020
	acatcagagt gcatctggag cctttggacc	acaagaagga attggccaac agttcatctg	4080
15	atgatgaaga ttttttcgct tctttgaaac	cgacaacaca tgaagccagc aaagagttgg	4140
	atggatatct ggcctgtgtt tcagacacca	gggagtctct gctcacgttt cctgctat	ttt 4200
20	gcagcctctc tatcaagact aatacacctc	ttcccgcatc ggctgcctgt gagaggcttt	4260
	tcagcactgc aggattgctt ttcagcccca	aaagagctag gcttgacact aacaattttg	4320
	agaatcagct tctactgaag ttaa	aatctga ggttttacia ctttgagtag	cgtgtactgg 4380
25	cattagattg tctgtcttat agtttgataa	ttaaatacaa acagttctaa agcaggataa	4440
	aaccttgtat gcatttcatt taatgtttt	tgagattaaa agcttaaaca aga	atctcta 4500
30	gttttctttc ttgcttttac ttttacttcc	ttaatactca agtacaattt taatggagta	4560
	cttttttact tttactcaag taagattcta	gccagatact tttactttta attgagtaaa	4620
	attttcccta agtacttgta ctttcacttg	agtaaaattt ttgagtactt tttacacctc	4680
35	tg		4682

<210> 7

<211> 321

<212> DNA

40 <213> Artificial

<220>

<223> Description of artificial sequence; Cycloheximide resistant gene

45 <220>

<221> CDS

<222> (1)..(321)

<400> 7

50

55

EP 2 441 831 B9

	atg gtc aac gta cct aaa acc cga aga acc ttc tgt aag aag tgt ggc	48
	Met Val Asn Val Pro Lys Thr Arg Arg Thr Phe Cys Lys Lys Cys Gly	
	1 5 10 15	
5	aag cat cag cct cac aaa gtg aca cag tat aag aag ggc aag gat tct	96
	Lys His Gln Pro His Lys Val Thr Gln Tyr Lys Lys Gly Lys Asp Ser	
	20 25 30	
10	ttg tat gcc cag gga agg agg cgc tat gat cgg aag cag agt ggc tat	144
	Leu Tyr Ala Gln Gly Arg Arg Arg Tyr Asp Arg Lys Gln Ser Gly Tyr	
	35 40 45	
15	ggt ggg cag aca aag caa att ttc cgg aag aag gct aag acc aca aag	192
	Gly Gly Gln Thr Lys Gln Ile Phe Arg Lys Lys Ala Lys Thr Thr Lys	
	50 55 60	
20	aag att gtg cta agg ctg gaa tgt gtt gag cct aac tgc aga tcc aag	240
	Lys Ile Val Leu Arg Leu Glu Cys Val Glu Pro Asn Cys Arg Ser Lys	
	65 70 75 80	
25	agg atg ctg gcc att aag aga tgc aag cat ttt gaa ctg gga gga gat	288
	Arg Met Leu Ala Ile Lys Arg Cys Lys His Phe Glu Leu Gly Gly Asp	
	85 90 95	
30	aag aag aga aag ggc caa gtg atc cag ttc taa	321
	Lys Lys Arg Lys Gly Gln Val Ile Gln Phe	
	100 105	
35	<210> 8	
	<211> 106	
	<212> PRT	
	<213> Artificial	
40	<220>	
	<223> Synthetic Construct	
45	<400> 8	
50		
55		

EP 2 441 831 B9

	Met	Val	Asn	Val	Pro	Lys	Thr	Arg	Arg	Thr	Phe	Cys	Lys	Lys	Cys	Gly		
	1				5					10					15			
5		Lys	His	Gln	Pro	His	Lys	Val	Thr	Gln	Tyr	Lys	Lys	Gly	Lys	Asp	Ser	
				20						25					30			
	Leu	Tyr	Ala	Gln	Gly	Arg	Arg	Arg	Tyr	Asp	Arg	Lys	Gln	Ser	Gly	Tyr		
10			35					40					45					
	Gly	Gly	Gln	Thr	Lys	Gln	Ile	Phe	Arg	Lys	Lys	Ala	Lys	Thr	Thr	Lys		
		50					55					60						
15		Lys	Ile	Val	Leu	Arg	Leu	Glu	Cys	Val	Glu	Pro	Asn	Cys	Arg	Ser	Lys	
		65					70					75					80	
	Arg	Met	Leu	Ala	Ile	Lys	Arg	Cys	Lys	His	Phe	Glu	Leu	Gly	Gly	Asp		
20					85					90					95			
	Lys	Lys	Arg	Lys	Gly	Gln	Val	Ile	Gln	Phe								
25				100					105									

<210> 9

<211> 1404

<212> DNA

30 <213> Artificial

<220>

<223> M2Z3 Heavy chain

35 <220>

<221> CDS

<222> (1)..(1404)

<400> 9

40

45

50

55

EP 2 441 831 B9

	atg gac tgg acc tgg agc atc ctt ttc ttg gtg gca gca gca aca ggt	48
	Met Asp Trp Thr Trp Ser Ile Leu Phe Leu Val Ala Ala Ala Thr Gly	
	1 5 10 15	
5	gcc cac tcc cag gtt cag ctg gtg cag tct gga gct gag gtg aag aag	96
	Ala His Ser Gln Val Gln Leu Val Gln Ser Gly Ala Glu Val Lys Lys	
	20 25 30	
10	cct ggg gcc tca gtg aag gtc tcc tgc aag gct tct ggt tac acc ttt	144
	Pro Gly Ala Ser Val Lys Val Ser Cys Lys Ala Ser Gly Tyr Thr Phe	
	35 40 45	
15	acc agc tat ggt atc agc tgg gtg cga cag gcc cct gga caa ggg ctt	192
	Thr Ser Tyr Gly Ile Ser Trp Val Arg Gln Ala Pro Gly Gln Gly Leu	
	50 55 60	
	gag tgg atg gga tgg atc agc gct tac aat ggt aac aca aac tat gca	240
	Glu Trp Met Gly Trp Ile Ser Ala Tyr Asn Gly Asn Thr Asn Tyr Ala	
	65 70 75 80	
20	cag aag ctc cag ggc aga gtc acc atg acc aca gac aca tcc acg agc	288
	Gln Lys Leu Gln Gly Arg Val Thr Met Thr Thr Asp Thr Ser Thr Ser	
	85 90 95	
25	aca gcc tac atg gag ctg agg agc ctg aga tct gac gac acg gcc gtg	336
	Thr Ala Tyr Met Glu Leu Arg Ser Leu Arg Ser Asp Asp Thr Ala Val	
	100 105 110	
30	tat tac tgt gcg agg gca gca gct ggc gga tac ttc cag cac tgg ggc	384
	Tyr Tyr Cys Ala Arg Ala Ala Gly Gly Tyr Phe Gln His Trp Gly	
	115 120 125	
	cag ggc acc ctg gtc acc gtc tcc tca gct agc acc aag ggc cca tcg	432
	Gln Gly Thr Leu Val Thr Val Ser Ser Ala Ser Thr Lys Gly Pro Ser	
	130 135 140	
35	gtc ttc ccc ctg gca ccc tcc tcc aag agc acc tct ggg ggc aca gcg	480
	Val Phe Pro Leu Ala Pro Ser Ser Lys Ser Thr Ser Gly Gly Thr Ala	
	145 150 155 160	
40	gcc ctg ggc tgc ctg gtc aag gac tac ttc ccc gaa ccg gtg acg gtg	528
	Ala Leu Gly Cys Leu Val Lys Asp Tyr Phe Pro Glu Pro Val Thr Val	
	165 170 175	
45	tcg tgg aac tca ggc gcc ctg acc agc ggc gtg cac acc ttc ccg gct	576
	Ser Trp Asn Ser Gly Ala Leu Thr Ser Gly Val His Thr Phe Pro Ala	
	180 185 190	
	gtc cta cag tcc tca gga ctc tac tcc ctc agc agc gtg gtg acc gtg	624
	Val Leu Gln Ser Ser Gly Leu Tyr Ser Leu Ser Ser Val Val Thr Val	
	195 200 205	
50	ccc tcc agc agc ttg ggc acc cag acc tac atc tgc aac gtg aat cac	672
	Pro Ser Ser Ser Leu Gly Thr Gln Thr Tyr Ile Cys Asn Val Asn His	
	210 215 220	
55	aag ccc agc aac acc aag gtg gac aag aaa gtt gag ccc aaa tct tgt	720
	Lys Pro Ser Asn Thr Lys Val Asp Lys Lys Val Glu Pro Lys Ser Cys	
	225 230 235 240	

EP 2 441 831 B9

		gac	aaa	act	cac	aca	tgc	cca	ccg	tgc	cca	gca	cct	gaa	ctc	ctg	ggg	768
		Asp	Lys	Thr	His	Thr	Cys	Pro	Pro	Cys	Pro	Ala	Pro	Glu	Leu	Leu	Gly	
						245					250					255		
5		gga	ccg	tca	gtc	ttc	ctc	ttc	ccc	cca	aaa	ccc	aag	gac	acc	ctc	atg	816
		Gly	Pro	Ser	Val	Phe	Leu	Phe	Pro	Pro	Lys	Pro	Lys	Asp	Thr	Leu	Met	
					260					265					270			
10		atc	tcc	cgg	acc	cct	gag	gtc	aca	tgc	gtg	gtg	gtg	gac	gtg	agc	cac	864
		Ile	Ser	Arg	Thr	Pro	Glu	Val	Thr	Cys	Val	Val	Val	Asp	Val	Ser	His	
				275					280					285				
15		gaa	gac	cct	gag	gtc	aag	ttc	aac	tgg	tac	gtg	gac	ggc	gtg	gag	gtg	912
		Glu	Asp	Pro	Glu	Val	Lys	Phe	Asn	Trp	Tyr	Val	Asp	Gly	Val	Glu	Val	
			290					295					300					
20		cat	aat	gcc	aag	aca	aag	ccg	cgg	gag	gag	cag	tac	aac	agc	acg	tac	960
		His	Asn	Ala	Lys	Thr	Lys	Pro	Arg	Glu	Glu	Gln	Tyr	Asn	Ser	Thr	Tyr	
		305					310					315					320	
25		cgt	gtg	gtc	agc	gtc	ctc	acc	gtc	ctg	cac	cag	gac	tgg	ctg	aat	ggc	1008
		Arg	Val	Val	Ser	Val	Leu	Thr	Val	Leu	His	Gln	Asp	Trp	Leu	Asn	Gly	
						325					330					335		
30		aag	gag	tac	aag	tgc	aag	gtc	tcc	aac	aaa	gcc	ctc	cca	gcc	ccc	atc	1056
		Lys	Glu	Tyr	Lys	Cys	Lys	Val	Ser	Asn	Lys	Ala	Leu	Pro	Ala	Pro	Ile	
					340				345						350			
35		gag	aaa	acc	atc	tcc	aaa	gcc	aaa	ggg	cag	ccc	cga	gaa	cca	cag	gtg	1104
		Glu	Lys	Thr	Ile	Ser	Lys	Ala	Lys	Gly	Gln	Pro	Arg	Glu	Pro	Gln	Val	
				355				360						365				
40		tac	acc	ctg	ccc	cca	tcc	cgg	gat	gag	ctg	acc	aag	aac	cag	gtc	agc	1152
		Tyr	Thr	Leu	Pro	Pro	Ser	Arg	Asp	Glu	Leu	Thr	Lys	Asn	Gln	Val	Ser	
			370					375					380					
45		ctg	acc	tgc	ctg	gtc	aaa	ggc	ttc	tat	ccc	agc	gac	atc	gcc	gtg	gag	1200
		Leu	Thr	Cys	Leu	Val	Lys	Gly	Phe	Tyr	Pro	Ser	Asp	Ile	Ala	Val	Glu	
						385		390				395					400	
50		tgg	gag	agc	aat	ggg	cag	ccg	gag	aac	aac	tac	aag	acc	acg	cct	ccc	1248
		Trp	Glu	Ser	Asn	Gly	Gln	Pro	Glu	Asn	Asn	Tyr	Lys	Thr	Thr	Pro	Pro	
					405					410						415		
55		gtg	ctg	gac	tcc	gac	ggc	tcc	ttc	ttc	ctc	tac	agc	aag	ctc	acc	gtg	1296
		Val	Leu	Asp	Ser	Asp	Gly	Ser	Phe	Phe	Leu	Tyr	Ser	Lys	Leu	Thr	Val	
					420					425					430			
60		gac	aag	agc	agg	tgg	cag	cag	ggg	aac	gtc	ttc	tca	tgc	tcc	gtg	atg	1344
		Asp	Lys	Ser	Arg	Trp	Gln	Gln	Gly	Asn	Val	Phe	Ser	Cys	Ser	Val	Met	
				435					440					445				
65		cat	gag	gct	ctg	cac	aac	cac	tac	acg	cag	aag	agc	ctc	tcc	ctg	tct	1392
		His	Glu	Ala	Leu	His	Asn	His	Tyr	Thr	Gln	Lys	Ser	Leu	Ser	Leu	Ser	
				450				455					460					
70		ccg	ggt	aaa	tga													1404
		Pro	Gly	Lys														
				465														

