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(54) **ANTI-PD1 ANTIBODIES AND THEIR USE AS THERAPEUTICS AND DIAGNOSTICS**

ANTI-PD1-ANTIKÖRPER SOWIE VERWENDUNG DAVON IN THERAPEUTIKA UND DIAGNOSTIKA

ANTICORPS ANTI-PD1 ET LEUR UTILISATION COMME PRODUITS THÉRAPEUTIQUES ET
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- **M. J. FULLER ET AL: "Immunotherapy of chronic hepatitis C virus infection with antibodies against programmed cell death-1 (PD-1)", PROCEEDINGS OF THE NATIONAL ACADEMY OF SCIENCES, vol. 110, no. 37, 26 August 2013 (2013-08-26), pages 15001-15006, XP055343159, US ISSN: 0027-8424, DOI: 10.1073/pnas.1312772110**

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Description**INTRODUCTION**

[0001] Programmed Death-1 (PD-1, also termed as CD279) is a 55 KD receptor protein related to CD28/CTLA4 co-stimulatory/inhibitory receptor family (Blank et al., 2005 Cancer Immunol Immunother 54:307-314). The genes and cDNAs coding for PD-1 were cloned and characterized in mouse and human (Ishida et al., 1992 EMBO J 11:3887-3395; Shinohara et al., 1994 Genomics 23:704-706). The full length PD-1 contains 288 amino acid residues (NCBI accession number: NP_005009). Its extracellular domain consists of amino acid residues 1-167, and the cytoplasmic C-terminal tail comprises residues 191-288, which has two hypothetical immune-regulatory motifs, an immunoreceptor tyrosine-based inhibitory motif (ITIM; Vivier et al., 1997 Immunol Today 18:286-291) and an immunoreceptor tyrosine switch motif (ITSM; Chemnitz et al., 2004 J Immunol 173:945-954).

[0002] To date, two sequence-related ligands, PD-L1 (B7-H1) and PD-L2 (B7-DC), have been identified to specifically interact with PD-1, inducing intracellular signal transduction that inhibits CD3 and CD28 mediated T-cell activation (Riley, 2009 Immunol Rev 229:114-125), which, in turn, attenuates T-cell activities, for example, reduction of cell proliferation, IL-2 and IFN- γ secretion, as well as other growth factor and cytokine secretion.

[0003] Expression of PD-1 was frequently found in immune cells such as T-cells, B-cells, monocytes and natural killer (NK) cells. It was rarely expressed in other human tissues, such as muscle, epithelium, neuronal tissues, etc. Furthermore, high level of PD-1 expression is often associated with activation of immune cells. For example, when human T-cell line, Jurkat, was activated by phytohaemagglutinin (PHA) or phorbol ester (12-O-tetradecanoylphorbol-13-acetate, or TPA), the expression of PD-1 was up-regulated visible in Western Blot (Vibharka et al., 1997 Exp Cell Res 232:25-28). The same phenomenon was observed in stimulated murine T- and B-lymphocytes and in primary human CD4⁺T-cells upon stimulation by anti-CD3 antibody (Agata et al., 1996 Int Immunol 8:765-772; Bennett et al., 2003 J Immunol 170:711-118). The increase of PD-1 expression following stimulation of T effector cells redirects the activated T-effector cells towards exhaustion and reduced immune activities. Therefore, PD-1 mediated inhibitory signal plays an important role in immune tolerance (Bour-Jordan et al., 2011 Immunol Rev 241:180-205).

[0004] Increase of PD-1 expression in tumor-infiltrating lymphocytes (TILs) and PD-1 ligand expression in tumor cells were reported in varieties of cancers involved in different types of tissues and organs such as lung (Konishi et al., 2004 Clin Cancer Res 10:5094-5100), liver (Shi et al., 2008 Int J Cancer 128:887-896; Gao et al., 2009 Clin Cancer Res 15:971-979), stomach (Wu et al., 2006 Acta Histochem 108:19-24), kidney (Thompson et al., 2004 Proc Natl Acad Sci 101:17174-17179; Thompson et al., 2007 Clin Cancer Res 13:1757-1761), breast (Ghebeh et al., 2006 Neoplasia 8:190-198), ovary (Hamanishi et al. 2007 Proc Natl Acad Sci 104:3360-3365), pancreas (Nomi et al., 2007 Clin Cancer Res 13:2151-2157), melanocytes (Hino et al., 2010 Cancer 116:1757-1766) and esophagus (Ohigashi et al., 2005 Clin Cancer Res 11:2947-2953). More frequently, the increased expression of PD-1 and PD-L1 in those cancers is associated with poor prognosis of patient survival outcome. Transgenic mice with PD-1 gene knockout inhibiting xenograft cancer cell growth further elucidated the significance of PD-1 signaling in the modulation of immune system for cancer eradication or tolerance (Zhang et al., 2009 Blood 114:1545-1552).

[0005] Not only does up-regulation of PD-1 signaling leads to immune tolerance to cancerous growth, but also to viral infection and expansion in human. The prevalent liver infection viruses, HBV and HCV, induce overexpression of PD-1 ligands in hepatocytes and activate PD-1 signaling in T-effector cells, resulting in T-cell exhaustion and tolerance to the viral infection (Boni et al., 2007 J Virol 81:4215-4225; Golden-Mason et al., 2008 J Immunol 180:3637-3641). Likewise, HIV infection frequently evades human immune system by similar mechanisms. Therapeutic modulation of PD-1 signaling by antagonist molecules may revert immune cells from tolerance, and reactivated to eradicate cancer and chronic viral infection (Blank et al., 2005 Cancer Immunol Immunother 54:307-314; Okazaki et al., 2007 Int Immunol 19:813-824). WO2006/121168 and WO2008156712 disclose high affinity anti-human PD-1 antibodies, relating to Nivolumab and Pembrolizumab, respectively.

[0006] The invention is defined by the claims and any other aspects or embodiments set forth herein not falling within the scope of the claims are for information only.

SUMMARY OF THE INVENTION

[0007] The invention provides methods and compositions for immune-inhibition of PD-1. The invention provides an antibody as defined in the claims. In one aspect, the disclosure provides an antibody antigen binding domain which specifically binds human PD-1, and comprises a complementarity determining region (CDR) having a sequence selected from SEQ ID NOS 11-22, 31-42 and 59-63.

[0008] The CDRs are amenable to recombination into heavy chain variable region (Vh) and light chain variable regions (Vk) which comprise (CDR-H1, CDR-H2 and CDR-H3) and (CDR-L1, CDR-L2 and CDR-L3) sequences, respectively and retain PD-1-specific binding and/or functionality.

[0009] In particular embodiments, the domain comprises a heavy chain variable region (Vh) or a light chain variable region (Vk) comprising:

a) CDR-H1 (SEQ ID NO:11, 17, 31, or 37), b) CDR-H2 (SEQ ID NO:12, 18, 32, or 38), c) CDR-H3 (SEQ ID NO:13, 18, 33, or 39);	d) CDR-L1 (SEQ ID NO:14, 20, 34, or 40), e) CDR-L2 (SEQ ID NO:15, 21, 35, or 41), or f) CDR-L3 (SEQ ID NO:16, 22, 36, or 42).
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[0010] In particular embodiments, the domain comprises a heavy chain variable region (Vh) and/or a light chain variable region (Vk) comprising:

a) mu317	CDR-H1, CDR-H2 and CDR-H3 (SEQ ID NOS:11-13); CDR-L1, CDR-L2 and CDR-L3 (SEQ ID NOS:14-16);
b) mu326	CDR-H1, CDR-H2 and CDR-H3 (SEQ ID NOS:17-19); CDR-L1, CDR-L2 and CDR-L3 (SEQ ID NOS:20-22);
c) 317-4B6	CDR-H1, CDR-H2 and CDR-H3 (SEQ ID NOS:31-33); CDR-L1, CDR-L2 and CDR-L3 (SEQ ID NOS:34-36);
d) 326-4A3	CDR-H1, CDR-H2 and CDR-H3 (SEQ ID NOS:37-39); CDR-L1, CDR-L2 and CDR-L3 (SEQ ID NOS:40-42);
e) 317-1	CDR-H1, CDR-H2 and CDR-H3 (SEQ ID NOS:11, 59, 13); CDR-L1, CDR-L2 and CDR-L3 (SEQ ID NOS:14-16);
f) 317-4B2	CDR-H1, CDR-H2 and CDR-H3 (SEQ ID NOS:11, 60, 13); CDR-L1, CDR-L2 and CDR-L3 (SEQ ID NOS:61, 15, 16);
g) 317-4B5	CDR-H1, CDR-H2 and CDR-H3 (SEQ ID NOS:11, 60, 13); CDR-L1, CDR-L2 and CDR-L3 (SEQ ID NOS:61, 15, 16);
h) 317-4B6	CDR-H1, CDR-H2 and CDR-H3 (SEQ ID NOS:11, 32, 13); CDR-L1, CDR-L2 and CDR-L3 (SEQ ID NOS:61, 15, 16);
i) 326-1	CDR-H1, CDR-H2 and CDR-H3 (SEQ ID NOS:17, 62, 19); CDR-L1, CDR-L2 and CDR-L3 (SEQ ID NOS:20-22);
j) 326-3B1	CDR-H1, CDR-H2 and CDR-H3 (SEQ ID NOS:17, 62, 19); CDR-L1, CDR-L2 and CDR-L3 (SEQ ID NOS:20-22);
k) 326-3G1	CDR-H1, CDR-H2 and CDR-H3 (SEQ ID NOS:17, 62, 19); or CDR-L1, CDR-L2 and CDR-L3 (SEQ ID NOS:20-22).

[0011] In particular embodiments, the domain comprises a heavy chain variable region (Vh) and a light chain variable region (Vk) comprising:

(a) CDR-H1 (SEQ ID NO 31), CDR-H2 (SEQ ID NO 12, 32, 59 or 60) and CDR-H3 (SEQ ID NO 33),
CDR-L1 (SEQ ID NO 14, 34 or 61), CDR-L2 (SEQ ID NO 35) and CDR-L3 (SEQ ID NO 36); or

(b) CDR-H1 (SEQ ID NO 37), CDR-H2 (SEQ ID NO 18, 38 or 62) and CDR-H3 (SEQ ID NO 39),
CDR-L1 (SEQ ID NO 40), CDR-L2 (SEQ ID NO 41) and CDR-L3 (SEQ ID NO 42).

[0012] In particular embodiments, the domain comprises a heavy chain variable region (Vh) or a light chain variable region (Vk) comprising:

a) mu317 (SEQ ID NOS:4 or 6);	p) 317-3H1 (SEQ ID NOS:69);
b) mu326 (SEQ ID NOS:8 or 10);	q) 317-311 (SEQ ID NOS:70);
c) 317-4B6 (SEQ ID NOS:24 or 26);	
d) 326-4A3 (SEQ ID NOS:28 or 30);	r) 317-4B1 (SEQ ID NOS:71);
e) 317-4B2 (SEQ ID NOS:43 or 44);	s) 317-4B3 (SEQ ID NOS:72);
f) 317-4B5 (SEQ ID NOS:45 or 46);	t) 317-4B4 (SEQ ID NOS:73);
g) 317-1 (SEQ ID NOS:48 or 50);	u) 317-4A2 (SEQ ID NOS:74);
h) 326-3B1 (SEQ ID NOS:51 or 52);	v) 326-3A1 (SEQ ID NOS:75);

(continued)

i) 326-3G1	(SEQ ID NOS:53 or 54);	w) 326-3C1	(SEQ ID NOS:76);
j) 326-1	(SEQ ID NOS:56 or 58);	x) 326-3D1	(SEQ ID NOS:77);
k) 317-3A1	(SEQ ID NOS:64);	y) 326-3E1	(SEQ ID NOS:78);
l) 317-3C1	(SEQ ID NOS:65);	z) 326-3F1	(SEQ ID NOS:79);
m) 317-3E1	(SEQ ID NOS:66);	aa) 326-3B N 55D	(SEQ ID NOS:80);
n) 317-3F1	(SEQ ID NOS:67);	ab) 326-4A1	(SEQ ID NOS: 81); or
o) 317-3G1	(SEQ ID NOS:68);	ac) 326-4A2	(SEQ ID NOS: 82).

[0013] In particular embodiments, the domain comprises a heavy chain variable region (Vh) and a light chain variable region (Vk) comprising:

a) mu317	(SEQ ID NOS:4 and 6);	p) 317-3H1	(SEQ ID NOS:69 and 26);
b) mu326	(SEQ ID NOS:8 and 10);	q) 317-3I1	(SEQ ID NOS:70 and 26);
c) 317-4B6	(SEQ ID NOS:24 and 26);		
d) 326-4A3	(SEQ ID NOS:28 and 30);	r) 317-4B1	(SEQ ID NOS:71 and 26);
e) 317-4B2	(SEQ ID NOS:43 and 44);	s) 317-4B3	(SEQ ID NOS:72 and 26);
f) 317-4B5	(SEQ ID NOS:45 and 46);	t) 317-4B4	(SEQ ID NOS:73 and 26);
g) 317-1	(SEQ ID NOS:48 and 50);	u) 317-4A2	(SEQ ID NOS:74 and 26);
h) 326-3B1	(SEQ ID NOS:51 and 52);	v) 326-3A1	(SEQ ID NOS:75 and 30);
i) 326-3G1	(SEQ ID NOS:53 and 54);	w) 326-3C1	(SEQ ID NOS:76 and 30);
j) 326-1	(SEQ ID NOS:56 and 58);	x) 326-3D1	(SEQ ID NOS:77 and 30);
k) 317-3A1	(SEQ ID NOS:64 and 26);	y) 326-3E1	(SEQ ID NOS:78 and 30);
l) 317-3C1	(SEQ ID NOS:65 and 26);	z) 326-3F1	(SEQ ID NOS:79 and 30);
m) 317-3E1	(SEQ ID NOS:66 and 26);	aa) 326-3B N55D	(SEQ ID NOS:80 and 30);
n) 317-3F1	(SEQ ID NOS:67 and 26);	ab) 326-4A1	(SEQ ID NOS:28 and 81); or
o) 317-3G1	(SEQ ID NOS:68 and 26);	ac) 326-4A2	(SEQ ID NOS:28 and 82).

[0014] In particular embodiments, the domain specifically binds PD1 residues: (a) K45 and I93 (AA numbering based on 2008 PNAS, 105:10483; equivalent to K58 and I106 in SEQ ID NO 2); or (b) I93, L95 and P97 (AA numbering based on 2008 PNAS, 105:10483; equivalent to I106, L108 and P110 in SEQ ID NO 2).

[0015] In particular embodiments, the domain induces IL-2 release in HuT78/PD-1 cells co-cultured with HEK293/OS8/PD-L1 cells or with EK293/OS8/PD-L2 cells, and/or inhibits IL-2 secretion in HuT78/P3Z cells co-cultured with HEK293/PD-L1 cells or with HEK293/PD-L2 cells.

[0016] The invention also provides an antibody IgG4 heavy chain effector or constant domain comprising any of SEQ ID NO:83-88, particularly SEQ ID NO 87 or 88..

[0017] The invention also provides antibodies, F(ab) or F(ab)2 comprising a subject PD-1 binding domain.

[0018] The invention also provides antibodies comprising a subject PD-1 binding domain and a IgG4 heavy chain effector or constant domain comprising any of SEQ ID NO:83-88, particularly SEQ ID NO 87 or 88.

[0019] The disclosure also provides a polynucleotide encoding a subject PD-1 binding domain, particularly cDNA sequences.

[0020] The disclosure provides methods of using the subject domains by administering the domain to a person determined to have cancer or a viral infection or to otherwise be in need of PD-1 antagonism.

[0021] The disclosure also provides fusion proteins comprising: (a) a single chain variable fragment (scFv) of an anti-human CD3 mAb OKT3 fused to the C-terminal domain (113-220) of mouse CD8 α (SEQ ID NO:89); or (b) the extracellular and transmembrane domains of human PD-1 fused to the cytoplasmic domain of human CD3 ζ chain (SEQ ID NO: 90).

[0022] The disclosure also provides methods of using the subject fusion proteins, comprising assaying, screening or selecting anti-PD-1 antibodies with a cell line expressing the fusion protein.

BREIF DESCRIPTION OF THE DRAWINGS

[0023]

Fig. 1. Schematic presentation of PD-1/Fc (top) and PD-1/His (bottom). ECD: extracellular domain. L: linker. H: His

tag. Fc: γ 4Fc fragment from human IgG4. N: N-terminus. C: C-terminus.

Fig. 2. Dose-dependent reaction curves of murine mAbs binding to human PD-1 in ELISA. The murine mAbs were indicated at top-left corner of each figure. MAb 317 and 517 share high degree of homology the variable region of heavy and light chains. The binding signal strength was indicated by direct OD₄₅₀ readings. The antigen, PD-1/His, was coated at increasing concentrations up to 70 nanograms per well in a volume of 50 microliters. The method was described in Example 1.

Fig. 3. Dose-dependent reaction curve of murine mAbs binding to human PD-1 expressed on live cells by FACS analyses. Murine antibody codes and EC₅₀ were indicated on each panel. MFI stands for mean fluorescence intensity. HuT78/PD-1 cells were suspended in 96-well plate at 5×10^4 cells per well for FACS. PD-1 mAbs binding to the cell surface target and FACS detection were performed as described in Example 1.

Fig. 4. Schematic presentation of the cell co-culture systems used for assaying functional activities of anti-PD-1 mAbs. T-cells (either CD4⁺ or CD8⁺) represent HuT78/PD-1 or primary T-cells in PBMCs. TCR: T-cell receptor. N: nucleus. C: cytoplasm

Fig. 5. Dose-dependent reaction curve of murine mAb-induced IL-2 secretion in HuT78/PD-1 cells co-cultured with HEK293/OS8/PD-L1 cells. Baseline: Average IL-2 release induced by mIgGs at all tested concentrations. Top line: Highest IL-2 release based on regression calculation by Prism Software.

Fig. 6. (A) Histograms showing IFN- γ secretion induced by anti-PD-1 mAbs in PBMCs (Donor-19) co-cultured with cell line HEK293/OS8/PD-L1. (B) Histograms showing IFN- γ secretion induced by anti-PD-1 mAbs in PBMCs (Donor-20) co-cultured with cell line HEK293/OS8/PD-L1.

Fig. 7. (A) and (B) ADCC activities of anti-PD-1 mAbs by co-culture of effector cells (NK92MI/PD-1) and target cells (HuT78/PD-1). Means were calculated from two data points of the representative experiments. The mAbs were added to concentration of 10 μ g/ml. Experiment performed as described in Example 9.

Fig. 8. Mapping the binding epitopes of anti-PD-1 mAbs by ELISA (up-panel) and Western Blot (lower panel). Conditioned media containing WT or Mt PD-1 were used to assess binding activity by ELISA and Western Blot. ** indicates the AA residues to which the mAb binding activity reduced to 25-50% of WT PD-1. *** indicates the AA residues to which the mAb binding activity reduced below 25% of WT PD-1.

Fig. 9. IFN- γ release induced by humanized anti-PD-1 mAbs in primary human PBMCs from different healthy donors, which were co-cultured with HEK293/OS8/PD-L1 cells.

Fig. 10. Cytotoxicity of NK92MI/PD-1 cells enhanced by humanized anti-PD-1 mAbs, hu317 (A) and hu326 (B). The target lung cancer cells, SK-MES-1/PD-L1, were co-cultured with the effector cells at the (T to E) ratio of 1 to 2, and assayed as described in Example 12.

Fig. 11. Individual tumor growth curves in three treatment groups, vehicle (PBS), human IgGs (hulgGs) and anti-PD-1 mAb (hu317-1/IgG4mt2). Each curve represents a tumor growth path, the tumor-bearing mice coded by numbers indicated on the right of each panel. Hep3B/OS8/PD-L1 cells (established from hepatocellular carcinoma line Hep3B) were seeded at Day 1, PBMCs were implanted at Day 15 and three doses of hu317-1/IgG4mt2 were injected at Day 18, 28 and 38, respectively. Methods described in Example 12.

DESCRIPTION OF PARTICULAR EMBODIMENTS OF THE INVENTION

[0024] PD-1 initiates inhibitory signaling in immune cells when engaged by its ligands, PD-L1 or PD-L2. In the cases of cancer outgrowth and viral infection, the activation of PD-1 signaling promotes immune tolerance, leading to the cancers or virus-infected cells escaping from immune surveillance and cancer metastasis or viral load increase. Inhibition of PD-1 mediated cellular signaling by therapeutic agents can activate immune cells including T-cells, B-cells and NK cells, and therefore enhance immune cell functions inhibiting cancer cell growth or viral infection, and restore immune surveillance and immune memory function to treat such human diseases.

[0025] The invention provides antibodies whose functions are antagonistic to the ligand-induced and PD-1-mediated cellular signaling in immune cells. Murine anti-PD-1 antibodies were humanized to a high degree of similarity to human antibodies in the framework regions. The full antibodies made in the modified human IgG4 variant format have a unique set of features in the aspects of effector functions and physicochemical properties. The disclosed anti-PD-1 antibodies are suitable for therapeutic uses in cancer treatment, controlling viral infections and other human diseases that are mechanistically involved in exacerbated immune tolerance.

Definitions

[0026] Unless the context indicates otherwise, the term "antibody" is used in the broadest sense and specifically covers antibodies (including full length monoclonal antibodies) and antibody fragments so long as they recognize PD-1. An antibody molecule is usually monospecific, but may also be described as idiospecific, heterospecific, or polyspecific. Antibody molecules bind by means of specific binding sites to specific antigenic determinants or epitopes on antigens.

"Antibody fragments" comprise a portion of a full length antibody, generally the antigen binding or variable region thereof. Examples of antibody fragments include Fab, Fab', F(ab')₂, and Fv fragments; diabodies; linear antibodies; single-chain antibody molecules; and multispecific antibodies formed from antibody fragments.

[0027] Monoclonal antibodies (MAbs) may be obtained by methods known to those skilled in the art. See, for example Kohler et al (1975); U.S. Pat. No. 4,376,110; Ausubel et al (1987-1999); Harlow et al (1988); and Colligan et al (1993). The mAbs of the invention may be of any immunoglobulin class including IgG, IgM, IgE, IgA, and any subclass thereof. A hybridoma producing a mAb may be cultivated in vitro or in vivo. High titers of mAbs can be obtained in in vivo production where cells from the individual hybridomas are injected intraperitoneally into mice, such as pristine-primed Balb/c mice to produce ascites fluid containing high concentrations of the desired mAbs. MAbs of isotype IgM or IgG may be purified from such ascites fluids, or from culture supernatants, using column chromatography methods well known to those of skill in the art.

[0028] An "isolated polynucleotide" refers to a polynucleotide segment or fragment which has been separated from sequences which flank it in a naturally occurring state, e.g., a DNA fragment which has been removed from the sequences which are normally adjacent to the fragment, e.g., the sequences adjacent to the fragment in a genome in which it naturally occurs. The term therefore includes, for example, a recombinant DNA which is incorporated into a vector, into an autonomously replicating plasmid or virus, or into the genomic DNA of a prokaryote or eukaryote, or which exists as a separate molecule (e.g., as a cDNA or a genomic or cDNA fragment produced by PCR or restriction enzyme digestion) independent of other sequences. It also includes a recombinant DNA, which is part of a hybrid gene encoding additional polypeptide sequence.

[0029] A "construct" means any recombinant polynucleotide molecule such as a plasmid, cosmid, virus, autonomously replicating polynucleotide molecule, phage, or linear or circular single-stranded or double-stranded DNA or RNA polynucleotide molecule, derived from any source, capable of genomic integration or autonomous replication, comprising a polynucleotide molecule where one or more polynucleotide molecule has been linked in a functionally operative manner, i.e. operably linked. A recombinant construct will typically comprise the polynucleotides of the invention operably linked to transcriptional initiation regulatory sequences that will direct the transcription of the polynucleotide in the intended host cell. Both heterologous and non-heterologous (i.e., endogenous) promoters can be employed to direct expression of the nucleic acids of the invention.

[0030] A "vector" refers any recombinant polynucleotide construct that may be used for the purpose of transformation, i.e. the introduction of heterologous DNA into a host cell. One type of vector is a "plasmid", which refers to a circular double stranded DNA loop into which additional DNA segments can be ligated. Another type of vector is a viral vector, wherein additional DNA segments can be ligated into the viral genome. Certain vectors are capable of autonomous replication in a host cell into which they are introduced (e.g., bacterial vectors having a bacterial origin of replication and episomal mammalian vectors). Other vectors (e.g., non-episomal mammalian vectors) are integrated into the genome of a host cell upon introduction into the host cell, and thereby are replicated along with the host genome. Moreover, certain vectors are capable of directing the expression of genes to which they are operatively linked. Such vectors are referred to herein as "expression vectors".

[0031] An "expression vector" as used herein refers to a nucleic acid molecule capable of replication and expressing a gene of interest when transformed, transfected or transduced into a host cell. The expression vectors comprise one or more phenotypic selectable markers and an origin of replication to ensure maintenance of the vector and to, if desired, provide amplification within the host. The expression vector further comprises a promoter to drive the expression of the polypeptide within the cells. Suitable expression vectors may be plasmids derived, for example, from pBR322 or various pUC plasmids, which are commercially available. Other expression vectors may be derived from bacteriophage, phagemid, or cosmid expression vectors.

