

(19)



(11)

EP 3 521 304 B9

(12)

CORRECTED EUROPEAN PATENT SPECIFICATION

(15) Correction information:
Corrected version no 1 (W1 B1)
Corrections, see
Sequence listing
Remarks
Sequence listing replaced or added

(51) International Patent Classification (IPC):
C07K 14/31 (2006.01) B01D 15/38 (2006.01)

(52) Cooperative Patent Classification (CPC):
C07K 14/31; B01D 15/3809; B01J 20/289

(48) Corrigendum issued on:
04.10.2023 Bulletin 2023/40

(45) Date of publication and mention
of the grant of the patent:
07.06.2023 Bulletin 2023/23

(21) Application number: **18154731.6**

(22) Date of filing: **01.02.2018**

(54) **FC BINDING PROTEINS WITH CYSTEINE IN THE C-TERMINAL HELICAL REGION**
FC-BINDUNGSPROTEINE MIT CYSTEIN IN DER C-TERMINALEN HELIKALEN REGION
PROTÉINES DE LIAISON FC AVEC LA CYSTÉINE DANS LA RÉGION HÉLICOÏDALE C-TERMINALE

(84) Designated Contracting States:
AL AT BE BG CH CY CZ DE DK EE ES FI FR GB
GR HR HU IE IS IT LI LT LU LV MC MK MT NL NO
PL PT RO RS SE SI SK SM TR

(43) Date of publication of application:
07.08.2019 Bulletin 2019/32

(60) Divisional application:
23176990.2 / 4 249 097

(73) Proprietor: **Repligen Corporation**
Waltham, MA 02453 (US)

(72) Inventors:

- **Fiedler, Erik**
06120 Halle/Saale (DE)

- **Haupts, Ulrich**
06120 Halle/Saale (DE)

(74) Representative: **Duxbury, Stephen et al**
Arnold & Siedsma
Bezuidenhoutseweg 57
2594 AC The Hague (NL)

(56) References cited:
EP-A1- 2 690 173 WO-A1-2015/005859
WO-A1-2016/079033 WO-A1-2017/009421
WO-A1-2018/029157 WO-A1-2019/030156

Remarks:
The complete document including Reference Table(s) and the Sequence Listing(s) can be downloaded from the EPO website

EP 3 521 304 B9

Note: Within nine months of the publication of the mention of the grant of the European patent in the European Patent Bulletin, any person may give notice to the European Patent Office of opposition to that patent, in accordance with the Implementing Regulations. Notice of opposition shall not be deemed to have been filed until the opposition fee has been paid. (Art. 99(1) European Patent Convention).

Description**FIELD OF THE INVENTION**

5 [0001] The present invention relates to Fc binding proteins comprising one or more domains having a Cysteine in the C-terminal helical region. The invention further relates to affinity matrices comprising the Fc binding proteins of the invention. The invention also relates to a use of these Fc binding proteins or affinity matrices for affinity purification of immunoglobulins and to methods of affinity purification using the Fc binding proteins of the invention.

BACKGROUND OF THE INVENTION

10 [0002] Many biotechnological and pharmaceutical applications require the removal of contaminants from a sample containing antibodies. An established procedure for capturing and purifying antibodies is affinity chromatography using the bacterial cell surface Protein A from *Staphylococcus aureus* as selective ligand for immunoglobulins (see, for example, review by Huse et al., J. Biochem. Biophys. Methods 51, 2002: 217-231). Wild-type Protein A binds to the Fc region of IgG molecules with high affinity and selectivity and is stable at high temperatures and in a wide range of pH values. Variants of Protein A with improved properties such as alkaline stability are available for purifying antibodies and various chromatographic matrices comprising Protein A ligands are commercially available. However, in particular wild-type Protein A based chromatography matrices show a loss of binding capacity for immunoglobulins following exposure to alkaline conditions.

TECHNICAL PROBLEMS UNDERLYING THE INVENTION

25 [0003] Most large scale production processes for antibodies or Fc-containing fusion proteins use Protein A for affinity purification. However, due to limitations of Protein A applications in affinity chromatography there is a need in the art to provide novel Fc binding proteins with improved properties that specifically bind to immunoglobulins in order to facilitate affinity purification of immunoglobulins. To maximally exploit the value of the chromatographic matrices comprising Fc binding proteins it is desirable to use the affinity ligand matrices multiple times. Between chromatography cycles, a thorough cleaning procedure is required for sanitization and removal of residual contaminants on the matrix. In this procedure, it is general practice to apply alkaline solutions with high concentrations of NaOH to the affinity ligand matrices. Wild-type Protein A domains cannot withstand such harsh alkaline conditions for an extended time and quickly lose binding capacity for immunoglobulin. Accordingly, there is an ongoing need in this field to obtain novel proteins capable of binding proteins comprising an Fc sequence.

30 [0004] The present invention provides Fc binding proteins that are particularly well-suited for affinity purification of immunoglobulins but overcome the disadvantages of the prior art. In particular, a significant advantage of the Fc binding proteins of the invention is their improved stability at high pH for a prolonged time period without reducing the Fc binding capacities in combination with high dynamic binding capacities. Further, the novel Fc binding proteins allow an elution of more than 95 % at pH of 3.5 or higher of the bound Fc protein from the Fc binding protein immobilized to a matrix.

35 [0005] The above overview does not necessarily describe all problems solved by the present invention.

SUMMARY OF THE INVENTION

40 [0006] A first aspect of the present invention is to provide an Fc binding protein suitable for affinity purification. This is achieved with an Fc binding protein comprising one or more Fc binding domains, wherein at least the amino acid in position 50, 51, or 53 corresponding to SEQ ID NO: 2 is Cysteine. In some embodiments, the Fc binding protein is comprising one or more Fc binding domains, wherein at least one domain comprises or essentially consists of or consists of an amino acid sequence of SEQ ID NO: 2 or an amino acid sequence with at least 89.5 % identity thereto wherein at least the amino acid in position, 50, 51, or 53 corresponding to SEQ ID NO: 2 is Cysteine. In some embodiments, at least one domain comprises or essentially consists of or consists of an amino acid sequence of SEQ ID NOs: 3-16, or an amino acid sequence with at least 89.5 % identity thereto, respectively, wherein at least the amino acid in position 50, 51, or 53 corresponding to SEQ ID NOs: 3-16 is Cysteine. In some embodiments, at least one domain comprises or essentially consists of or consists of an amino acid sequence of SEQ ID NOs: 7-8, or an amino acid sequence with at least 89.5 % identity thereto, respectively, wherein at least the amino acid in position, 50, 51, or 53 corresponding to SEQ ID NOs: 7-8 is Cysteine. In some embodiments, at least one domain comprises or essentially consists of or consists of an amino acid sequence selected from the group of SEQ ID NOs: 77, 90-95 or an amino acid sequence with at least 89.5 % identity thereto.

55 [0007] In a second aspect the present invention relates to an affinity separation matrix comprising the Fc binding protein of the first aspect.

[0008] In a third aspect the present invention relates to a use of the Fc binding protein of the first aspect or of the affinity separation matrix of the second aspect for affinity purification of immunoglobulins or proteins comprising an Fc part of immunoglobulins.