<210> 10

<211> 467
<212> PRT
<213> Artificial

5 <220>
 <223> Synthetic Construct

<400> 10

10

15

20

25

30

35

40

45

50

55

EP 2 441 831 B9

	Met	Asp	Trp	Thr	Trp	Ser	Ile	Leu	Phe	Leu	Val	Ala	Ala	Ala	Thr	Gly	
	1				5					10					15		
5	Ala	His	Ser	Gln	Val	Gln	Leu	Val	Gln	Ser	Gly	Ala	Glu	Val	Lys	Lys	
				20					25					30			
10	Pro	Gly	Ala	Ser	Val	Lys	Val	Ser	Cys	Lys	Ala	Ser	Gly	Tyr	Thr	Phe	
			35					40					45				
15	Thr	Ser	Tyr	Gly	Ile	Ser	Trp	Val	Arg	Gln	Ala	Pro	Gly	Gln	Gly	Leu	
		50					55					60					
20	Glu	Trp	Met	Gly	Trp	Ile	Ser	Ala	Tyr	Asn	Gly	Asn	Thr	Asn	Tyr	Ala	
	65					70					75					80	
25	Gln	Lys	Leu	Gln	Gly	Arg	Val	Thr	Met	Thr	Thr	Asp	Thr	Ser	Thr	Ser	
					85					90					95		
30	Thr	Ala	Tyr	Met	Glu	Leu	Arg	Ser	Leu	Arg	Ser	Asp	Asp	Thr	Ala	Val	
				100					105					110			
35	Tyr	Tyr	Cys	Ala	Arg	Ala	Ala	Ala	Gly	Gly	Tyr	Phe	Gln	His	Trp	Gly	
			115					120					125				
40	Gln	Gly	Thr	Leu	Val	Thr	Val	Ser	Ser	Ala	Ser	Thr	Lys	Gly	Pro	Ser	
		130					135					140					
45	Val	Phe	Pro	Leu	Ala	Pro	Ser	Ser	Lys	Ser	Thr	Ser	Gly	Gly	Thr	Ala	
	145					150					155					160	
50	Ala	Leu	Gly	Cys	Leu	Val	Lys	Asp	Tyr	Phe	Pro	Glu	Pro	Val	Thr	Val	
					165					170					175		
55	Ser	Trp	Asn	Ser	Gly	Ala	Leu	Thr	Ser	Gly	Val	His	Thr	Phe	Pro	Ala	
				180					185					190			
60	Val	Leu	Gln	Ser	Ser	Gly	Leu	Tyr	Ser	Leu	Ser	Ser	Val	Val	Thr	Val	
			195				200						205				
65	Pro	Ser	Ser	Ser	Leu	Gly	Thr	Gln	Thr	Tyr	Ile	Cys	Asn	Val	Asn	His	
		210					215					220					
70	Lys	Pro	Ser	Asn	Thr	Lys	Val	Asp	Lys	Lys	Val	Glu	Pro	Lys	Ser	Cys	
	225					230					235					240	

EP 2 441 831 B9

	Asp	Lys	Thr	His	Thr	Cys	Pro	Pro	Cys	Pro	Ala	Pro	Glu	Leu	Leu	Gly	
					245					250					255		
5	Gly	Pro	Ser	Val	Phe	Leu	Phe	Pro	Pro	Lys	Pro	Lys	Asp	Thr	Leu	Met	
				260						265				270			
10	Ile	Ser	Arg	Thr	Pro	Glu	Val	Thr	Cys	Val	Val	Val	Asp	Val	Ser	His	
			275					280					285				
15	Glu	Asp	Pro	Glu	Val	Lys	Phe	Asn	Trp	Tyr	Val	Asp	Gly	Val	Glu	Val	
		290					295					300					
20	His	Asn	Ala	Lys	Thr	Lys	Pro	Arg	Glu	Glu	Gln	Tyr	Asn	Ser	Thr	Tyr	
	305					310					315					320	
25	Arg	Val	Val	Ser	Val	Leu	Thr	Val	Leu	His	Gln	Asp	Trp	Leu	Asn	Gly	
					325					330					335		
30	Lys	Glu	Tyr	Lys	Cys	Lys	Val	Ser	Asn	Lys	Ala	Leu	Pro	Ala	Pro	Ile	
				340					345					350			
35	Glu	Lys	Thr	Ile	Ser	Lys	Ala	Lys	Gly	Gln	Pro	Arg	Glu	Pro	Gln	Val	
			355					360					365				
40	Tyr	Thr	Leu	Pro	Pro	Ser	Arg	Asp	Glu	Leu	Thr	Lys	Asn	Gln	Val	Ser	
		370					375					380					
45	Leu	Thr	Cys	Leu	Val	Lys	Gly	Phe	Tyr	Pro	Ser	Asp	Ile	Ala	Val	Glu	
	385					390					395					400	
50	Trp	Glu	Ser	Asn	Gly	Gln	Pro	Glu	Asn	Asn	Tyr	Lys	Thr	Thr	Pro	Pro	
				405						410					415		
55	Val	Leu	Asp	Ser	Asp	Gly	Ser	Phe	Phe	Leu	Tyr	Ser	Lys	Leu	Thr	Val	
				420					425					430			
60	Asp	Lys	Ser	Arg	Trp	Gln	Gln	Gly	Asn	Val	Phe	Ser	Cys	Ser	Val	Met	
			435					440					445				
65	His	Glu	Ala	Leu	His	Asn	His	Tyr	Thr	Gln	Lys	Ser	Leu	Ser	Leu	Ser	
		450					455					460					
70	Pro	Gly	Lys														
	465																

<210> 11
<211> 708

<212> DNA
<213> Artificial

<220>
<223> M2Z3 Light chian

<220>
<221> CDS
<222> (1)..(708)

<400> 11

EP 2 441 831 B9

	atg gcc agc ttc cct ctc ctc ctc acc ctc ctc act cac tgt gca ggg	48
	Met Ala Ser Phe Pro Leu Leu Leu Thr Leu Leu Thr His Cys Ala Gly	
	1 5 10 15	
5	tcc tgg gcc cag tct gtg ctg act cag cca ccc tca gcg tct ggg acc	96
	Ser Trp Ala Gln Ser Val Leu Thr Gln Pro Pro Ser Ala Ser Gly Thr	
	20 25 30	
10	ccc ggg cag agg gtc acc atc tct tgt tct gga agc aac tcc aac atc	144
	Pro Gly Gln Arg Val Thr Ile Ser Cys Ser Gly Ser Asn Ser Asn Ile	
	35 40 45	
15	gga agt aaa act gta aac tgg tac cag cag ctc cca gga acg gcc ccc	192
	Gly Ser Lys Thr Val Asn Trp Tyr Gln Gln Leu Pro Gly Thr Ala Pro	
	50 55 60	
	aaa ctc ctc atc tct agt aat aat cag cgg ccc tca ggg gtc cct gac	240
	Lys Leu Leu Ile Ser Ser Asn Asn Gln Arg Pro Ser Gly Val Pro Asp	
	65 70 75 80	
20	cga ttc tct ggc tcc aag tct ggc acc tca gcc tcc ctg gcc atc agt	288
	Arg Phe Ser Gly Ser Lys Ser Gly Thr Ser Ala Ser Leu Ala Ile Ser	
	85 90 95	
25	ggg ctc cag tct gag gat gag gct gat tat tac tgt gca gca tgg gat	336
	Gly Leu Gln Ser Glu Asp Glu Ala Asp Tyr Tyr Cys Ala Ala Trp Asp	
	100 105 110	
	gac agc ctg aat ggt gtg gta ttc ggc gga ggg acc aag ctg acc gtc	384
	Asp Ser Leu Asn Gly Val Val Phe Gly Gly Gly Thr Lys Leu Thr Val	
	115 120 125	
30	cta ggt cag ccc aag gct gcc ccc tcg gtc act ctg ttc cca ccc tcc	432
	Leu Gly Gln Pro Lys Ala Ala Pro Ser Val Thr Leu Phe Pro Pro Ser	
	130 135 140	
35	tct gag gag ctt caa gcc aac aag gcc aca ctg gtg tgt ctc ata agt	480
	Ser Glu Glu Leu Gln Ala Asn Lys Ala Thr Leu Val Cys Leu Ile Ser	
	145 150 155 160	
40	gac ttc tac ccg gga gcc gtg aca gtg gcc tgg aag gca gat agc agc	528
	Asp Phe Tyr Pro Gly Ala Val Thr Val Ala Trp Lys Ala Asp Ser Ser	
	165 170 175	
	ccc gtc aag gcg gga gtg gag acc acc aca ccc tcc aaa caa agc aac	576
	Pro Val Lys Ala Gly Val Glu Thr Thr Pro Ser Lys Gln Ser Asn	
	180 185 190	
45	aac aag tac gcg gcc agc agc tac ctg agc ctg acg cct gag cag tgg	624
	Asn Lys Tyr Ala Ala Ser Ser Tyr Leu Ser Leu Thr Pro Glu Gln Trp	
	195 200 205	
50	aag tcc cac aaa agc tac agc tgc cag gtc acg cat gaa ggg agc acc	672
	Lys Ser His Lys Ser Tyr Ser Cys Gln Val Thr His Glu Gly Ser Thr	
	210 215 220	
55	gtg gag aag aca gtg gcc cct aca gaa tgt tca tag	708
	Val Glu Lys Thr Val Ala Pro Thr Glu Cys Ser	
	225 230 235	

<210> 12

EP 2 441 831 B9

<211> 235
<212> PRT
<213> Artificial

5 <220>
<223> Synthetic Construct

<400> 12

10	Met	Ala	Ser	Phe	Pro	Leu	Leu	Leu	Thr	Leu	Leu	Thr	His	Cys	Ala	Gly
	1				5					10					15	
15	Ser	Trp	Ala	Gln	Ser	Val	Leu	Thr	Gln	Pro	Pro	Ser	Ala	Ser	Gly	Thr
				20					25					30		
20	Pro	Gly	Gln	Arg	Val	Thr	Ile	Ser	Cys	Ser	Gly	Ser	Asn	Ser	Asn	Ile
			35					40					45			
25	Gly	Ser	Lys	Thr	Val	Asn	Trp	Tyr	Gln	Gln	Leu	Pro	Gly	Thr	Ala	Pro
	50						55					60				
30	Lys	Leu	Leu	Ile	Ser	Ser	Asn	Asn	Gln	Arg	Pro	Ser	Gly	Val	Pro	Asp
	65					70					75					80
35	Arg	Phe	Ser	Gly	Ser	Lys	Ser	Gly	Thr	Ser	Ala	Ser	Leu	Ala	Ile	Ser
					85					90					95	
40	Gly	Leu	Gln	Ser	Glu	Asp	Glu	Ala	Asp	Tyr	Tyr	Cys	Ala	Ala	Trp	Asp
				100					105					110		
45	Asp	Ser	Leu	Asn	Gly	Val	Val	Phe	Gly	Gly	Gly	Thr	Lys	Leu	Thr	Val
			115					120					125			
50	Leu	Gly	Gln	Pro	Lys	Ala	Ala	Pro	Ser	Val	Thr	Leu	Phe	Pro	Pro	Ser
		130					135					140				
55	Ser	Glu	Glu	Leu	Gln	Ala	Asn	Lys	Ala	Thr	Leu	Val	Cys	Leu	Ile	Ser
	145					150					155					160
	Asp	Phe	Tyr	Pro	Gly	Ala	Val	Thr	Val	Ala	Trp	Lys	Ala	Asp	Ser	Ser
					165					170					175	
	Pro	Val	Lys	Ala	Gly	Val	Glu	Thr	Thr	Thr	Pro	Ser	Lys	Gln	Ser	Asn
				180					185					190		
	Asn	Lys	Tyr	Ala	Ala	Ser	Ser	Tyr	Leu	Ser	Leu	Thr	Pro	Glu	Gln	Trp