Additional Embodiments of the Disclosure

[0032] In specific embodiments the disclosure provides mouse monoclonal antibodies identified from screening murine hybridoma clones as disclosed herein.

[0033] In other embodiments the disclosure provides compositions of the following polynucleotide and protein sequences:

- a) The cDNA sequence, SEQ ID NO 3, encoding the heavy chain variable region of murine mAb 317;
- b) The protein sequence of the heavy chain variable region of murine mAb 317 or mu317_Vh (SEQ ID NO 4);
- c) The cDNA sequence, SEQ ID NO 5, encoding the light chain variable region of murine mAb 317;
- d) The protein sequence of the light chain variable region of murine mAb 317 or mu317_Vk (SEQ ID NO 6);
- e) The cDNA sequence, SEQ ID NO 7, encoding the heavy chain variable region of murine mAb 326;
- f) The protein sequence of the heavy chain variable region of murine mAb 326 or mu326_Vh (SEQ ID NO 8);
- g) The cDNA sequence, SEQ ID NO 9, encoding the light chain variable region of murine mAb 326;

h) The protein sequence of the light chain variable region of murine mAb 326 or mu326_Vk (SEQ ID NO 10).

[0034] In one aspect, the disclosure provides compositions comprising complement determinant region (CDR) sequences, which mediate binding to the target antigens, PD-1, including the CDR sequences of mu317 and m326:

- a) The CDR1 of mu317 heavy chain (mu317 H-CDR1) contains amino acid sequence of GFSLTSGYGVH (SEQ ID NO 11);
- b) The mu317 H-CDR2 contains amino acid sequence of VIWAGGSTNYNSALMS (SEQ ID NO 12);
- c) The mu317 H-CDR3 contains amino acid sequence of ARAYGNYWYIDV (SEQ ID NO 13);
- d) The CDR1 of mu317 light chain (mu317 L-CDR1) contains amino acid sequence of KASQSVSNDVA (SEQ ID NO 14);
- e) The mu317 L-CDR2 contains amino acid sequence of YAFHRFT (SEQ ID NO 15);
- f) The mu317 L-CDR3 contains amino acid sequence of HQAYSSPYT (SEQ ID NO 16);
- g) The mu326 H-CDR1 contains amino acid sequence of GYTFTNYGMN (SEQ ID NO 17);
- h) The mu326 H-CDR2 contains amino acid sequence of WINNNNGEPTYAEFFKG (SEQ ID NO 18);
- i) The mu326 H-CDR3 contains amino acid sequence of ARDVMDY (SEQ ID NO 19);
- j) The mu326 L-CDR1 contains amino acid sequence of RASESVDNYGYSFMH (SEQ ID NO 20);
- k) The mu326 L-CDR2 contains amino acid sequence of RASNLES (SEQ ID NO 21);
- l) The mu326 L-CDR3 contains amino acid sequence of QQSKEYPT (SEQ ID NO 22).

[0035] In another embodiment, the disclosure provides compositions comprising the sequences of the humanization monoclonal antibodies emanated from murine mAbs mu317 and mu326, including:

- a) The humanization mAb hu317-4B6 comprises protein sequence of heavy chain variable region (Vh) as SEQ ID NO 24, which is encoded by
- b) the cDNA of hu317-4B6_Vh (SEQ ID NO 23);
- c) The humanization mAb hu317-4B6 also comprises protein sequence of light chain variable region (Vk) as SEQ ID NO 26, which is encoded by
- d) the cDNA of hu317-4B6 (SEQ ID NO 25);
- e) the humanization mAb hu326-4A3 comprises protein sequence of Vh as SEQ ID NO 28, which is encoded by
- f) the cDNA of hu326-4A3-Vh (SEQ ID NO 27);
- g) The humanization mAb hu326-4A3 also comprises protein sequence of Vk as SEQ ID NO 30, which is encoded by
- h) the cDNA of hu326-4A3_Vk (SEQ ID NO 29);
- i) The protein sequences of hu317-4B2_Vh (SEQ ID NO 43) and hu317-4B2_Vk (SEQ ID NO 44);
- j) The protein sequences of hu317-4B5_Vh (SEQ ID NO 45) and hu317-4B5_Vk (SEQ ID NO 46);
- k) The protein sequence of hu317-1_Vh (SEQ ID NO 48) and the cDNA encoding for hu317-1_Vh (SEQ ID NO 47);
- l) The protein sequence of hu317-1_Vk (SEQ ID NO 50) and the cDNA encoding for hu317-1_Vk (SEQ ID NO 49);
- m) The protein sequences of hu326-3B1_Vh (SEQ ID NO 51) and hu326-3B1_Vk (SEQ ID NO 52);
- n) The protein sequences of hu326-3G1_Vh (SEQ ID NO 53) and hu326-3G1_Vk (SEQ ID NO 54);
- o) The protein sequence of hu326-1_Vh (SEQ ID NO 56) and the cDNA encoding for hu326-1_Vh (SEQ ID NO 55);
- p) The protein sequence of hu326-1_Vk (SEQ ID NO 58) and the cDNA encoding for hu326-1_Vk (SEQ ID NO 57);
- q) The protein sequences of other humanization mAbs emanated from mu317 (SEQ ID NO 63-74);
- r) The protein sequences of other humanization mAbs emanated from mu326 (SEQ ID NO 75-82);

[0036] In one aspect, the disclosure provides compositions comprising the CDR sequences of the humanization monoclonal antibodies. The CDRs may be shared among the same series of humanization mAbs, such as hu317 or hu326 (see Table 15-16). Non-redundant CDRs are listed below:

- a) H-CDR1 sequence of GFSLTSGYGVH (SEQ ID NO 31), shared throughout humanization mAbs hu317 and mu317 in the heavy chains;
- b) H-CDR3 sequence of ARAYGNYWYIDV (SEQ ID NO 33), shared throughout humanization mAbs hu317 and mu317 in the heavy chains;
- c) L-CDR1 sequence of KSSSVSNDVA (SEQ ID NO 34), shared throughout humanization mAbs hu317-4B2, hu317-4B5 and hu317-4B6 in the light chains;
- d) L-CDR2 sequence of YAFHRFT (SEQ ID NO 35), shared throughout humanization mAbs hu317 and mu317 in the light chains;
- e) L-CDR3 sequence of HQAYSSPYT (SEQ ID NO 36), shared throughout humanization mAbs hu317 and mu317 in the light chains;

- f) H-CDR2 sequence of VIYADGSTNYPNPSLKS (SEQ ID NO 32) in hu317-4B6_Vh;
- g) H-CDR2 sequence of VIYAGGSTNYPNPSLKS (SEQ ID NO 60) in hu317-4B2_Vh and hu317-4B5_Vh;
- h) H-CDR2 sequence of VIWAGGSTNYPNPSLKS (SEQ ID NO 59) in hu317-1_Vh;
- i) L-CDR1 sequence of KASQSVSNDVA (SEQ ID NO 11) in hu317-1_Vk;
- j) H-CDR1 sequence of GYTFTNYGMN (SEQ ID NO 37), shared throughout humanization mAbs hu326 and mu326 in the heavy chains;
- k) H-CDR3 sequence of ARDVMDY (SEQ ID NO 39), shared throughout humanization mAbs hu326 and mu326 in the heavy chains;
- l) L-CDR1 sequence of RASESVDNYGYFSFMH (SEQ ID NO 40), shared throughout humanization mAbs hu326 and mu326 in the light chains;
- m) L-CDR2 sequence of RASNLES (SEQ ID NO 41), shared throughout humanization mAbs hu326 and mu326 in the light chains;
- n) L-CDR3 sequence of QQSKEYPT (SEQ ID NO 42), shared throughout humanization mAbs hu326 and mu326 in the light chains;
- o) H-CDR2 sequence of WINNNNAEPTYAQDFRG (SEQ ID NO 38) in hu326_4A3_Vh;
- p) H-CDR2 sequence of WINNNNGEPTYAQGFRG (SEQ ID NO 62) in the Vh of hu326_1 and other hu317 mAbs.

[0037] In another aspect, the disclosure provides particular binding epitopes of the humanized anti-PD-1 mAbs on the antigen, and functional use thereof. Six critical amino acid (AA) residues in PD-1 required for the ligand binding were mutated individually, and mutant and wild-type PD-1 proteins were used to assess the binding epitopes. The residue whose mutation significantly impaired the antibody binding is recognized as a key or significant binding epitope. Significant binding epitopes of mAbs hu317-4B5 and hu317-4B6 are K45 and I93 (AA numbering based on 2008 PNAS, 105:10483; equivalent to K58 and I106 in SEQ ID NO 2); and significant binding epitopes of mAbs hu326-3B1 and hu317-4A3 are I93, L95 and P97 (AA numbering based on 2008 PNAS, 105:10483; equivalent to I106, L108 and P110 in SEQ ID NO 2).

[0038] In a further aspect, the disclosure provides compositions comprising the constant region sequences of recombinant human IgG4 variants, which may be linked to the variable regions of the subject antibodies, including the humanized anti-PD-1 mAbs, which showed preferred effector functions and physicochemical properties. The sequences are as follows:

The constant region sequence of IgG4mt10 (SEQ ID NO 88);

- a) A reference sequence of IgG4mt1 (SEQ ID NO 83);
- b) A reference sequence of IgG4mt2 (SEQ ID NO 84);
- c) A reference sequence of IgG4mt6 (SEQ ID NO 85);
- d) A reference sequence of IgG4mt8 (SEQ ID NO 86);
- e) A reference sequence of IgG4mt9 (SEQ ID NO 87).

[0039] In another embodiment, the disclosure provides methods for assaying anti-PD-1 antibody functions, using a plasmid expressing the recombinant fusion protein, OS8, to generate stable cell lines, HEK293/OS8/PD-L1 or HEK293/OS8/PD-L2, which co-expresses OS8 (a T cell-activating molecule) and a PD-1 ligand. The cell lines were used to engage T-cells and PBMCs by co-culture to assess the functionality of anti-PD-1 mAbs (see Example 3 and Example 4). Alternatively, another plasmid expressing the recombinant fusion protein, P3Z, was used to generate stable cell line, HuT78/P3Z, in which P3Z functions as molecular sensor and signal transduction mediator. When P3Z is engaged by PD-1 ligand, it will transmit intracellular signal to activate IL-2 release in the HuT78 cells. The systems may be used to assess inhibitory effect of anti-PD-1 mAbs (see Example 3).

[0040] In one aspect, the disclosure provides compositions comprising the amino acid sequences of the recombinant fusion proteins as follows:

- a) Protein sequence of OS8 (SEQ ID NO 89);
- b) Protein sequence of P3Z (SEQ ID NO 90).

[0041] In another aspect, the disclosure provides methods of generating the stable cell lines that express the recombinant fusion proteins described herein, and methods of using the system to quantitatively assay the functional activities of anti-PD-1 mAbs.

[0042] In another embodiment the disclosure provides polynucleotides encoding the subject proteins. The polynucleotides may be operably linked to a heterologous transcription regulating sequence for expression, and may be incorporated into vectors, cells, etc.

[0043] In another embodiment, the disclosure provides the murine anti-PD-1 antibodies and humanized version anti-PD-1 antibodies, including hu317-4B6, hu317-4B5, hu317-4B2, etc., and hu326-4A3, hu326-3B1, hu326-3G1, etc.,

having functions to suppress PD-1 mediated signal transduction, and to activate immune cells, which trigger a cascade of immune responses including cytokine secretion and cytotoxicity towards target cells such as cancer cells, and such functional use of the antibodies.

[0044] In one aspect, the invention provides humanized anti-PD-1 antibodies as defined in the claims that activate several types of immune cells that express PD-1, including human T-cells, NK-cells and PBMCs, whose functions are to amplify the immune response signals, to mobilize immune system and to act as immune effector cells for clearance of cancer cells and viral infections, and such functional use of the antibodies.

[0045] In another aspect, the humanized anti-PD-1 mAbs are used as therapeutic agents to treat human diseases that are involved in suppression of immune cells by PD-1 mediated intracellular signaling, leading to disease progression, particularly cancers and viral infections.

[0046] The compositions of the disclosure are useful for the treatment of cancer, neurodegenerative and infectious, particularly viral, diseases and other conditions in which inappropriate or detrimental expression of the human PD-1 and/or is a component of the etiology or pathology of the condition. Hence, the disclosure provides methods for treating cancer or inhibiting tumor progression in a subject in need thereof with a subject anti-PD-1 protein. The disclosure further provides the use of subject polynucleotides for the manufacture of a medicament for treating cancer or inhibiting tumor progression in a subject.

[0047] The disclosure includes all combinations of the recited particular embodiments.

EXAMPLES

Example 1. Generation of anti-PD-1 monoclonal antibody

[0048] Anti-PD-1 monoclonal antibodies (mAbs) were generated based on conventional hybridoma fusion technology (Kohler and Milstein 1976 Eur J Immunol 6:511-519; de St Groth and Sheidegger 1980, J Immunol Methods 35:1-21; Mechetner 2007 Methods Mol Biol 378:1-13) with minor modifications. MAbs with high binding activities in enzyme-linked immunosorbent assay (ELISA) and fluorescence-activated cell sorting (FACS) assay were selected for further characterization

PD-1 recombinant protein for immunization and binding assays

[0049] Expression plasmid containing full-length human PD-1 cDNA was obtained from Origene (Cat. No. SC117011, NCBI Accession No: NM_005018.1, Beijing, China). The extracellular domain consisting of amino acid (AA) 1-168 of PD-1 (SEQ NO.1, SEQ NO.2) was PCR-amplified, and subcloned in pcDNA3.1-based expression vector (Invitrogen, Carlsbad, CA, USA) with C-terminus fused either to a His₆ tag or to the γ Fc domain of human IgG4 heavy chain, which resulted in two recombinant fusion protein expression plasmids, PD-1-EC/His and PD-1-EC/Fc (abbreviated as PD-1/His and PD-1/Fc). The schematic presentation of immunogen/antigen proteins were shown in Fig 1. For the recombinant fusion protein production, PD-1/His and PD-1/Fc plasmids were transiently transfected into 293-F cells in 1-3 liters of medium (Invitrogen), and cultured for 5-7 days in a CO₂ incubator equipped with rotating shaker. The supernatant containing the recombinant protein was collected and cleared by centrifugation at 15000g for 30 minutes. PD-1/His was purified through immobilized metal affinity chromatography using Ni-Sepharose Fast Flow (Cat. No. 17531801, GE Lifesciences, Shanghai, China), followed by size exclusion chromatography using a HiLoad 16/60 Superdex 200 column (Cat. No. 17106901, GE Lifesciences, Shanghai, China). PD-1/Fc was purified using a Protein G Sepharose Fast Flow column (Cat. No. 17061805, GE Lifesciences). Both PD-1/His and PD-1/Fc proteins were dialyzed against phosphate buffered saline (PBS) and stored in -80°C freezer in small aliquots.

[0050] The cDNA coding for human PD-L1 was chemically synthesized by Genescript (Nanjing, China) based on the published sequence (NCBI Accession No. NM_014143). The PD-L2 expression plasmid was purchased from Origene (Cat. No. SC108873, NCBI Accession No. NM_025239.2, Beijing, China). Both cDNAs were cloned in pcDNA3.1/Hygromycin (Cat. No. V870-20, Invitrogen), and pcDNA3.1/V5-His (Cat. No. V810-20, Invitrogen), respectively.

Stable expression cell line

[0051] Stable cell lines expressing human PD-1, PD-L1 or PD-L2 were established by transfection of pcDNA3.1 plasmids containing PD-1, PD-L1 and PD-L2 to HUT78 (ATCC, Manassas, VA, USA) and HEK293 (ATCC), respectively, and followed by selection with medium containing 200 micrograms of hygromycin (Cat. No. 10687-010, Invitrogen) or 1 mg of G418 (Sigma) per milliliter. Single clones were isolated by conventional method, either limited dilution or picking up single colonies from culture-well surface. All clones were screened by Western blot and FACS analysis using anti-PD-1, PD-L1 and PD-L2 antibodies (Cat. No. 12-9969, 17-5983, 12-5888, eBioscience, San Diego, USA), respectively, and the top expression clones were selected for FACS binding assay to screen hybridoma monoclonal antibodies, or

used in functional assays.

Immunization, hybridoma fusion and cloning

- 5 **[0052]** Eight to twelve week-old Balb/c mice (from BEIJING HFK BIOCSIENCE CO.,LTD, Beijing, China) were immunized subcutaneously with 100ul of adjuvant (Cat. No. KX0210041, KangBiQuan, Beijing, China) containing 5 micrograms of PD-1/Fc. The immunization was conducted by two injections of the above immunogen with three weeks apart. Two weeks after the 2nd immunization, the mice sera were evaluated for PD-1 binding by FACS (following sections). The mice with high anti-PD-1 antibody titers in sera were selected and boosted intraperitoneally with 50 micrograms of PD-1/Fc in the absence of any adjuvant. Three days after boosting, the splenocytes were isolated and fused with the murine myeloma cell line, SP2/0 cells (ATCC), using standard techniques (Geftter, M.L. et al., 1977 Somat Cell Genet, 3:231-236).
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Assess PD-1 binding activity of antibodies by ELISA and FACS

- 15 **[0053]** The supernatants of hybridoma clones were initially screened by Enzyme-Linked Immuno-Sorbent Assay (ELISA) as described in "Flanagan, M.L. et al. 2007 Methods in Molecular Biology 378:33-52" with some modifications. Briefly, 50-200 nanograms of PD-1/His or PD-1/Fc protein in 50 microliters of phosphate buffered saline (PBS) were coated in 96-well plate (Shenzhen JinCanHua Industry Co., Ltd, Shenzhen, China) on per well base. The HRP-linked anti-mouse IgG antibody (Cat. No. 7076S, Cell Signaling Technology, USA and Shanghai, China) and chemiluminescent reagent (Cat. No. PA107-01, TIANGEN, China) were used to detect and develop the ELISA signal, which were read out by a plate reader (PHREastar FS, BMG LABTECH, Germany) at wavelength of 450 nm. The ELISA-positive antibody producer clones were further verified by fluorescence-activated cell sorting (FACS) using a conventional method. PD-1 stable expression cell lines, HuT78/PD-1 (10^5 cells/well), described above, was stained with supernatants from anti-PD-1 hybridomas in V-bottom 96-well plates (Cat. No. 3897, Corning, USA and Shanghai, China). To block human Fc receptors, cells were pre-incubated with human IgG (20 μ g/ml) (Cat. No. H11296, LifeHolder, USA and Shanghai, China). PD-1 antibodies were detected with Dylight™ 649-labelled goat anti-mouse IgG antibody (Cat. No. 405312, Biolegend, San Diego, USA) and cell fluorescence was monitored using a flow cytometer (Guava easyCyte 8HT, Merck-Millipore, USA and Shanghai, China).
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25 **[0054]** The conditioned media of hybridoma cells that showed positive signal in both ELISA and FACS assay were subjected to functional assays to identify antibodies with good functional activity in human immune cell-based assays (herein). The antibodies with positive functional activity were further subcloned and characterized.
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Subcloning and adaptation to serum-free or low serum medium

- 35 **[0055]** The positive hybridoma clones from primary screening through ELISA, FACS and functional assays were subcloned by the conventional method of limited dilution. Each of the positive clones was plated out in a 96-well plate, cultured in RPMI1640 medium (Cat. No. SH30809.01B, Hyclone, Shanghai, China) with 10% fetal bovine serum (FBS, Cat. No. SH30084.03, Hyclone, Beijing, China) in CO₂ incubator. Three subclones from each limited dilution plate were selected and characterized by FACS and functional assays. The subclones selected through functional assays were defined as monoclonal antibody. The top subclones were adapted for growth in the CDM4MAb medium (Cat. No. SH30801.02, Hyclone) with 1-3% FBS.
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Expression and purification of monoclonal antibodies

- 45 **[0056]** Either murine monoclonal antibody-producing hybridoma cells or recombinant antibody plasmids-transfected 293-F cells (Cat. No. R79007, Invitrogen) were cultured in CDM4MAb medium (Cat. No. SH30801.02, Hyclone) or Freestyle293 Expression medium (Cat. No. 12338018, Invitrogen), respectively, in a CO₂ incubator at 37°C for 5 to 7 days. The conditioned medium was collected through centrifugation at 10,000g for 30 minutes to remove all cells and cell debris, and filtrated through a 0.22 μ m membrane before purification. Murine or recombinant antibodies were applied and bound to a Protein A column (Cat. No. 17127901, GE Life Sciences) following the manufacturer's guidance, washed with PBS, eluted in the buffer containing 20mM citrate, 150mM NaCl, pH3.5. The eluted materials were neutralized with 1M Tris pH8.0, and usually contained antibodies of above 90% purity. The Protein A-affinity purified antibodies were either dialyzed against PBS or further purified using a HiLoad 16/60 Superdex200 column (Cat. No. 17531801, GE Life Sciences) to remove aggregates. Protein concentrations were determined by measuring absorbance at 280nm or by Bradford assay (Cat. No. 1856210, Thermo Scientific, Rockford, IL, USA) using bovine IgG of defined concentration (Cat. No. 23212, Thermo Scientific) as the standards. The purified antibodies were stored in aliquots in -80°C freezer.
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Example 2. Comparison of binding activities among anti-PD-1 antibodies

[0057] Through screening thousands of hybridoma clones we identified some top monoclonal antibodies (mAb), which bind to human PD-1 with high specificity and strength. As shown in ELISA assay (Figure 2), three of the top antibodies elicited such binding strength and specificity. FACS analysis results demonstrated the selected monoclonal antibodies bind to the native PD-1 proteins expressed on cell surface. Murine mAb317 (mu317), mu326 and mu150 showed concentration-dependent binding activity, and their binding EC_{50} (Effective concentration at 50% activity) was significantly lower than that of the control mu55 (Figure 3).

Assess mAb binding affinity by Surface Plasmon Resonance (SPR)

[0058] The mAbs with high binding activities in ELISA and FACS, as well as with potent functional activities in the cell-based assays (herein) were examined for their binding kinetic constant in real time binding reactions. Murine anti-PD-1 mAbs were purified from hybridoma supernatants using protein A Flow column (Cat. No. 17531801, GE Life Sciences) followed by exclusion chromatography using a HiLoad 16/60 Superdex200 column (Cat. No. 17106901, GE Life Sciences). The purified anti-PD-1 antibodies were concentrated to 0.5-1 mg/mL in PBS and stored in aliquots in -80°C freezer.

[0059] For determining binding affinities of PD-1 mAbs, SPR measurements were performed in HBS-N buffer (10 mM HEPES pH 7.4, 0.15 M NaCl, 3 mM EDTA, 0.005% v/v surfactant P20, GE Healthcare) using the BIAcore™ T-200 instrument (GE Life Sciences). Anti-mouse Fc CM5 biosensor chip (GE Healthcare) was generated using a standard primary amine coupling protocol. PD-1 mAbs at 0.3 µg/ml were captured on anti-mouse Fc surface for 1 min at 10 µl/min. PD-1/Fc in a serial dilutions from 3.3nM to 120nM was injected over antibody-bound surface for 3 min at 30 µl/min followed by a 10 min dissociation phase. Association rates (K_a or k_{on}) and dissociation rates (K_d or k_{off}) were calculated using the one-to-one Langmuir binding model (BIA Evaluation Software, GE Life Sciences). The equilibrium dissociation constant (K_D) was calculated as the ratio k_{off}/k_{on} .

[0060] As shown in Table 1, both mu326 and mu517, a cognate sequence family member related to mu317, have a sub-nanomolar K_D equaling to 0.324 nM and 0.289 nM, respectively, which is significantly better than that of mu134. The K_{on} rate was similar among the three mAbs listed in Table 1, yet the K_{off} rate was significantly different, much faster dissociation rate was observed in mu134.

Table 1. Binding constant of certain top antibodies

mAbs	K_{on} (M^{-1}, s^{-1})	K_{off} (s)	K_D (M)
mu326	2.4×10^5	7.79×10^{-5}	3.24×10^{-10}
mu517	1.96×10^5	5.66×10^{-5}	2.89×10^{-10}
mu134	1.1×10^5	3.69×10^{-4}	3.35×10^{-9}

Affinity determination of anti-PD-1 Fabs by SPR

[0061] Anti-PD-1 mAbs were converted into Fab version by PCR to fuse the variable regions of heavy and light chains to the N-terminus of human IgG2-CH1 and constant region of kappa chain, respectively, and subcloned in pcDNA3.1 vector (Invitrogen). Both expression vectors were co-expressed in 293-F cells using a transient transfection protocol similar to the transient expression of whole antibodies. Briefly, the Fab kappa chain was PCR amplified and subcloned in pcDNA3.1-based expression vector (Invitrogen, Carlsbad, CA, USA). In a separate plasmid, the heavy chain variable region (VH) together with the CH1 coding sequence from human IgG2 was fused with a C-terminal c-Myc-His8 tag by overlapping PCR, and then subcloned in the expression vector. The C232S and C233S (Kabat residue numbering, Kabat et al. Sequence of proteins of immunologic interest, 5th ed Bethesda, MD, NIH 1991) mutations were introduced in the IgG2 heavy chain to prevent disulfide bond exchange and stabilize human IgG2 in the IgG2-A conformation (Lightle et al. 2010 Protein Sci 19(4): 753-762). Both constructs contained a signal peptide upstream of the Fab mature sequences. Secreted expression of Fab was achieved by co-transfection of above 2 plasmids into 293-F cells and cell culture supernatants were harvested 6-7 days post transfection. His8-tagged Fabs were purified from cell culture supernatants using a Ni-sepharose Fast Flow column (Cat. No. 17531801, GE Life Sciences) followed by size exclusion chromatography using a HiLoad 16/60 Superdex200 column (Cat. No. 17106901, GE Life Sciences). The purified Fabs were concentrated to 0.5-5 mg/mL in PBS and stored in aliquots in -80°C freezer.