[0009] In a fourth aspect the present invention relates to a method of affinity purification of immunoglobulins or proteins comprising an Fc part of immunoglobulins comprising the steps of (a) providing a liquid containing an immunoglobulin; (b) providing an affinity separation matrix comprising an immobilized Fc binding protein of the first aspect coupled to said affinity separation matrix; (c) contacting said liquid and said affinity separation matrix, wherein said immunoglobulin binds to said immobilized Fc binding protein; and (d) eluting said immunoglobulin from said matrix, thereby obtaining an eluate containing said immunoglobulin. This summary of the invention does not necessarily describe all features of the present invention. Other embodiments will become apparent from a review of the ensuing detailed description.

BRIEF DESCRIPTION OF THE FIGURES.

[0010]

Figure 1. Amino acid sequences of Fc binding domains with Cysteine in the c-terminal helical region. The numbers in the top row refer to the corresponding amino acid position in the Fc binding domain. **Figure 1A.** Fc binding domains of SEQ ID NO: 2. **Figure 1B.** Examples for Fc binding domains with Cys in helix 3.

Figure 2. Immobilization of Fc binding domains with Cysteine in the helix 3. **Figure 2A.** Coupling efficiency of IB14 proteins with Cysteine in positions 43, 46, 47, 50, or 58 on Epoxy-matrix. Y-axis: coupled amount of protein in nmol/ml. **Figure 2B.** Coupling of cs14 43C and cs14 46C to Epoxy-activated Sepharose 6B matrix. Y-axis: n(protein per ml resin) in nmol **Figure 3.** Caustic stability of Fc binding proteins with Cysteine in the c-terminal region 3.

Figure 3A. Analysis of the alkaline stability of IB14 with Cysteine in positions 43, 46, 47, 50, or 58. Y-axis: remaining IgG binding activity of the Fc binding protein IB14 and Cys Variants in %. Grey column: IgG binding activity after 6 h of continuous 0.5 M NaOH treatment. Black column: IgG binding activity without NaOH. **Figure 3B.** Caustic stability and DBC 10% of cs14 with Cysteine in positions 43 or 46. Y-axis: DBC 10 % (mg/ml). Black: DBC 10% (mg/ml), 0 h NaOH, light grey: DBC 10% (mg/ml), 18 h NaOH, dark grey: DBC 10% (mg/ml), 72 h NaOH, The Cys variants were compared to cs14 with C-terminal Cysteine and to Protein A.

Figure 4. The remaining binding capacity (in %) after 6, 24, and 36 hours of continuous 0.5 M NaOH treatment is shown for cs26 46C, compared to cs26 without Cys in position 46 and compared to wildtype Protein A domain C. The x-axis shows the time of 0.5 M NaOH incubation in hours.

Figure 5. The dynamic binding capacity (DBC; mg/ml) is shown for cs26 46C (monomer and dimer), compared to cs26 without Cys in position 46, and compared to recombinant wildtype Protein A.

DETAILED DESCRIPTION OF THE INVENTION

Definitions

[0011] Before the present invention is described in detail below, it is to be understood that this invention is not limited to the particular methodology, protocols and reagents described herein as these may vary. It is also to be understood that the terminology used herein is for the purpose of describing particular embodiments only, and is not intended to limit the scope of the present invention which will be limited only by the appended claims. Unless defined otherwise, all technical and scientific terms used herein have the same meanings as commonly understood by one of ordinary skill in the art to which this invention belongs.

[0012] Preferably, the terms used herein are consistent with the definitions provided in "A multilingual glossary of biotechnological terms: (IUPAC Recommendations)", Leuenberger, H.G.W, Nagel, B. and Kölbl, H. eds. (1995), Helvetica Chimica Acta, CH-4010 Basel, Switzerland). Throughout this specification and the claims which follow, unless the context requires otherwise, the word "comprise", and variations such as "comprises" and "comprising", will be understood to imply the inclusion of a stated member, integer or step or group of members, integers or steps but not the exclusion of any other member, integer or step or group of members, integers or steps.

[0013] As used in the description of the invention and the appended claims, the singular forms "a", "an" and "the" are used interchangeably and intended to include the plural forms as well and fall within each meaning, unless the context clearly indicates otherwise. Also, as used herein, "and/or" refers to and encompasses any and all possible combinations of one or more of the listed items, as well as the lack of combinations when interpreted in the alternative ("or").

[0014] The term "about", as used herein, encompasses the explicitly recited amounts as well as deviations therefrom of $\pm 10\%$. More preferably, a deviation 5% is encompassed by the term "about".

[0015] All sequences referred to herein are disclosed in the attached sequence listing that, with its whole content and disclosure, is a part of this specification.

[0016] In the context of the present invention, the term "Fc binding protein" or "immunoglobulin-binding protein" or "Ig binding protein" is used to describe proteins that are capable to specifically bind to the Fc region of an immunoglobulin. The "immunoglobulin" as understood herein can include, but is not necessarily limited to, mammalian IgG, such as for example human IgG₁, human IgG₂, human IgG₄, mouse IgG, rat IgG, goat IgG, bovine IgG, guinea pig IgG, rabbit IgG; human IgM, human IgA; and immunoglobulin fragments comprising a Fc region. Due to a specific binding to the Fc region, the "Fc binding proteins" or "Ig binding proteins" of the invention are capable of binding to entire immunoglobulins, and to immunoglobulin fragments comprising a Fc region, fusion proteins comprising an Fc region of an immunoglobulin, and conjugates comprising an Fc region of an immunoglobulin. While the Fc binding proteins of the invention herein exhibit specific binding to the Fc region of an immunoglobulin, it is not excluded that Fc binding proteins can additionally bind with reduced affinity to other regions, such as Fab regions of immunoglobulins.

[0017] The term "binding" according to the invention preferably relates to a specific binding. "Specific binding" means that an Fc binding protein or an Fc binding domain binds stronger to an immunoglobulin for which it is specific compared to the binding to another non-immunoglobulin target.

[0018] The term "binding activity" refers to the ability of an Fc binding protein of the invention to bind to immunoglobulin. For example, the binding activity can be determined before and/or after alkaline treatment. The binding activity can be determined for an Fc binding protein or for an Fc binding protein coupled to a matrix, i.e. for an immobilized binding protein. The term "artificial" refers to an object that is not naturally occurring, i.e. the term refers to an object that has been produced or modified by man. For example, a polypeptide or polynucleotide sequence that has been generated by man (e.g. for example in a laboratory by genetic engineering, by shuffling methods, or by chemical reactions, etc.) or intentionally modified is artificial.

[0019] The term "dissociation constant" or "K_D" defines the specific binding affinity. As used herein, the term "K_D" (usually measured in "mol/L", sometimes abbreviated as "M") is intended to refer to the dissociation equilibrium constant of the particular interaction between a first protein and a second protein. In the context of the present invention, the term K_D is particularly used to describe the binding affinity between an Fc binding protein and an immunoglobulin. An Fc binding protein of the invention is considered to bind to an immunoglobulin, if it has a dissociation constant K_D to immunoglobulin of at least 1 μM or less, or preferably 100 nM or less, more preferably 50 nM or less, even more preferably 10 nM or less.