EP 2 441 831 B9

195

200

205

5

Lys Ser His Lys Ser Tyr Ser Cys Gln Val Thr His Glu Gly Ser Thr
210 215 220

10

Val Glu Lys Thr Val Ala Pro Thr Glu Cys Ser
225 230 235

<210> 13

<211> 1855

<212> DNA

<213> artificial

15

<220>

<223> nonautologus Tol1 transposon

20

<400> 13

25

30

35

40

45

50

55

EP 2 441 831 B9

	cagtagcggg	tctaggcacg	ggccgtccgg	gcggtggcct	ggggcgga	actgaagggg	60
	ggcggcaccg	gcggctcagc	cctttgta	atattaatat	gcaccactat	tggtttactt	120
5	atgtcacagt	ttgtaagttt	gtaacagcct	gaacctggcc	gcgccgccgc	cctcgccccg	180
	cagctgcgct	ctcctgtctt	tgagaagtag	acacaaatgt	gtgtgaagaa	ggagaaggga	240
10	gggggcgcgg	ggtgagcacg	gagcgtcgcc	gcgtttgcgc	atgcgcaaaa	cctggctggc	300
	tcatctttca	ggggagggca	cggtcgcggg	cttgatgaaa	aaaataaaag	taaaaactgc	360
	gactgcgccg	tcatgtagcg	aatcagcgcc	cctggctgta	gctgcacgcg	ctcctgctgg	420
15	aaatgtgtga	agaggggggg	gggggggggg	gctgcgggga	atcagttcaa	ttgtgggacg	480
	cttccaaatt	aagtggctag	gtggggacaa	gggcgggggt	ttgaatctac	ttcataaaac	540
	ctttttatat	tataagtcag	tcataagggtg	acattctata	acctacattt	taataaaggt	600
20	ataaaaaata	tattctgctt	tttttgggtt	aattttgtgt	gaaatgtcca	aataaaaaaa	660
	atggcaacac	aaaacaatgc	tgtcactaag	gtgacagttg	gttcagtcga	cggacttgat	720
25	gccttcttcg	tgacgtgagg	acatttatgc	caaacaaacg	ccaataaaca	tctaaaatat	780
	ggaaaagaaa	aggtcaaagc	catctgggtc	ccaatttaga	aagaaaagaa	aagaagaaga	840
	ggagaaaaga	gataaagaaa	agggtaaagtc	ctcacagctt	gatgcatggt	ttttctaaat	900
30	tctaatagcta	cctgccctac	aacaacgttg	ccgatgaaaa	ctttattttg	gtcgatgacc	960
	aacactgaat	taggcccaca	tgttgcaaat	agcgtcattt	tttttttttt	tttttagattt	1020
	tattcttaaa	aatttgctct	gccttaactt	gtaacattag	ttatgattca	tgtgtctgtc	1080
35	tgctctgctg	taacacaaaag	gttttggttg	gttttgctgt	tgtataactag	ctcataatgt	1140
	taaaaaagct	gtgatgggtta	cacagcatgc	tggtgctgcc	ataagatgct	aatggggcaa	1200
40	ataatttgag	attgggtcatt	aatttaataa	tcatttggtg	cagcctaaac	gttttcacaa	1260
	tgtttttttg	acattttaact	ggggatttag	gggttaattt	tgagcctgca	tatgaagttt	1320
	attttttatt	tgttttacaa	atgtgggatt	atatttttag	ccaatagaat	ttccataaat	1380

EP 2 441 831 B9

ctgtaggtag ttttaaaaat gaatattttac cattttactgc aactctatgg ggacaaaaca 1440
 taatgtaaca ggtcataact aaaaatgtgc caatcaaagg attgaagacg gaaaacatga 1500
 5 gttaatTTTT cttctctgaa gtagagatcg atatagaaca tgacaattta aatttccaat 1560
 tcataaatgt ttttaaaaata tttattttat attattttatt taacattgag tttgattcaa 1620
 10 tattttctta gctaactgta tttttgccat gcttatggtc ttttattttt tgtgttctga 1680
 taacttttat aatgcttttc agaattttga catcttttgt atccacttct taatttcaat 1740
 gacaataaaa catttcagtt gacgaagaca aacaaagttc tgttgtgact atgggggggg 1800
 15 ggggcgccctg gggatggtct cgcccgggga gtaattcagg gtagaaccgc cactg 1855

<210> 14
 <211> 200
 <212> DNA
 20 <213> artificial
 <220>
 <223> Tol1-L transposon sequence
 25 <400> 14

cagtagcggg tctaggcacg ggccgtccgg gcggtggcct ggggcggaaa actgaagggg 60
 30 ggcggcacccg gcggctcagc cctttgtaat atattaatat gcaccactat tggtttactt 120
 atgtcacagt ttgtaagttt gtaacagcct gaacctggcc gcgccgccgc cctcgccccg 180
 cagctgcgct ctcctgtctt 200

35 <210> 15
 <211> 505
 <212> DNA
 40 <213> artificial
 <220>
 <223> Tol1-R transoposon sequence
 45 <400> 15

50

55

EP 2 441 831 B9

	atatttttag ccaatagaat ttccataaat ctgtaggtag ttttaaaaat gaatatttac	60
	catttactgc aactctatgg ggacaaaaca taatgtaaca ggtcataact aaaaatgtgc	120
5	caatcaaagg attgaagacg gaaaacatga gttaattttt cttctctgaa gtagagatcg	180
	atatagaaca tgacaattta aattttccaat tcataaatgt ttttaaaata tttattttat	240
10	attatttatt taacattgag ttgattcaa tattttctta gctaactgta tttttgccat	300
	gcttatggtc ttttattttt tgtgttctga taacttttat aatgcttttc agaattttga	360
	catcttttgt atccacttct taattttcaat gacaataaaa catttcagtt gacgaagaca	420
15	aacaaagttc tgttgtgact atgggggggg ggggcgcctg gggatggtct cgcccgggga	480
	gtaattcagg gtagaaccgc cactg	505

20
 <210> 16
 <211> 2745
 <212> DNA
 <213> Oryzias latipes

25
 <220>
 <221> CDS
 <222> (30)..(2585)

30
 <400> 16

35

40

45

50

55

EP 2 441 831 B9

	gccaaacaaa	cgccaaaaaac	atctaaaaat	atg	gag	aaa	aaa	agg	tca	aag	cca		53				
				Met	Glu	Lys	Lys	Arg	Ser	Lys	Pro						
				1				5									
5	tct	ggt	gcc	caa	ttt	aga	aag	aaa	aga	aaa	gaa	gaa	gag	gag	aaa	aga	101
	Ser	Gly	Ala	Gln	Phe	Arg	Lys	Lys	Arg	Lys	Glu	Glu	Glu	Glu	Lys	Arg	
	10						15				20						
10	gat	aaa	gaa	aag	ggg	gca	ctt	cta	aga	tat	ttt	gga	tcg	tct	acc	act	149
	Asp	Lys	Glu	Lys	Gly	Ala	Leu	Leu	Arg	Tyr	Phe	Gly	Ser	Ser	Thr	Thr	
	25					30					35					40	
15	gct	caa	gat	gag	aca	tct	acc	tcc	ctg	cca	gct	atc	tca	tca	gcc	aca	197
	Ala	Gln	Asp	Glu	Thr	Ser	Thr	Ser	Leu	Pro	Ala	Ile	Ser	Ser	Ala	Thr	
					45					50					55		
20	gtc	aca	gtc	tca	ccc	cct	cag	gat	gag	cta	cca	tct	aca	tcc	tct	gct	245
	Val	Thr	Val	Ser	Pro	Pro	Gln	Asp	Glu	Leu	Pro	Ser	Thr	Ser	Ser	Ala	
				60					65					70			
25	act	cat	gta	gtt	cca	cag	ttg	tta	cct	gag	caa	agt	ttt	gat	agt	gag	293
	Thr	His	Val	Val	Pro	Gln	Leu	Leu	Pro	Glu	Gln	Ser	Phe	Asp	Ser	Glu	
			75				80						85				
30	gct	gaa	gac	gtt	gtt	cca	tct	acg	tct	acc	cag	ctt	gag	act	tca	gaa	341
	Ala	Glu	Asp	Val	Val	Pro	Ser	Thr	Ser	Thr	Gln	Leu	Glu	Thr	Ser	Glu	
		90					95					100					
35	atg	cct	ggt	gat	gaa	acc	cca	ctg	acc	ccg	act	gct	gag	gac	cag	cct	389
	Met	Pro	Gly	Asp	Glu	Thr	Pro	Leu	Thr	Pro	Thr	Ala	Glu	Asp	Gln	Pro	
	105					110					115					120	
40	cta	cca	act	gac	cct	gca	aag	tgg	ccc	tca	cct	ctg	act	gac	agg	ata	437
	Leu	Pro	Thr	Asp	Pro	Ala	Lys	Trp	Pro	Ser	Pro	Leu	Thr	Asp	Arg	Ile	
					125					130					135		
45	cgg	atg	gag	ctg	gtt	cga	aga	gga	cca	agt	agc	ata	cca	cct	gac	ttt	485
	Arg	Met	Glu	Leu	Val	Arg	Arg	Gly	Pro	Ser	Ser	Ile	Pro	Pro	Asp	Phe	
				140					145					150			
50	gtt	ttc	cca	aga	aat	gac	agt	gat	ggg	aga	agt	tgt	cat	cac	cac	tat	533
	Val	Phe	Pro	Arg	Asn	Asp	Ser	Asp	Gly	Arg	Ser	Cys	His	His	His	Tyr	
			155					160					165				
55	ttc	agg	aag	aca	cta	gta	agt	ggt	gaa	aaa	ata	gca	aga	act	tgg	ttg	581
	Phe	Arg	Lys	Thr	Leu	Val	Ser	Gly	Glu	Lys	Ile	Ala	Arg	Thr	Trp	Leu	
		170					175					180					
60	atg	tat	tca	aaa	gtg	aag	aac	agc	ctc	ttt	tgc	ttt	tgt	tgc	aaa	ttg	629
	Met	Tyr	Ser	Lys	Val	Lys	Asn	Ser	Leu	Phe	Cys	Phe	Cys	Cys	Lys	Leu	
		185				190					195					200	
65	ttt	tcc	aac	aaa	aac	att	aat	tta	aca	act	tct	ggt	aca	gca	aac	tgg	677
	Phe	Ser	Asn	Lys	Asn	Ile	Asn	Leu	Thr	Thr	Ser	Gly	Thr	Ala	Asn	Trp	
				205						210					215		

EP 2 441 831 B9

	aaa cat gca agc aca tac ctc aca gca cac gaa aaa agc cca gaa cac	725
	Lys His Ala Ser Thr Tyr Leu Thr Ala His Glu Lys Ser Pro Glu His	
	220 225 230	
5	ctc aat tgt atg aaa gca tgg aag gaa ctg tca ggg agg atc aga agt	773
	Leu Asn Cys Met Lys Ala Trp Lys Glu Leu Ser Gly Arg Ile Arg Ser	
	235 240 245	
10	ggg aaa aca att gat aag cag gag atg gca ctt ctg gaa gag gag cgg	821
	Gly Lys Thr Ile Asp Lys Gln Glu Met Ala Leu Leu Glu Glu Glu Arg	
	250 255 260	
15	gtg aga tgg aga gca gtg cta acc cgt ctc att gct att gtg cag tca	869
	Val Arg Trp Arg Ala Val Leu Thr Arg Leu Ile Ala Ile Val Gln Ser	
	265 270 275 280	
20	ctg gca gtt cgg aat ttg gct cta agg gga cac aca gaa aca ctg ttc	917
	Leu Ala Val Arg Asn Leu Ala Leu Arg Gly His Thr Glu Thr Leu Phe	
	285 290 295	
25	aca tca tca aat ggg aat ttt ttg aaa gag gtt gaa ctg atg gcc agg	965
	Thr Ser Ser Asn Gly Asn Phe Leu Lys Glu Val Glu Leu Met Ala Arg	
	300 305 310	
30	ttt gat ccc ata atg aaa gat cat ctt aac cgt gta tta aga gga aca	1013
	Phe Asp Pro Ile Met Lys Asp His Leu Asn Arg Val Leu Arg Gly Thr	
	315 320 325	
35	gca agt cac aac agc tac ata ggc cat cat gtg cag aat gaa ctt att	1061
	Ala Ser His Asn Ser Tyr Ile Gly His His Val Gln Asn Glu Leu Ile	
	330 335 340	
40	gat ttg ttg agc agc aaa atc cta tcc gct ata gtg gat gac atc aaa	1109
	Asp Leu Leu Ser Ser Lys Ile Leu Ser Ala Ile Val Asp Asp Ile Lys	
	345 350 355 360	
45	aag gca aaa tat ttt tca ata att ctg gac tgc act ctg gat ata agc	1157
	Lys Ala Lys Tyr Phe Ser Ile Ile Leu Asp Cys Thr Leu Asp Ile Ser	
	365 370 375	
50	cac aca gaa cag ttg tca gtt ata att aga gtg gtg tca ctg atg gag	1205
	His Thr Glu Gln Leu Ser Val Ile Ile Arg Val Val Ser Leu Met Glu	
	380 385 390	
55	aag cct cag atc agg gaa cat ttt atg ggg ttt ttg gag gca gag gag	1253
	Lys Pro Gln Ile Arg Glu His Phe Met Gly Phe Leu Glu Ala Glu Glu	
	395 400 405	
60	tcc aca ggc cag cac ttg gca tcc atg atc tta aac aga ctt gag gag	1301
	Ser Thr Gly Gln His Leu Ala Ser Met Ile Leu Asn Arg Leu Glu Glu	
	410 415 420	
65	tta gga att tct ttt gaa gac tgc aga gga caa tca tat gat aat ggg	1349
	Leu Gly Ile Ser Phe Glu Asp Cys Arg Gly Gln Ser Tyr Asp Asn Gly	
	425 430 435 440	
70	gca aat atg aaa ggc aaa aat aag gga gta caa gcc agg ctc tta gaa	1397
	Ala Asn Met Lys Gly Lys Asn Lys Gly Val Gln Ala Arg Leu Leu Glu	
	445 450 455	
75	aag aat ccc cgt gct ctg ttt ttg cca tgc ggt gca cac aca ttg aat	1445
	Lys Asn Pro Arg Ala Leu Phe Leu Pro Cys Gly Ala His Thr Leu Asn	
	460 465 470	