[0062] For affinity determinations of anti-PD-1 Fabs, SPR assays were used with the BIAcore™ T-200 instrument (GE Life Sciences). Briefly, human PD-1/His or cynomolgus monkey PD-1/His was coupled to activated CM5 biosensor chips (Cat. No. BR100530, GE Life Sciences) to achieve approximately 100-200 response units (RU), followed by blocking

un-reacted groups with 1M ethanolamine. Fab samples of increasing concentration from 0.12nM to 90nM were injected in the SPR running buffer (10 mM HEPES, 150 mM NaCl, 0.05% Tween20, pH7.4) at 30 μ L/minute, and binding responses on human PD-1/His or monkey PD-1/His were calculated by subtracting of RU from a blank flow-cell. Association rates (k_{on}) and dissociation rates (k_{off}) were calculated using the one-to-one Langmuir binding model (BIA Evaluation Software, GE Life Sciences). The equilibrium dissociation constant (K_d) was calculated as the ratio k_{off}/k_{on} .

[0063] The SPR-determined binding affinities of anti-PD-1 Fabs were listed in Table 18. Each anti-PD-1 Fab bound with high affinity ($K_d = 0.15$ -1 nM) to human PD-1. All Fabs, except 326-3G1, bound with slightly lower but comparable (within 5 fold in K_d) affinities to cynomolgus monkey PD-1.

Example 3. Functional activity of anti-PD-1 antibodies in human T cells.

Generation of stable cell lines

[0064] Retroviral packaging cell line PT67, human T cell lines HuT78 and HEK293 were obtained from the American Type Culture Collection (ATCC, Rockville, MD). A HuT78 subline HuT78/PD-1 that expresses PD-1 was generated by retroviral transduction using pFB-neo vector (Stratagene/Agilent Tech, Santa Clara, CA) containing the *PD-1* gene, according to the protocol described previously (Zhang et al. 2005 Blood 106: 1544-1551). The T cell engager, a membrane-anchored chimeric Ab (OS8), was constructed by fusing the single chain variable fragment (scFv) of an anti-human CD3 mAb OKT3 (Kipriyanov et al. 1997, PEDS 10:445-453) to the C-terminal domain (113-220) of mouse CD8 α (NCBI Accession No: NP_001074579.1) which includes hinge, transmembrane and cytoplasmic domains. By doing so, anti-CD3 scFv is anchored to cell surface as a T cell activator. Human PD-L1, PD-L2 and OS8 cDNAs were sub-cloned into pcDNA3.1 vector. Stable cell lines HEK293/OS8/PD-L1, Hep3B/OS8/PD-L1 and HEK293/OS8/PD-L2 that co-express both OS8 and PD-L1 or PD-L2 cDNAs were generated by co-transfection of HEK293 and Hep3B cells (ATCC) with the paired plasmids, followed by hygromycin or G418 selection for 10-14 days. Cell lines were then cloned by limiting dilution as described previously (Fuller SA, et al. Curr Protoc Mol Biol. Chapter 11:Unit11.8., 2001). Chimeric PD-1 receptor, named P3Z, was constructed by fusing the extracellular and transmembrane domains) of human PD-1 to the cytoplasmic domain of human CD3 ζ chain (NCBI Accession No. NP_932170.1). P3Z-coding cDNA sequence was cloned into pFB-neo and delivered into HuT78 cells via retroviral transduction to generate HuT78/P3Z cells.

Determination of PD-1 antibody functions by IL-2 release in HuT78/PD-1 cells

[0065] To determine whether anti-PD-1 antibodies can block the interaction of PD-L1-induced PD-1 signaling, HuT78/PD-1 cells (1.5x10⁴ cells per well in 96-well plate) were pre-incubated with hybridoma supernatants or PD-1 antibodies for 15 minutes prior to co-culture with HEK293/OS8/PD-L1 or HEK293/OS8/PD-L2 cells (4x10⁴ per well) in a flat bottom plate fed with 200 μ l of RPMI1640 growth medium per well at 37°C. After 16-18 hours, supernatants of the co-culture were collected. IL-2 was assayed by ELISA using human IL-2 Ready-Set-Go! ELISA kits (Cat. No. 88-7025, eBiosciences, San Diego, CA). In this assay, blockade of PD-1 signaling with anti-PD-1 antibodies resulted in enhanced TCR signaling and IL-2 production (Fig. 4).

[0066] As shown in Fig. 5 and Table 2, murine anti-PD-1 mAb, mu317 and mu326, elicited significantly higher functional activity than mu30, inhibiting PD-L1-induced PD-1 signaling which leads to increased IL-2 secretion. Both had higher IL-2 secretion (top line, Table 2), 675 and 634 pg/ml, respectively, and both had lower EC₅₀ (Effective concentration of mAb at 50% level of IL-2 secretion induction) than mu30 antibody.

Table 2. IL-2 release induced by anti-PD-1 mAbs in HuT78/PD-1 cells co-cultured with HEK293/OS8/PD-L1 cells

Antibody	Baseline (pg/ml)	Top line (pg/ml)	EC ₅₀ (μ g/ml)
mu30	95	527	0.229
mu317	95	675	0.083
mu326	95	634	0.053
mIgGs	95	N/A	N/A
Baseline: Average IL-2 release induced by mIgGs at all tested concentrations, see Fig. 4.			
Top line: Highest IL-2 release based on regression calculation by Prism Software, Fig. 4.			
N/A: Not applicable			

[0067] Not only did the engagement of HuT78/PD-1 cells by anti-PD-1 mAbs block PD-L1 induced T-cell activation,

but also blocked PD-L2 induced IL-2 release. Table 3 presented the data showing mu317 and mu326 had much higher potency in activating the T-cells as indicated by the parameters (EC_{50}) of IL-2 secretion than those of mu476 .

Table 3. IL-2 release induced by anti-PD-1 mAbs in HuT78/PD-1 cells co-cultured with HEK293/OS8/PD-L2 cells

Antibody	Baseline (pg/ml)	Top line (pg/ml)	EC_{50} (μ g/ml)
476	180	599	0.183
317	192	563	0.032
326	218	635	0.038
Baseline: Average IL-2 release induced in the lower tail part of the sigmoid reaction curve. Top line: Average IL-2 release induced at the plateau part of the sigmoid reaction curve			

Determination of PD-1 antibody functions by reverse signaling of IL-2 release in HuT78/P3Z cells

[0068] In chimeric receptor P3Z, PD-1 signaling domain was replaced with the cytoplasmic domain of CD3 ζ . Therefore, P3Z mediates activation upon engagement with dPD-L1, rather than inhibition as original PD-1 receptor. In this assay, HuT78/P3Z cells (3×10^4 /well) were pre-incubated with hybridoma supernatants or PD-1 antibodies for 15 minutes prior to co-culture with HEK293/PD-L1 or HEK293/PD-L2 cells (5×10^4 /well) in 96-well flat bottom plates (a total volume of 200 μ l/well) at 37°C. After 16-18 hours, supernatants were collected and IL-2 production was assayed by ELISA as described above.

[0069] The functional activity of murine anti-PD-1 mAbs was further confirmed by direct read-out of T-cell activation in reverse signaling assay described above. Consistent to the result described above, mu317 and mu326 had best functional activity among the mAbs we screened. As shown in Table 4 and Table 5, mu317 and mu326 were much more potent than one of the low activity mAbs, mu37, both in terms of IC_{50} and maximum inhibition.

Table 4. Inhibition of IL-2 secretion by anti-PD-1 mAbs in HuT78/P3Z cells co-cultured with HEK293/PD-L1 cells

Antibody	IC_{50} (μ g/ml)	Max inhibition, %
37	0.287	86.9
317	0.083	99.3
326	0.039	97.6
Maximum inhibition was calculated as percentage (%) of inhibition with anti-PD-1 mAbs added to the highest level of 10 μ g/ml in culture.		

Table 5. Inhibition of IL-2 secretion by anti-PD-1 mAbs in HuT78/P3Z cells co-cultured with HEK293/PD-L2 cells

Antibody	IC_{50} (μ g/ml)	Max inhibition, %
37	0.127	43.3
317	0.020	94.3
326	0.018	93.4
Maximum inhibition was calculated as percentage (%) of inhibition with anti-PD-1 mAbs added to the highest level of 10 μ g/ml in culture.		

Example 4. Activation of IFN- γ secretion by anti-PD-1 mAb in primary human PBMCs co-cultured with HEK293/OS8/PD-L1 cells

[0070] To verify if the selected top mAbs against PD-1 also exert functional effect on primary human immune cells, we assayed the antibody function by using freshly isolated peripheral blood mononuclear cells (PBMCs), which are mainly consisted of T-cells (50-70%), B-cells and NK cells (15-30%), and monocytes (2-10%). Human PBMCs were isolated from healthy donors by density gradient centrifugation using ficoll lymphocyte separation medium (Histopaque-1077; Sigma-Aldrich, MO) according to the manufacturer's instructions. All the human blood collection followed the

Internal Procedure of Beigene. PBMCs were then stimulated with anti-CD3 mAb (40 ng/mL) OKT3 (Cat. No. 16-0037, eBioscience, CA) for 3 days prior to assay. FACS analysis (Example 1) showed that PD-1 expression on the activated PBMCs (primarily T cells) was increased to variable degree dependent on individual donors (Table 6). To determine the response of pre-activated T cells to PD-1 ligand-positive tumor cells upon engagement TCR/CD3 complex, PBMCs (1x10⁴) were co-cultured with either HEK293/OS8/PD-L1 or HEK293/OS8/PD-L2 cells (3x10⁴) in 96-well flat-bottom plates for 15-18 hours. Cell-free supernatants were assayed for IFN- γ level by ELISA using Ready-Set-Go! ELISA kits (Cat. No. 88-7316, eBiosciences), which is the most prominent indicator of T-cell activation, as well as of other immune cell activation (Thakur A. et al. 2012 Vaccine, 30:4907-4920).

PBMCs and treatment	Percent gated PD-1 staining positive cells versus total PMBCs stained	
	Donor-3	Donor-4
PBMCs, not stimulated / stained by PD-1 Ab	12.0%	3.2%
PBMCs, stimulated / stained by PD-1 Ab	40.0%	38.1%
PBMCs, not stimulated / stained by control Ab	$\leq 0.5\%$	$\leq 0.5\%$
PBMCs, stimulated / stained by control Ab	$\leq 0.5\%$	$\leq 0.5\%$
Stimulation: freshly isolated PBMCs were cultured for 3 days in presence of anti-CD3 antibody, OKT3, and IL-2. Without stimulation: fresh PBMCs subjected to antibody staining and FACS analysis.		

[0071] Fig. 6 demonstrated that presence of mAbs mu317 and mu326 in the co-culture of pre-activated PBMCs and HEK293/OS8/PD-L1 cells resulted in increasing IFN- γ accumulation in a dose-dependent manner. Although the base level of IFN- γ with control murine IgG treatment varies among different donors, the increase of IFN- γ secretion in PBMCs treated by mu317 or mu326 is statistically significant in the range of 0.1 to 10 μ g/ml of antibody treatment. Comparing to the corresponding level of mIgG-treated PBMCs, IFN- γ secretion induced by mu317 and mu326 between the 0.1 to 10 μ g/ml concentration levels increased 2.5 to 3.2 fold in PBMCs from Donor-19, and increased 1.4 to 2.3 fold in PBMCs of Donor-20, respectively.

Example 5. Activation of human NK cells by anti-PD1 mAbs

Stable cell lines for functional assay in NK cells

[0072] Primary human NK cells were reported previously to express PD-1 protein in response to IL-2 treatment and inhibiting PD-1-mediated signaling enhanced cytotoxicity of NK cells (2010 Blood, 116: 2286). For quantitative assay of functional effect exerted by anti-PD-1 mAbs in NK cells, human NK cell line NK92MI (ATCC) and lung cancer cell line SK-Mes-1 (ATCC) were engineered to stably express human PD-1 and PD-L1, respectively, by retroviral transduction according to the protocols described previously (Zhang et al. 2005, Blood 106: 1544-1551, Zhang et al. 2006, Cancer Res, 66: 5927). The two stable cell lines were named as NK92MI/PD-1 and SK-Mes-1/PD-L1

Anti-PD-1 Abs promote IFN- γ production and secretion in NK92MI/PD-1 cells

[0073] Functional activity of the anti-PD-1 mAbs on NK cells was assayed by quantitative measurement of IFN- γ production and secretion in NK92MI/PD-1 cells which were co-cultured with lung cancer cell line SK-MES-1/PD-L1 at ratio of 1 to 2 in 96-well flat-bottom plate with total of 6 x 10⁴ cells per well. The anti-PD-1 mAbs were added to NK92MI/PD-1 cells 15 minutes before the co-culture started, then the cells were co-cultured for overnight in CO₂ incubator. Cell-free supernatants were assayed for IFN- γ level by ELISA as described in Example 4.

[0074] All anti-PD-1 mAbs triggered significant increase of IFN- γ production from the baseline with low concentration of antibody treatment to top line with high concentration of antibody treatment. The two top antibodies, mu317 and mu326, had lower EC₅₀, than the comparison antibody 5C, indicating they have more potent activating effect to the NK cells (Table 7).

Table 7. IFN- γ secreted in medium (pg/ml) by NK92MI/PD-1 cells in presence of anti-PD-1 mAb and SK-MES-1/PD-L1 cells

Antibody	Baseline (pg/ml)	Top line (pg/ml)	EC ₅₀ (μ g/ml)
317	28	532	0.40

(continued)

Antibody	Baseline (pg/ml)	Top line (pg/ml)	EC ₅₀ (μg/ml)
326	15	509	0.20
5C	20	535	1.17
Baseline: Average IFN-γ release induced in the lower tail part of the sigmoid reaction curve. Top line: Average IFN-γ release induced at the plateau part of the sigmoid reaction curve			

Anti-PD-1 antibody enhances cancer cell killing mediated by NK92MI/PD-1 cells

[0075] Cytotoxicity of NK92MI/PD-1 cells against SK-MES-1/PD-L1 cells was determined by lactate dehydrogenase (LDH) release assay using the CytoTox 96 Non-Radioactive Cytotoxicity Assay kit (Promega, Madison, WI). In brief, NK92MI/PD-1 cells (10⁵) were pre-incubated with anti-PD-1 mAbs at final concentrations within the range of 0.004-10 μg/ml for 15 minutes, and SK-MES-1/PD-L1 cells (2x10⁴) were added to the immune cell culture in a 96-well V-bottom plate at an effector to tumor cell (E:T) ratio of 5:1, then co-cultured for 5 hours. The complete tumor cell lysis was set as maximum cell killing, the LDH-release assay readout of each sample was calculated as percentage of maximum cell killing. The cell killings (%) of all samples were normalized cross the plates using 10% of baseline as the common standard.

[0076] In the specific cytotoxicity assay set as above, the selected anti-PD-1 mAbs caused a net tumor cell killing (= top line - baseline) ranging from 19% to 20.2% at high concentration of mAb input. Mu317 and mu326 had lower EC₅₀ than mu336, indicating better potency to trigger NK92MI/PD-1 cell-mediated tumor cell killing (Table 8).

Table 8. Cytotoxicity of NK92MI/PD-1 cells towards tumor cells induced by anti-PD-1 mAb

Antibody	Baseline (%)	Top line (%)	EC ₅₀ (μg/ml)
317	10	29.06	0.50
326	10	30.19	0.37
336	10	29.72	1.52
Baseline: Percent of tumor cells killed not due to the effect of anti-PD-1 mAbs, normalized to 10% cross plates. Top line: Average percent of tumor killed in presence of highest concentrations of mAbs, i.e. 3 μg/ml and 10 μg/ml			

Example 6. Cloning and sequence analyses of PD-1 mAbs

[0077] The murine hybridoma clones secreting a specific mAb were cultured to a density of 3 to 10 X 10⁶ cells in a 100mm-tissue culture dish, and the cells were harvested through centrifugation at 1500 rpm in a swing bucket rotor. Total cellular RNA was isolated using Ultrapure RNA kit (Cat. No. CW0581, CWBIOTECH, Beijing, China) following the manufacturer's protocol. The RNA was resuspended in double-deionized water, concentration measured by NanoDrop (ThermoFisher, Shanghai, China).

[0078] PCR primers used for mAb cDNA cloning were synthesized by Invitrogen (Beijing, China) based on the sequences reported previously (Brocks et al. 2001 Mol Med 7:461-469). The 1st strand cDNA was synthesized using reverse transcriptase (Cat. No. AH301-02, Transgen Biotech, Beijing, China). PCR amplification of specific mAb cDNA was performed using PCR reagent kit (Cat. No. Ap221-12, TransGen Biotech, Beijing, China) and following manufacturer's protocol. The PCR product was either directly sequenced by service provider (GeneWiz, Beijing, China) or subcloned into a pCR vector (Invitrogen), subsequently sequenced (GeneWiz).

[0079] The protein sequences of murine mAbs were analyzed by sequence homology alignment. MAbs were grouped based on sequence homology and epitope-mapping results (Example 13). Complement determinant regions (CDRs) were identified based on Kabat (Wu, T.T. and Kabat, E.A., 1970 J. Exp. Med. 132:211-250) and IMGT system (Lefranc M.-P. et al., 1999 Nucleic Acids Research, 27, 209-212) by sequence annotation and by internet-based sequence analysis (http://www.imgt.org/IMGT_vquest,share/textes/index.html and <http://www.ncbi.nlm.nih.gov/igblast/>). As shown in Table 9, the CDRs of mu317 and mu326 are very different in sequence length and identity.

Table 9. CDRs of mu317 and mu326

MAbs	CDR1	SEQ ID NO	CDR2	SEQ ID NO	CDR3	SEQ ID NO
mu317, HC	<u>GFSLT</u> SYGVH	11	VIWAGGSTNYNSALMS	12	<u>ARAYGNYWYIDV</u>	13
mu317, LC	KASQSVSNDVA	14	<u>YAFHRFT</u>	15	<u>HQAYSSPYT</u>	16
mu326, HC	<u>GYTFTNYGMN</u>	17	<u>WINNNNGEPTYAEEFKG</u>	18	<u>ARDVMDY</u>	19
mu326, LC	RASESVDNYGYSFMH	20	<u>RASNLES</u>	21	<u>QQSKEYPT</u>	22
Note: CDRs in bold face are based on Kabat system; CDRs underlined are based IMGT system.						

Example 7. Humanization of the murine mAbs*Simulation of antibody 3D structure*

[0080] The three dimensional structures were simulated for variable domains of mu317 and mu326 in order to identify framework residues that might be important for supporting CDR loop structures. Potentially important framework residues were kept as the original murine residues in the first round antibody humanization. The previously established structural modeling method for antibodies (Morea et al. Methods 2000 20:267-279) was adopted to simulate 3D structure of anti-PD-1 mAbs based on the known canonical structures of antibodies (Al-Lazikani et al. 1997 Journal of Molecular Biology 273:927-948). Briefly, the sequence of each variable domain (Vk and Vh) of murine antibody was blasted in the PDB database (Protein Data Bank, <http://blast.ncbi.nlm.nih.gov/>) to identify the most homologous antibody sequence with known high resolution structure (resolution less than 2.5 angstrom). Selected structure templates for modeling mu317 and mu326 (listed in Table 10) had the same classes of canonical loop structures in L-CDR1, L-CDR2, L-CDR3, H-CDR1, and H-CDR2 to the target antibodies to be modeled. If the templates for the Vk and the Vh came from different immunoglobulins, they were packed together by a least-squares fit of the main chain atoms to form a hybrid structure of Vk-Vh interface residues, which was used as the templates for structural homology modeling by Swiss-model program (Kiefer et al. 2009 Nucleic Acids Research 37, D387-D392). Certain side chain conformation was adjusted while the main chain conformations were retained. At the sites where the parent structure and the modeled structure had the same residue, the side chain conformation was retained. At sites where the residues were different, side chain conformations were modeled on the basis of template structure, rotamer libraries and packing considerations. After homology modeling, PLOP program (Jacobson et al. 2002 Journal of Physical Chemistry 106:11673-11680) was used to refine the homology models to minimize all-atom energy and optimize Vk and Vh interface. This step was performed to improve the stereochemistry, especially in those regions where segments of structures coming from different antibodies had been joined together.

Table 10. Structure templates used in antibody structure simulations

Antibody chain	PDB code of template structure (PDB template for H-CDR3)	Sequence identity	Sequence similarity
mu317 Vk	3MXV	87%	92%
mu317 Vh	3VFG	83%	91%
mu326 Vk	1EJO	92%	94%
mu326 Vh	1NCA	88%	90%
317-1 Vk	4HJJ	90%	95%
317-1 Vh	3VFG (1AY1)	75%	87%
326-1 Vk	1EJO	87%	92%
326-1 Vh	3T2N (3CXD)	84%	86%

[0081] The structures were also simulated for CDR-grafted 317-1 and 326-1 in order to guide further rounds of antibody

engineering to enhance the extents of humanization and/or enhance antibody stabilities. The selected structure templates are also listed in Table 10. The structure simulations were done in a similar way to above procedure, except that the possible conformations of H-CDR3 were taken from PDB templates 1AY1 for 317-1 and 3CXD for 326-1, respectively, which contained H-CDR3s of similar size and torso region. Energy minimization for grafted H-CDR3 residues was done using PLOP.

Humanization

[0082] For humanization of the anti-PD-1 mAbs, we searched human germline IgG genes homologous to the cDNA sequences of mu317 and mu326 variable regions by blasting the human immunoglobulin gene database in IMGT (http://www.imgt.org/IMGT_vquest/share/textes/index.html) and NCBI (<http://www.ncbi.nlm.nih.gov/igblast/>) websites. The human IGVH and IGVK with high homology to the PD-1 mAbs were selected as the template for humanization.

[0083] Humanization was carried out in principle by CDR-grafting. In the 1st round of humanization, mutations from murine to human amino acid residues in framework sequences of variable regions was guided by the simulated 3D structures, and only the murine amino acid residues whose changes retain the overall antibody and CDR loop structure were mutated to human sequence as described above. The initial versions of humanized mAbs were hu317-1 (SEQ NO 47-50) and hu326-1 (SEQ NO 55-58), which comprise a heavy chain with humanized variable heavy chain (Vh) fused to human IgG2 constant region (NCBI accession No. P01859) and a light chain with humanized variable light chain kappa (Vk) fused to human Ig kappa C-region (NCBI Accession No. P01834). Likewise, we generated chimeric antibodies from mu317 and mu326, which are consisted of a murine VH fused to human IgG2 constant region and a murine VK fused to human Ig kappa C-region. The full chimeric antibodies were named as ch317 and ch326, respectively. All recombinant mAbs were expressed and purified as described in Example 1.

[0084] FACS and functional assays demonstrated that mAb hu317-1 almost retained the same binding and functional activity as the mu317 and ch317. The EC₅₀ difference in FACS analysis between mu317 versus ch317 and hu317-1 may be interpreted by the fact that two different detection antibodies, a goat anti-mouse IgG and a goat anti-human IgG, were used in FACS. In the two functional assays, all three versions of 317 were treated more equal, and the results also close to each other (Table 11).

[0085] As result of the initial round of humanization for mu326, mAb hu326-1 retained similar functional feature to the parental ch326 and mu326 although functional activity in FACS binding assay and in HuT78/PD-1 cell-based IL-2 release assay may be slightly weaker than ch326 (Table 12).

Table 11. Comparison of mu317, ch317 and hu317-1 by FACS and functional

assays

Assay/Parameter		mu317	ch317	hu317-1
FACS	EC ₅₀ (μg/ml)	0.11	0.36	0.46
	Top MFI*	205	217	203
Assay-1	EC ₅₀ (μg/ml)	0.11	0.08	0.09
	Top line (pg/ml)	346	294	386
	Baseline (pg/ml)	98	82	91
Assay-2	IC ₅₀ (μg/ml)	0.11	0.10	0.11
	Max inhibition	99.5%	99.0%	99.8%
*MFI: mean fluorescence intensity from FACS analysis				
Assay-1: IL-2 release induced by the mAbs in HuT78/PD-1 cells co-cultured with HEK293/OS8/PD-L1 cells				
Assay-2: IL-2 release induced by the mAbs in HuT78/P3Z cells co-cultured with HEK293/PD-L1 cells				

Table 12. Comparison of mu317, ch317 and hu317-1 by FACS and functional assays

Assay/Parameter		mu326	ch326	hu326-1
FACS	EC ₅₀ (μg/ml)	0.126	0.072	0.117
	Top MFI	195	163	129
Assay-1	EC ₅₀ (μg/ml)	0.038	0.074	0.112
	Top line (pg/ml)	1149	1057	1143
	Baseline (pg/ml)	242	250	283
Assay-2	IC ₅₀ (μg/ml)	0.14	0.12	0.10
	Max inhibition	96.9%	81.0%	84.4%
Assay-1: IL-2 release induced by the mAbs in HuT78/PD-1 cells co-cultured with HEK293/OS8/PD-L1 cells				
Assay-2: IL-2 release induced by the mAbs in HuT78/P3Z cells co-cultured with HEK293/PD-L1 cells				

[0086] Based on the 1st round of humanization, we further mutated the other murine amino acid (AA) residues in the framework (FR) of hu317-1_Vh and _Vκ individually to assess the impact on the antibody function. As shown in Table 13, the seven individual mutations in Vh and one mutation in VK of hu317-1 all have similar functional activities. Only minor changes were observed in some Vh mutation, such as hu317-2_K71V with slightly weaker inhibitory function among the mutations. However, when all the murine amino acid residues mutated together to human (hu317-3A), the function is clearly weaker than the rest mutations in FACS and IL-2 release assays.