[0020] The terms "protein" and "polypeptide" refer to any linear molecular chain of two or more amino acids linked by peptide bonds and does not refer to a specific length of the product. Thus, "peptides", "protein", "amino acid chain," or any other term used to refer to a chain of two or more amino acids, are included within the definition of "polypeptide," and the term "polypeptide" may be used instead of, or interchangeably with any of these terms. The term "polypeptide" is also intended to refer to the products of post-translational modifications of the polypeptide, including without limitation glycosylation, acetylation, phosphorylation, amidation, proteolytic cleavage, modification by non-naturally occurring amino acids and similar modifications which are well-known in the art. Thus, Fc binding proteins comprising two or more protein domains also fall under the definition of the term "protein" or "polypeptides".

[0021] The terms "alkaline stable" or "alkaline stability" or "caustic stable" or "caustic stability" (also abbreviated as "cs" herein) refer to the ability of the Fc binding protein of the invention to withstand alkaline conditions without significantly losing the ability to bind to immunoglobulins. The skilled person in this field can easily test alkaline stability by incubating an Fc binding protein with sodium hydroxide solutions, e.g., as described in the Examples, and subsequent testing of the binding activity to immunoglobulin by routine experiments known to someone skilled in the art, for example, by chromatographic approaches.

[0022] Fc binding proteins of the invention as well as matrices comprising Fc binding proteins of the invention exhibit an "increased" or "improved" alkaline stability, meaning that the molecules and matrices incorporating said Fc binding proteins are stable under alkaline conditions for an extended period of time relative to a reference protein.

[0023] The term "parental" in the term "parental protein" or "parental domain" as used herein refers to an Fc binding protein that is subsequently modified to generate a variant of said parental protein or domain. Said parental protein or domain may be an artificial domain (for example, but not limited to, SEQ ID NO: 78-83), a naturally occurring *Staphylococcus aureus* Protein A domain (for example, SEQ ID NO: 84 for domain C, SEQ ID NO: 85 for domain B), or a variant or engineered version of a naturally occurring *Staphylococcus aureus* Protein A domain (for example, SEQ ID NO: 86 for domain Z).

[0024] The term "variant" as used herein includes an amino acid sequence of an Fc binding protein or domain that differs from another amino acid sequence by at least one amino acid substitution, deletion or insertion. These modifications may be generated by genetic engineering or by chemical synthesis or chemical reactions carried out by man. For example, SEQ ID NO: 27 (cs26 A46C) is a variant of SEQ ID NO: 7 (cs26).

[0025] The term "conjugate" as used herein relates to a molecule comprising or essentially consisting of at least a first protein attached chemically to other substances such as to a second protein or a non-proteinaceous moiety.

[0026] The term "modification" or "amino acid modification" refers to an exchange, a deletion, or an insertion of an amino acid at a particular position in a parent polypeptide sequence by another amino acid. Given the known genetic

code, and recombinant and synthetic DNA techniques, the skilled scientist can readily construct DNAs encoding the amino acid variants.

[0027] The term "substitution" or "amino acid substitution" refers to an exchange of an amino acid at a particular position in a parent polypeptide sequence by another amino acid. For example, the substitution G46C refers to a Fc binding protein, in which the glycine at position 46 is replaced by a cysteine. For the preceding example, 46C refers to a cysteine at position 46. For the purposes herein, multiple substitutions are typically separated by a slash. For example, A11/S11A/K35R/A46C refers to a variant comprising the combination of substitutions A11, S11A, K35R, and A46C.

[0028] The term "deletion" or "amino acid deletion" refers to the removal of an amino acid at a particular position in a parent polypeptide sequence.

[0029] The term "insertions" or "amino acid insertion" refers to the addition of amino acids to the parent polypeptide sequence.

[0030] Throughout this description, the amino acid residue position numbering convention of FIG. 1 is used, and the position numbers are designated as corresponding to those for example in SEQ ID NOs: 1-16.

[0031] The term "amino acid sequence identity" refers to a quantitative comparison of the identity (or differences) of the amino acid sequences of two or more proteins. "Percent (%) amino acid sequence identity" or "percent identical" or "percent identity" with respect to a reference polypeptide sequence is defined as the percentage of amino acid residues in a sequence that are identical with the amino acid residues in the reference polypeptide sequence, after aligning the sequences and introducing gaps, if necessary, to achieve the maximum percent sequence identity.

[0032] To determine the sequence identity, the sequence of a query protein is aligned to the sequence of a reference protein. Methods for alignment are well-known in the art. For example, for determining the extent of an amino acid sequence identity of an arbitrary polypeptide relative to a reference amino acid sequence, the SIM Local similarity program is preferably employed (Xiaoquin Huang and Webb Miller (1991), *Advances in Applied Mathematics*, vol. 12: 337-357), that is freely available (see also: <http://www.expasy.org/tools/sim-prot.html>). For multiple alignment analysis ClustalW is preferably used (Thompson et al. (1994) *Nucleic Acids Res.*, 22(22): 4673-4680). Preferably, the default parameters of the SIM Local similarity program or of ClustalW are used, when calculating sequence identity percentages.

[0033] In the context of the present invention, the extent of sequence identity is generally calculated with respect to the total length of the unmodified sequence, if not explicitly stated otherwise. Each amino acid of the query sequence that differs from the reference amino acid sequence at a given position is counted as one difference. The sum of differences is then related to the length of the reference sequence to yield a percentage of non-identity. The quantitative percentage of identity is calculated as 100 minus the percentage of non-identity.

[0034] As used herein, the phrases "percent identical" or "percent (%) amino acid sequence identity" or "percent identity", in the context of two polypeptide sequences, refer to two or more sequences or subsequences that have in some embodiments at least 89.5 %, in some embodiments at least 91 %, in some embodiments at least 93 %, in some embodiments at least 94 %, in some embodiments at least 96 %, in in some embodiments at least 98 %, and in some embodiments 100 % amino acid residue identity, when compared and aligned for maximum correspondence, as measured using one of the following sequence comparison algorithms or by visual inspection. For clarity reasons, for example a sequence with at least 89.5 % identity includes all sequences with identities higher than 89.5% identity, e.g. embodiments with at least 91 %, at least 93 %, at least 94 %, at least 96 %, at least 98 %, 100 % amino acid identity.

[0035] The percent identity exists in some embodiments over a region of at least 50 residue, at least 52 residues, in some embodiments over a region of at least 53 residues, in some embodiments over a region of at least 54 residues, in some embodiments over a region of at least 55 residues, in some embodiments over a region of at least 56 residues, in some embodiments over a region of at least 57 residues, and in some embodiments over a region of at least 58 residues.

[0036] The term "fused" means that the components are linked by peptide bonds, either directly or via peptide linkers.