EP 2 441 831 B9

	tta gtt gtg tgt gat gct gct aag aga tct gtt gat gct atg agc tac	1493
	Leu Val Val Cys Asp Ala Ala Lys Arg Ser Val Asp Ala Met Ser Tyr	
	475 480 485	
5	ttt ggt gtc ctg caa aag ctt tac act tta ttt tca gcc tct gcc caa	1541
	Phe Gly Val Leu Gln Lys Leu Tyr Thr Leu Phe Ser Ala Ser Ala Gln	
	490 495 500	
10	cga tgg gcc ata ctg aag agt cag gtg agc atc act cta aag tcg tgg	1589
	Arg Trp Ala Ile Leu Lys Ser Gln Val Ser Ile Thr Leu Lys Ser Trp	
	505 510 515 520	
	aca gaa aca agg tgg gag agc aaa atc aaa agc atc gag ccc atg agg	1637
	Thr Glu Thr Arg Trp Glu Ser Lys Ile Lys Ser Ile Glu Pro Met Arg	
	525 530 535	
15	tac cag gga gct gca gtg aga gag gct tta ata gaa gtg aga gac aag	1685
	Tyr Gln Gly Ala Ala Val Arg Glu Ala Leu Ile Glu Val Arg Asp Lys	
	540 545 550	
20	acc aaa gac cca gtt ata aag gct gag gcc cag tct ttg tct gaa gag	1733
	Thr Lys Asp Pro Val Ile Lys Ala Glu Ala Gln Ser Leu Ser Glu Glu	
	555 560 565	
	gta ggg tcg tac cgc ttc aac atc tgc aca gtc gta tgg cat gac att	1781
	Val Gly Ser Tyr Arg Phe Asn Ile Cys Thr Val Val Trp His Asp Ile	
	570 575 580	
25	cta tct aca ata aag cat gtc agc aaa ctc atg cag tct cca aat atg	1829
	Leu Ser Thr Ile Lys His Val Ser Lys Leu Met Gln Ser Pro Asn Met	
	585 590 595 600	
30	cat gtg gac cta gct gtg agt ctt ttg aag aag act gaa caa agt ctc	1877
	His Val Asp Leu Ala Val Ser Leu Leu Lys Lys Thr Glu Gln Ser Leu	
	605 610 615	
	cag agc tac agg gca aat ggc ttt gtg aat gca cag atg gca gcc aaa	1925
	Gln Ser Tyr Arg Ala Asn Gly Phe Val Asn Ala Gln Met Ala Ala Lys	
	620 625 630	
35	gaa atg tgc aag gaa atg aat gtc gag gct att ttg aaa caa aaa aga	1973
	Glu Met Cys Lys Glu Met Asn Val Glu Ala Ile Leu Lys Gln Lys Arg	
	635 640 645	
40	ata aga tcc aca aag tgc caa ttc tcg tat gaa tca cac gat gag cct	2021
	Ile Arg Ser Thr Lys Cys Gln Phe Ser Tyr Glu Ser His Asp Glu Pro	
	650 655 660	
45	ttc agt gac gca ctt aaa aag ttg gag gtt gaa ttt ttc aat gtt gtt	2069
	Phe Ser Asp Ala Leu Lys Lys Leu Glu Val Glu Phe Phe Asn Val Val	
	665 670 675 680	
	gtt gat gaa gcc ttg tca gcc atc gcg gag agg ttt tcc aca ttg gaa	2117
	Val Asp Glu Ala Leu Ser Ala Ile Ala Glu Arg Phe Ser Thr Leu Glu	
	685 690 695	
50	gtt gta caa aac aga ttt ggg gtt ttg acc aat ttc cca agc ctt gga	2165
	Val Val Gln Asn Arg Phe Gly Val Leu Thr Asn Phe Pro Ser Leu Gly	
	700 705 710	
55	gac gag gag ctg acg gag caa tgc gag gca cta ggc aac ata ctc cat	2213
	Asp Glu Glu Leu Thr Glu Gln Cys Glu Ala Leu Gly Asn Ile Leu His	
	715 720 725	
	ttt gag aag aac tgg gat ttg gac agt aga gag ctt gtt cag gaa atc	2261

EP 2 441 831 B9

	Phe	Glu	Lys	Asn	Trp	Asp	Leu	Asp	Ser	Arg	Glu	Leu	Val	Gln	Glu	Ile	
	730						735					740					
5	aag	aac	ttg	cct	aac	tta	cca	tca	acg	act	cca	agt	ctc	ctt	gag	ctc	2309
	Lys	Asn	Leu	Pro	Asn	Leu	Pro	Ser	Thr	Thr	Pro	Ser	Leu	Leu	Glu	Leu	
	745					750					755				760		
	atc	tct	ttc	atg	tct	gat	aag	gat	cta	tca	gaa	atc	tat	ccg	aac	ttt	2357
10	Ile	Ser	Phe	Met	Ser	Asp	Lys	Asp	Leu	Ser	Glu	Ile	Tyr	Pro	Asn	Phe	
					765					770					775		
	tgg	act	gct	ctc	agg	att	gca	ctc	acc	ttg	cca	gtc	act	gtg	gct	caa	2405
	Trp	Thr	Ala	Leu	Arg	Ile	Ala	Leu	Thr	Leu	Pro	Val	Thr	Val	Ala	Gln	
					780					785					790		
15	gca	gag	agg	agc	ttt	tca	aaa	cta	aaa	ttg	atc	aag	tcg	tac	ctg	agg	2453
	Ala	Glu	Arg	Ser	Phe	Ser	Lys	Leu	Lys	Leu	Ile	Lys	Ser	Tyr	Leu	Arg	
			795					800					805				
	tca	aca	atg	tca	cag	gag	cga	ctc	act	aac	ctt	gcc	ggt	ggt	agc	atc	2501
20	Ser	Thr	Met	Ser	Gln	Glu	Arg	Leu	Thr	Asn	Leu	Ala	Val	Val	Ser	Ile	
		810					815					820					
	aat	cac	tca	gta	ggg	gag	cag	ata	tca	tat	gat	gat	ggt	att	gac	gag	2549
25	Asn	His	Ser	Val	Gly	Glu	Gln	Ile	Ser	Tyr	Asp	Asp	Val	Ile	Asp	Glu	
	825					830					835					840	
	ttt	gca	tca	aga	aag	gct	agg	aag	ggt	agg	ttt	tag	ttggtggttt				2595
	Phe	Ala	Ser	Arg	Lys	Ala	Arg	Lys	Val	Arg	Phe						
					845					850							
30	ctggttattgt	attggtgctg	cagttatatt	tatttttagcg	tgtcatttgt	gtgataaaag											2655
	gtttgtgctt	tataatatatt	atatttatatt	atttattcaa	tattgagttt	gattcaatat											2715
	tttcttagct	aactgtatatt	ttgccatgct														2745
35																	
	<210>	17															
	<211>	851															
	<212>	PRT															
	<213>	Oryzias latipes															
40																	
	<400>	17															
45																	
50																	
55																	

EP 2 441 831 B9

	Met	Glu	Lys	Lys	Arg	Ser	Lys	Pro	Ser	Gly	Ala	Gln	Phe	Arg	Lys	Lys	
	1				5					10					15		
5																	
	Arg	Lys	Glu	Glu	Glu	Glu	Lys	Arg	Asp	Lys	Glu	Lys	Gly	Ala	Leu	Leu	
				20					25					30			
10	Arg	Tyr	Phe	Gly	Ser	Ser	Thr	Thr	Ala	Gln	Asp	Glu	Thr	Ser	Thr	Ser	
			35					40					45				
	Leu	Pro	Ala	Ile	Ser	Ser	Ala	Thr	Val	Thr	Val	Ser	Pro	Pro	Gln	Asp	
		50					55					60					
15																	
	Glu	Leu	Pro	Ser	Thr	Ser	Ser	Ala	Thr	His	Val	Val	Pro	Gln	Leu	Leu	
	65					70					75					80	
20																	
25																	
30																	
35																	
40																	
45																	
50																	
55																	

EP 2 441 831 B9

	Pro	Glu	Gln	Ser	Phe	Asp	Ser	Glu	Ala	Glu	Asp	Val	Val	Pro	Ser	Thr	
					85					90					95		
5	Ser	Thr	Gln	Leu	Glu	Thr	Ser	Glu	Met	Pro	Gly	Asp	Glu	Thr	Pro	Leu	
				100					105					110			
	Thr	Pro	Thr	Ala	Glu	Asp	Gln	Pro	Leu	Pro	Thr	Asp	Pro	Ala	Lys	Trp	
10			115					120					125				
	Pro	Ser	Pro	Leu	Thr	Asp	Arg	Ile	Arg	Met	Glu	Leu	Val	Arg	Arg	Gly	
		130					135					140					
15	Pro	Ser	Ser	Ile	Pro	Pro	Asp	Phe	Val	Phe	Pro	Arg	Asn	Asp	Ser	Asp	
	145					150					155					160	
	Gly	Arg	Ser	Cys	His	His	His	Tyr	Phe	Arg	Lys	Thr	Leu	Val	Ser	Gly	
20					165					170					175		
	Glu	Lys	Ile	Ala	Arg	Thr	Trp	Leu	Met	Tyr	Ser	Lys	Val	Lys	Asn	Ser	
				180					185					190			
25	Leu	Phe	Cys	Phe	Cys	Cys	Lys	Leu	Phe	Ser	Asn	Lys	Asn	Ile	Asn	Leu	
			195					200					205				
	Thr	Thr	Ser	Gly	Thr	Ala	Asn	Trp	Lys	His	Ala	Ser	Thr	Tyr	Leu	Thr	
30			210				215						220				
	Ala	His	Glu	Lys	Ser	Pro	Glu	His	Leu	Asn	Cys	Met	Lys	Ala	Trp	Lys	
	225					230					235					240	
35	Glu	Leu	Ser	Gly	Arg	Ile	Arg	Ser	Gly	Lys	Thr	Ile	Asp	Lys	Gln	Glu	
					245					250					255		
	Met	Ala	Leu	Leu	Glu	Glu	Glu	Arg	Val	Arg	Trp	Arg	Ala	Val	Leu	Thr	
40				260					265					270			
	Arg	Leu	Ile	Ala	Ile	Val	Gln	Ser	Leu	Ala	Val	Arg	Asn	Leu	Ala	Leu	
			275					280					285				
45	Arg	Gly	His	Thr	Glu	Thr	Leu	Phe	Thr	Ser	Ser	Asn	Gly	Asn	Phe	Leu	
		290					295					300					
	Lys	Glu	Val	Glu	Leu	Met	Ala	Arg	Phe	Asp	Pro	Ile	Met	Lys	Asp	His	
50	305					310					315					320	
	Leu	Asn	Arg	Val	Leu	Arg	Gly	Thr	Ala	Ser	His	Asn	Ser	Tyr	Ile	Gly	
				325						330					335		
55	His	His	Val	Gln	Asn	Glu	Leu	Ile	Asp	Leu	Leu	Ser	Ser	Lys	Ile	Leu	