[0087] In the initial trial described above, hu326-1 reached significant humanization level in the FR except for a few of murine AA residues left. Yet, it has weaker function than the mu326. Therefore, we made more individual mutations either back to murine residues or forward to human residues to explore the contribution of each individual AA to mAb326 function. Table 14 presented all single AA mutations made based on hu326-1_Vh template (SEQ NO 56, SEQ NO 57) and their functional assay results. Majority of the mutations showed better functional activity than those of hu326-1, matching the original mu326 mAb. A couple of mutations (E46K and F95Y) showed slightly less potency in the EC₅₀ or IC₅₀, indicating the role of those residues in the antibody structure and function.

Table 13. Comparison of functional activity of Fabs with humanization mutations in hu317-1 framework

Fab and composition		FACS, EC ₅₀	IL-2 release in HuT78/P3Z	
Vh	Vκ		Max inhibition, %	EC ₅₀
hu317-1_Vh	hu317-1_Vκ	0.19	98.78	0.30
hu317-2_L48I	hu317-1_Vκ	0.14	98.51	0.37
hu317-2_L67V	hu317-1_Vκ	0.15	98.57	0.30
hu317-2_K71V	hu317-1_Vκ	0.18	96.55	0.48
hu317-2_N73T	hu317-1_Vκ	0.15	98.29	0.31
hu317-2_S76N	hu317-1_Vκ	0.13	98.56	0.28
hu317-2_V78F	hu317-1_Vκ	0.18	98.03	0.38
hu317-2_M82L	hu317-1_Vκ	0.13	98.47	0.27
hu317-1_Vh	HU317-2_G100Q	0.21	98.86	0.27
hu317-3A	hu317-1_Vκ	0.32	79.66	0.35

(continued)

Fab and composition		FACS, EC ₅₀	IL-2 release in HuT78/P3Z	
V _h	V _κ		Max inhibition, %	EC ₅₀

Note: Unit for is μg/ml; mutated amino acid residue numbering is same as in the listed sequences for hu317-1; hu317-3A has all the framework sequence mutated to human.

Table 14. Comparison of functional activity of mAbs with mutations in hu326-1 framework

Antibody	FACS, EC ₅₀ μg/ml	IL-2 release in HuT78/P3Z		IL-2 release in HuT78/PD-1	
		Max inhibition, %	IC ₅₀ , μg/ml	Top line, pg/ml	EC ₅₀ , μg/ml
ch326	0.118	93.05	0.074	993	0.135
hu326-1	0.317	92.38	0.087	987	0.213
hu326-2 S9P ^B	0.145	96.04	0.075	1022	0.136
hu326-2 A16E ^B	0.155	96.33	0.078	1048	0.126
hu326-2 E46K ^B	0.132	95.25	0.079	1244	0.259
hu326-2 G63D ^B	0.139	96.44	0.064	1069	0.120
hu326-2 A76V ^F	0.102	96.65	0.071	1002	0.112
hu326-2 S84N ^B	0.131	96.52	0.060	1015	0.126
hu326-2 S85N ^B	0.110	95.62	0.093	932	0.104
hu326-2 T88N ^B	0.098	95.85	0.102		
hu326-2 F95Y ^F	0.097	95.62	0.166	1028	0.135

^B: Back mutation to murine amino acid; ^F: Forward mutation to human amino acid.
All of the mutations were made in hu326-1_Vh (SEQ NO 56), which were paired with hu326-1_Vk (SEQ NO 58).

[0088] To explore the best possible Vh and VK sequence composition for mAbs 317 and 326 that could be used as therapeutics in human, we made a variety of combination mutations (including some mutations in the CDR sequences) in considerations of the antibody features, such as humanization level in FR, functional activities, physicochemical properties, antibody-dependent cell-mediated cytotoxicity (ADCC) and complement-dependent cytotoxicity (CDC). Most of the mutations were deemed not passing the qualification standards. Through the engineering process, six of the humanized, recombinant mAbs were selected for their potential therapeutic utility: hu317-4B2 (SEQ ID NO 43-44), hu317-4B5 (SEQ ID NO 45-46), hu317-4B6 (SEQ ID NO 23-26), hu326-3B1 (SEQ ID NO 51-52), hu326-3G1 (SEQ ID NO 53-54) and hu326-4A3 (SEQ ID NO 27-30). The CDRs of the mAb were compared to those of original murine antibodies, shown in Table 15 and Table 16.

[0089] Among the six mAbs, hu317-4B2, hu317-4B5 and hu317-4B6 are closely related to each other in sequences and very similar in their functional activities and strength. On the other hand, hu326-3B1, hu326-3G1 and hu326-4A3 are quite close to each other in sequences and functionalities (Table 17-18). Within each of the two groups of mAbs, they also shared many other features in addition to sequences and function, such as physicochemical properties and binding epitopes (described in Examples 10 and 11) though some minor differences do exist.

Table 15. Comparison of CDRs among different versions of mAbs 317

mAbs	CDR1	SEQ ID NO	CDR2	SEQ ID NO	CDR3	SEQ ID NO
mu317, HC	GFSLTSYGVH	11	VIWAGGSTNYNSALMS	12	ARAYGNYWYIDV	13
hu317-1, HC	GFSLTSYGVH	11	VIWAGGSTNYN <u>PSLKS</u>	59	ARAYGNYWYIDV	13
hu317-4B2, HC	GFSLTSYGVH	11	VI <u>Y</u> AGGSTNYN <u>PSLKS</u>	60	ARAYGNYWYIDV	13

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(continued)

mAbs	CDR1	SEQ ID NO	CDR2	SEQ ID NO	CDR3	SEQ ID NO
hu317-4B5, HC	GFSLTSYGVH	11	VIYAGGSTNYN <u>PSLKS</u>	60	ARAYGNYWYIDV	13
hu317-4B6, HC	GFSLTSYGVH	11	VIY <u>AD</u> GSTNYN <u>PSLKS</u>	32	ARAYGNYWYIDV	13
mu317, LC	KASQSVSNDVA	14	YAFHRFT	15	HQAYSSPYT	16
hu317-1, LC	KASQSVSNDVA	14	YAFHRFT	15	HQAYSSPYT	16
hu317-4B2, LC	K <u>SS</u> <u>ES</u> SVSNDVA	61	YAFHRFT	15	HQAYSSPYT	16
hu317-4B5, LC	K <u>SS</u> <u>ES</u> SVSNDVA	61	YAFHRFT	15	HQAYSSPYT	16
hu317-4B6, LC	K <u>SS</u> <u>ES</u> SVSNDVA	61	YAFHRFT	15	HQAYSSPYT	16
Note: AA residues underlined are changed from murine sequence to human antibody sequences or for improvement of physicochemical properties.						

Table 16. Comparison of CDRs among different versions of mAbs 326

mAbs	CDR1	SEQ ID NO	CDR2	SEQ ID NO	CDR3	SEQ ID NO
mu326, HC	GYTFTNYGMN	17	WINNNNGEPTYAEEFKG	18	ARDVMDY	19
hu326-1, HC	GYTFTNYGMN	17	WINNNNGEPTYA <u>QG</u> FRG	62	ARDVMDY	19
hu326-3B1, HC	GYTFTNYGMN	17	WINNNNGEPTYA <u>QD</u> FRG	62	ARDVMDY	19
hu326-3G1, HC	GYTFTNYGMN	17	WINNNNGEPTYA <u>QD</u> FRG	62	ARDVMDY	19
hu326-4A3, HC	GYTFTNYGMN	17	WINNNNA <u>E</u> PTYA <u>QD</u> FRG	38	ARDVMDY	19
mu326, LC	RASESVDNYGYFSMH	20	RASNLES	21	QQSKEYPT	22
hu326-1, LC	RASESVDNYGYFSMH	20	RASNLES	21	QQSKEYPT	22
hu326-3B1, LC	RASESVDNYGYFSMH	20	RASNLES	21	QQSKEYPT	22
hu326-3G1, LC	RASESVDNYGYFSMH	20	RASNLES	21	QQSKEYPT	22
hu326-4A3, LC	RASESVDNYGYFSMH	20	RASNLES	21	QQSKEYPT	22
Note: AA residues underlined are changed from murine sequence to human antibody sequences or for improvement of physicochemical properties.						

Table 17. Binding activities of humanized mAbs assayed by ELISA and FACS

mAbs	ELISA, EC ₅₀ µg/ml	FACS, EC ₅₀ µg/ml
hu317-4B2	0.066	0.129*
hu317-4B5	0.057	0.115*

(continued)

mAbs	ELISA, EC ₅₀ µg/ml	FACS, EC ₅₀ µg/ml
hu317-4B6	0.061	0.092*
hu326-3B1	0.092	0.165
hu326-3G1	0.088	0.190
hu326-4A3	0.091*	0.142*
* FACS data by using Fab version of antibodies without normalization. ** Data from bridging study and normalized.		

Table 18. Binding affinity of Fabs assayed by SPR

Fab	K _{on} (M ⁻¹ , S ⁻¹)	K _{off} (s)	K _D (M)
hu317-4B5	3.89 x 10 ⁵	9.07 x 10 ⁻⁵	2.33 x 10 ⁻¹⁰
hu317-4B6	5.71 x 10 ⁵	8.37 x 10 ⁻⁵	1.47 x 10 ⁻¹⁰
hu326-3B1	2.18 x 10 ⁵	1.90 x 10 ⁻⁴	8.70 x 10 ⁻¹⁰
hu326-3G1	2.00 x 10 ⁵	2.01 x 10 ⁻⁴	1.00 x 10 ⁻⁹

Affinity determination of humanized anti-PD-1 Fabs by SPR

[0090] Anti-PD-1 mAbs were converted into Fab version by PCR to fuse the variable regions of heavy and light chains to the N-terminus of human IgG2-CH1 and constant region of kappa chain, respectively, and subcloned in pcDNA3.1 vector (Invitrogen). Both expression vectors were co-expressed in 293-F cells using a transient transfection protocol similar to the transient expression of whole antibodies. Briefly, the Fab kappa chain was PCR amplified and subcloned in pcDNA3.1-based expression vector (Invitrogen, Carlsbad, CA, USA). In a separate plasmid, the heavy chain variable region (VH) together with the CH1 coding sequence from human IgG2 was fused with a C-terminal c-Myc-His8 tag by overlapping PCR, and then subcloned in the expression vector. The C232S and C233S (Kabat residue numbering, Kabat et al. Sequence of proteins of immunologic interest, 5th ed Bethesda, MD, NIH 1991) mutations were introduced in the IgG2 heavy chain to prevent disulfide bond exchange and stabilize human IgG2 in the IgG2-A conformation (Lightle et al. 2010 Protein Sci 19(4): 753-762). Both constructs contained a signal peptide upstream of the Fab mature sequences. Secreted expression of Fab was achieved by co-transfection of above 2 plasmids into 293-F cells and cell culture supernatants were harvested 6-7 days post transfection. His8-tagged Fabs were purified from cell culture supernatants using a Ni-sepharose Fast Flow column (Cat. No. 17531801, GE Life Sciences) followed by size exclusion chromatography using a HiLoad 16/60 Superdex200 column (Cat. No. 17106901, GE Life Sciences). The purified Fabs were concentrated to 0.5-5 mg/mL in PBS and stored in aliquots in -80°C freezer.

[0091] For affinity determinations of anti-PD-1 Fabs, SPR assays were used with the BIAcore™ T-200 instrument (GE Life Sciences). Briefly, human PD-1/His or cynomolgus monkey PD-1/His was coupled to activated CM5 biosensor chips (Cat. No. BR100530, GE Life Sciences) to achieve approximately 100-200 response units (RU), followed by blocking un-reacted groups with 1M ethanolamine. Fab samples of increasing concentration from 0.12nM to 90nM were injected in the SPR running buffer (10 mM HEPES, 150 mM NaCl, 0.05% Tween20, pH7.4) at 30µL/minute, and binding responses on human PD-1/His or monkey PD-1/His were calculated by subtracting of RU from a blank flow-cell. Association rates (k_{on}) and dissociation rates (k_{off}) were calculated using the one-to-one Langmuir binding model (BIA Evaluation Software, GE Life Sciences). The equilibrium dissociation constant (K_D) was calculated as the ratio k_{off}/k_{on} .

[0092] The SPR-determined binding affinities of anti-PD-1 Fabs were listed in Table 18. Each anti-PD-1 Fab bound with high affinity (K_D = 0.15-1 nM) to human PD-1. All Fabs, except 326-3G1, bound with slightly lower but comparable (within 5 fold in K_D) affinities to cynomolgus monkey PD-1.

Example 8. Generation and expression of **recombinant anti-PD-1** mAbs with **modified human IgG4 constant region**

[0093] Since PD-1 is primarily expressed in activated T cells, PD-1 blocking antibodies linked to naturally occurring type of IgG- Fc moieties are expected to induce Fc-mediated effector functions, such as ADCC and CDC, to a variable degree depending on the IgG subclasses, which results in elimination of activated T cells (Natsume A, et al, 2009 Drug

Des Devel Ther. 3: 7-16). Human antibody subclass IgG4 was shown in many previous reports that it has modest ADCC and almost no CDC effector function (Moore GL, et al. 2010 MAbs, 2:181-189). On the other hand, natural IgG4 was found less stable in stress conditions such as in acidic buffer or under increasing temperature (Angal, S. 1993 Mol Immunol, 30:105-108; Dall'Acqua, W. et al, 1998 Biochemistry, 37:9266-9273; Aalberse et al. 2002 Immunol, 105:9-19). In order to spare PD-1⁺ T cells from being killed and to improve physicochemical properties of the anti-PD-1 antibodies, the humanized mAbs were linked to IgG4 engineered by combinations of mutations to have reduced or null FcγR binding or C1q binding activities, therefore, attenuating or eliminating ADCC and CDC effector functions. Considering physicochemical properties of antibody as a biological drug, one of the less desirable, intrinsic properties of IgG4 is dynamic separation of its two heavy chains in solution to form half antibody, which lead to bi-specific antibodies generated *in vivo* via a process called "Fab arm exchange" (Van der Neut Kolfshoten M, et al. 2007 Science, 317:1554-157). The mutation of serine to proline at position 228 (EU numbering system) appeared inhibitory to the IgG4 heavy chain separation (Angal, S. 1993 Mol Immunol, 30:105-108; Aalberse et al. 2002 Immunol, 105:9-19). Some of the amino acid residues in the hinge and γFc region were reported to have impact on antibody interaction with Fcγ receptors (Chappel SM, et al. 1991 Proc. Natl. Acad. Sci. USA, 88:9036-9040; Mukherjee, J. et al., 1995 FASEB J, 9:115-119; Armour, K.L. et al., 1999 Eur J Immunol, 29:2613-2624; Clynes, R.A. et al., 2000 Nature Medicine, 6:443-446; Arnold J.N., 2007 Annu Rev Immunol, 25:21-50). Furthermore, some rarely occurring IgG4 isoforms in human population may also elicit different physicochemical properties (Brusco, A. et al. 1998 Eur J Immunogenet, 25:349-55; Aalberse et al. 2002 Immunol, 105:9-19). However, lumping all the mutations and isoforms previously discovered into a specific antibody does not warrant for an ideal antibody molecule to share all the features for therapeutics such as described above, which may be resulted from contradictory effect of the combined mutations and from impact of variable region to the effector function and physicochemical properties of an antibody (Igawa T. et al., 2010 Prot Eng Design Select, 23:385-392; Perchiacca J.M. and Tessier P.M., 2012 Ann Rev Biomol Eng 3:263-286).

[0094] To generate anti-PD-1 mAbs with least ADCC, CDC and instability, we modified the hinge and γFc region of human IgG4 by introduce a number of combinations of mutations, which created IgG4mt1 to IgG4mt12. Some of the modified IgG4 variants were clearly less desirable as indicated by our assay results, several relevant IgG4 variants and modified sequences were listed in Table 19. The assessment of these antibodies is described herein.

Table 19. Sequence modifications of IgG4 variants

IgG4 and variants	Amino acid residues*																
	...	228	229	230	231	232	233	234	235	236	...	265	...	309	...	409	...
IgG4	...	S	C	P	A	P	E	F	L	G	...	D	...	L	...	R	...
IgG4mt1	...	<u>P</u>	C	P	A	P	E	F	L	G	...	D	...	L	...	R	...
IgG4mt2	...	<u>P</u>	C	P	A	P	<u>P</u>	<u>V</u>	<u>A</u>	G	...	D	...	L	...	R	...
IgG4mt6	...	<u>P</u>	C	P	A	P	<u>P</u>	<u>V</u>	<u>A</u>	G	...	<u>A</u>	...	L	...	R	...
IgG4mt8	...	<u>P</u>	C	P	A	P	<u>P</u>	<u>V</u>	<u>A</u>	G	...	<u>T</u>	...	L	...	R	...
IgG4mt9	...	<u>P</u>	C	P	A	P	<u>P</u>	<u>V</u>	<u>A</u>	G	...	<u>A</u>	...	L	...	<u>K</u>	...
IgG4mt10	...	<u>P</u>	C	P	A	P	<u>P</u>	<u>V</u>	<u>A</u>	G	...	<u>A</u>	...	<u>V</u>	...	<u>K</u>	...
* Amino acid numbering is based on EU system. Changes are highlighted by underline.																	

Example 9. IgG4mt10 has no FcγR binding, lowest ADCC and CDC effector function

[0095] ADCC is initiated when an antibody binds to cell surface target protein followed by ligation to Fcγ receptors (FcγRs) expressed on effector cells. It was well documented that human IgG1 has significantly higher binding affinity to FcγRs than IgG2 and IgG4, specially, binding to FcγR-I and PcγR-IIIA, which correlated to the strength of IgG1 to activate ADCC. Reminiscent of ADCC, CDC is activated when an antibody cross-links a cell surface target and C1q protein, which followed by a cascade reaction of complement complex formation and target cell lysis. As proxy of ADCC and CDC, assays for antibody binding to FcγRs and C1q may serve as the fundamental indicator of ADCC and CDC. We therefore systematically assessed the mAbs binding to all the major FcγRs.

FcγR binding

[0096] Binding of various IgG4 mutants to FcγRs was determined by flow cytometry. In brief, a series of HEK293 transfectants expressing human FcγRs were established. These transfectants expressed FcγRI, FcγRIIA, FcγRIIB or

Fc γ RIIIA. Multi-subunit Fc γ Rs (i.e., Fc γ RI and Fc γ RIIIA) were co-expressed with FcR γ . Polymorphic variants (i.e., Fc γ RIIA H131 and R131, Fc γ RIIA F158 and V158) were also included. A secondary antibody (goat anti-human IgG F(ab)²-Alexa Fluor 488, Jackson ImmunoResearch, West Grove, PA, USA) was used to detect the binding of anti-PD-1 mAbs with modified IgG4 variants (Table 19) to Fc γ R⁺ HEK293 cells. As expected, anti-PD-1 mAbs in IgG1 format (hu317-1/IgG1 and hu317-4B6/IgG1) bind strongly to all Fc γ Rs including Fc γ RI, Fc γ RIIA (H131 and R131 alleles), Fc γ RIIB, and Fc γ RIIIA (V158 and F158 alleles) (Table 20). Interestingly, when the two different version of humanization mAbs, hu317-1 and hu317-4B6 (with differences in both Vh and VK), were generated in the same IgG4 variant format, such as either in IgG4mt1 or in IgG4mt6 format, their binding strength (MFI) vary by an range from a couple fold to close to 100 fold (e.g. 455.2/115.7 = 3.9 fold; 13.6/1.0 = 13.6 fold; 434.6/4.9 = 88.7 fold; and etc., see Table 20). It is consistent to the previous findings by other that the variable regions of antibodies do have significant impact on the binding to FcRs, therefore, exerting the impact on effector function such as ADCC (Igawa T. et al., 2010 Prot Eng Design Select, 23:385-392; Perchiacca J.M. and Tessier P.M., 2012 Ann Rev Biomol Eng 3:263-286).

[0097] As demonstrated in Table 20, when hu317-4B6 and hu326-4A3 were made in IgG4mt10 format, they have the lowest binding activity to Fc γ Rs among the PD-1 mAbs and IgG variant formats listed in the table, as well as many other humanization mAbs and IgG formats we have tested in the study. The uniqueness of hu317-4B6 and hu326-4A3 in IgG4mt10 format in this regard may not be extended to the same family of humanization mAbs with somewhat distant sequence homology, such as hu317-1, as described above.

Table 20. Binding strength (MFI*) of anti-PD-1 mAbs to Fc γ Rs determined by FACS

mAbs	Fc γ RI	Fc γ RIIA (H131)	Fc γ RIIA (R131)	Fc γ RIIB	Fc γ RIIA (F158)	Fc γ RIIA (V158)
hu317-1 /IgG1	2152.9	168.7	139.6	442.4	99.7	277.2
hu317-4B6 /IgG1	2771.7	1.7	0.6	1.9	28.0	293.7
hu317-1 /IgG4mt1	455.2	21.3	21.9	434.6	0.6	20.7
hu317-4B6 /IgG4mt1	115.7	0.2	0.0	4.9	0	6.1
hu317-1 /IgG4mt6	13.6	1.0	0.8	1.8	0.9	1.1
hu317-4B6 /IgG4mt6	1.0	0	0	0	0	0
hu317-4B6 /IgG4mt10	0.4	0	0	0	0	0
hu326-4A3 /IgG4mt10	0.5	0	0	0	0	0
* MFI: mean fluorescence intensity from FACS analysis						

ADCC

[0098] Classical ADCC involves activation of NK cells by antibodies engaging to Fc γ RIIIA or CD16. To verify whether humanized anti-PD-1 mAbs induce ADCC, NK92MI/CD16V cells, which were generated from NK92MI cells (ATCC) by co-transducing expression plasmids containing CD16 (V158 allele) and FcR γ genes, were used as effector cells, and PD-1-expressing T cell line, HuT78/PD-1, was used as target cells. NK92MI/CD16V cells (4x10⁴) were co-cultured with equal number of HuT78/PD-1 cells in 96-well V-bottom plates for 5h. Cytotoxicity was determined by LDH release assay described in previous section. The results confirmed that hu317-4B2/IgG4mt6, hu317-4B6/IgG4mt6, hu317-4B6/IgG4mt10 and hu326-4A3/IgG4mt10 all have base level of ADCC comparing to the positive controls (Fig. 7). The minor difference in ADCC between those 4 mAbs may be attributable to experimental error (see error bars in Fig. 7).

CDC

[0099] Human IgG4 antibodies, in general, do not induce any CDC via classical pathway. Whether anti-PD-1 mAbs in IgG4mt10 format will trigger CDC was evaluated using a PD-1-expressing T cell line, HuT78/PD-1, and fresh human serum from healthy donors. Cell lysis by CDC was determined by Celltiter glo assay kits (Promega, Beijing, China). In brief, HuT78/PD-1 cells (2x10⁴) were incubated in serum-free RPMI1640 (Invitrogen) with anti-PD-1 Abs (10 μ g/ml) at 37°C for 15 minutes before adding normal human serum (NHS) to the final concentration of 15% or 50% in 96-well flat-bottom plates in a total volume of 120 μ l. After overnight incubation at 37°C, cells were lysed and assayed for ATP concentration. To test whether humanized anti-PD-1 mAbs in IgG4mt10 can kill PD-1⁺ primary T cells via CDC, PBMCs isolated from healthy donors were pre-activated with anti-CD3 Ab OKT3 (40 ng/ml) for 3 days before co-culture with

anti-PD-1 Abs plus NHS. The amount of ATP is directly proportional to the number of cells present in culture. Fluorescence was read using a 96-well fluorometer (PHERA Star FS, BMG LABTECH). The results are expressed in relative fluorescence units (RFU) that are proportional to the number of viable cells. The percent CDC activity was calculated as follows: % CDC activity = $[(\text{RFU test} - \text{RFU background}) / (\text{RFU at total cell lysis} - \text{RFU background})] \times 100$. In general, we were not able to detect any ADCC mediated by anti-PD-1 mAbs in IgG4mt10 format that bind to activated PBMCs. In hypersensitive experimental conditions, such as using PD-1 highly-expressing cell line, high serum and antibody concentration, we detected very low level of CDC in some occasions, and there is not much differences between different versions and anti-PD-1 mAbs, indicating the anti-PD-1 mAbs in IgG4 variant formats retained the feature of low or no CDC activity as the common form of IgG4.