[0037] The term "fusion protein" relates to a protein comprising at least a first protein joined genetically to at least a second protein. A fusion protein is created through joining of two or more genes that originally coded for separate proteins. Thus, a fusion protein may comprise a multimer of identical or different proteins which are expressed as a single, linear polypeptide. As used herein, the term "linker" refers in its broadest meaning to a molecule that covalently joins at least two other molecules. In typical embodiments of the present invention, a "linker" is to be understood as a moiety that connects an Fc binding domain with at least one further Fc binding domain, i.e. a moiety linking two protein domains to each other to generate a multimer. In preferred embodiments, the "linker" is a peptide linker, i.e. the moiety linking the two protein domains is one single amino acid or a peptide comprising two or more amino acids.

[0038] The term "chromatography" refers to separation technologies which employ a mobile phase and a stationary phase to separate one type of molecules (e.g., immunoglobulins) from other molecules (e.g. contaminants) in the sample. The liquid mobile phase contains a mixture of molecules and transports these across or through a stationary phase (such as a solid matrix). Due to the differential interaction of the different molecules in the mobile phase with the stationary phase, molecules in the mobile phase can be separated.

[0039] The term "affinity chromatography" refers to a specific mode of chromatography in which a ligand coupled to a stationary phase interacts with a molecule (i.e. immunoglobulin) in the mobile phase (the sample) i.e. the ligand has

a specific binding affinity for the molecule to be purified. As understood in the context of the invention, affinity chromatography involves the addition of a sample containing an immunoglobulin to a stationary phase which comprises a chromatography ligand, such as an Fc binding protein of the invention.

[0040] The terms "solid support" or "solid matrix" are used interchangeably for the stationary phase. The terms "affinity matrix" or "affinity separation matrix" or "affinity chromatography matrix", as used interchangeably herein, refer to a matrix, e.g. a chromatographic matrix, onto which an affinity ligand e.g., an Fc binding protein of the invention is attached. The ligand (e.g., Fc binding protein) is capable of specific binding to a molecule of interest (e.g., an immunoglobulin as defined above) which is to be purified or removed from a mixture.

[0041] The term "affinity purification" as used herein refers to a method of purifying immunoglobulins as defined above from a liquid by binding the immunoglobulins as defined above to an Fc binding protein that is immobilized to a matrix. Thereby, all other components of the mixture except immunoglobulins are removed. In a further step, the bound immunoglobulins is eluted in purified form.

EMBODIMENTS OF THE INVENTION

[0042] The present invention will now be further described. In the following passages different aspects of the invention are defined in more detail. Each aspect defined below may be combined with any other aspect or aspects unless clearly indicated to the contrary. In particular, any feature indicated as being preferred or advantageous may be combined with any other feature or features indicated as being preferred or advantageous.

[0043] In a first aspect the present invention is directed to a Fc binding protein, wherein the amino acid in position 50, the amino acid in position 51, or the amino acid in position 53 corresponding to SEQ ID NO: 2 is Cysteine. In a first aspect, the Fc protein comprises one or more domains, wherein at least one domain comprises or essentially consists of or consists of an amino acid sequence of SEQ ID NO: 2 or an amino acid with at least 89.5 %, at least 91 %, at least 93 %, at least 94 %, at least 96 %, at least 98 %, or 100 % identity wherein the amino acid in position 50, the amino acid in position 51, or the amino acid in position 53 corresponding to SEQ ID NO: 2 is Cysteine.

[0044] The surprising advantage of the Fc binding proteins having a Cysteine in position 50, 51, or 53 is that they confer an alkali stability for a prolonged period of time, without impairing the Fc-binding properties, and with site directed coupling efficiencies to a matrix providing a high binding capacity. The Fc binding domains are three-helix bundles of 58 amino acids with helix 1 from amino acid residues 7-19, helix 2 from amino acid residues 23-37, and helix 3 from amino acid residues 40-55. Amino acid positions 50 and 51 are part of helix 3 of the Fc-binding protein while Fc binding is mediated by helices one and two. The Fc binding proteins of the invention have less than a 20% reduction in binding capacity following an incubation in 0.5 M NaOH for at least 6 hours up to at least 72 hours. This feature is important for chromatography approaches with cleaning procedures using alkaline solutions with high NaOH concentrations to remove contaminants on the matrix so for example that the matrix can be used several times. Further, in addition to having high caustic stability, Fc binding proteins having Cys in helix 3 in position 50, 51, or 53 show high coupling efficiencies. For example, the dynamic binding capacity of variants with Cys in for example position 46 is superior compared to recombinant Protein A (see FIG. 5).

[0045] Preferred Fc binding proteins with Cysteine in helix 3. Disclosed are sequences with an amino acid selected from the group consisting of SEQ ID NOs: 1-83. Disclosed are amino acid sequences with at least 80 %, at least 81 %, at least 82 %, at least 84 %, at least 86 %, at least 87 %, at least 89.5 %, at least 91 %, at least 93 %, at least 94.5 %, at least 96 %, at least 98 %, or 100 % sequence identity to an amino acid selected from the group consisting of SEQ ID NOs: 1-86, 90-95 provided that they have a Cysteine in position 43, 46, 47, 50, 51, or 53. Preferred are embodiments with sequences with at least 89.5 % identity to an amino acid selected from the group consisting of SEQ ID NOs: 2,3-16, 90-95 having a Cysteine in position 50, 51, or 53.

[0046] The amino acid sequences of SEQ ID NOs: 1-86, 90-95 might comprise further modifications, such as insertions, deletions, or further substitutions. In some embodiments, Fc binding domains have 1, 2, 3, 4, 5, or 6 further substitutions. In some embodiments, Fc binding domains have a deletion of 1, 2, 3, or 4 amino acids within the first 4 amino acids of its N-terminus and/or a deletion of 1 or 2 amino acids at the C-terminus. In some embodiments, Fc binding domains have deletions at the N-terminus, for example in positions 1, 2, and 4, or in positions 1, 2, and 3. Disclosed are Fc binding domains having deletions at the C-terminus, for example in positions 57 and/or 58 (as shown for example, SEQ ID NOs: 33-34). Disclosed are amino acid sequences with at least 89.5 % sequence identity to the amino acid sequence to any of the aforementioned SEQ ID NOs (for example SEQ ID NOs: 1-86, 90-95). Examples for Fc binding domains with Cys in position 43, 46, 47, 50, 51, or 53 include for example the amino acid sequences shown in Figure 1.

[0047] SEQ ID NO: 7 (cs26) and variants. In some embodiments, the Fc binding protein is comprising one or more domains of an amino acid sequence of SEQ ID NO: 7 or of an amino acid sequence with at least 89.5 % identity thereto. For example, amino acid sequences with at least 89.5% identity to SEQ ID NO: 7 include cs24 (SEQ ID NO: 8). Examples for variants of cs26 having Cys in position, 50, 51, or 53 include for example SEQ ID NOs: 90-95.

[0048] SEQ ID NO: 8 (cs24) and variants. In some embodiments, the Fc binding protein is comprising one or more

domains of an amino acid sequence of SEQ ID NO: 8 or of an amino acid sequence with at least 89.5 % identity thereto. For example, amino acid sequences with at least 89.5% identity to SEQ ID NO: 8 include cs26 (SEQ ID NO: 7). Examples for variants of cs24 having Cys in position 50, 51, or 53 include for example SEQ ID NOs: 90-95.