EP 2 441 831 B9

	340		345		350
5	Ser Ala Ile Val Asp Asp Ile Lys Lys Ala Lys Tyr Phe Ser Ile Ile	355	360		365
10	Leu Asp Cys Thr Leu Asp Ile Ser His Thr Glu Gln Leu Ser Val Ile	370	375		380
15	Ile Arg Val Val Ser Leu Met Glu Lys Pro Gln Ile Arg Glu His Phe	385	390	395	400
20	Met Gly Phe Leu Glu Ala Glu Glu Ser Thr Gly Gln His Leu Ala Ser	405	410		415
25	Met Ile Leu Asn Arg Leu Glu Glu Leu Gly Ile Ser Phe Glu Asp Cys	420	425		430
30	Arg Gly Gln Ser Tyr Asp Asn Gly Ala Asn Met Lys Gly Lys Asn Lys	435	440		445
35	Gly Val Gln Ala Arg Leu Leu Glu Lys Asn Pro Arg Ala Leu Phe Leu	450	455	460	
40	Pro Cys Gly Ala His Thr Leu Asn Leu Val Val Cys Asp Ala Ala Lys	465	470	475	480
45	Arg Ser Val Asp Ala Met Ser Tyr Phe Gly Val Leu Gln Lys Leu Tyr	485	490		495
50	Thr Leu Phe Ser Ala Ser Ala Gln Arg Trp Ala Ile Leu Lys Ser Gln	500	505		510
55	Val Ser Ile Thr Leu Lys Ser Trp Thr Glu Thr Arg Trp Glu Ser Lys	515	520		525
	Ile Lys Ser Ile Glu Pro Met Arg Tyr Gln Gly Ala Ala Val Arg Glu	530	535	540	
	Ala Leu Ile Glu Val Arg Asp Lys Thr Lys Asp Pro Val Ile Lys Ala	545	550	555	560
	Glu Ala Gln Ser Leu Ser Glu Glu Val Gly Ser Tyr Arg Phe Asn Ile	565	570		575
	Cys Thr Val Val Trp His Asp Ile Leu Ser Thr Ile Lys His Val Ser	580	585		590
	Lys Leu Met Gln Ser Pro Asn Met His Val Asp Leu Ala Val Ser Leu	595	600	605	

EP 2 441 831 B9

	Leu	Lys	Lys	Thr	Glu	Gln	Ser	Leu	Gln	Ser	Tyr	Arg	Ala	Asn	Gly	Phe	
	610						615					620					
5	Val	Asn	Ala	Gln	Met	Ala	Ala	Lys	Glu	Met	Cys	Lys	Glu	Met	Asn	Val	
	625					630					635					640	
	Glu	Ala	Ile	Leu	Lys	Gln	Lys	Arg	Ile	Arg	Ser	Thr	Lys	Cys	Gln	Phe	
10					645					650					655		
	Ser	Tyr	Glu	Ser	His	Asp	Glu	Pro	Phe	Ser	Asp	Ala	Leu	Lys	Lys	Leu	
				660					665					670			
15	Glu	Val	Glu	Phe	Phe	Asn	Val	Val	Val	Asp	Glu	Ala	Leu	Ser	Ala	Ile	
			675					680					685				
	Ala	Glu	Arg	Phe	Ser	Thr	Leu	Glu	Val	Val	Gln	Asn	Arg	Phe	Gly	Val	
20		690					695					700					
	Leu	Thr	Asn	Phe	Pro	Ser	Leu	Gly	Asp	Glu	Glu	Leu	Thr	Glu	Gln	Cys	
25	705					710					715					720	
	Glu	Ala	Leu	Gly	Asn	Ile	Leu	His	Phe	Glu	Lys	Asn	Trp	Asp	Leu	Asp	
					725					730					735		
30	Ser	Arg	Glu	Leu	Val	Gln	Glu	Ile	Lys	Asn	Leu	Pro	Asn	Leu	Pro	Ser	
				740					745					750			
	Thr	Thr	Pro	Ser	Leu	Leu	Glu	Leu	Ile	Ser	Phe	Met	Ser	Asp	Lys	Asp	
35			755					760					765				
	Leu	Ser	Glu	Ile	Tyr	Pro	Asn	Phe	Trp	Thr	Ala	Leu	Arg	Ile	Ala	Leu	
	770						775					780					
40	Thr	Leu	Pro	Val	Thr	Val	Ala	Gln	Ala	Glu	Arg	Ser	Phe	Ser	Lys	Leu	
	785					790					795					800	
	Lys	Leu	Ile	Lys	Ser	Tyr	Leu	Arg	Ser	Thr	Met	Ser	Gln	Glu	Arg	Leu	
45					805					810					815		
	Thr	Asn	Leu	Ala	Val	Val	Ser	Ile	Asn	His	Ser	Val	Gly	Glu	Gln	Ile	
50				820					825					830			
	Ser	Tyr	Asp	Asp	Val	Ile	Asp	Glu	Phe	Ala	Ser	Arg	Lys	Ala	Arg	Lys	
			835					840					845				
55	Val	Arg	Phe														
	850																

Claims

1. A method for producing a protein of interest, comprising introducing a protein expression vector (a) which comprises a gene fragment comprising a DNA encoding a protein of interest and a selectable marker gene and, at both terminals of the gene fragment, a pair of transposon sequences which are the Tol1 nucleotide sequences shown in SEQ ID NO:14 and SEQ ID NO:15 or the Tol2 nucleotide sequences shown in SEQ ID NO:2 and SEQ ID NO:3, into a suspension CHO cell capable of surviving and proliferating in a serum-free medium; introducing an expression vector (b) which comprises a DNA encoding a transposase which recognizes the transposon sequences and has activity of transferring a gene fragment inserted between the transposon sequences into a chromosome into the CHO cell; integrating the gene fragment inserted between the transposon sequences into a chromosome of the CHO cell to obtain a said CHO cell capable of expressing the protein of interest; and suspension-culturing the CHO cell.
2. A method according to claim 1, comprising:
 - (A) simultaneously introducing the expression vectors (a) and (b) into the CHO cell,
 - (B) expressing transiently the transposase from the expression vector introduced in the step (A) to integrate the gene fragment inserted between the transposon sequences into a chromosome of the CHO cell to obtain a said suspension CHO cell capable of expressing the protein of interest, and
 - (C) suspension-culturing the suspension CHO cell capable of expressing the protein of interest obtained in the step (B) to produce the protein of interest.
3. A method for obtaining a suspension CHO cell capable of expressing a protein of interest, comprising introducing a protein expression vector which comprises a gene fragment comprising a DNA encoding a protein of interest and a selectable marker gene and, at both terminals of the gene fragment, a pair of transposon sequences which are the Tol1 nucleotide sequences shown in SEQ ID NO:14 and SEQ ID NO:15 or the Tol2 nucleotide sequences shown in SEQ ID NO:2 and SEQ ID NO:3, into a suspension CHO cell capable of surviving and proliferating in a serum-free medium; introducing an expression vector (b) which comprises a DNA encoding a transposase which recognizes the transposon sequences and has activity of transferring a gene fragment inserted between the transposon sequences into a chromosome into the CHO cell; and integrating the gene fragment inserted between the transposon sequences into a chromosome of the CHO cell.
4. The method according to any one of claims 1 to 3, wherein the CHO cell is at least one selected from CHO-K1, CHO-K1SV, DUKXB11, CHO/DG44, Pro-3 and CHO-S.
5. The method according to any one of the preceding claims, wherein the selectable marker gene is a cycloheximide resistance gene.
6. The method according to claim 5, wherein the cycloheximide resistance gene is a gene encoding a mutant of human ribosomal protein L36a.
7. The method according to claim 6, wherein the mutant is a mutant in which proline at position 54 of the human ribosomal protein L36a is substituted with another amino acid.
8. The method according to claim 7, wherein the other amino acid is glutamine.
9. A suspension CHO cell capable of surviving and proliferating in a serum-free medium and of producing a protein of interest, which cell comprises an expression vector (a) comprising a gene fragment comprising a DNA encoding a protein of interest and a selectable marker gene and, at both terminals of the gene fragment, a pair of transposon sequences which are the Tol1 nucleotide sequences shown in SEQ ID NO:14 and SEQ ID NO:15 or the Tol2 nucleotide sequences shown in SEQ ID NO:2 and SEQ ID NO:3, and an expression vector (b) comprising a DNA encoding a transposase (a transferase) which recognizes the transposon sequences and has activity of transferring the gene fragment inserted between the transposon sequences into a chromosome to integrate the gene fragment inserted between the transposon sequences into a chromosome of the CHO cell.
10. The cell according to claim 9, wherein the CHO cell is at least one selected from CHO-K1, CHO-K1SV, DUKXB11, CHO/DG44, Pro-3 and CHO-S.

11. The cell according to claim 9 or 10, wherein the selectable marker gene is a cycloheximide resistance gene.
12. The cell according to claim 11, wherein the cycloheximide resistance gene is a gene encoding a mutant of human ribosomal protein L36a.
13. The cell according to claim 12, wherein the mutant is a mutant in which proline at position 54 of the human ribosomal protein L36a is substituted with another amino acid.
14. The cell according to claim 13, wherein the other amino acid is glutamine.
15. Use of a protein expression vector (a) comprising a gene fragment comprising a DNA encoding a protein of interest and a selectable marker gene and, at both terminals of the gene fragment, a pair of transposon sequences which are the Tol1 nucleotide sequences shown in SEQ ID NO:14 and SEQ ID NO:15 or the Tol2 nucleotide sequences shown in SEQ ID NO:2 and SEQ ID NO:3 and an expression vector (b) comprising a DNA encoding a transposase which recognizes the transposon sequences and has activity of transferring a gene fragment inserted between the transposon sequences into a chromosome, to integrate the gene fragment inserted between the transposon sequences into a chromosome of a suspension CHO cell capable of surviving and proliferating in a serum-free medium.

Patentansprüche

1. Verfahren zum Herstellen eines Proteins von Interesse, umfassend Einführen eines Proteinexpressionsvektors (a), der ein Genfragment umfasst, das eine DNA umfasst, die ein Protein von Interesse und ein selektierbares Markergen und an beiden Endstellen des Genfragments ein Paar von Transposonsequenzen kodiert, die die Tol1-Nukleotidsequenzen gezeigt in SEQ ID NO:14 und SEQ ID NO:15 oder die Tol2-Nukleotidsequenzen gezeigt in SEQ ID NO:2 und SEQ ID NO:3 sind, in eine CHO-Suspensionszelle, die in der Lage ist, in einem serumfreien Medium zu überleben und sich zu vermehren; Einführen eines Expressionsvektors (b), umfassend eine DNA, die eine Transposase kodiert, die die Transposonsequenzen erkennt und Aktivität des Übertragens eines Genfragments hat, das zwischen den Transposonsequenzen in ein Chromosom in die CHO-Zelle eingesetzt ist; Integrieren des Genfragments, das zwischen den Transposonsequenzen in ein Chromosom der CHO-Zelle eingesetzt ist, um eine solche CHO-Zelle zu erhalten, die in der Lage ist, das Protein von Interesse zu exprimieren; und Suspensionskultivieren der CHO-Zelle.
2. Verfahren nach Anspruch 1, umfassend:
 - (A) simultanes Einführen der Expressionsvektoren (a) und (b) in die CHO-Zelle,
 - (B) vorübergehendes Exprimieren der Transposase aus dem Expressionsvektor, der in dem Schritt (A) eingeführt wird, um das Genfragment zu integrieren, das zwischen den Transposonsequenzen in ein Chromosom der CHO-Zelle eingesetzt ist, um eine solche CHO-Suspensionszelle zu erhalten, die in der Lage ist, das Protein von Interesse zu exprimieren, und
 - (C) Suspensionskultivieren der CHO-Suspensionszelle, die in der Lage ist, das Protein von Interesse, das in dem Schritt (B) erhalten wird, zu exprimieren, um das Protein von Interesse herzustellen.
3. Verfahren zum Erhalten einer CHO-Suspensionszelle, die in der Lage ist, ein Protein von Interesse zu exprimieren, umfassend Einführen eines Proteinexpressionsvektors, der ein Genfragment umfasst, das eine DNA umfasst, die ein Protein von Interesse und ein selektierbares Markergen und an beiden Endstellen des Genfragments ein Paar von Transposonsequenzen kodiert, die die Tol1-Nukleotidsequenzen gezeigt in SEQ ID NO:14 und SEQ ID NO:15 oder die Tol2-Nukleotidsequenzen gezeigt in SEQ ID NO:2 und SEQ ID NO:3 sind, in eine CHO-Suspensionszelle, die in der Lage ist, in einem serumfreien Medium zu überleben und sich zu vermehren; Einführen eines Expressionsvektors (b), umfassend eine DNA, die eine Transposase kodiert, die die Transposonsequenzen erkennt und Aktivität des Übertragens eines Genfragments hat, das zwischen den Transposonsequenzen in ein Chromosom in die CHO-Zelle eingesetzt ist; und Integrieren des Genfragments, das zwischen den Transposonsequenzen in ein Chromosom der CHO-Zelle eingesetzt ist.
4. Verfahren nach einem der Ansprüche 1 bis 3, wobei die CHO-Zelle mindestens eine ausgewählt aus CHO-K1, CHO-K1SV, DUKXB11, CHO/DG44, Pro-3 und CHO-S ist.
5. Verfahren nach einem der vorhergehenden Ansprüche, wobei das selektierbare Markergen ein Cycloheximid-Resistenzgen ist.