Example 10. Humanized anti-PD-1 mAbs in IgG4mt10 format have enhanced stability under stress conditions

Stability of anti-PD-1 antibodies in high temperature and acidic conditions

[0100] Anti-PD-1 antibodies used in stability studies were all purified from protein A column followed by size exclusion chromatography (SEC) as described in previous sections. Following purification, the aggregate contents of purified antibody samples were monitored in analytical size exclusion chromatography-high performance liquid chromatography (SEC-HPLC), which fell within the range of 0%-0.5%.

[0101] For SEC-HPLC analysis, the antibody samples were analyzed using a TSKgel G3000 SWXL column (7.8x300 mm, Cat. No. 08541, Tosoh Bioscience, Shanghai, China) under isocratic elution condition (elution buffer 0.2 M sodium phosphate, pH7.2), and subsequent detection at UV-215 nm. In each run, 10 microliters of antibody sample was loaded onto the column and eluted at a flow rate of 1mL/minute. The dimer or larger aggregate species of antibody were separated from monomeric species and the percentages of dimers and aggregates were determined based on the integrated peak areas from UV traces.

[0102] For speed-enhanced shelf stability study, anti-PD-1 antibodies (10-40mg/mL in PBS) were kept in incubators at 40-50°C for 4-7 days in order to test the stability of antibodies in high temperature condition. The antibody samples were then analyzed for heat-induced formation of dimer and aggregates in SEC-HPLC. For each of the anti-PD-1 antibodies analyzed, less than 2% became higher molecular weight species (dimers and aggregates), indicating the anti-PD-1 antibodies had good stability in high temperature conditions.

[0103] Antibody's stability in acidic condition has been a key challenge in the downstream manufacturing process (Liu et al. 2010 mAbs 2:480-499). Antibody elution from protein A and inactivation of virus usually require incubation of antibody in low pH (2.5-4) conditions. However, such acidic conditions could potentially cause antibody denaturation and aggregation. Human IgG4 has been known to be less stable than IgG1 and IgG2 (2002 Immunology 105:9). Therefore, we assayed the humanized mAbs made with various IgG4 mutant forms. Briefly, Antibody stabilities in low pH conditions were studied by 1:1 volume of each antibody sample (10 mg/mL in PBS) mixed with low pH buffers containing 50 mM sodium citrate, 100mM NaCl at pH3.6, 3.3, 3.0 or 2.7, respectively. After 1 hour incubation at room temperature, the antibody samples in low pH conditions were neutralized by 1:5 dilution into SEC-HPLC elution buffer containing 0.2M sodium phosphate, pH7.2. SEC-HPLC analyses were done as described above and percentages of dimers and aggregates induced by low pH conditions were quantified. The anti-PD-1 mAb 317-4B6 in IgG1 format was most stable in bioprocessing-relevant acidic conditions even when pH value get as low as 2.7. Among the anti-PD-1 mAbs made in several IgG4 variants, hu317-4B6/IgG4mt10 and hu326-4A3/IgG4mt10 were the most stable under the acidic buffer condition (Table 21) as the acid-induced aggregates were significantly reduced to a level that was comparable to that of the IgG1 format of anti-PD-1 mAbs, 317-4B6 and 326-4A3, i.e. the soluble aggregate is less than 2% (Table 21).

Table 21. Dimer and soluble aggregates formed in acidic buffers and assayed by SEC-HPLC

anti-PD-1 mAbs	% of dimer and aggregates				
	pH7.2	pH3.6	pH3.3	pH3.0	pH2.7
317-4B6/IgG1	0.0%	0.0%	0.2%	0.1%	0.2%
317-4B6/IgG4mt1	0.0%	1.0%	11.0%	49.0%	48.0%
317-4B6/IgG4mt3	0.0%	13.0%	31.0%	>50%	>50%
317-4B6/IgG4mt6	0.0%	4.0%	41.0%	>50%	>50%
317-4B6/IgG4mt9	0.0%	0.5%	2.1%	3.3%	2.0%
317-4B6/IgG4mt10	0.0%	0.2%	0.6%	0.6%	1.4%

(continued)

anti-PD-1 mAbs	% of dimer and aggregates				
	pH7.2	pH3.6	pH3.3	pH3.0	pH2.7
326-4A3/IgG4mt10	0.0%	0.0%	0.4%	0.5%	1.2%

Example 11. Mapping the binding epitopes of anti-PD-1 mAbs

[0104] Previous reports about the crystal structures of PD-1/PD-L1 and PD-1/PD-L2 complexes had shed light to understanding critical amino acid (AA) residues on PD-1 which are required for the ligand-binding (Zhang et al. 2004 Immunity, 20:337-347; Lin D.Y. et al. 2008 PNAS 105:3011-3016; Lazar-Molnar E. et al. 2008 PNAS, 105:10483-10488). In fact, six of such AA residues were identified on the receptor through point mutation analysis required for PD-L1 binding. Five of the six AA residues were also required for PD-L2 binding (Lin D.Y. et al. 2008 PNAS 105:3011-3016). Based on the information from the structure-guided mutation analysis we hypothesized that most effective way for functional mAbs to block PD-1 mediated signaling is to compete with PD-1 ligands by binding to the six critical AA residues, therefore, occupying the binding epitopes required for the ligand binding. To explore the hypothesis and to understand the mechanism of action by functional PD-1 antibodies, we have made six mutants of PD-1 by replacing each of the six critical AAs to Ala, individually, i.e. K45A, I93A, L95A, P97A, I101A and E103A (AA residue numbering based on Lin D.Y. et al. 2008 PNAS 105:3011-3016). The mutant PD-1/Fc and PD-1/His (Fig. 1) were used as templates for PCR-guided mutagenesis or rolling-circle mutagenesis using Fast Mutagenesis System (Cat. No. FM111, Transgen Biotech, Beijing, China). All mutants were sub-cloned in our pcDNA-based expression vectors, and verified by sequencing. The mutated and wild-type PD-1 proteins were expressed by transient transfection (described in Example 1), and prepared after 4 to 6 days of culture. The conditioned media (CM) were analyzed by Western blot to verify the PD-1 protein expression in terms of quality and quantity. The supernatants (CM), after clearing cell debris, were directly used in ELISA analysis or Western blot for epitope-mapping.

[0105] To study the binding epitopes of humanized anti-PD-1 mAbs, ELISA assays using the wild-type (WT) and mutant (Mt) PD-1 were performed to assess the binding activities of hu317-4B5, hu317-4B6, hu326-3B1 and hu326-4A3. For comparison to check the uniqueness of the antibody binding signature, two reference antibody (Reference Ab-1 and Reference Ab-2 from US8008449B2 and US8168757B2, respectively) were included in the study. Equal volume of CM containing WT or Mt PD-1 was coated in 96-well plate for all mAbs in the same ELISA assay. All ELISA results were normalized using the mean ELISA readings of WT PD-1 binding signals as the standard. ELISA binding signals to a specific Mt PD-1 were further normalized against the highest antibody binding read-out (set as 100%) to the specific Mt PD-1. For convenience of data analysis, When a mAb's ELISA binding signal for a specific mutant dropped below 50% relative to WT PD-1, it is defined that the amino acid residue is a significant binding epitope because whose mutation significantly abrogated the antibody binding. Likewise, if a mAb's ELISA binding signal for a specific mutant dropped below 25%, it is defined as very significant. As shown in Fig. 8, two of the critical AA residues in PD-1, K45 and I93, are significant or very significant epitopes for mAbs hu317-4B5 and hu317-4B6 binding, and three AA residues, I93, L95 and P97, are either significant or very significant epitopes for hu326-3B1 and hu326-4A3. On the other hand, the two reference antibodies have distinctive binding epitopes, P97 is significant for Reference Ab-1, while L95 and P97 are significant for Reference Ab-2.

[0106] Interestingly, when the PD-1 protein is denatured in Western Blot, mAb hu317-4B5 and -4B6 were still capable of binding to WT PD-1 though the critical binding epitopes (K45 and I93) are not close to each other (non-linear). It indicated that the PD-1 protein became renatured to some degree after denaturation in SDS-PAGE of Western Blot process, which allows the anti-PD-1 mAbs to recognize and bind to it. Taking the advantage of this observation, we performed Western Blot analysis for all six antibodies used in above ELISA study. The overall results from Western Blot corroborated very well to the ELISA results, i.e. the significant or very significant epitopes, whose mutations resulted in low binding signals in ELISA, also gave weakest Western Blot band comparing to the binding to other mutant PD-1 (Fig. 8). Some minor differences between ELISA and Western Blot were also observed, e.g., the ELISA binding signals on I93A and E103A by reference Ab-2 were relatively stronger than those in Western Blot. It may be indicative of that those AA residues may also contribute to the binding because whose mutations impacted the binding though only under stress condition (i.e. denaturation or losing native conformation). As summarized in Table 22, the anti-PD-1 mAbs in this invention have identifiable binding epitopes differing from other anti-PD-1 antibody.

Table 22. Summary* of key epitopes by anti-PD-1 mAbs

	K45A	I93A	L95A	P97A	I101A	E103A
hu317-4B5	***	**				
hu317-4B6	***	**				
hu326-3B1		**	**	**		
hu326-4A3		***	**	**		
Ref. Ab-1				**		
Ref. Ab-2			**	**		
* based on Fig.8						

Example 12. Anti-PD-1 mAbs activate primary human PBMCs and inhibit tumor growth in xenograft mouse models

Humanized anti-PD-1 mAbs activate human PBMCs

[0107] Throughout the humanization processes, the humanized anti-PD-1 mAbs at various stages retained similar functional activities as assessed by ELISA, FACS and immune cell-based cytokine release assays. To confirm the function of final versions of humanized mAbs, we assayed the activating functions of hu317-4B5, hu317-4B6, hu326-3B1 and hu326-4A3 using primary human PBMCs. The results demonstrated that those mAbs throughout the humanization maintained the original murine mAb functions to activate primary PBMCs although the degree of activation differs among the four donors due to the variance of individual's genetic background (Fig. 9).

Humanized anti-PD-1 mAbs enhance NK cell-based cytotoxicity against cancer cells

[0108] Reminiscent of the original murine mAbs, the humanized anti-PD-1 mAbs, hu317-4B5, hu317-4B6, hu326-3B1 and hu326-3G1, enhance NK92MI/PD-1 cell-mediated cytotoxicity against the target lung cancer cells, SK-MES-1/PD-L1, in a dose-dependent manner (Fig. 10, Table 23). It appeared evident that in principle the humanized anti-PD-1 mAbs might function to break immune cell tolerance mediated by PD-1 signaling, enhancing the cancer killing activity by immune cells, e.g. NK cells and cytotoxic T-lymphocytes.

Humanized anti-PD-1 mAb activates human PBMCs and inhibits tumor growth in a mouse xenograft cancer model in vivo

[0109] All above experimental evidences indicated that the anti-PD-1 mAbs might work in mouse cancer models utilizing immune-compromised mice xenografted with human cancer cells, subsequently implanting human PBMCs and applying the mAb treatment to inhibit cancer cell growth *in vivo*. The experiment was designed as follows. Seven-eight week old SCID-male mice (Vital River Laboratories, China) were inoculated subcutaneously at right flank with 3×10^6 Hep3B/OS8-PD-L1 cells in 50% Matrigel (BD Biosciences, New Jersey, USA). Fifteen days after tumor inoculation, the mice bearing tumor size between 100-250 mm³ were randomized and divided into three treatment groups. One hundred microliters of pooled PBMCs (5×10^5) from 2 healthy donors were injected intratumorally. Three days post PBMC-implanting, anti-PD-1 antibodies (Hu317-IgG4mt2) and human IgG were administered via s.c. at a dose of 10 mg/kg, respectively. Antibody treatment was repeated once every 10 days for a total of 3 times. PBS was injected in a parallel group as negative control. Tumors were measured twice a week using a caliper starting on day 7. Tumor volumes were calculated using the following formula: $[D \times (d^2)]/2$, in which D represents the large diameter of the tumor, and d represents the small diameter. All animal studies were performed following Beigene Animal Care and Use Procedure.

[0110] In the *in vivo* study, although 60% of tumors in the control groups were auto-regressed, the rest of *in vivo* experiment is still quite informative, which were presented in Fig. 11. In the control groups, either vehicle-treated or human IgG (hulgG)-treated group, each has 40% tumors (2 of 5 mice) outgrowing larger than the baseline at starting point. The two tumors in PBS-treated group grew much larger (above 2,000 mm³, one tumor-bearing mouse was terminated earlier due to passing tumor size limit by protocol). The two tumors in hulgG-treated group grew to the size of 800 and 1,370 mm³, significantly above the average baseline of 164 mm³ though smaller than the PBS-treated tumors. On the other hand, in the anti-PD-1 mAb (hu317-1/IgG4mt2)-treated group, tumors were either completely regressed or close to baseline size (one tumor = 200 mm³, which grew back slowly after regressed to 50% of baseline at two weeks from PBMC implanting). The results indicated that the anti-PD-1 mAb described above can activate human immune cells inhibiting tumor cells growth in the mouse *in vivo* cancer model, which is consistent to the *in vitro* experimental

results described above.

SEQUENCE LISTING

5 **[0111]**

<110> BeiGene
 Li, Kang
 Zhang, Tong
 10 Song, Jing
 Xu, Lanlan
 Liu, Qi
 Peng, Hao

15 <120> ANTI-PD1 ANTIBODIES AND THEIR USE AS THERAPEUTICS AND DIAGNOSIS

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gagagcttcg tgctaaactg gtaccgcatg agccccagca accagacgga caagctggcc 180

35 gccttccccg aggaccgcag ccagcccggc caggactgcc gcttccgtgt cacacaactg 240

cccaacgggc gtgacttcca catgagcgtg gtcagggccc ggcgcaatga cagcggcacc 300

40 tacctctgtg gggccatctc cctggccccc aaggcgcaga tcaaagagag cctgcgggca 360

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aggccagccg gccagttcca aacc 444

45 <210> 2
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 <213> Homo sapiens

50 <400> 2

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Pro Gly Trp Phe Leu Asp Ser Pro Asp Arg Pro Trp Asn Pro Pro Thr
1 5 10 15

Phe Ser Pro Ala Leu Leu Val Val Thr Glu Gly Asp Asn Ala Thr Phe
20 25 30

Thr Cys Ser Phe Ser Asn Thr Ser Glu Ser Phe Val Leu Asn Trp Tyr
35 40 45

Arg Met Ser Pro Ser Asn Gln Thr Asp Lys Leu Ala Ala Phe Pro Glu
50 55 60

Asp Arg Ser Gln Pro Gly Gln Asp Cys Arg Phe Arg Val Thr Gln Leu
65 70 75 80

Pro Asn Gly Arg Asp Phe His Met Ser Val Val Arg Ala Arg Arg Asn
85 90 95

Asp Ser Gly Thr Tyr Leu Cys Gly Ala Ile Ser Leu Ala Pro Lys Ala
100 105 110

Gln Ile Lys Glu Ser Leu Arg Ala Glu Leu Arg Val Thr Glu Arg Arg
115 120 125

Ala Glu Val Pro Thr Ala His Pro Ser Pro Ser Pro Arg Pro Ala Gly
130 135 140

Gln Phe Gln Thr
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<212> DNA
<213> Mus musculus

<400> 3

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ccaggaaagg gactggaatg gctgggagta atatgggccc gtggaagcac aaattataat 180
tcggctctca tgtccagact gagcatcagc aaagacaact ccaggagcca agttttctta 240
agaatgaaca gtctgcaaac tgatgacaca gccatgtact actgtgccag agcctatggt 300
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<212> PRT

<213> Mus musculus

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1 5 10 15

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Asn Leu Ser Ile Thr Cys Thr Val Ser Gly Phe Ser Leu Thr Ser Tyr
20 25 30

15

Gly Val His Trp Ile Arg Gln Pro Pro Gly Lys Gly Leu Glu Trp Leu
35 40 45

Gly Val Ile Trp Ala Gly Gly Ser Thr Asn Tyr Asn Ser Ala Leu Met
50 55 60

20

Ser Arg Leu Ser Ile Ser Lys Asp Asn Ser Arg Ser Gln Val Phe Leu
65 70 75 80

25

Arg Met Asn Ser Leu Gln Thr Asp Asp Thr Ala Met Tyr Tyr Cys Ala
85 90 95

30

Arg Ala Tyr Gly Asn Tyr Trp Tyr Ile Asp Val Trp Gly Ala Gly Thr
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Thr Val Thr Val Ser Ser
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35

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gggcagtctc ctaaactgct gataaactat gcatttcac gcttcactgg agtccctgat 180

50

cgtttcactg gcagtggata tgggacggat ttcattttca ccatcagcac tgtgcaggct 240

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<212> PRT

<213> Mus musculus

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				20					25					30			
15	Val	Ala	Trp	Tyr	Gln	Gln	Lys	Pro	Gly	Gln	Ser	Pro	Lys	Leu	Leu	Ile	
			35					40					45				
20	Asn	Tyr	Ala	Phe	His	Arg	Phe	Thr	Gly	Val	Pro	Asp	Arg	Phe	Thr	Gly	
		50					55					60					
25	Ser	Gly	Tyr	Gly	Thr	Asp	Phe	Ile	Phe	Thr	Ile	Ser	Thr	Val	Gln	Ala	
30																	
	65					70					75					80	
35	Glu	Asp	Leu	Ala	Val	Tyr	Phe	Cys	His	Gln	Ala	Tyr	Ser	Ser	Pro	Tyr	
					85					90					95		
40	Thr	Phe	Gly	Gly	Gly	Thr	Lys	Leu	Glu	Met	Lys						
				100					105								

<210> 7

<211> 342

<212> DNA

<213> Mus musculus

<400> 7

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	tcctgcaagg	cttctgggta	taccttcaca	aactatggaa	tgaactgggt	gaagcaggct	120
	ccaggaaagg	gtttaaagtg	gatgggctgg	ataaacaata	ataatggaga	gccaacatat	180
45	gctgaagagt	tcaagggacg	gtttgccttc	tctttggaaa	cctctgccag	cactgcctat	240
	ttgcagatca	acaacctcaa	aatgaggac	acggctacat	atttctgtgc	aagagatgtt	300
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<210> 8

<211> 114

<212> PRT

<213> Mus musculus

<400> 8

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5	Thr	Val	Lys	Ile	Ser	Cys	Lys	Ala	Ser	Gly	Tyr	Thr	Phe	Thr	Asn	Tyr
				20					25					30		
10	Gly	Met	Asn	Trp	Val	Lys	Gln	Ala	Pro	Gly	Lys	Gly	Leu	Lys	Trp	Met
			35					40					45			
15	Gly	Trp	Ile	Asn	Asn	Asn	Asn	Gly	Glu	Pro	Thr	Tyr	Ala	Glu	Glu	Phe
	50						55					60				
20	Lys	Gly	Arg	Phe	Ala	Phe	Ser	Leu	Glu	Thr	Ser	Ala	Ser	Thr	Ala	Tyr
	65					70					75					80
25	Leu	Gln	Ile	Asn	Asn	Leu	Lys	Asn	Glu	Asp	Thr	Ala	Thr	Tyr	Phe	Cys
					85					90					95	
30	Ala	Arg	Asp	Val	Met	Asp	Tyr	Trp	Gly	Gln	Gly	Thr	Ser	Val	Thr	Val
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Ser Ser

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 <211> 330
 <212> DNA
 <213> Mus musculus

35 <400> 9

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	cagcagaaac	caggacagcc	acccaactc	ctcatctatc	gtgcatccaa	cctagaatct	180
	gggatccctg	ccaggttcag	tggcagtggg	tctaggacag	gcttcaccct	caccattaat	240
45	cctgtggagg	ctgatgatgt	tgcaacctat	tactgtcagc	aaagtaaaga	atatccgacg	300
	ttcgggtggag	gcaccaagct	ggaagtcaaa				330

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 <212> PRT
 <213> Mus musculus

55 <400> 10

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5	Gln	Arg	Ala	Thr	Ile	Ser	Cys	Arg	Ala	Ser	Glu	Ser	Val	Asp	Asn	Tyr	
				20					25					30			
10	Gly	Tyr	Ser	Phe	Met	His	Trp	Tyr	Gln	Gln	Lys	Pro	Gly	Gln	Pro	Pro	
			35					40					45				
15	Gln	Leu	Leu	Ile	Tyr	Arg	Ala	Ser	Asn	Leu	Glu	Ser	Gly	Ile	Pro	Ala	
		50					55					60					
20	Arg	Phe	Ser	Gly	Ser	Gly	Ser	Arg	Thr	Gly	Phe	Thr	Leu	Thr	Ile	Asn	
	65					70					75				80		
25	Pro	Val	Glu	Ala	Asp	Asp	Val	Ala	Thr	Tyr	Tyr	Cys	Gln	Gln	Ser	Lys	
					85					90					95		
30	Glu	Tyr	Pro	Thr	Phe	Gly	Gly	Gly	Thr	Lys	Leu	Glu	Val	Lys			
				100					105					110			
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	<213> Mus musculus																
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	<211> 16																
40	<212> PRT																
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	1				5					10				15			
50	<210> 13																
	<211> 12																
	<212> PRT																
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55	<400> 13																
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	1				5					10							

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<210> 14
 <211> 11
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 <213> Mus musculus
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 Lys Ala Ser Gln Ser Val Ser Asn Asp Val Ala
 10 1 5 10

 <210> 15
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 <213> Mus musculus
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 <400> 15

 Tyr Ala Phe His Arg Phe Thr
 20 1 5

 <210> 16
 <211> 9
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 25
 <400> 16

 His Gln Ala Tyr Ser Ser Pro Tyr Thr
 30 1 5

 <210> 17
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 Gly Tyr Thr Phe Thr Asn Tyr Gly Met Asn
 1 5 10

 <210> 18
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 1 5 10 15
 55
 Gly

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<210> 19
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 <400> 19

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 1 5
 10

 <210> 20
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 <213> Mus musculus
 15
 <400> 20

 Arg Ala Ser Glu Ser Val Asp Asn Tyr Gly Tyr Ser Phe Met His
 1 5 10 15
 20

 <210> 21
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 <400> 21

 30
 Arg Ala Ser Asn Leu Glu Ser
 1 5

 <210> 22
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 <400> 22

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 Gln Gln Ser Lys Glu Tyr Pro Thr
 1 5

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 <210> 23
 <211> 354
 <212> DNA
 <213> Artificial Sequence

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 <220>
 <223> 317-4B6 cDNA-Vh

 <400> 23
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acctgcactg tctctgggtt ttcattaacc agctatggtg tacactggat ccggcagccc 120
5 ccaggaagg gactggagtg gatcggggtc atatacgccg atggaagcac aaattataat 180
ccctccctca agagtcgagt gaccatatca aaagacacct ccaagaacca ggtttccctg 240
aagctgagct ctgtgaccgc tgcggacacg gccgtgtatt actgtgagag agcctatggt 300
10 aactactggt acatcgatgt ctggggccaa gggaccacgg tcaccgtctc ctca 354

<210> 24

<211> 118

<212> PRT

<213> Artificial Sequence

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<223> 317-4B6 pro-Vh

<400> 24

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20 25 30
30
Gly Val His Trp Ile Arg Gln Pro Pro Gly Lys Gly Leu Glu Trp Ile
35 40 45
35
Gly Val Ile Tyr Ala Asp Gly Ser Thr Asn Tyr Asn Pro Ser Leu Lys
50 55 60
40
Ser Arg Val Thr Ile Ser Lys Asp Thr Ser Lys Asn Gln Val Ser Leu
65 70 75 80
40
Lys Leu Ser Ser Val Thr Ala Ala Asp Thr Ala Val Tyr Tyr Cys Ala
85 90 95
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Arg Ala Tyr Gly Asn Tyr Trp Tyr Ile Asp Val Trp Gly Gln Gly Thr
100 105 110
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Thr Val Thr Val Ser Ser
115

<210> 25

<211> 321

<212> DNA

<213> Artificial Sequence

<220>

<223> 317-4B6 cDNA-Vk

EP 3 044 234 B9

<400> 25

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ggacagcctc ctaagctgct cattaactat gcatttcata gcttcactgg ggtccctgac      180
10 cgattcagtg gcagcgggta tgggacagat ttcactctca ccatcagcag cctgcaggct      240
gaagatgtgg cagtttatta ctgtcaccag gcttatagtt ctccgtacac gtttggccag      300
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 <211> 107
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 <213> Artificial Sequence

20 <220>
 <223> 317-4B6 pro-Vk

<400> 26

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    Glu Arg Ala Thr Ile Asn Cys Lys Ser Ser Glu Ser Val Ser Asn Asp
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    Val Ala Trp Tyr Gln Gln Lys Pro Gly Gln Pro Pro Lys Leu Leu Ile
    35          40             45

    Asn Tyr Ala Phe His Arg Phe Thr Gly Val Pro Asp Arg Phe Ser Gly
    50          55             60

    Ser Gly Tyr Gly Thr Asp Phe Thr Leu Thr Ile Ser Ser Leu Gln Ala
40          65             70             75             80

    Glu Asp Val Ala Val Tyr Tyr Cys His Gln Ala Tyr Ser Ser Pro Tyr
45          85             90             95