[0049] SEQ ID NO: 3 (cs14) and variants. In some embodiments, the Fc binding protein is comprising one or more domains of an amino acid sequence of SEQ ID NO: 3 or of a sequence with at least 89.5 % sequence identity thereto suitable for modifications in position 50, 51, or 53 with Cysteine. Disclosed are, for example, amino acid sequences with at least 89.5% identity to SEQ ID NO: 3 include SEQ ID NO: 10 (cs25), SEQ ID NOs: 11 (cs47h3), SEQ ID NO: 12 (cs47h4), SEQ ID NO: 13 (cs74h1), and SEQ ID NO: 14 (cs74h2). Disclosed are, examples for variants of cs14 having Cys in position 43, 46, 47, 50, 51, or 53 include for example SEQ ID NOs: 17-20, 40-52, 64, 65, 70, 71, 74-76.

[0050] SEQ ID NO: 4 (cs27) and variants. In some embodiments, the Fc binding protein is comprising one or more domains of an amino acid sequence of SEQ ID NO: 4 or of an amino acid sequence with at least 89.5 % identity thereto suitable for modifications in position 50, 51, or 53 with Cysteine. Disclosed are, examples for variants of cs27 having Cys in position 43, 46, 47, 50, 51, or 53 include for example SEQ ID NOs: 21-25.

[0051] SEQ ID NO: 5 (cs20) and variants. In some embodiments, the Fc binding protein is comprising one or more domains of an amino acid sequence of SEQ ID NO: 5 or of an amino acid sequence with at least 89.5 % identity thereto suitable for modifications in position 50, 51, or 53 with Cysteine. Disclosed are, for example, amino acid sequences with at least 89.5% identity to SEQ ID NO: 5 include SEQ ID NO: 9 (cs17). Disclosed are examples for variants of cs20 having Cys in position 43, 46, 47, 50, 51, or 53 include for example SEQ ID NOs: 53-57.

[0052] SEQ ID NO: 6 (cs42) and variants. In some embodiments, the Fc binding protein is comprising one or more domains of an amino acid sequence of SEQ ID NO: 6 or of an amino acid sequence with at least 89.5 % identity thereto suitable for modifications in position 50, 51, or 53 with Cysteine. Disclosed are, for example, amino acid sequences with at least 89.5% identity to SEQ ID NO: 6 include cs28 (different in position 4) or cs41 (SEQ ID NO: 15). Disclosed are examples for variants of cs42 having Cys in position 43, 46, 47, 50, 51, or 53 include for example SEQ ID NOs: 58-63.

[0053] SEQ ID NO: 16 (cs43) and variants. In some embodiments, the Fc binding protein is comprising one or more domains of an amino acid sequence of SEQ ID NO: 16 or of an amino acid sequence with at least 89.5 % identity thereto suitable for modifications in position 50, 51, or 53 with Cysteine. Disclosed are, for example, amino acid sequences with at least 89.5% identity to SEQ ID NO: 16 include cs44 (different in position 44), cs31 (different in positions 25 and 54), or cs45 (different in positions 25 and 26). Examples for variants of cs43 with Cys in position 43, 46, 47, 50, 51, or 53 include for example SEQ ID NOs: 66-69, 72.

[0054] Sequences of Fc binding proteins; preferred amino acid positions.

[0055] Surprisingly, a Cysteine in helix 3, particularly in amino acid positions selected from the group consisting of positions 50, 51, or 53 of Fc binding domains increases the alkaline stability of Fc binding domain compared to a domain without Cysteine in position 50, 51, or 53 as shown in the Figures and in the Examples, and improves the site-specific coupling of the Fc binding protein to a matrix which improves capacity.

[0056] Provided that the Fc binding domain has the cysteines in position 50, 51 or 53, the Fc binding domain includes amino acids at other positions as follows: In some embodiments, said Fc binding domain comprises an Isoleucine at position 1. It is preferred that the amino acid in position 1 of the Fc binding domain is not Threonine (T). It is preferred that the amino acid in position 1 is Isoleucine (I) or Alanine (A). Alternatively, position 1 may be deleted. In some embodiments, said Fc binding domain comprises an Alanine (A), Glutamic Acid (E), or Isoleucine (I) at position 11. It is preferred that the amino acid in position 11 is not Asparagine (N) or Lysine (K). It is preferred that the amino acid in position 11 is Alanine (A), Isoleucine (I), Glutamic acid (E), Histidine (H), or Proline (P), more preferred A, I, or E, most preferred A. Alternatively, the amino acid in position 11 is Serine (S). In some embodiments, said Fc binding domain comprises an Arginine (R) or Isoleucine (I) at position 35. It is preferred that the amino acid in position 35 is not Proline (P), Asparagine (N), Glycine (G), Tryptophan (W), Alanine (A), Glutamine (Q), or Methionine (M). In some embodiment, said Fc binding domain comprises a Leucine (L) at position 42. It is preferred that the amino acid in position 42 is not Tyrosine (Y). In some embodiments, in addition to 50C, 51C, or 53C, the Fc binding domain comprises 2, 3 or 4 of the amino acid positions selected from the group consisting of 11, 11A, 11S, 35R, and 42L. In some embodiments, at least 90 % of positions 11, 3A, 6D, 9Q, 10Q, 12A, 13F, 14Y, 15E, 16I, 17L, 18H, 19L, 20P, 21N, 22L, 23T, 24E, 26Q, 27R, 28N, 29A, 30F, 31I, 32Q, 33S, 34L, 36D, 37D, 38P, 39S, 41S, 42L, 45L, 48A, 52N, 55Q, 56A, 57P are identical in Fc binding domains of the invention. It is preferred that position 2 is A or D, position 4 is K or Q, position 5 is H or F, position 7 is K or E, position 8 is D, A or E, position 11 is A, S, I, or E, position 25 is D or E, position 35 is R or I, position 40 is V, T or Q, position 43 is E, S or C, position 44 is I, V or L, position 46 is C, A or G, position 47 is E or C, position 49 is K or Q, position 50 is K or C, position 51 is L or C, position 53 is D or E or C, position 54 is A or S, and position 58 is P or K. An Fc binding protein of the invention comprises one or more Fc binding domains that comprises or essentially consists or consists of the amino acid sequence of SEQ ID NO: 2 or at least 89.5 % identical amino acid sequences thereto wherein an amino acid in position 50, an amino acid in position 51, or an amino acid in position 53 corresponding to SEQ ID NO: 2 is Cysteine. The amino acid sequence SEQ ID NO: 2 is the following amino acid sequence: IX₂AX₄X₅DX₇X₈QX₁₁AFYEILHLPNLTEX₂₅QRNAFIQSLX₃₅DDPSX₄₀SLX₄₃X₄₄LX₄₆X₄₇AX₄₉X₅₀