6. Verfahren nach Anspruch 5, wobei das Cycloheximid-Resistenzgen ein Gen ist, das eine Mutante von humanem ribosomalem Protein L36a ist.
7. Verfahren nach Anspruch 6, wobei die Mutante eine Mutante ist, in der Prolin an Position 54 des humanen ribosomalen Proteins L36a mit einer anderen Aminosäure substituiert ist.
8. Verfahren nach Anspruch 7, wobei die andere Aminosäure Glutamin ist.
9. CHO-Suspensionszelle, die in der Lage ist, in einem serumfreien Medium zu überleben und sich zu vermehren, und ein Protein von Interesse herzustellen, wobei die Zelle einen Expressionsvektor (a) umfasst, der ein Genfragment umfasst, das eine DNA umfasst, die ein Protein von Interesse und ein selektierbares Markergen und an beiden Endstellen des Genfragments ein Paar von Transposonsequenzen kodiert, die die Tol1-Nukleotidsequenzen gezeigt in SEQ ID NO:14 und SEQ ID NO:15 oder die Tol2-Nukleotidsequenzen gezeigt in SEQ ID NO:2 und SEQ ID NO:3 sind, und einen Expressionsvektor (b), umfassend eine DNA, die eine Transposase (eine Transferase) kodiert, die die Transposonsequenzen erkennt und Aktivität des Übertragens des Genfragments hat, das zwischen den Transposonsequenzen in ein Chromosom eingesetzt ist, um das Genfragment, das zwischen den Transposonsequenzen in ein Chromosom der CHO-Zelle eingesetzt ist, zu integrieren.
10. Zelle nach Anspruch 9, wobei die CHO-Zelle mindestens eine ausgewählt aus CHO-K1, CHO-K1SV, DUKXB11, CHO/DG44, Pro-3 und CHO-S ist.
11. Zelle nach Anspruch 9 oder 10, wobei das selektierbare Markergen ein Cycloheximid-Resistenzgen ist.
12. Zelle nach Anspruch 11, wobei das Cycloheximid-Resistenzgen ein Gen ist, das eine Mutante von humanem ribosomalem Protein L36a ist.
13. Zelle nach Anspruch 12, wobei die Mutante eine Mutante ist, in der Prolin an Position 54 des humanen ribosomalen Proteins L36a mit einer anderen Aminosäure substituiert ist.
14. Zelle nach Anspruch 13, wobei die andere Aminosäure Glutamin ist.
15. Verwendung eines Proteinexpressionsvektors (a), der ein Genfragment umfasst, das eine DNA umfasst, die ein Protein von Interesse und ein selektierbares Markergen und an beiden Endstellen des Genfragments ein Paar von Transposonsequenzen kodiert, die die Tol1-Nukleotidsequenzen gezeigt in SEQ ID NO:14 und SEQ ID NO:15 oder die Tol2-Nukleotidsequenzen gezeigt in SEQ ID NO:2 und SEQ ID NO:3 sind, und eines Expressionsvektors (b), umfassend eine DNA, die eine Transposase kodiert, die die Transposonsequenzen erkennt und Aktivität des Übertragens eines Genfragments hat, das zwischen den Transposonsequenzen in ein Chromosom eingesetzt ist, um das Genfragment zu integrieren, das zwischen den Transposonsequenzen in ein Chromosom einer CHO-Suspensionszelle eingesetzt ist, die in der Lage ist, in einem serumfreien Medium zu überleben und sich zu vermehren.

Revendications

1. Procédé pour produire une protéine d'intérêt, comprenant l'introduction d'un vecteur d'expression protéique (a) qui comprend un fragment de gène comprenant un ADN encodant une protéine d'intérêt et un gène marqueur sélectionnable et, aux deux extrémités du fragment de gène, une paire de séquences de transposons qui sont les séquences de nucléotides Tol1 représentées dans SEQ ID n° : 14 et SEQ ID n° : 15 ou les séquences de nucléotides Tol2 représentées dans SEQ ID n° : 2 et SEQ ID n° : 3, dans une cellule CHO en suspension capable de survivre et de se proliférer dans un milieu dépourvu de sérum ; l'introduction d'un vecteur d'expression (b) qui comprend un ADN encodant une transposase qui reconnaît les séquences de transposons et présente l'activité de transfert d'un fragment de gène inséré entre les séquences de transposons dans un chromosome dans la cellule CHO ; l'intégration du fragment de gène inséré entre les séquences de transposons dans un chromosome de la cellule CHO pour obtenir une dite cellule CHO capable d'exprimer la protéine d'intérêt ; et la culture en suspension de la cellule CHO.
2. Procédé selon la revendication 1, comprenant :
 - (A) l'introduction simultanée des vecteurs d'expression (a) et (b) dans la cellule CHO,
 - (B) l'expression transitoire de la transposase à partir du vecteur d'expression introduit dans l'étape (A) pour

intégrer le fragment de gène inséré entre les séquences de transposons dans un chromosome de la cellule CHO pour obtenir une dite cellule CHO en suspension capable d'exprimer la protéine d'intérêt, et (C) la culture en suspension de la cellule CHO en suspension capable d'exprimer la protéine d'intérêt obtenue dans l'étape (B) pour produire la protéine d'intérêt.

3. Procédé pour obtenir une cellule CHO en suspension capable d'exprimer une protéine d'intérêt, comprenant l'introduction d'un vecteur d'expression protéique qui comprend un fragment de gène comprenant un ADN encodant une protéine d'intérêt et un gène marqueur sélectionnable et, aux deux extrémités du fragment de gène, une paire de séquences de transposons qui sont les séquences de nucléotides Tol1 représentées dans SEQ ID n° : 14 et SEQ ID n° : 15 ou les séquences de nucléotides Tol2 représentées dans SEQ ID n° : 2 et SEQ ID n° : 3, dans une cellule CHO en suspension capable de survivre et de se proliférer dans un milieu dépourvu de sérum ; l'introduction d'un vecteur d'expression (b) qui comprend un ADN encodant une transposase qui reconnaît les séquences de transposons et présente l'activité de transfert d'un fragment de gène inséré entre les séquences de transposons dans un chromosome dans la cellule CHO ; et l'intégration du fragment de gène inséré entre les séquences de transposons dans un chromosome de la cellule CHO.
4. Procédé selon l'une quelconque des revendications 1 à 3, dans lequel la cellule CHO est au moins l'une sélectionnée parmi CHO-K1, CHO-K1SV, DUKXB11, CHO/DG44, Pro-3 et CHO-S.
5. Procédé selon l'une quelconque des revendications précédentes, dans lequel le gène marqueur sélectionnable est un gène de résistance au cycloheximide.
6. Procédé selon la revendication 5, dans lequel le gène de résistance au cycloheximide est un gène encodant un mutant de protéine ribosomale humaine L36a.
7. Procédé selon la revendication 6, dans lequel le mutant est un mutant dans lequel la proline à la position 54 de la protéine ribosomale humaine L36a est remplacée avec un autre acide aminé.
8. Procédé selon la revendication 7, dans lequel l'autre acide aminé est la glutamine.
9. Cellule CHO en suspension capable de survivre et de se proliférer dans un milieu dépourvu de sérum et de produire une protéine d'intérêt, laquelle cellule comprend un vecteur d'expression (a) comprenant un fragment de gène comprenant un ADN encodant une protéine d'intérêt et un gène marqueur sélectionnable et, aux deux extrémités du fragment de gène, une paire de séquences de transposons qui sont les séquences de nucléotides Tol1 représentées dans SEQ ID n° : 14 et SEQ ID n° : 15 ou les séquences de nucléotides Tol2 représentées dans SEQ ID n° : 2 et SEQ ID n° : 3, et un vecteur d'expression (b) comprenant un ADN encodant une transposase (une transférase) qui reconnaît les séquences de transposons et présente une activité de transfert du fragment de gène inséré entre les séquences de transposons dans un chromosome pour intégrer le fragment de gène inséré entre les séquences de transposons dans un chromosome de la cellule CHO.
10. Cellule selon la revendication 9, dans laquelle la cellule CHO est au moins l'une sélectionnée parmi CHO-K1, CHO-K1SV, DUKXB11, CHO/DG44, Pro-3 et CHO-S.
11. Cellule selon la revendication 9 ou 10, dans laquelle le gène marqueur sélectionnable est un gène de résistance au cycloheximide.
12. Cellule selon la revendication 11, dans laquelle le gène de résistance au cycloheximide est un gène encodant un mutant de protéine ribosomale humaine L36a.
13. Cellule selon la revendication 12, dans laquelle le mutant est un mutant dans lequel la proline à la position 54 de la protéine ribosomale humaine L36a est remplacée avec un autre acide aminé.
14. Cellule selon la revendication 13, dans laquelle l'autre acide aminé est la glutamine.
15. Utilisation d'un vecteur d'expression protéique (a) comprenant un fragment de gène comprenant un ADN encodant une protéine d'intérêt et un gène marqueur sélectionnable et, aux deux extrémités du fragment de gène, une paire de séquences de transposons qui sont les séquences de nucléotides Tol1 représentées dans SEQ ID n° : 14 et SEQ ID n° : 15 ou les séquences de nucléotides Tol2 représentées dans SEQ ID n° : 2 et SEQ ID n° : 3 et un vecteur

EP 2 441 831 B9

d'expression (b) comprenant un ADN encodant une transposase qui reconnaît les séquences de transposons et présente l'activité de transfert d'un fragment de gène inséré entre les séquences de transposons dans un chromosome, pour intégrer le fragment de gène inséré entre les séquences de transposons dans un chromosome d'une cellule CHO en suspension capable de survivre et de se proliférer dans un milieu dépourvu de sérum.