          Thr Phe Gly Gln Gly Thr Lys Leu Glu Ile Lys
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50 <210> 27
 <211> 342
 <212> DNA
 <213> Artificial Sequence

55 <220>
 <223> 326-4A3 cDNA-Vh

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<400> 27

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 cccggccagg gcctgaagtg gatgggctgg atcaacaaca acaacgccga gccacacctac 180
 10 gccaggact tcagaggcag attcgtgttc agcctggaca ccagcgccag caccgcctac 240
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 atggactact ggggccaggg caccctggtg accgtgagca gc 342

15 <210> 28
 <211> 114
 <212> PRT
 <213> Artificial Sequence

20 <220>
 <223> 326-4A3 pro-Vh

<400> 28

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30 Ser Val Lys Val Ser Cys Lys Ala Ser Gly Tyr Thr Phe Thr Asn Tyr
 20 25 30

Gly Met Asn Trp Val Arg Gln Ala Pro Gly Gln Gly Leu Lys Trp Met
 35 40 45

Gly Trp Ile Asn Asn Asn Asn Ala Glu Pro Thr Tyr Ala Gln Asp Phe
 50 55 60

40 Arg Gly Arg Phe Val Phe Ser Leu Asp Thr Ser Ala Ser Thr Ala Tyr
 65 70 75 80

45 Leu Gln Ile Ser Ser Leu Lys Thr Glu Asp Thr Ala Val Tyr Tyr Cys
 85 90 95

Ala Arg Asp Val Met Asp Tyr Trp Gly Gln Gly Thr Leu Val Thr Val
 100 105 110

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Ser Ser

55 <210> 29
 <211> 330
 <212> DNA
 <213> Artificial Sequence

EP 3 044 234 B9

<220>

<223> 326-4A3 cDNA-Vk

<400> 29

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10 cagcagaaac caggacaacc tcctaaactc ctgatttacc gtgcatccaa cctagaatct 180
ggggtcccag ccaggttcag cggcagtggg tctgggaccg atttcaccct cacaattaat 240
cctgtggaag ctgaggatac tgcaaattat tactgtcagc aaagtaaaga atatccgacg 300
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<210> 30

<211> 110

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<212> PRT

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<223> 326-4A3 pro-Vk

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Gln Arg Ala Thr Ile Thr Cys Arg Ala Ser Glu Ser Val Asp Asn Tyr
20 25 30

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Gly Tyr Ser Phe Met His Trp Tyr Gln Gln Lys Pro Gly Gln Pro Pro
35 40 45

40

Lys Leu Leu Ile Tyr Arg Ala Ser Asn Leu Glu Ser Gly Val Pro Ala
50 55 60

45

Arg Phe Ser Gly Ser Gly Ser Gly Thr Asp Phe Thr Leu Thr Ile Asn
65 70 75 80

Pro Val Glu Ala Glu Asp Thr Ala Asn Tyr Tyr Cys Gln Gln Ser Lys
85 90 95

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Glu Tyr Pro Thr Phe Gly Gly Gly Thr Lys Val Glu Ile Lys

100

105

110

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<210> 31

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<212> PRT

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<213> Mus musculus

<400> 31

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<210> 32

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<211> 16

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<223> 317-4B6 H-CDR2 or CDR-H2

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<212> PRT

<213> Mus musculus

<400> 33

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<212> PRT

<213> Mus musculus

<400> 35

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Tyr Ala Phe His Arg Phe Thr
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<210> 36
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 <210> 37
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 15 <213> Mus musculus

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 Gly Tyr Thr Phe Thr Asn Tyr Gly Met Asn
 20 1 5 10

 <210> 38
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 25 <212> PRT
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 <223> 326-4A3 H-CDR2 or CDR-H2
 30 <400> 38

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 Gly

 40 <210> 39
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 45 <400> 39

 Ala Arg Asp Val Met Asp Tyr
 50 1 5

 <210> 40
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5 <210> 41
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15 <210> 42
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20 <400> 42

Gln	Gln	Ser	Lys	Glu	Tyr	Pro	Thr
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<220>
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35 <400> 43

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	1				5					10					15	
5	Thr	Leu	Ser	Leu	Thr	Cys	Thr	Val	Ser	Gly	Phe	Ser	Leu	Thr	Ser	Tyr
				20					25					30		
10	Gly	Val	His	Trp	Ile	Arg	Gln	Pro	Pro	Gly	Lys	Gly	Leu	Glu	Trp	Ile
			35					40					45			
15	Gly	Val	Ile	Tyr	Ala	Gly	Gly	Ser	Thr	Asn	Tyr	Asn	Pro	Ser	Leu	Lys
		50					55					60				
20	Ser	Arg	Val	Thr	Ile	Ser	Lys	Asp	Thr	Ser	Lys	Asn	Gln	Phe	Ser	Leu
	65					70					75					80
25	Lys	Leu	Ser	Ser	Val	Thr	Ala	Ala	Asp	Thr	Ala	Val	Tyr	Tyr	Cys	Ala
					85					90					95	
30	Arg	Ala	Tyr	Gly	Asn	Tyr	Trp	Tyr	Ile	Asp	Val	Trp	Gly	Gln	Gly	Thr
				100					105					110		
35	Thr	Val	Thr	Val	Ser	Ser										
				115												
40	<210> 44															
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5	Glu	Arg	Ala	Thr	Ile	Asn	Cys	Lys	Ser	Ser	Glu	Ser	Val	Ser	Asn	Asp
				20					25					30		
10	Val	Ala	Trp	Tyr	Gln	Gln	Lys	Pro	Gly	Gln	Pro	Pro	Lys	Leu	Leu	Ile
			35					40					45			
15	Asn	Tyr	Ala	Phe	His	Arg	Phe	Thr	Gly	Val	Pro	Asp	Arg	Phe	Ser	Gly
		50					55					60				
20	Ser	Gly	Tyr	Gly	Thr	Asp	Phe	Thr	Leu	Thr	Ile	Ser	Ser	Leu	Gln	Ala
	65					70					75				80	
25	Glu	Asp	Val	Ala	Val	Tyr	Tyr	Cys	His	Gln	Ala	Tyr	Ser	Ser	Pro	Tyr
					85					90					95	
30	Thr	Phe	Gly	Gln	Gly	Thr	Lys	Leu	Glu	Ile	Lys					
				100					105							

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<212> PRT

<213> Artificial Sequence

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<223> 317-4B5 pro-Vh

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40	Thr	Leu	Ser	Leu	Thr	Cys	Thr	Val	Ser	Gly	Phe	Ser	Leu	Thr	Ser	Tyr
				20					25					30		
45	Gly	Val	His	Trp	Ile	Arg	Gln	Pro	Pro	Gly	Lys	Gly	Leu	Glu	Trp	Ile
			35					40					45			
50	Gly	Val	Ile	Tyr	Ala	Gly	Gly	Ser	Thr	Asn	Tyr	Asn	Pro	Ser	Leu	Lys
		50					55					60				

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	Ser	Arg	Val	Thr	Ile	Ser	Lys	Asp	Thr	Ser	Lys	Asn	Gln	Val	Ser	Leu
	65					70					75					80
5	Lys	Leu	Ser	Ser	Val	Thr	Ala	Ala	Asp	Thr	Ala	Val	Tyr	Tyr	Cys	Ala
					85					90					95	
10	Arg	Ala	Tyr	Gly	Asn	Tyr	Trp	Tyr	Ile	Asp	Val	Trp	Gly	Gln	Gly	Thr
				100					105					110		
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30	Glu	Arg	Ala	Thr	Ile	Asn	Cys	Lys	Ser	Ser	Glu	Ser	Val	Ser	Asn	Asp
				20					25					30		
35	Val	Ala	Trp	Tyr	Gln	Gln	Lys	Pro	Gly	Gln	Pro	Pro	Lys	Leu	Leu	Ile
			35					40					45			
40	Asn	Tyr	Ala	Phe	His	Arg	Phe	Thr	Gly	Val	Pro	Asp	Arg	Phe	Ser	Gly
	50						55					60				
45	Ser	Gly	Tyr	Gly	Thr	Asp	Phe	Thr	Leu	Thr	Ile	Ser	Ser	Leu	Gln	Ala
	65					70					75					80
50	Glu	Asp	Val	Ala	Val	Tyr	Tyr	Cys	His	Gln	Ala	Tyr	Ser	Ser	Pro	Tyr
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<400> 47

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ccaggaagg gactggagtg gctgggggtc atatgggccg gtggaagcac aaattataat      180
10 ccctccctca agagtcgact gaccatatca aaagacaact ccaagagcca ggtttccctg      240
aagatgagct ctgtgaccgc tgcggacacg gccgtgtatt actgtgcgag agcctatggt      300
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<400> 48

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```

30      Thr Leu Ser Leu Thr Cys Thr Val Ser Gly Phe Ser Leu Thr Ser Tyr
      20              25              30

```

```

35      Gly Val His Trp Ile Arg Gln Pro Pro Gly Lys Gly Leu Glu Trp Leu
      35              40              45

```

```

40      Gly Val Ile Trp Ala Gly Gly Ser Thr Asn Tyr Asn Pro Ser Leu Lys
      50              55              60

```

```

45      Ser Arg Leu Thr Ile Ser Lys Asp Asn Ser Lys Ser Gln Val Ser Leu
      65              70              75              80

```

```

50      Lys Met Ser Ser Val Thr Ala Ala Asp Thr Ala Val Tyr Tyr Cys Ala
      85              90              95

```

```

55      Arg Ala Tyr Gly Asn Tyr Trp Tyr Ile Asp Val Trp Gly Gln Gly Thr
      100              105              110

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      Thr Val Thr Val Ser Ser
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atcaactgca aggccagcca gagtgtgagt aatgatgtag cttggtacca gcagaaacca      120
10 ggacagcctc ctaagctgct cattaactat gcatttcata gtttcaactgg ggtccctgac      180
cgattcagtg gcagcgggta tgggacagat ttcactctca ccatcagcag cctgcaggct      240
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<400> 50

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```

```
Glu Arg Ala Thr Ile Asn Cys Lys Ala Ser Gln Ser Val Ser Asn Asp
          20           25           30
```

35

```
Val Ala Trp Tyr Gln Gln Lys Pro Gly Gln Pro Pro Lys Leu Leu Ile
          35           40           45
```

40

```
Asn Tyr Ala Phe His Arg Phe Thr Gly Val Pro Asp Arg Phe Ser Gly
          50           55           60
```