$X_{51}NX_{53}X_{54}QAPX_{58}$ wherein the amino acid at position 1 is selected from I, or is deleted, the amino acid at position 2 (X_2) is selected from A or D, or is deleted, the amino acid at position 3 is selected from A, or is deleted, the amino acid at position 4 (X_4) is selected from K or Q, or is deleted, the amino acid at position 5 (X_5) is selected from H or F, the amino acid at position 7 (X_7) is selected from K or E, the amino acid at position 8 (X_8) is selected from D, A, or E, the amino acid at position 11 (X_{11}) is selected from A, S, I, or E, the amino acid at position 25 (X_{25}) is selected from D or E, the amino acid at position 35 (X_{35}) is selected from R or I, the amino acid at position 40 (X_{40}) is selected from Q, T, or V, the amino acid at position 43 (X_{43}) is selected from E, or S, , the amino acid at position 44 (X_{44}) is selected from I, L, or V, the amino acid at position 46 (X_{46}) is selected from G, or A, the amino acid at position 47 (X_{47}) is E , the amino acid at position 49 (X_{49}) is selected from K or Q, the amino acid at position 50 (X_{50}) is K or C, the amino acid at position 51 (X_{51}) is L or C, the amino acid at position 53 (X_{53}) is selected from D, E or C, the amino acid at position 54 (X_{54}) is selected from A or S, the amino acid at position 57 is selected from P, or deleted, and the amino acid at position 58 (X_{58}) is selected from P or K, or deleted. Hence, Fc binding proteins of the invention have a cysteine in one of the following positions: 50, 51, or 53. Selected examples for Fc binding domains of the invention include SEQ ID NOs: 90-95.

[0057] High alkaline stability as result of Cysteine in the c-terminal region of Fc binding proteins. In some embodiments, Fc binding proteins with Cysteine in position 50, 51, or 53 provide surprisingly particularly good alkaline stability of the Fc binding protein, as shown in the Examples and in the Figures. It was most surprising and unexpected that the Fc binding proteins with Cysteine in helix 3 are able to bind to Ig even after alkaline treatment for several hours. The alkaline stability of the Fc binding protein is determined by comparing the loss in Ig binding activity after at least 6 h incubation in 0.5 M NaOH (see FIG. 3). For example, the binding capacity of cs26 46C remains at least 20% higher than wildtype domain C after prolonged (for example, 36 h) incubation with 0.5 M NaOH (see FIG. 4).

[0058] Affinity to Immunoglobulin. All Fc binding proteins of the invention bind to Immunoglobulin with a dissociation constant K_D preferably below 500 nM, or below 100 nM, even more preferably 10 nM or less. Methods for determining binding affinities of Fc binding proteins or domains, i.e. for determining the dissociation constant K_D , are known to a person of ordinary skill in the art and can be selected for instance from the following methods known in the art: Surface Plasmon Resonance (SPR) based technology, Bio-layer interferometry (BLI), enzyme-linked immunosorbent assay (ELISA), flow cytometry, isothermal titration calorimetry (ITC), analytical ultracentrifugation, radioimmunoassay (RIA or IRMA) and enhanced chemiluminescence (ECL). Some of the methods are described further in the Examples. Typically, the dissociation constant K_D is determined at 20 °C, 25 °C, or 30 °C. If not specifically indicated otherwise, the K_D values recited herein are determined at 22 °C +/- 3 °C by surface plasmon resonance. In an embodiment of the first aspect, the Fc binding protein has a dissociation constant K_D to human IgG₁ in the range between 0.1 nM and 100 nM, preferably between 0.1 nM and 50 nM.

[0059] Multimers. In one embodiment of the invention, the Fc binding protein comprises 1, 2, 3, 4, 5, 6, 7, or 8, preferably 2, 3, 4, 5, or 6, Fc binding domains linked to each other, i.e. the Fc binding protein can be, for example, a monomer, a dimer, a trimer, a tetramer, a pentamer, or a hexamer. A multimer may comprise two, three, four, or even more binding domains.

[0060] Multimers of the invention are fusion proteins generated artificially, generally by recombinant DNA technology well-known to a skilled person.

[0061] In some embodiments, the multimer is a homo-multimer, e.g. the amino acid sequences of all Fc binding domains of the Fc binding protein are identical.

[0062] A multimer may comprise two or more Fc binding domains, wherein said Fc binding domains preferably comprise or essentially consist of a sequence as described above, provided that they have a Cysteine in position 50, 51, or 53.

[0063] Disclosed are, for example, SEQ ID NO: 27 or SEQ ID NO: 22 were used to generate the homo-multimeric fusion constructs described herein in Example 1, see for example SEQ ID NO: 32 , SEQ ID NO: 35, or SEQ ID NO: 25.

[0064] In some embodiments, the multimer is a hetero-multimer, e.g. at least one alkaline stable Fc binding domain has a different amino acid sequence than the other Fc binding domains within the immunoglobulin-binding protein.

[0065] Linker. In some embodiments of the first aspect, the one or more Fc binding domains are directly linked to each other. In other embodiments, the one or more Fc binding domains are linked to each other with one or more linkers. Preferred in these typical embodiments are peptide linkers. This means that the peptide linker is an amino acid sequence that connects a first Fc binding domain with a second Fc binding domain. The peptide linker is connected to the first Fc binding domain and to the second Fc binding domain by a peptide bond between the C-terminal and N-terminal ends of the domains, thereby generating a single, linear polypeptide chain. The length and composition of a linker may vary between at least one and up to about 30 amino acids. More specifically, a peptide linker has a length of between 1 and 30 amino acids; e.g., 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 11, 12, 13, 14, 15, 16, 17, 18, 19, 20, 21, 22, 23, 24, 25, 26, 27, 28, 29, 30 amino acids. It is preferred that the amino acid sequence of the peptide linker is stable against caustic conditions and proteases. Linkers should not destabilize the conformation of the domains in the Fc binding protein. Well-known are linkers that comprise or consist of small amino acids such as glycine and serine. The linkers can be glycine-rich (e.g., more than 50% of the residues in the linker can be glycine residues). Also preferred are linkers that comprise further amino acids. Other embodiments of the invention comprise linkers consisting of alanine, proline, and serine.

Other linkers for the fusion of proteins are known in the art and can be used. In some embodiments, the multimer of Fc binding proteins comprises one or more linkers connecting the Fc binding domains wherein the linker are identical or different.

[0066] Conjugation to a solid support. In some embodiments of the invention, the Fc binding protein is conjugated to a solid support. Cysteine in helix 3 in position 50, 51, or 53 of the Fc binding domain comprises an attachment site for site-specific covalent coupling of the Fc binding protein to a solid support. Cysteine in position 50, 51, or 53 enables specific chemical reactions with a reactive group of the solid phase or a linker between the solid phase and the protein, for example selected from N-hydroxysuccinimide, iodacetamide, maleimide, epoxy, or alkene groups.

[0067] In some embodiments of the invention, the Fc binding protein may also comprise additional amino acid residues at the N- and/or C-terminal end, such as for example an additional sequence with or without a tag at the N- and/or C-terminal end.

[0068] Affinity separation matrix. In another aspect the present invention is directed to an affinity separation matrix, comprising an Fc binding protein of the first aspect.

[0069] In preferred embodiments of the second aspect, the affinity separation matrix is a solid support. The affinity separation matrix comprises at least one Fc binding protein of the invention.