5

10

15

20

25

30

35

40

45

50

55

Fig.1

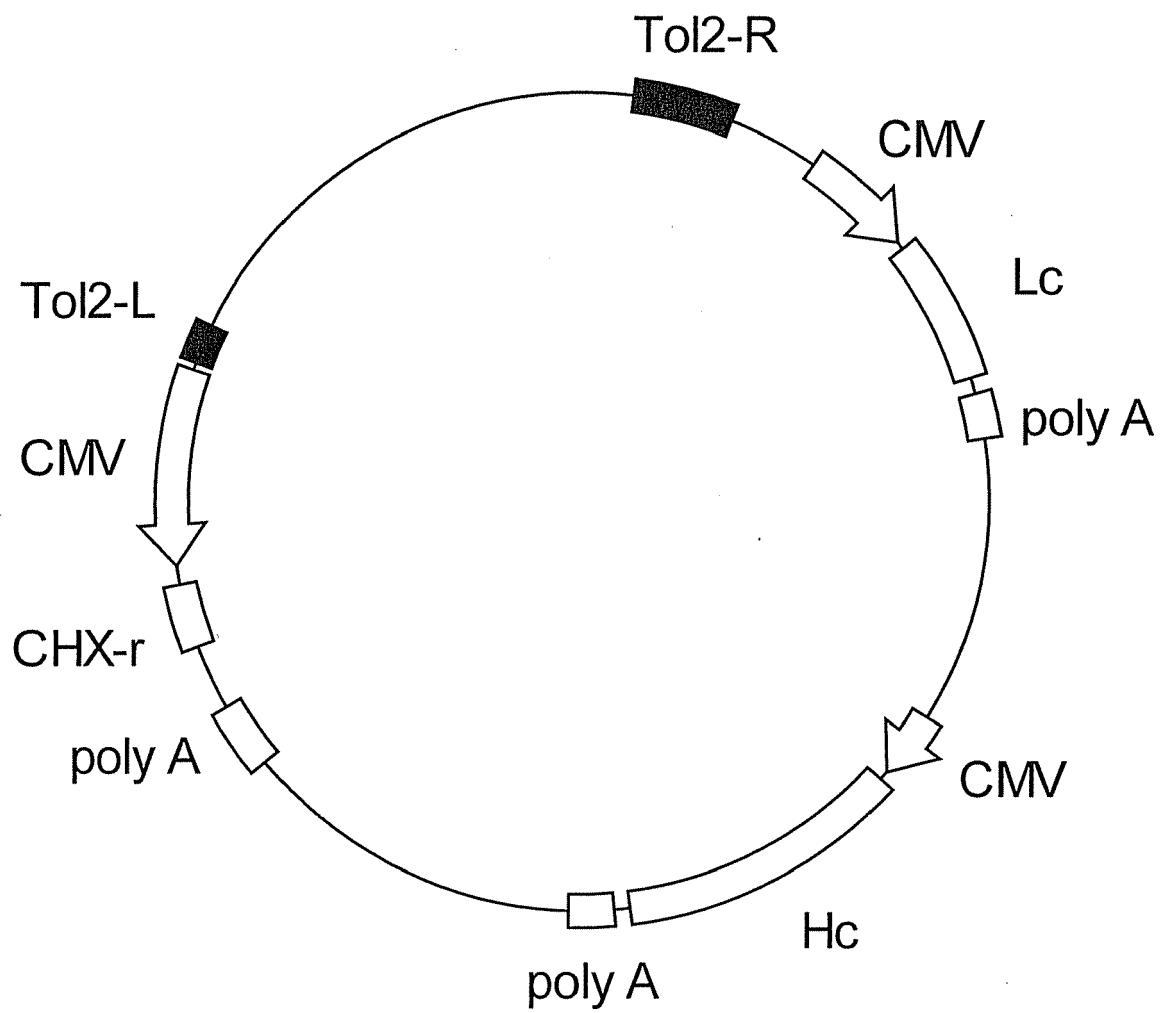


Fig.2

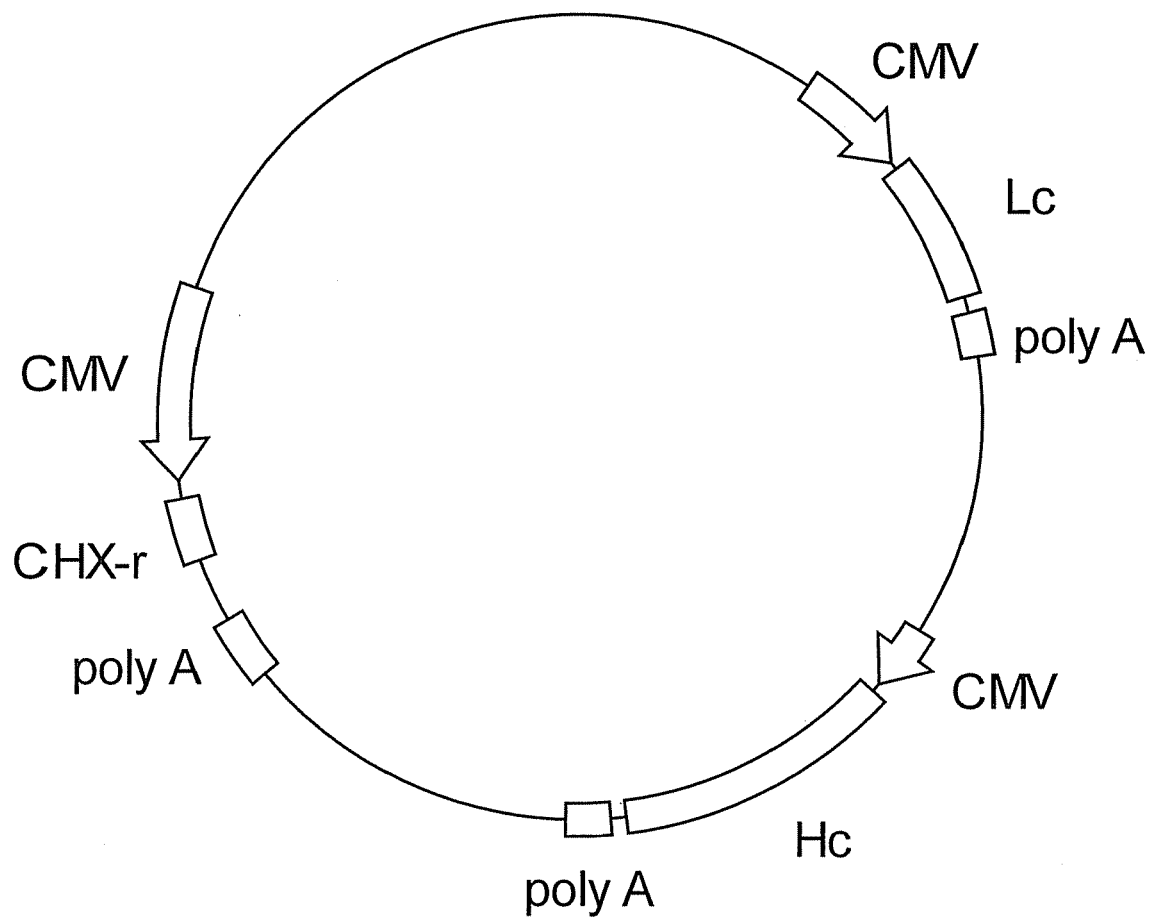


Fig.3

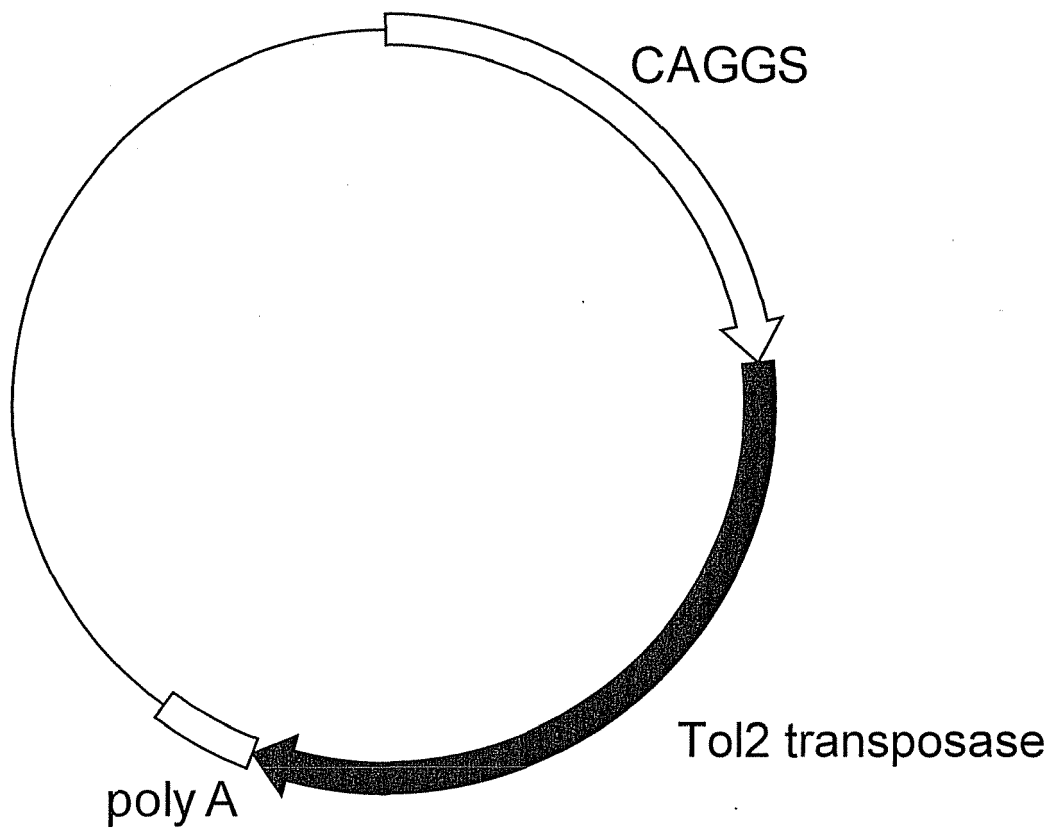


Fig. 4A

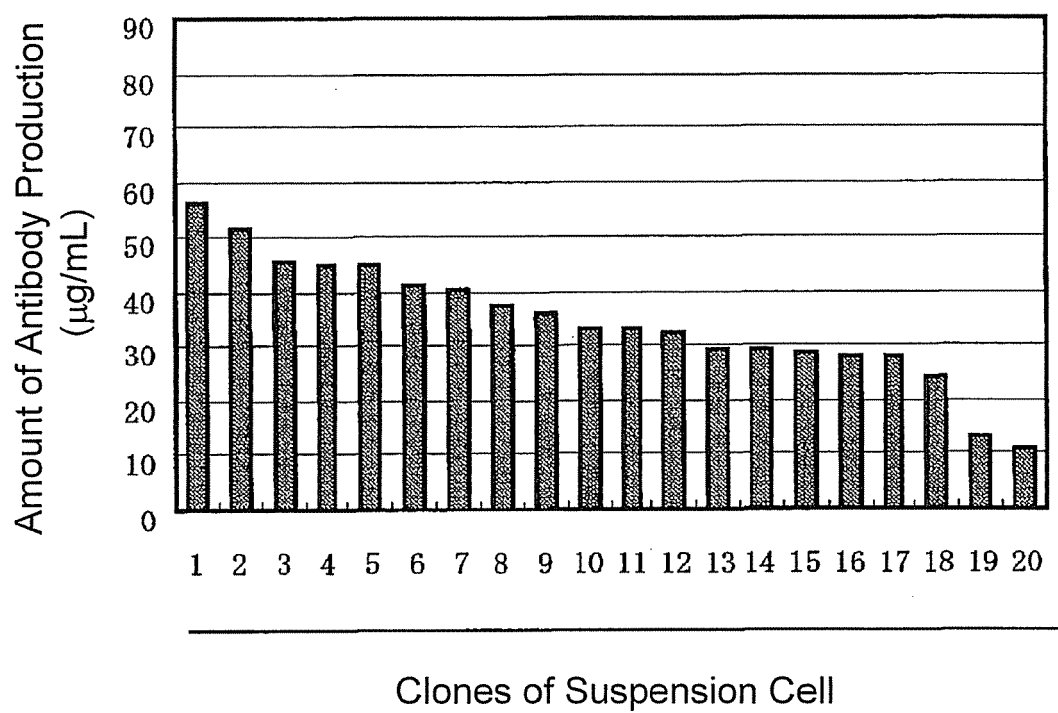


Fig.4B

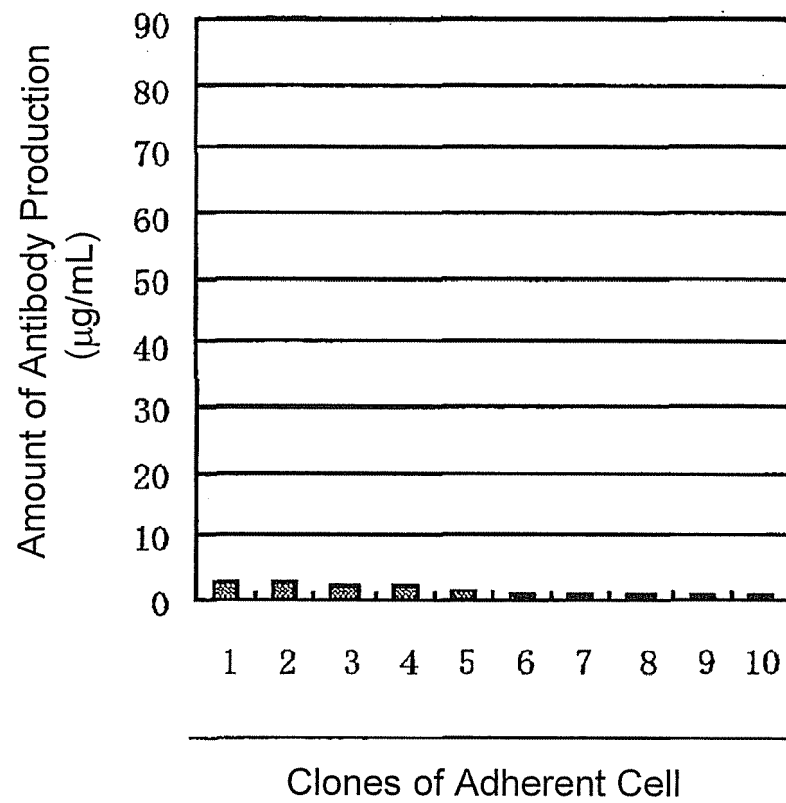


Fig.5

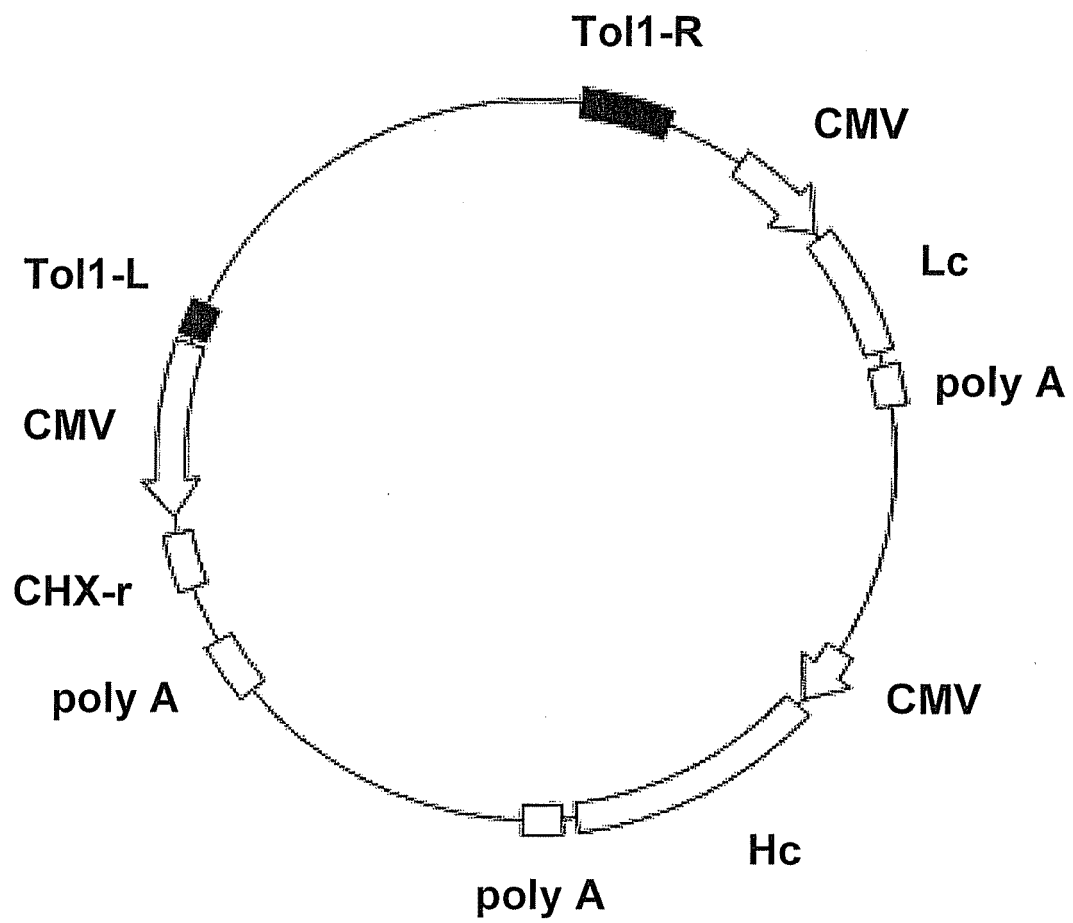


Fig.6

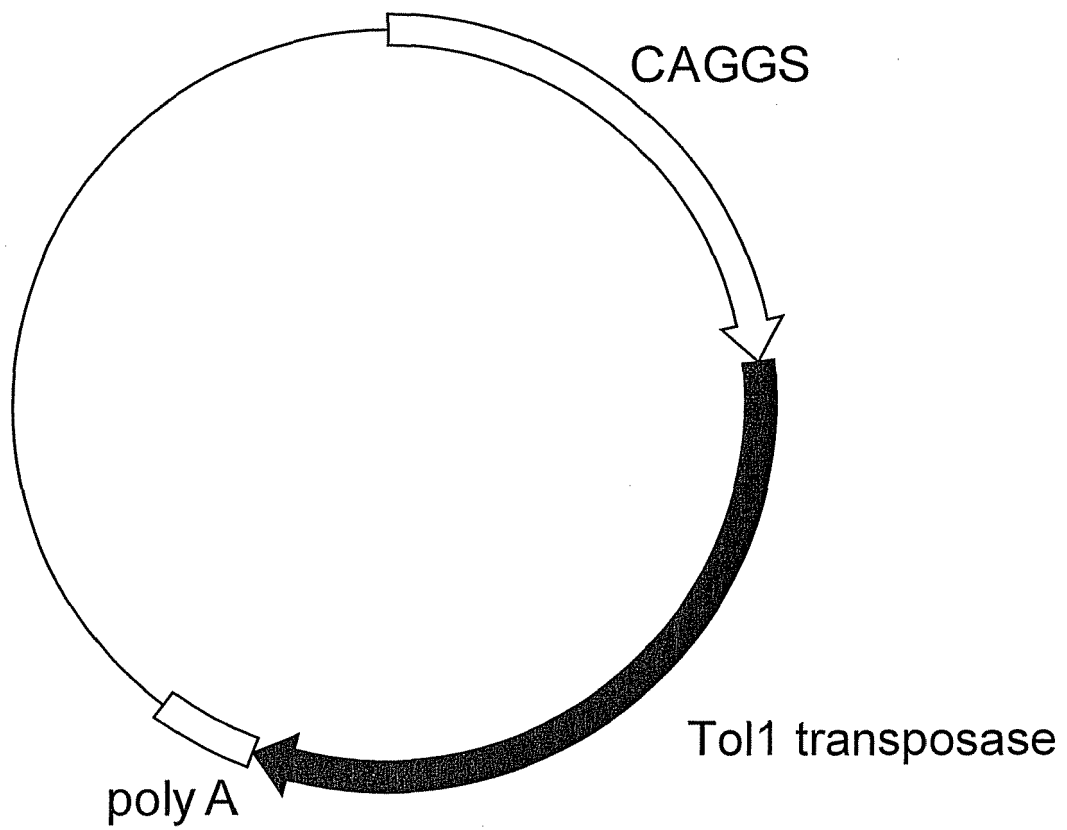
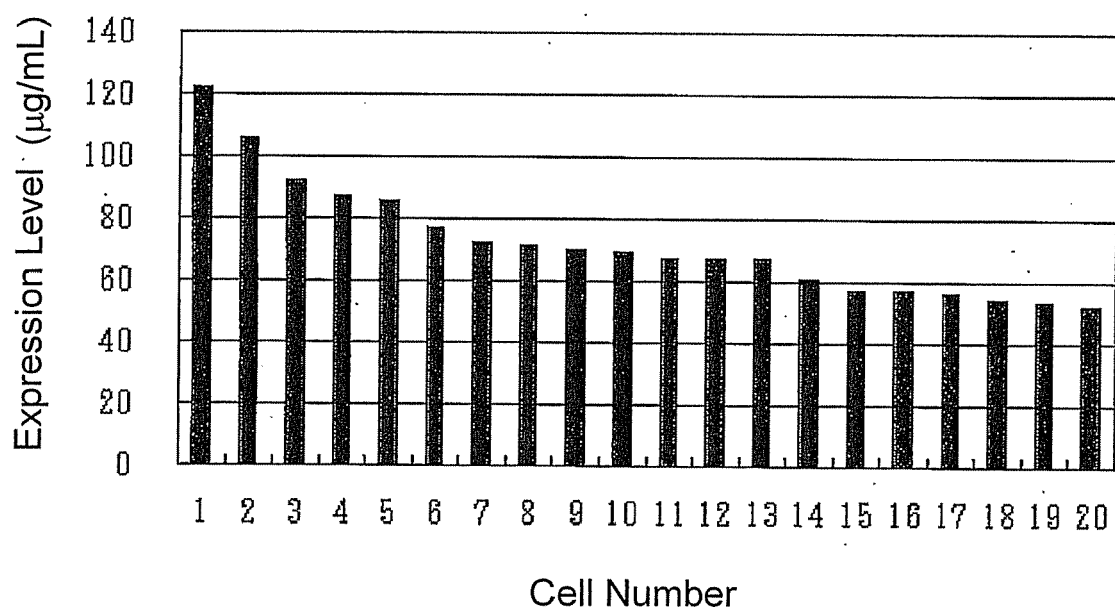


Fig.7



REFERENCES CITED IN THE DESCRIPTION

This list of references cited by the applicant is for the reader's convenience only. It does not form part of the European patent document. Even though great care has been taken in compiling the references, errors or omissions cannot be excluded and the EPO disclaims all liability in this regard.

Patent documents cited in the description

- WO 2008072540 A [0017] [0136]
- JP 2001532188 A [0017]
- JP 2002262879 A [0017] [0060] [0062]
- JP 2003235575 A [0046] [0102]
- WO 200535586 A [0068]
- WO 200231140 A [0068] [0071]
- WO 2005035586 A [0071]
- WO 0061739 A [0071]
- US 20070148165 A [0076]
- US 6737056 B [0077]
- US 7297775 B [0077]
- US 7317091 B [0077]
- WO 2006061723 A [0104] [0105]
- JP 2009140626 A [0153] [0156]
- US 61186138 A [0153]
- WO 69874 A [0156]
- US 61186138 B [0156]

Non-patent literature cited in the description

- *Nature*, 1996, vol. 383, 30 [0018]
- *Cell*, 1997, vol. 91, 501-510 [0018]
- *Nucleic Acids Res.*, 2003, vol. 31, 6873-6881 [0018]
- *Insect Mol. Biol.*, 1996, vol. 5, 141-151 [0018]
- *Genetics.*, 2004, vol. 166, 895-899 [0018]
- *PLoS Genet.*, 2006, vol. 2, e169 [0018]
- *Proc. Natl. Acad. Sci. USA*, 1998, vol. 95, 10769-10773 [0018]
- *Proc. Natl. Acad. Sci. USA*, 2001, vol. 98, 6759-6764 [0018]