45

```
Ser Gly Tyr Gly Thr Asp Phe Thr Leu Thr Ile Ser Ser Leu Gln Ala
65           70           75           80
```

```
Glu Asp Val Ala Val Tyr Tyr Cys His Gln Ala Tyr Ser Ser Pro Tyr
          85           90           95
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50

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Thr Phe Gly Gly Gly Thr Lys Leu Glu Ile Lys
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<400> 51

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10

Ser Val Lys Val Ser Cys Lys Ala Ser Gly Tyr Thr Phe Thr Asn Tyr
20 25 30

15

Gly Met Asn Trp Val Arg Gln Ala Pro Gly Gln Gly Leu Lys Trp Met
35 40 45

20

Gly Trp Ile Asn Asn Asn Asn Gly Glu Pro Thr Tyr Ala Gln Asp Phe
50 55 60

Arg Gly Arg Phe Val Phe Ser Leu Asp Thr Ser Ala Ser Thr Ala Tyr
65 70 75 80

25

Leu Gln Ile Ser Ser Leu Lys Thr Glu Asp Thr Ala Val Tyr Tyr Cys
85 90 95

30

Ala Arg Asp Val Met Asp Tyr Trp Gly Gln Gly Thr Leu Val Thr Val
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<400> 52

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5	Gln	Arg	Ala	Thr	Ile	Thr	Cys	Arg	Ala	Ser	Glu	Ser	Val	Asp	Asn	Tyr	
				20					25					30			
10	Gly	Tyr	Ser	Phe	Met	His	Trp	Tyr	Gln	Gln	Lys	Pro	Gly	Gln	Pro	Pro	
			35					40					45				
15	Lys	Leu	Leu	Ile	Tyr	Arg	Ala	Ser	Asn	Leu	Glu	Ser	Gly	Val	Pro	Ala	
		50					55					60					
20	Arg	Phe	Ser	Gly	Ser	Gly	Ser	Arg	Thr	Asp	Phe	Thr	Leu	Thr	Ile	Asn	
	65					70					75					80	
25	Pro	Val	Glu	Ala	Asn	Asp	Thr	Ala	Asn	Tyr	Tyr	Cys	Gln	Gln	Ser	Lys	
					85					90					95		
30	Glu	Tyr	Pro	Thr	Phe	Gly	Gly	Gly	Thr	Lys	Val	Glu	Ile	Lys			
				100					105					110			

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<400> 53

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	1				5					10					15		
5	Ser	Val	Lys	Val	Ser	Cys	Lys	Ala	Ser	Gly	Tyr	Thr	Phe	Thr	Asn	Tyr	
				20					25					30			
10	Gly	Met	Asn	Trp	Val	Arg	Gln	Ala	Pro	Gly	Gln	Gly	Leu	Lys	Trp	Met	
			35					40					45				
15	Gly	Trp	Ile	Asn	Asn	Asn	Asn	Gly	Glu	Pro	Thr	Tyr	Ala	Gln	Asp	Phe	
		50					55					60					
20	Arg	Gly	Arg	Phe	Val	Phe	Ser	Leu	Asp	Thr	Ser	Ala	Ser	Thr	Ala	Tyr	
	65					70					75					80	
25	Leu	Gln	Ile	Ser	Ser	Leu	Lys	Thr	Glu	Asp	Thr	Ala	Val	Tyr	Tyr	Cys	
					85					90					95		
30	Ala	Arg	Asp	Val	Met	Asp	Tyr	Trp	Gly	Gln	Gly	Thr	Leu	Val	Thr	Val	
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	Gln	Arg	Ala	Thr	Ile	Thr	Cys	Arg	Ala	Ser	Glu	Ser	Val	Asp	Asn	Tyr	
				20					25					30			
5	Gly	Tyr	Ser	Phe	Met	His	Trp	Tyr	Gln	Gln	Lys	Pro	Gly	Gln	Pro	Pro	
			35					40					45				
10	Lys	Leu	Leu	Ile	Tyr	Arg	Ala	Ser	Asn	Leu	Glu	Ser	Gly	Val	Pro	Ala	
		50					55					60					
15	Arg	Phe	Ser	Gly	Ser	Gly	Ser	Arg	Thr	Asp	Phe	Thr	Leu	Thr	Ile	Asn	
	65					70					75					80	
20	Pro	Val	Glu	Ala	Asn	Asp	Thr	Ala	Asn	Tyr	Tyr	Cys	Gln	Gln	Ser	Lys	
					85					90					95		
25	Glu	Tyr	Pro	Thr	Phe	Gly	Gly	Gly	Thr	Lys	Val	Glu	Ile	Lys			
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35	agctgcaagg	ccagcggcta	caccttcacc	aactacggca	tgaactgggt	gagacaggcc											120
	cccggccagg	gcctggagtg	gatgggctgg	atcaacaaca	acaacggcga	gccacctac											180
	gccaggggct	tcagaggcag	attcgtgttc	agcctggaca	ccagcgccag	caccgcctac											240
40	ctgcagatca	gcagcctgaa	gaccgaggac	accgccgtgt	acttctgcgc	cagagacgtg											300
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5	Ser	Val	Lys	Val	Ser	Cys	Lys	Ala	Ser	Gly	Tyr	Thr	Phe	Thr	Asn	Tyr	
				20					25					30			
	Gly	Met	Asn	Trp	Val	Arg	Gln	Ala	Pro	Gly	Gln	Gly	Leu	Glu	Trp	Met	
10			35					40					45				
	Gly	Trp	Ile	Asn	Asn	Asn	Asn	Gly	Glu	Pro	Thr	Tyr	Ala	Gln	Gly	Phe	
		50					55					60					
15	Arg	Gly	Arg	Phe	Val	Phe	Ser	Leu	Asp	Thr	Ser	Ala	Ser	Thr	Ala	Tyr	
	65					70					75					80	
	Leu	Gln	Ile	Ser	Ser	Leu	Lys	Thr	Glu	Asp	Thr	Ala	Val	Tyr	Phe	Cys	
20					85				90						95		
	Ala	Arg	Asp	Val	Met	Asp	Tyr	Trp	Gly	Gln	Gly	Thr	Thr	Val	Thr	Val	
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	atcacctgca	gagccagtga	aagtgttgat	aattatggct	atagttttat	gcactggtat											120
	cagcagaaac	caggacaacc	tcctaaactc	ctgattttacc	gtgcatccaa	cctagaatct											180
45	gggggtcccag	ccaggttcag	cggcagtggg	tctaggaccg	atttcaccct	cacaattaat											240
	cctgtggaag	ctaataatgac	tgcaaattat	tactgtcagc	aaagtaaaga	atatccgacg											300
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				20					25					30		
15	Gly	Tyr	Ser	Phe	Met	His	Trp	Tyr	Gln	Gln	Lys	Pro	Gly	Gln	Pro	Pro
			35					40					45			
20	Lys	Leu	Leu	Ile	Tyr	Arg	Ala	Ser	Asn	Leu	Glu	Ser	Gly	Val	Pro	Ala
		50					55					60				
25	Arg	Phe	Ser	Gly	Ser	Gly	Ser	Arg	Thr	Asp	Phe	Thr	Leu	Thr	Ile	Asn
	65					70					75					80
30	Pro	Val	Glu	Ala	Asn	Asp	Thr	Ala	Asn	Tyr	Tyr	Cys	Gln	Gln	Ser	Lys
					85					90					95	
35	Glu	Tyr	Pro	Thr	Phe	Gly	Gly	Gly	Thr	Lys	Val	Glu	Ile	Lys		
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<211> 16

<212> PRT

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<400> 59

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<223> 317-4B2 H-CDR2 or CDR-H2

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 1 5 10 15
 45
 Gly
 <210> 64
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 55
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5	Thr	Leu	Ser	Leu	Thr	Cys	Thr	Val	Ser	Gly	Phe	Ser	Leu	Thr	Ser	Tyr
				20					25					30		
10	Gly	Val	His	Trp	Ile	Arg	Gln	Pro	Pro	Gly	Lys	Gly	Leu	Glu	Trp	Ile
			35					40					45			
15	Gly	Val	Ile	Trp	Ala	Gly	Gly	Ser	Thr	Asn	Tyr	Asn	Pro	Ser	Leu	Lys
		50					55					60				
20	Ser	Arg	Val	Thr	Ile	Ser	Val	Asp	Thr	Ser	Lys	Asn	Gln	Phe	Ser	Leu
	65					70					75					80
25	Lys	Leu	Ser	Ser	Val	Thr	Ala	Ala	Asp	Thr	Ala	Val	Tyr	Tyr	Cys	Ala
					85					90					95	
30	Arg	Ala	Tyr	Gly	Asn	Tyr	Trp	Tyr	Ile	Asp	Val	Trp	Gly	Gln	Gly	Thr
				100					105					110		
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5	Thr	Leu	Ser	Leu	Thr	Cys	Thr	Val	Ser	Gly	Phe	Ser	Leu	Thr	Ser	Tyr
				20					25					30		
10	Gly	Val	His	Trp	Ile	Arg	Gln	Pro	Pro	Gly	Lys	Gly	Leu	Glu	Trp	Leu
			35					40					45			
15	Gly	Val	Ile	Trp	Ala	Gly	Gly	Ser	Thr	Asn	Tyr	Asn	Pro	Ser	Leu	Lys
		50					55					60				
20	Ser	Arg	Val	Thr	Ile	Ser	Val	Asp	Thr	Ser	Lys	Asn	Gln	Phe	Ser	Leu
	65					70					75					80
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30	Arg	Ala	Tyr	Gly	Asn	Tyr	Trp	Tyr	Ile	Asp	Val	Trp	Gly	Gln	Gly	Thr
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10	Gly	Val	His	Trp	Ile	Arg	Gln	Pro	Pro	Gly	Lys	Gly	Leu	Glu	Trp	Ile
			35					40					45			
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		50					55					60				
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	65					70					75					80
25	Lys	Leu	Ser	Ser	Val	Thr	Ala	Ala	Asp	Thr	Ala	Val	Tyr	Tyr	Cys	Ala
					85					90					95	
30	Arg	Ala	Tyr	Gly	Asn	Tyr	Trp	Tyr	Ile	Asp	Val	Trp	Gly	Gln	Gly	Thr
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	1				5					10					15	
60	Thr	Leu	Ser	Leu	Thr	Cys	Thr	Val	Ser	Gly	Phe	Ser	Leu	Thr	Ser	Tyr
				20					25					30		
65	Gly	Val	His	Trp	Ile	Arg	Gln	Pro	Pro	Gly	Lys	Gly	Leu	Glu	Trp	Ile
			35					40					45			
70	Gly	Val	Ile	Trp	Ala	Gly	Gly	Ser	Thr	Asn	Tyr	Asn	Pro	Ser	Leu	Lys
		50					55					60				

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	Ser	Arg	Val	Thr	Ile	Ser	Val	Asp	Thr	Ser	Lys	Asn	Gln	Val	Ser	Leu
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5	Lys	Leu	Ser	Ser	Val	Thr	Ala	Ala	Asp	Thr	Ala	Val	Tyr	Tyr	Cys	Ala
					85					90					95	
10	Arg	Ala	Tyr	Gly	Asn	Tyr	Trp	Tyr	Ile	Asp	Val	Trp	Gly	Gln	Gly	Thr
				100					105					110		
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				20					25					30		
35	Gly	Val	His	Trp	Ile	Arg	Gln	Pro	Pro	Gly	Lys	Gly	Leu	Glu	Trp	Ile
			35				40						45			
40	Gly	Val	Ile	Trp	Ala	Gly	Gly	Ser	Thr	Asn	Tyr	Asn	Pro	Ser	Leu	Lys
	50					55						60				
45	Ser	Arg	Val	Thr	Ile	Ser	Val	Asp	Thr	Ser	Lys	Ser	Gln	Val	Ser	Leu
	65				70						75					80
50	Lys	Leu	Ser	Ser	Val	Thr	Ala	Ala	Asp	Thr	Ala	Val	Tyr	Tyr	Cys	Ala
					85					90					95	
55	Arg	Ala	Tyr	Gly	Asn	Tyr	Trp	Tyr	Ile	Asp	Val	Trp	Gly	Gln	Gly	Thr
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<223> 317-3H1 pro-Vh

<400> 69

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Thr Leu Ser Leu Thr Cys Thr Val Ser Gly Phe Ser Leu Thr Ser Tyr
20 25 30

Gly Val His Trp Ile Arg Gln Pro Pro Gly Lys Gly Leu Glu Trp Ile
35 40 45

Gly Val Ile Trp Ala Gly Gly Ser Thr Asn Tyr Asn Pro Ser Leu Lys
50 55 60

Ser Arg Val Thr Ile Ser Lys Asp Thr Ser Lys Ser Gln Val Ser Leu
65 70 75 80

Lys Leu Ser Ser Val Thr Ala Ala Asp Thr Ala Val Tyr Tyr Cys Ala
85 90 95

Arg Ala Tyr Gly Asn Tyr Trp Tyr Ile Asp Val Trp Gly Gln Gly Thr
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Thr Val Thr Val Ser Ser
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5	Thr	Leu	Ser	Leu	Thr	Cys	Thr	Val	Ser	Gly	Phe	Ser	Leu	Thr	Ser	Tyr
				20					25					30		
10	Gly	Val	His	Trp	Ile	Arg	Gln	Pro	Pro	Gly	Lys	Gly	Leu	Glu	Trp	Ile
			35					40					45			
15	Gly	Val	Ile	Trp	Ala	Gly	Gly	Ser	Thr	Asn	Tyr	Asn	Pro	Ser	Leu	Lys
		50					55					60				
20	Ser	Arg	Val	Thr	Ile	Ser	Lys	Asp	Thr	Ser	Lys	Asn	Gln	Val	Ser	Leu
	65					70					75					80
25	Lys	Leu	Ser	Ser	Val	Thr	Ala	Ala	Asp	Thr	Ala	Val	Tyr	Tyr	Cys	Ala
					85					90					95	
30	Arg	Ala	Tyr	Gly	Asn	Tyr	Trp	Tyr	Ile	Asp	Val	Trp	Gly	Gln	Gly	Thr
				100					105					110		
35	Thr	Val	Thr	Val	Ser	Ser										
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5	Thr	Leu	Ser	Leu	Thr	Cys	Thr	Val	Ser	Gly	Phe	Ser	Leu	Thr	Ser	Tyr
				20					25					30		
10	Gly	Val	His	Trp	Ile	Arg	Gln	Pro	Pro	Gly	Lys	Gly	Leu	Glu	Trp	Ile
			35					40					45			
15	Gly	Val	Ile	Trp	Ala	Gly	Gly	Ser	Thr	Asn	Tyr	Asn	Pro	Ser	Leu	Lys
		50					55					60				
20	Ser	Arg	Val	Thr	Ile	Ser	Lys	Asp	Thr	Ser	Lys	Asn	Gln	Phe	Ser	Leu
	65					70					75					80
25	Lys	Leu	Ser	Ser	Val	Thr	Ala	Ala	Asp	Thr	Ala	Val	Tyr	Tyr	Cys	Ala
					85					90					95	
30	Arg	Ala	Tyr	Gly	Asn	Tyr	Trp	Tyr	Ile	Asp	Val	Trp	Gly	Gln	Gly	Thr
				100					105					110		
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 5 Thr Leu Ser Leu Thr Cys Thr Val Ser Gly Phe Ser Leu Thr Ser Tyr
 20 25 30
 10 Gly Val His Trp Ile Arg Gln Pro Pro Gly Lys Gly Leu Glu Trp Ile
 35 40 45
 15 Gly Val Ile Asn Ala Gly Gly Ser Thr Asn Tyr Asn Pro Ser Leu Lys
 50 55 60
 20 Ser Arg Val Thr Ile Ser Lys Asp Thr Ser Lys Asn Gln Phe Ser Leu
 65 70 75 80
 25 Lys Leu Ser Ser Val Thr Ala Ala Asp Thr Ala Val Tyr Tyr Cys Ala
 85 90 95
 30 Arg Ala Tyr Gly Asn Tyr Trp Tyr Ile Asp Val Trp Gly Gln Gly Thr
 100 105 110
 35 Thr Val Thr Val Ser Ser
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 50 <400> 73
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 1 5 10 15
 45 Thr Leu Ser Leu Thr Cys Thr Val Ser Gly Phe Ser Leu Thr Ser Tyr
 20 25 30
 50 Gly Val His Trp Ile Arg Gln Pro Pro Gly Lys Gly Leu Glu Trp Ile
 35 40 45
 55 Gly Val Ile Trp Ala Gly Gly Ser Thr Asn Tyr Asn Pro Ser Leu Lys
 50 55 60

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	Ser	Arg	Val	Thr	Ile	Ser	Lys	Asp	Thr	Ser	Lys	Asn	Gln	Phe	Ser	Leu
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5	Lys	Leu	Ser	Ser	Val	Thr	Ala	Ala	Asp	Thr	Ala	Val	Tyr	Tyr	Cys	Ala
					85					90					95	
10	Arg	Ala	Tyr	Gly	Asn	Tyr	Pro	Tyr	Ile	Asp	Val	Trp	Gly	Gln	Gly	Thr
				100					105					110		
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30	Glu	Arg	Ala	Thr	Ile	Asn	Cys	Lys	Ser	Ser	Gln	Ser	Val	Ser	Asn	Asp
				20					25					30		
35	Val	Ala	Trp	Tyr	Gln	Gln	Lys	Pro	Gly	Gln	Pro	Pro	Lys	Leu	Leu	Ile
			35					40					45			
40	Asn	Tyr	Ala	Phe	His	Arg	Phe	Thr	Gly	Val	Pro	Asp	Arg	Phe	Ser	Gly
	50						55					60				
	Ser	Gly	Tyr	Gly	Thr	Asp	Phe	Thr	Leu	Thr	Ile	Ser	Ser	Leu	Gln	Ala
	65					70					75					80
45	Glu	Asp	Val	Ala	Val	Tyr	Tyr	Cys	His	Gln	Ala	Tyr	Ser	Ser	Pro	Tyr
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10	Ser Val Lys Val Ser Cys Lys Ala Ser Gly Tyr Thr Phe Thr Asn Tyr	20 25 30
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20	Gly Trp Ile Asn Asn Asn Asn Gly Glu Pro Thr Tyr Ala Gln Gly Phe	50 55 60
25	Arg Gly Arg Phe Val Phe Ser Leu Asp Thr Ser Ala Ser Thr Ala Tyr	65 70 75 80
30	Leu Gln Ile Ser Ser Leu Lys Thr Glu Asp Thr Ala Val Tyr Tyr Cys	85 90 95
35	Ala Arg Asp Val Met Asp Tyr Trp Gly Gln Gly Thr Leu Val Thr Val	100 105 110
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<400> 76

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5	Ser	Val	Lys	Val	Ser	Cys	Lys	Ala	Ser	Gly	Tyr	Thr	Phe	Thr	Asn	Tyr
				20					25					30		
10	Gly	Met	Asn	Trp	Val	Arg	Gln	Ala	Pro	Gly	Gln	Gly	Leu	Lys	Trp	Met
			35					40					45			
15	Gly	Trp	Ile	Asn	Asn	Asn	Asn	Gly	Glu	Pro	Thr	Tyr	Ala	Gln	Asp	Phe
		50					55					60				
20	Arg	Gly	Arg	Phe	Val	Phe	Ser	Leu	Asp	Thr	Ser	Ala	Ser	Thr	Ala	Tyr
	65					70					75					80
25	Leu	Gln	Ile	Ser	Ser	Leu	Lys	Thr	Glu	Asp	Thr	Ala	Val	Tyr	Tyr	Cys
					85					90					95	
30	Ala	Arg	Asp	Val	Met	Asp	Tyr	Trp	Gly	Gln	Gly	Thr	Thr	Val	Thr	Val
				100					105					110		
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	Gln	Val	Gln	Leu	Val	Gln	Ser	Gly	Ser	Glu	Leu	Lys	Lys	Pro	Gly	Ala
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5	Ser	Val	Lys	Val	Ser	Cys	Lys	Ala	Ser	Gly	Tyr	Thr	Phe	Thr	Asn	Tyr
				20					25					30		
10	Gly	Met	Asn	Trp	Val	Arg	Gln	Ala	Pro	Gly	Gln	Gly	Leu	Glu	Trp	Met
			35					40					45			
15	Gly	Trp	Ile	Asn	Asn	Asn	Asn	Gly	Glu	Pro	Thr	Tyr	Ala	Gln	Gly	Phe
		50					55					60				
20	Arg	Gly	Arg	Phe	Val	Phe	Ser	Leu	Asp	Thr	Ser	Ala	Ser	Thr	Ala	Tyr
	65					70					75					80
25	Leu	Gln	Ile	Ser	Ser	Leu	Lys	Thr	Glu	Asp	Thr	Ala	Val	Tyr	Tyr	Cys
					85					90					95	
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	1				5					10					15		
5	Ser	Val	Lys	Val	Ser	Cys	Lys	Ala	Ser	Gly	Tyr	Thr	Phe	Thr	Asn	Tyr	
				20					25					30			
10	Gly	Met	Asn	Trp	Val	Arg	Gln	Ala	Pro	Gly	Gln	Gly	Leu	Lys	Trp	Met	
			35					40					45				
15	Gly	Trp	Ile	Asn	Asn	Asn	Asn	Gly	Glu	Pro	Thr	Tyr	Ala	Gln	Asp	Phe	