[0070] An affinity matrix is useful for separation of immunoglobulins and should retain the Ig binding property even after highly alkaline conditions as applied during cleaning processes. Such cleaning of matrices is essential for long-term repeated use of matrices.

[0071] Solid support matrices for affinity chromatography are known in the art and include for example but are not limited to, agarose and stabilized derivatives of agarose (e.g. Sepharose 6B,

[0072] Praesto™ Pure; CaptivA®, rPROTEIN A Sepharose Fast Flow, Mabselect®, Prisma®, and other), cellulose or derivatives of cellulose, controlled pore glass (e.g. ProSep® vA resin), monolith (e.g. CIM® monoliths), silica, zirconium oxide (e.g. CM Zirconia or CPG®), titanium oxide, or synthetic polymers (e.g. polystyrene such as Poros 50A or Poros MabCapture® A resin, polyvinylether, polyvinyl alcohol, polyhydroxyalkyl acrylates, polyhydroxyalkyl methacrylates, polyacrylamides, polymethacrylamides etc) and hydrogels of various compositions. In certain embodiments the support comprises a polyhydroxy polymer, such as a polysaccharide. Examples of polysaccharides suitable for supports include but are not limited to agar, agarose, dextran, starch, cellulose, pullulan, etc, and stabilized variants of these.

[0073] The formats for solid support matrices can be of any suitable well-known kind. Such solid support matrix for coupling the Fc binding protein of the invention might comprise for example, one of the following: columns, capillaries, particles, membranes, filters, monoliths, fibers, pads, gels, slides, plates, cassettes, or any other format commonly used in chromatography and known to someone skilled in the art.

[0074] In one embodiment, the matrix is comprised of substantially spherical particles, also known as beads, for example Sepharose or Agarose beads. Suitable particle sizes may be in the diameter range of 5-500 μm, such as 10-100 μm, such as 20-80 μm, such as 40-70 μm. Matrices in particle form can be used as a packed bed or in a suspended form including expanded beds.

[0075] In an alternative embodiment, the solid support matrix is a membrane, for example a hydrogel membrane. In some embodiments, the affinity purification involves a membrane as matrix to which the Fc binding protein of the first aspect is covalently bound. The solid support can also be in the form of a membrane in a cartridge.

[0076] In some embodiments, the affinity purification involves a chromatography column containing a solid support matrix to which the Fc binding protein of the first aspect is covalently bound.

[0077] The Fc binding protein of the invention may be attached to a suitable solid support matrix via conventional coupling techniques. Methods for immobilization of protein ligands to solid supports are well-known in this field and easily performed by the skilled person in this field using standard techniques and equipment.

[0078] Use of the Fc binding protein. In a third aspect the present invention is directed to the use of the Fc binding protein of the first aspect or an affinity matrix of the second aspect for affinity purification of immunoglobulins or variants thereof, i.e. the Fc binding protein of the invention is used for affinity chromatography. In some embodiments, the Fc binding protein of the invention is immobilized onto a solid support as described in the second aspect of the invention.

[0079] Method of affinity purification of immunoglobulins. In a fourth aspect the present invention is directed to a method of affinity purification of immunoglobulins, the method comprising (a) providing a liquid containing an immunoglobulin; (b) providing an affinity separation matrix comprising an immobilized Fc binding protein of the first aspect coupled to said affinity separation matrix; (c) contacting said liquid with said affinity separation matrix, wherein said immunoglobulin binds to said immobilized Fc binding protein; and (d) eluting said immunoglobulin from said matrix, thereby obtaining an eluate containing said immunoglobulin. In some embodiments, the method of affinity purification may further comprising one or more washing steps carried out between steps (c) and (d) under conditions sufficient to remove from the affinity separation matrix some or all molecules that are non-specifically bound thereto. Non-specifically bound means any binding that does not involve an interaction between the at least one binding domain of the presently disclosed subject matter and an Immunoglobulin.

[0080] Affinity separation matrixes suitable for the disclosed uses and methods are those matrixes according to the

embodiments described above and as known to someone skilled in the art.

[0081] In some embodiments of the fourth aspect, the elution of the immunoglobulin from the matrix in step (d) is effected through a change in pH and/or a change in salt concentration. Any suitable solution can be used, for example by a solution with pH 5 or lower (see e.g. Table 3 in Example 9), or by a solution with pH 11 or higher.

[0082] In some embodiments, a further step (f) for efficient cleaning and sanitization of the affinity matrix is added, preferably by using an alkaline liquid, for example, with pH of 13 - 14. In certain embodiments, the cleaning liquid comprises 0.1 - 1.0 M NaOH or KOH, preferably 0.25 - 0.5 M NaOH or KOH. Due to the high alkaline stability of the Fc binding proteins of the invention, such strong alkaline solution can be used for cleaning purposes. In some embodiments, the affinity matrix can be re-used at least 10 times, at least 20 times, at least 30 times, at least 40 times, at least 50 times, at least 60 times, at least 70 times, at least 80 times, at least 90 times, or at least 100 times, due to a repetition of steps (a) to (e), optionally (a) to (f) can be repeated at least 10 times, at least 20 times, at least 30 times, at least 40 times, at least 50 times, at least 60 times, at least 70 times, at least 80 times, at least 90 times, or at least 100 times.

[0083] In general, suitable conditions for performing the method of affinity purification are well known to someone skilled in the art. In some embodiments, the disclosed uses or methods of affinity purification comprising the disclosed Fc binding domains may provide elution of at least about 90%, at least about 95 %, at least about 96 %, at least about 97 %, at least about 98 %, at least about 99 % of Fc containing proteins at a pH of greater than or equal to 3.5 (e.g., about 4.0, about 4.5, about 5.0, about 5.5, about 6.0, or about 6.5).

[0084] Nucleic acid molecule. In a fifth aspect, the present invention is directed to a nucleic acid molecule, preferably an isolated nucleic acid molecule, encoding a Fc binding protein of any embodiment disclosed above. In one embodiment, the present invention is directed to a vector comprising the nucleic acid molecule. A vector means any molecule or entity (e.g., nucleic acid, plasmid, bacteriophage or virus) that can be used to transfer protein coding information into a host cell. In one embodiment, the vector is an expression vector.

[0085] In a sixth aspect, the present invention is directed to an expression system which comprises a nucleic acid or a vector as disclosed above, for example a prokaryotic host cell, for example *E. coli*, or a eukaryotic host, for example yeast *Saccharomyces cerevisiae* or *Pichia pastoris* or mammalian cells such as CHO cells.

[0086] Method for the production of a Fc binding protein. In a seventh aspect the present invention is directed to a method for the production of a Fc binding protein of the invention, comprising the step(s): (a) culturing the host cell of the sixth aspect under suitable conditions for the expression of the binding protein in order to obtain said Fc binding protein; and (b) optionally isolating said Fc binding protein. Suitable conditions for culturing a prokaryotic or eukaryotic host are well-known to the person skilled in the art.