- *Nature*, 2005, vol. 436, 221-226 [0018]
- *Nucleic Acids Res.*, 2003, vol. 31, 6873-6881 [0018]
- *Nucleic Acids Res.*, 2007, vol. 35, e87 [0018]
- *Proc. Natl. Acad. Sci. USA*, 2006, vol. 103, 15008-15013 [0018]
- *Genome Biology*, 2007, vol. 8 (1), 7.1-7.10 [0018]
- *Genetics*, 2006, vol. 174, 639-649 [0018]
- KOZAK M. *Nucleic Acids Res.*, 1984, vol. 12, 857-872 [0049]
- KONDO K. *J. Bacteriol.*, 1995, vol. 177 (24), 7171-7177 [0062]
- Shin Idenshi Kogaku Handbook (New Genetic Engineering Handbook). Yodo-sha [0065]
- *Journal of Experimental Medicine*, 1958, vol. 108, 945 [0066]
- *Proc. Natl. Acad. Sci. USA.*, 1968, 601275 [0066]
- *Genetics*, 1968, vol. 55, 513 [0066]
- *Chromosoma*, 1973, vol. 41, 129 [0066]
- *Methods in Cell Science*, 1996, vol. 18, 115 [0066]
- *Radiation Research*, 1997, vol. 148, 260 [0066]
- *Proc. Natl. Acad. Sci. USA.*, 1980, vol. 77, 4216 [0066]
- *Proc. Natl. Acad. Sci.*, 1968, vol. 60, 1275 [0066]
- *Cell*, 1975, vol. 6, 121 [0066]
- *Molecular Cell Genetics*, 883-900 [0066]
- Somatic Cell and Molecular Genetics. 1986, vol. 12, 55 [0068]
- *Mol. Biotechnol.*, 2000, vol. 15 (3), 249-57 [0084]
- *J. Biotechnol.*, 2007, vol. 130 (3), 282-90 [0084]
- *Cytotechnology*, 2006, vol. 52, 199-207 [0094]
- Protein Experimentation Note (the first volume) - Extraction, Separation and Expression of Recombinant Protein (translation of a textbook written in Japanese). Yodo-sha [0097]
- *Molecular Cloning* [0099]
- Current Protocols in Molecular Biology. Current Protocols [0099]
- *Gene*, 1991, vol. 108, 193-200 [0109]
- KAWAKAMI K. ; NODA T. *Genetics*, 2004, vol. 166, 895-899 [0112]
- *Yeast Res.*, 2007, vol. 7, 1307-1316 [0130] [0151]