		50					55					60					
20	Arg	Gly	Arg	Phe	Val	Phe	Ser	Leu	Asp	Thr	Ser	Ala	Ser	Thr	Ala	Tyr	
	65					70					75					80	
25	Leu	Gln	Ile	Ser	Ser	Leu	Lys	Thr	Glu	Asp	Thr	Ala	Val	Tyr	Tyr	Cys	
					85					90					95		
30	Ala	Arg	Asp	Val	Met	Asp	Tyr	Trp	Gly	Gln	Gly	Thr	Thr	Val	Thr	Val	
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<400> 79

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45	Ser	Val	Lys	Val	Ser	Cys	Lys	Ala	Ser	Gly	Tyr	Thr	Phe	Thr	Asn	Tyr	
				20					25					30			
50	Gly	Met	Asn	Trp	Val	Arg	Gln	Ala	Pro	Gly	Gln	Gly	Leu	Glu	Trp	Met	
			35					40					45				
55	Gly	Trp	Ile	Asn	Asn	Asn	Asn	Gly	Glu	Pro	Thr	Tyr	Ala	Gln	Gly	Phe	
		50					55					60					
60	Arg	Gly	Arg	Phe	Val	Phe	Ser	Leu	Asp	Thr	Ser	Ala	Ser	Thr	Ala	Tyr	
	65					70					75					80	

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	Leu	Gln	Ile	Ser	Ser	Leu	Lys	Thr	Glu	Asp	Thr	Ala	Val	Tyr	Tyr	Cys
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25	Ser	Val	Lys	Val	Ser	Cys	Lys	Ala	Ser	Gly	Tyr	Thr	Phe	Thr	Asn	Tyr
				20					25					30		
30	Gly	Met	Asn	Trp	Val	Arg	Gln	Ala	Pro	Gly	Gln	Gly	Leu	Lys	Trp	Met
		35					40						45			
35	Gly	Trp	Ile	Asn	Asn	Asn	Asp	Gly	Glu	Pro	Thr	Tyr	Ala	Gln	Asp	Phe
	50					55						60				
40	Arg	Gly	Arg	Phe	Val	Phe	Ser	Leu	Asp	Thr	Ser	Ala	Ser	Thr	Ala	Tyr
	65					70					75					80
45	Leu	Gln	Ile	Ser	Ser	Leu	Lys	Thr	Glu	Asp	Thr	Ala	Val	Tyr	Tyr	Cys
				85					90					95		
50	Ala	Arg	Asp	Val	Met	Asp	Tyr	Trp	Gly	Gln	Gly	Thr	Leu	Val	Thr	Val
				100					105					110		
55	Ser	Ser														
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	1				5					10					15	
5	Gln	Arg	Ala	Thr	Ile	Thr	Cys	Arg	Ala	Ser	Glu	Ser	Val	Asp	Asn	Tyr
				20					25					30		
10	Gly	Tyr	Ser	Phe	Met	His	Trp	Tyr	Gln	Gln	Lys	Pro	Gly	Gln	Pro	Pro
			35					40					45			
15	Lys	Leu	Leu	Ile	Tyr	Arg	Ala	Ser	Asn	Leu	Glu	Ser	Gly	Val	Pro	Ala
	50						55					60				
20	Arg	Phe	Ser	Gly	Ser	Gly	Ser	Arg	Thr	Asp	Phe	Thr	Leu	Thr	Ile	Asn
	65					70					75					80
25	Pro	Val	Glu	Ala	Glu	Asp	Thr	Ala	Asn	Tyr	Tyr	Cys	Gln	Gln	Ser	Lys
					85					90					95	
30	Glu	Tyr	Pro	Thr	Phe	Gly	Gly	Gly	Thr	Lys	Val	Glu	Ile	Lys		
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35	<400> 82															
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45	Gln	Arg	Ala	Thr	Ile	Thr	Cys	Arg	Ala	Ser	Glu	Ser	Val	Asp	Asn	Tyr
				20					25					30		
50	Gly	Tyr	Ser	Phe	Met	His	Trp	Tyr	Gln	Gln	Lys	Pro	Gly	Gln	Pro	Pro
			35					40					45			
55	Lys	Leu	Leu	Ile	Tyr	Arg	Ala	Ser	Asn	Leu	Glu	Ser	Gly	Val	Pro	Ala
	50						55					60				
60	Arg	Phe	Ser	Gly	Ser	Gly	Ser	Gly	Thr	Asp	Phe	Thr	Leu	Thr	Ile	Asn
	65					70					75					80
65	Pro	Val	Glu	Ala	Asn	Asp	Thr	Ala	Asn	Tyr	Tyr	Cys	Gln	Gln	Ser	Lys
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Glu Tyr Pro Thr Phe Gly Gly Gly Thr Lys Val Glu Ile Lys
100 105 110

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20 Ser Thr Ser Glu Ser Thr Ala Ala Leu Gly Cys Leu Val Lys Asp Tyr
20 25 30

25 Phe Pro Glu Pro Val Thr Val Ser Trp Asn Ser Gly Ala Leu Thr Ser
35 40 45

Gly Val His Thr Phe Pro Ala Val Leu Gln Ser Ser Gly Leu Tyr Ser
50 55 60

30 Leu Ser Ser Val Val Thr Val Pro Ser Ser Ser Leu Gly Thr Lys Thr
65 70 75 80

35 Tyr Thr Cys Asn Val Asp His Lys Pro Ser Asn Thr Lys Val Asp Lys
85 90 95

40 Arg Val Glu Ser Lys Tyr Gly Pro Pro Cys Pro Pro Cys Pro Ala Pro
100 105 110

Glu Phe Glu Gly Gly Pro Ser Val Phe Leu Phe Pro Pro Lys Pro Lys
115 120 125

45 Asp Thr Leu Met Ile Ser Arg Thr Pro Glu Val Thr Cys Val Val Val
130 135 140

50 Asp Val Ser Gln Glu Asp Pro Glu Val Gln Phe Asn Trp Tyr Val Asp
145 150 155 160

Gly Val Glu Val His Asn Ala Lys Thr Lys Pro Arg Glu Glu Gln Phe
165 170 175

55 Asn Ser Thr Tyr Arg Val Val Ser Val Leu Thr Val Leu His Gln Asp
180 185 190

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	Trp	Leu	Asn	Gly	Lys	Glu	Tyr	Lys	Cys	Lys	Val	Ser	Asn	Lys	Gly	Leu	
			195					200					205				
5	Pro	Ser	Ser	Ile	Glu	Lys	Thr	Ile	Ser	Lys	Ala	Lys	Gly	Gln	Pro	Arg	
		210					215					220					
10	Glu	Pro	Gln	Val	Tyr	Thr	Leu	Pro	Pro	Ser	Gln	Glu	Glu	Met	Thr	Lys	
	225					230					235					240	
15	Asn	Gln	Val	Ser	Leu	Thr	Cys	Leu	Val	Lys	Gly	Phe	Tyr	Pro	Ser	Asp	
					245					250					255		
20	Ile	Ala	Val	Glu	Trp	Glu	Ser	Asn	Gly	Gln	Pro	Glu	Asn	Asn	Tyr	Lys	
				260					265					270			
25	Thr	Thr	Pro	Pro	Val	Leu	Asp	Ser	Asp	Gly	Ser	Phe	Phe	Leu	Tyr	Ser	
			275					280					285				
30	Arg	Leu	Thr	Val	Asp	Lys	Ser	Arg	Trp	Gln	Glu	Gly	Asn	Val	Phe	Ser	
	290						295					300					
35	Cys	Ser	Val	Met	His	Glu	Ala	Leu	His	Asn	His	Tyr	Thr	Gln	Lys	Ser	
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40	Leu	Ser	Leu	Ser	Leu	Gly	Lys										
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5	Ser	Thr	Ser	Glu	Ser	Thr	Ala	Ala	Leu	Gly	Cys	Leu	Val	Lys	Asp	Tyr
				20					25					30		
10	Phe	Pro	Glu	Pro	Val	Thr	Val	Ser	Trp	Asn	Ser	Gly	Ala	Leu	Thr	Ser
			35					40					45			
15	Gly	Val	His	Thr	Phe	Pro	Ala	Val	Leu	Gln	Ser	Ser	Gly	Leu	Tyr	Ser
		50					55						60			
	Leu	Ser	Ser	Val	Val	Thr	Val	Pro	Ser	Ser	Ser	Leu	Gly	Thr	Lys	Thr
20																
25																
30																
35																
40																
45																
50																
55																

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	65					70										75						80
5	Tyr	Thr	Cys	Asn	Val	Asp	His	Lys	Pro	Ser	Asn	Thr	Lys	Val	Asp	Lys						
					85					90					95							
	Arg	Val	Glu	Ser	Lys	Tyr	Gly	Pro	Pro	Cys	Pro	Pro	Cys	Pro	Ala	Pro						
10				100					105					110								
	Pro	Val	Ala	Gly	Gly	Pro	Ser	Val	Phe	Leu	Phe	Pro	Pro	Lys	Pro	Lys						
			115					120					125									
15	Asp	Thr	Leu	Met	Ile	Ser	Arg	Thr	Pro	Glu	Val	Thr	Cys	Val	Val	Val						
		130					135					140										
	Asp	Val	Ser	Gln	Glu	Asp	Pro	Glu	Val	Gln	Phe	Asn	Trp	Tyr	Val	Asp						
20	145					150					155					160						
	Gly	Val	Glu	Val	His	Asn	Ala	Lys	Thr	Lys	Pro	Arg	Glu	Glu	Gln	Phe						
					165					170					175							
25	Asn	Ser	Thr	Tyr	Arg	Val	Val	Ser	Val	Leu	Thr	Val	Leu	His	Gln	Asp						
				180					185					190								
	Trp	Leu	Asn	Gly	Lys	Glu	Tyr	Lys	Cys	Lys	Val	Ser	Asn	Lys	Gly	Leu						
30			195					200					205									
	Pro	Ser	Ser	Ile	Glu	Lys	Thr	Ile	Ser	Lys	Ala	Lys	Gly	Gln	Pro	Arg						
35		210					215					220										
	Glu	Pro	Gln	Val	Tyr	Thr	Leu	Pro	Pro	Ser	Gln	Glu	Glu	Met	Thr	Lys						
	225					230					235					240						
40	Asn	Gln	Val	Ser	Leu	Thr	Cys	Leu	Val	Lys	Gly	Phe	Tyr	Pro	Ser	Asp						
					245					250					255							
	Ile	Ala	Val	Glu	Trp	Glu	Ser	Asn	Gly	Gln	Pro	Glu	Asn	Asn	Tyr	Lys						
45				260					265					270								
	Thr	Thr	Pro	Pro	Val	Leu	Asp	Ser	Asp	Gly	Ser	Phe	Phe	Leu	Tyr	Ser						
50			275					280					285									
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	130						135					140					
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50	Gly	Val	Glu	Val	His	Asn	Ala	Lys	Thr	Lys	Pro	Arg	Glu	Glu	Gln	Phe	
					165					170					175		
55	Asn	Ser	Thr	Tyr	Arg	Val	Val	Ser	Val	Leu	Thr	Val	Leu	His	Gln	Asp	
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15	Asn Gln Val Ser Leu Thr Cys Leu Val Lys Gly Phe Tyr Pro Ser Asp			
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20	Ile Ala Val Glu Trp Glu Ser Asn Gly Gln Pro Glu Asn Asn Tyr Lys			
		260	265	270
25	Thr Thr Pro Pro Val Leu Asp Ser Asp Gly Ser Phe Phe Leu Tyr Ser			
		275	280	285
30	Arg Leu Thr Val Asp Lys Ser Arg Trp Gln Glu Gly Asn Val Phe Ser			
	290	295	300	
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40																
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Gln	Lys	Phe	Lys	Asp	Lys	Ala	Thr	Leu	Thr	Thr	Asp	Lys	Ser	Ser	Ser	85	90	95	
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Ser	Ala	Ser	Ser	Ser	Val	Ser	Tyr	Met	Asn	Trp	Tyr	Gln	Gln	Lys	Ser	180	185	190	
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	290						295					300					

Claims

1. An antibody comprising an antibody antigen binding domain which binds human PD-1 and comprises a heavy chain variable region (Vh) comprising the amino acid sequence SEQ ID NO:24; and a light chain variable region (Vk) comprising the amino acid sequence SEQ ID NO:26.
2. The antibody according claim 1 which binds PD1 residues: K45 and I93, equivalent to K58 and I106 in SEQ ID NO 2.
3. The antibody according to any one of claims 1-2 which comprises an IgG4 constant domain comprising any of SEQ ID NO: 83-88.
4. The antibody according to claim 3 which comprises an IgG4 constant domain comprising SEQ ID NO 87 or 88.
5. The antibody according to claim 1 which induces IL-2 release in PD-1-transformed HuT78 cells co-cultured with OS8/PD-L1 or OS8/PD-L2 cotransformed HEK293 cells, and/or inhibits IL-2 secretion in P3Z-transformed HuT78 cells co-cultured with PD-L1- or PD-L2-transformed HEK293 cells, wherein:

OS8 comprises an amino acid sequence of SEQ ID NO:89 and is a membrane-anchored chimeric Ab constructed by fusing the single chain variable fragment (scFv) of an anti-human CD3 mAb OKT3 to the C-terminal domain (113-220) of mouse CD8 α ; and
P3Z comprises an amino acid sequence of SEQ ID NO:90 and is a chimeric PD-1 receptor, constructed by fusing the extracellular and transmembrane domains of human PD-1 to the cytoplasmic domain of human CD3 ζ chain.
6. An F(ab) or F(ab)₂ of the antibody according to any one of claims 1-5.
7. The antibody according to any one of claims 1-5 for use in the treatment of cancer or viral infection.

Patentansprüche

1. Antikörper, der eine Antikörper-Antigen-Bindungsdomäne umfasst, die menschliches PD-1 bindet und Folgendes umfasst:

eine variable Schwerkettenregion (Vh), die die Aminosäuresequenz SEQ ID NO: 24 umfasst; und
eine variable Leichtkettenregion (Vk), die die Aminosäuresequenz SEQ ID NO: 26 umfasst.
2. Antikörper nach Anspruch 1, der die PD1-Reste K45 und I93 bindet, die äquivalent zu K58 und I106 in SEQ ID NO: 2 sind.
3. Antikörper nach einem der Ansprüche 1 bis 2, der eine konstante IgG4-Domäne umfasst, die beliebige der SEQ ID NO: 83-88 umfasst.
4. Antikörper nach Anspruch 3, der eine konstante IgG4-Domäne umfasst, die SEQ ID NO: 87 oder 88 umfasst.
5. Antikörper nach Anspruch 1, der die IL-2-Freisetzung in PD-1-transformierten HuT78-Zellen induziert, die mit OS8/PD-L1- oder OS8/PD-L2-cotransformierten HEK293-Zellen kokultiviert wurden, und/oder die IL-2-Sekretion in P3Z-transformierten HuT78-Zellen hemmt, die mit PD-L1- oder PD-L2-transformierten HEK293-Zellen kokultiviert wurden, wobei:

OS8 eine Aminosäuresequenz von SEQ ID NO: 89 umfasst und ein membranverankerter chimärer Ab ist, der durch Fusionieren des variablen Einkettenfragments (scFv) eines Anti-Mensch-CD3-mAb OKT3 an die C-terminale Domäne (113-220) von Maus-CD8 α konstruiert wird; und
P3Z eine Aminosäuresequenz von SEQ ID NO: 90 umfasst und ein chimärer PD-1-Rezeptor ist, der durch Fusionieren der extrazellulären Domäne und Transmembrandomäne von menschlichem PD-1 an die zytoplasmatische Domäne einer menschlichen CD3 ζ -Kette konstruiert wird.
6. F(ab) oder F(ab)₂ eines Antikörpers nach einem der Ansprüche 1 bis 5.

7. Antikörper nach einem der Ansprüche 1 bis 5 zur Verwendung bei der Behandlung von Krebs oder einer Virusinfektion.

5 Revendications

1. Anticorps comprenant un domaine de liaison antigène-anticorps qui se lie à la PD-1 humaine et comprend une région variable de chaîne lourde (Vh) comprenant la séquence d'acides aminés SEQ ID NO : 24 ; et une région variable de chaîne légère (Vk) comprenant la séquence d'acides aminés SEQ ID NO : 26.
2. Anticorps selon la revendication 1 qui se lie aux résidus de PD1 : K45 et I93, équivalent à K58 et I106 dans SEQ ID NO 2.
3. Anticorps selon l'une quelconque des revendications 1 à 2, qui comprend un domaine constant d'IgG4 comprenant l'une quelconque des SEQ ID NO : 83 à 88.
4. Anticorps selon la revendication 3, qui comprend un domaine constant d'IgG4 comprenant SEQ ID NO 87 ou 88.
5. Anticorps selon la revendication 1 qui induit une libération d'IL-2 dans des cellules HuT78 transformées par PD-1 co-cultivées avec des cellules HEK293 co-transformées par OS8/PD-L1 ou OS8/PD-L2, et/ou inhibe une sécrétion d'IL-2 dans des cellules HuT78 transformées par P3Z co-cultivées avec des cellules HEK293 transformées par PD-L1 ou PD-L2, dans lequel :

OS8 comprend une séquence d'acides aminés de SEQ ID NO : 89 et est un Ab chimérique à ancrage membranaire construit en fusionnant le fragment variable à chaîne unique (scFv) d'un OKT3 CD3 mAb anti-humain au domaine C-terminal (113 à 220) de CD8 α de souris ; et

P3Z comprend une séquence d'acides aminés de SEQ ID NO : 90 et est un récepteur PD-1 chimérique, construit en fusionnant les domaines extracellulaire et transmembranaire de PD-1 humaine au domaine cytoplasmique de chaîne CD3 ζ humaine.
6. F(ab) ou F(ab)₂ de l'anticorps selon l'une quelconque des revendications 1 à 5.
7. Anticorps selon l'une quelconque des revendications 1 à 5, destiné à être utilisé dans le traitement du cancer ou d'une infection virale.

Fig. 1

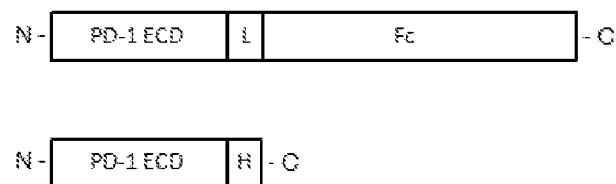


Fig. 2

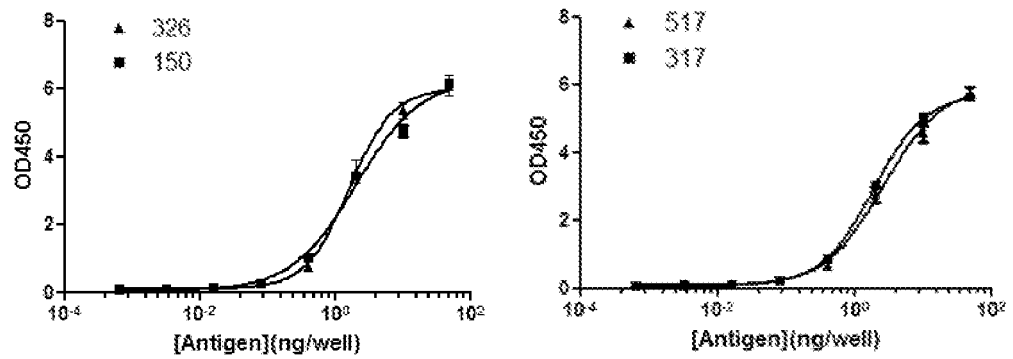


Fig. 3

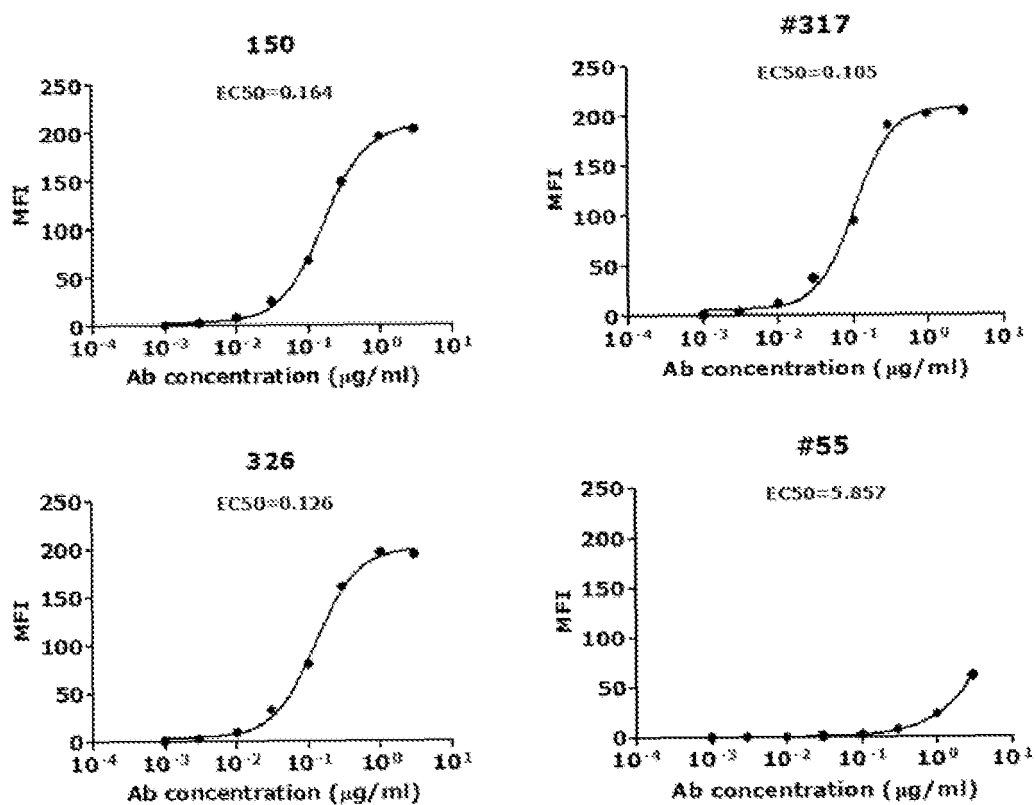


Fig. 4

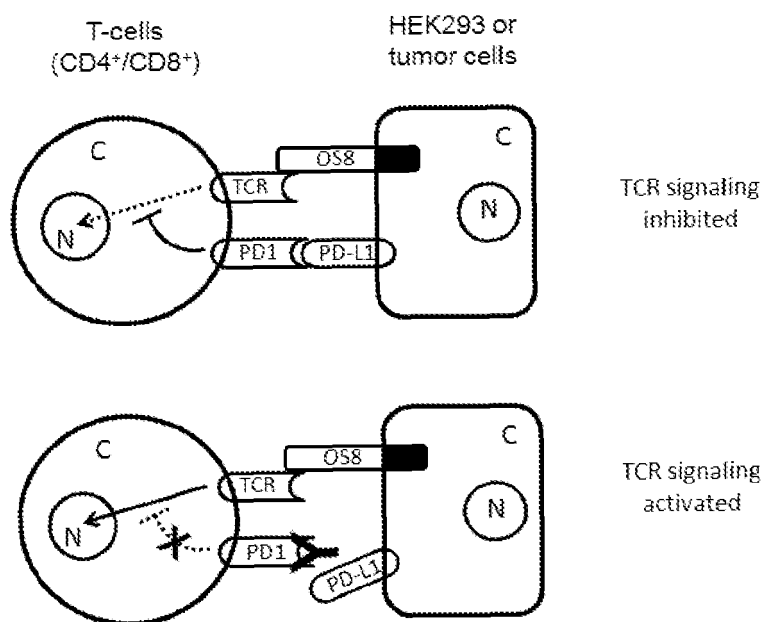


Fig. 5

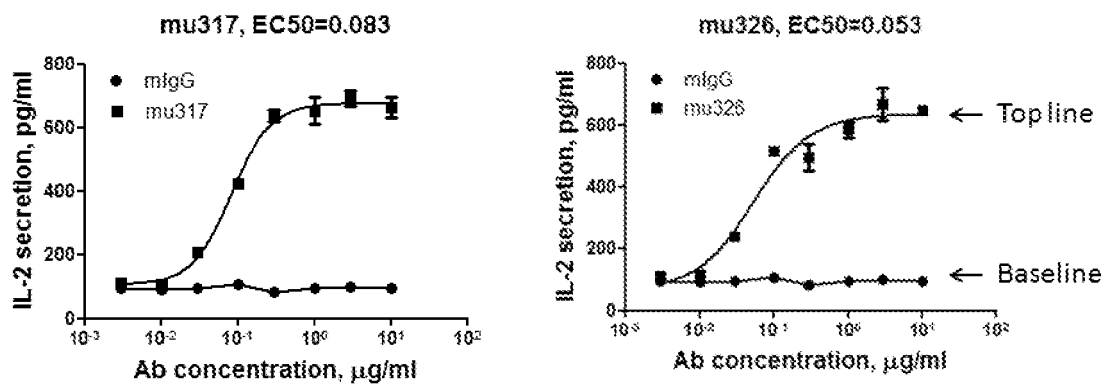
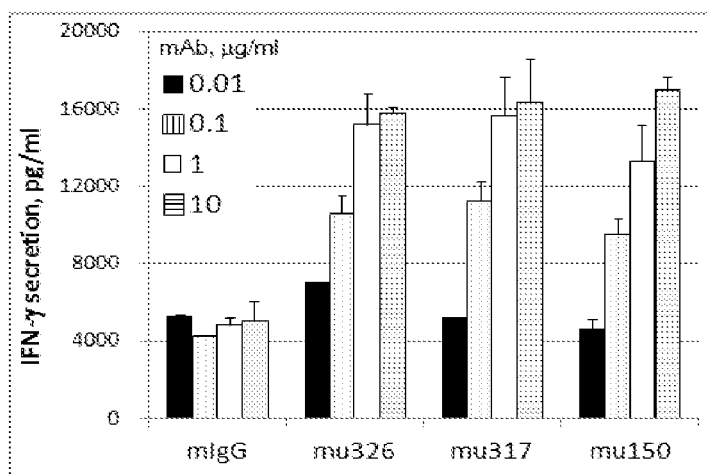


Fig. 6 (A)



(B)

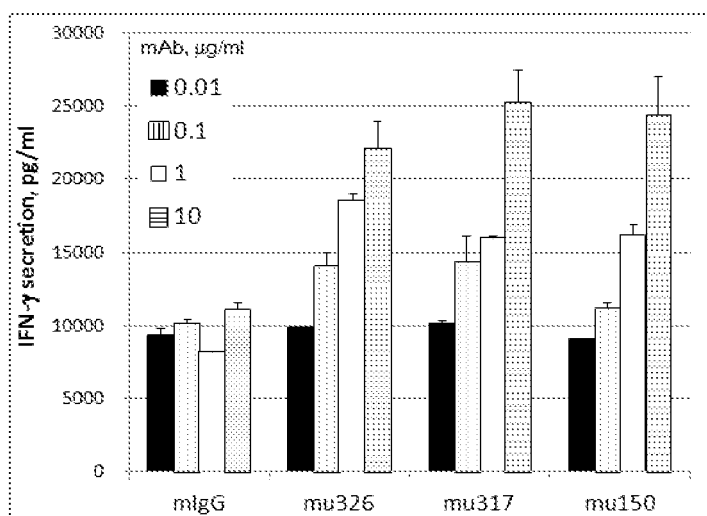


Fig. 7

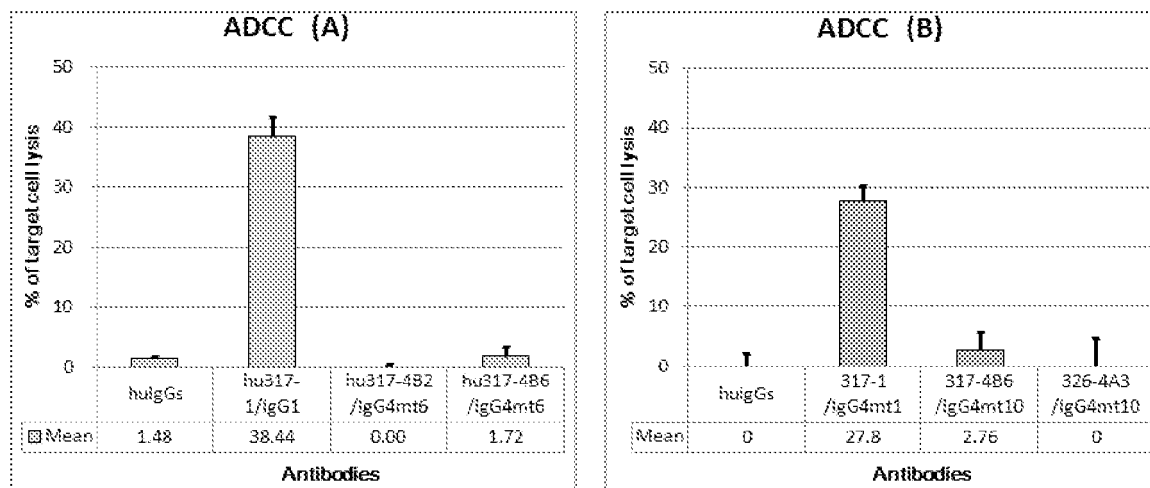


Fig. 8

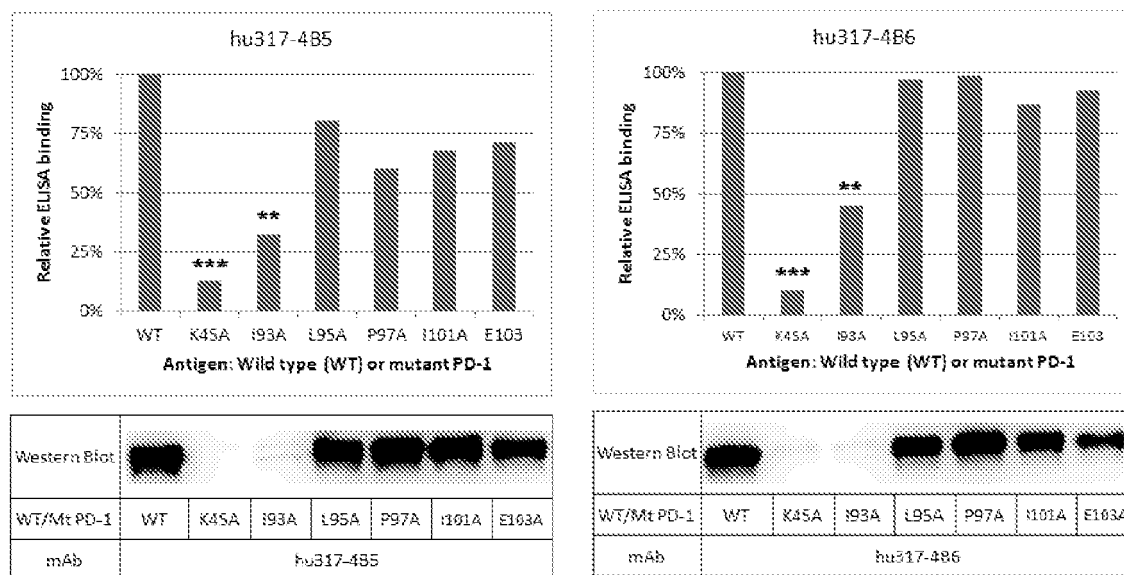


Fig. 8 (cont'd)

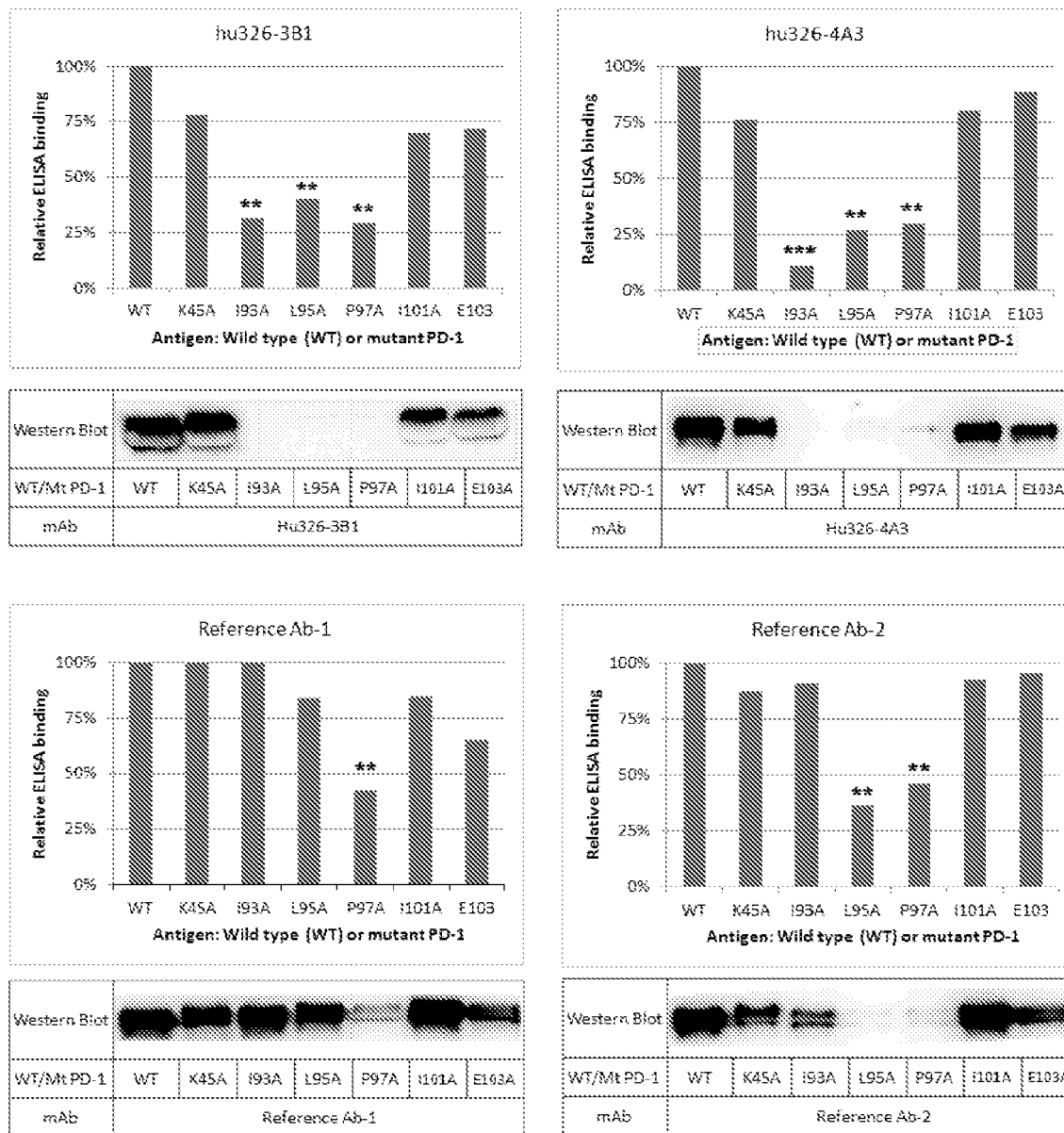


Fig. 9.

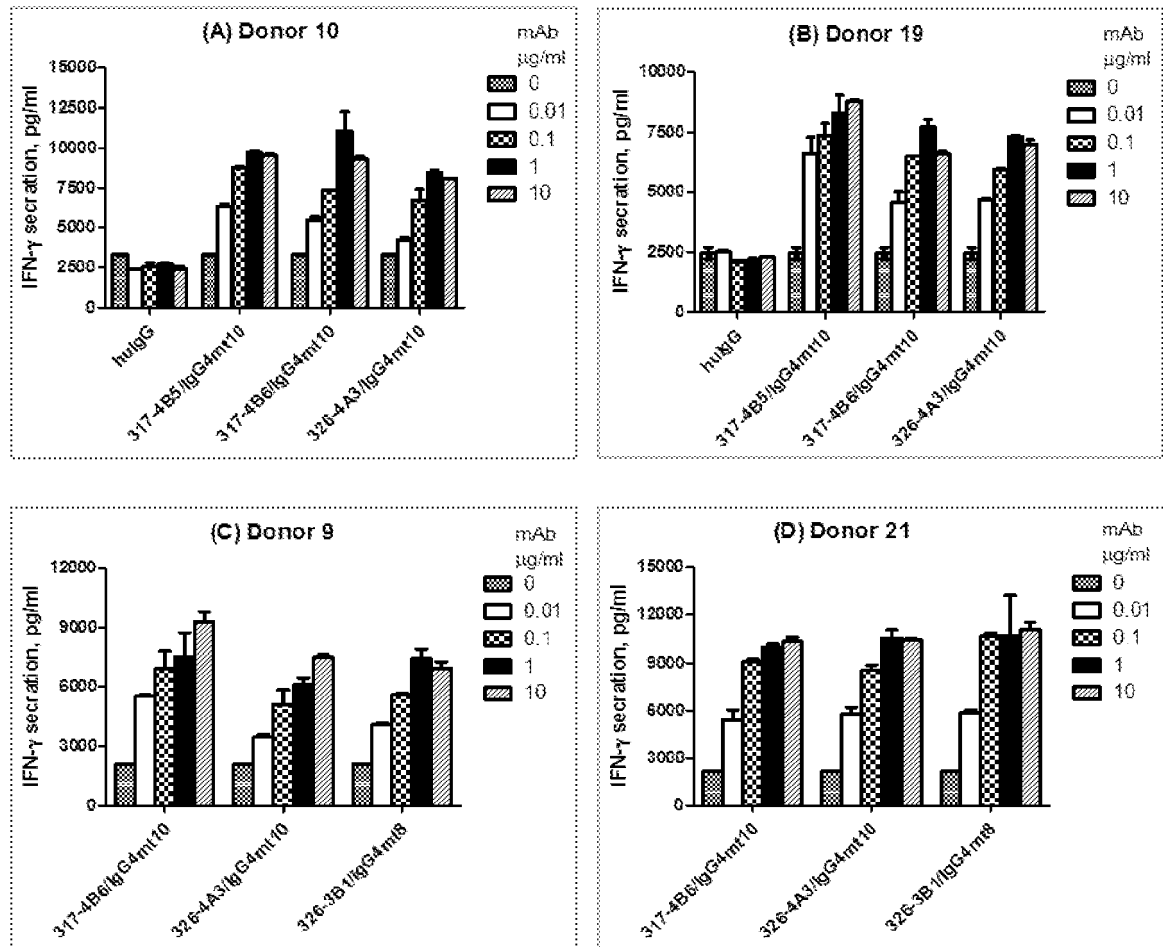


Fig. 10

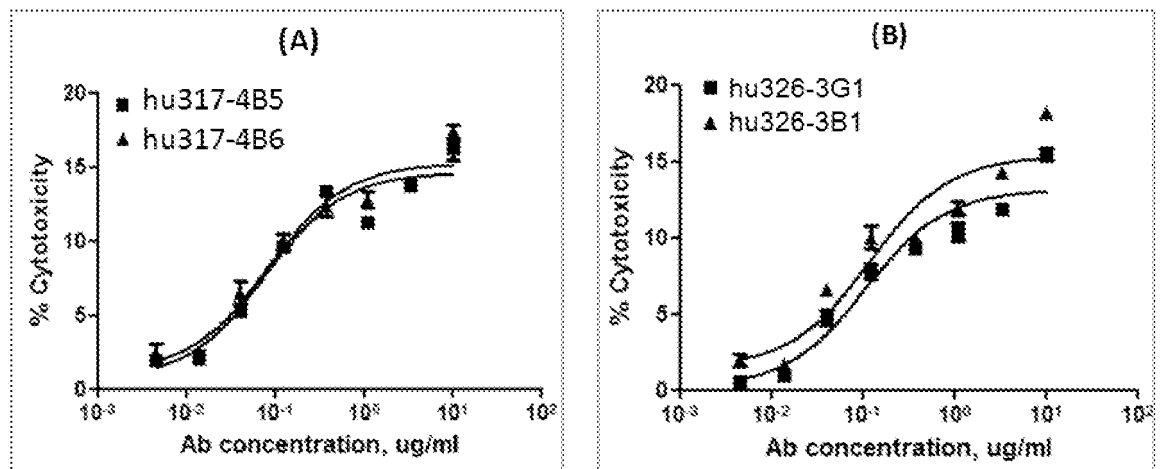
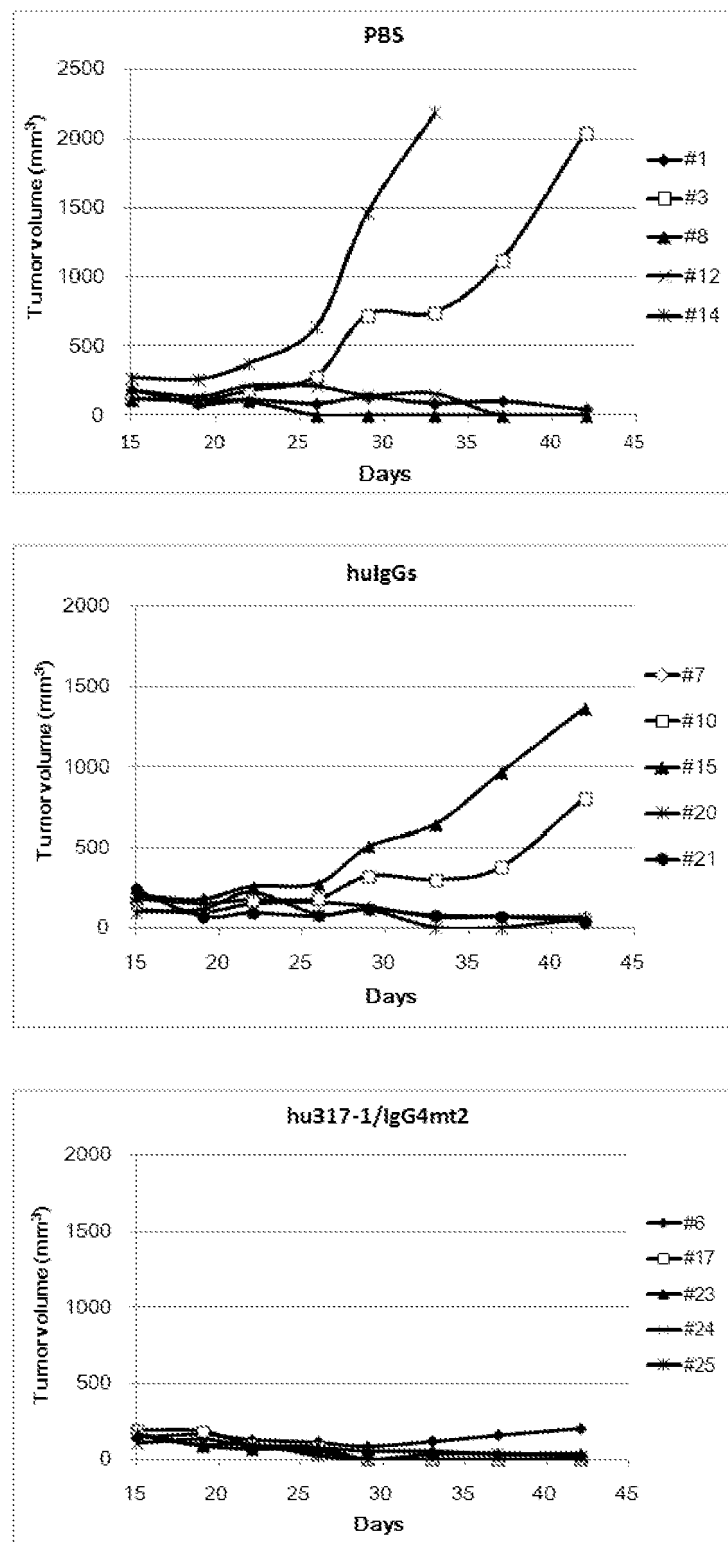


Fig. 11



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