[0087] Fc binding molecules of the invention may be prepared by any of the many conventional and well-known techniques such as plain organic synthetic strategies, solid phase-assisted synthesis techniques or by commercially available automated synthesizers. On the other hand, they may also be prepared by conventional recombinant techniques alone or in combination with conventional synthetic techniques.

[0088] One embodiment of the present invention is directed to a method for the preparation of a Fc binding protein according to the invention as detailed above, said method comprising the following steps: (a) preparing a nucleic acid encoding an Fc binding protein as defined above; (b) introducing said nucleic acid into an expression vector; (c) introducing said expression vector into a host cell; (d) cultivating the host cell; (e) subjecting the host cell to culturing conditions under which an Fc binding protein is expressed, thereby (e) producing an Fc binding protein as described above; optionally (f) isolating the protein produced in step (e); and (g) optionally conjugating the protein to solid matrices as described above.

[0089] In a further embodiment of the present invention the production of the Fc binding protein is performed by cell-free *in vitro* transcription / translation.

EXAMPLES

[0090] The following Examples are provided for further illustration of the invention. The invention, however, is not limited thereto, and the following Examples merely show the practicability of the invention on the basis of the above description.

Example 1. Generation of artificial mosaic proteins that bind to Fc

[0091] Fc binding proteins were initially generated by a shuffling process of naturally occurring Protein A domains (E, B, D, A, C, Z). In more detail, the shuffling process as understood herein is an assembly process resulting in artificial amino acid sequences starting from a set of non-identical known amino acid sequences. The shuffling process comprised the following steps: a) providing sequences of five naturally occurring Protein A domains E, B, D, A, and C, and Protein A variant domain Z; b) alignment of said sequences; c) statistical fragmentation *in silico* to identify subsequences that were recombined, and then d) assembly of new, artificial sequences of the various fragments to produce a mosaic

EP 3 521 304 B9

product, i.e. a novel and artificial amino acid sequence. The fragments generated in step c) were of any length, e.g. if the fragmented parent sequence had a length of n, the fragments was of length 1 to n-1.

[0092] The relative positions of the amino acids in the mosaic products were maintained with respect to the starting amino acid sequences. At least 90 % of positions Q9, Q10, A12, F13, Y14, L17, P20, L22, Q26, R27, F30, I31, Q32, S33, L34, D36, D37, P38, S39, S41, L45, E47, A48, K50, L51, Q55, A56, P57 are identical between the artificial amino acid sequences of for example IB14, IB27, IB24, IB26, IB28, IB20 (SEQ ID NOs: 78-83) and naturally occurring Protein A domains or Protein A domain variants. The overall amino acid sequence of, for example, Fc binding proteins IB14, IB26, and IB27 is artificial in that it is not more than 85 % identical to the overall amino acid sequence of any of the naturally occurring Protein A domains or domain Z. After the initial artificial Fc binding proteins was generated, in some cases the protein was further modified by site-specific randomization of the amino acid sequence to further modify biochemical properties. The further modifications were introduced by site-saturation mutagenesis of individual amino acid residues.

[0093] Genes for artificial Fc binding proteins were synthesized and cloned into an *E. coli* expression vector using standard methods known to a skilled person. DNA sequencing was used to verify the correct sequence of inserted fragments.

[0094] To generate multimeric Fc binding proteins, 2 or 3 identical Fc binding domains were genetically fused.

[0095] For specific purification, optionally a strep-tag (WSHPQFEK; SEQ ID NO: 89) was added to the C-terminus of the Fc binding proteins.

Example 2. Mutagenesis to generate variants

[0096] For site-directed mutagenesis, the Q5[®] site-directed Mutagenesis Kit (NEB; Cat. No. E0554S) was used according to the manufacturer's instructions. PCRs were carried out with oligonucleotides coding for each specific substitution respectively and a plasmid containing the template. Products were ligated and transformed into *E. coli* XL2-blue cells (Stratagene) via electroporation. Single colonies were isolated and DNA sequencing was used for insert containing clones to verify the correct sequences.

[0097] A combination of several point mutations was generated by GeneArt™ Strings™ synthesis (Thermo Fisher Scientific). The Strings DNA fragments corresponded to a purified PCR product and were cloned into a derivative of a pET28a vector. Ligation products were transformed into *E. coli* XL2-blue cells via electroporation. Single colonies were screened by PCR to identify constructs containing inserts of the right size. DNA sequencing was used to verify the correct sequences. Some variants with point mutations are shown for example in **Figure 1**.

Example 3. Expression of Fc binding proteins

[0098] BL21 (DE3) competent cells were transformed with an expression plasmid encoding Fc binding proteins. Cells were spread onto selective agar plates (Kanamycin) and incubated overnight at 37°C. Precultures were inoculated from single colony in 100 ml 2xYT medium and cultured for 16 hours at 37 °C at 160 rpm in a conventional orbital shaker in baffled 1 L Erlenmeyer flasks supplemented with 150 µg/ml Kanamycin without lactose and antifoam. The OD₆₀₀ readout should be in the range of 6-12. Main culture was inoculated from previous overnight culture with an adjusted start-OD₆₀₀ of 0.5 in 400 ml superrich medium (modified H15 medium 2% Glucose, 5% Yeast extract, 0.89% Glycerol, 0.76% Lactose, 250 mM MOPS, 202 mM TRIS, pH 7.4, Antifoam SE15) in 1 L thick-walled Erlenmeyer flasks that was supplemented with 150 µg/ml Kanamycin. Cultures were transferred to a resonant acoustic mixer (RAMbio) and incubated at 37 °C with 20 x g. Aeration was facilitated by Oxy-Pump stoppers. Recombinant protein expression was induced by metabolizing glucose and subsequently allowing lactose to enter the cells. At predefined time points OD₆₀₀ was measured, samples adjusted to 5/OD₆₀₀ were withdrawn, pelleted and frozen at -20 °C. Cells were grown overnight for approx. 24 hours to reach a final OD₆₀₀ of about 45-60. To collect biomass cells were centrifuged at 16000 x g for 10 min at 20 °C. Pellets were weighed (wet weight) and pH was measured in the supernatant. Cells were stored at -20 °C before processing.

Example 4: SDS-PAGE Analysis of expression and solubility of Fc binding proteins

[0099] Samples taken during fermentation were resuspended in 300 µl extraction buffer (PBS supplemented with 0.2 mg/ml Lysozyme, 0.5x BugBuster, 7.5 mM MgSO₄, 40 U Benzonase) and solubilized by agitation in a thermomixer at 700 rpm, rt for 15 min. Soluble proteins were separated from insoluble proteins by centrifugation (16000 x g, 2 min, rt). Supernatant was withdrawn (soluble fraction) and the pellet (insoluble fraction) was resuspended in equivalent amount of urea buffer (8 M urea, 0.2 M Tris, 2 mM EDTA, pH 8.5). 50 µl were taken both from the soluble and insoluble fraction, and 12 µl 5x sample buffer as well as 5 µl 0.5 M DTT were added. Samples were boiled at 95 °C for 5 min. Finally, 8 µl of those samples were applied to NuPage Novex 4-12 % Bis-Tris SDS gels which were run in accordance to the manufacturer's recommendations and stained with Coomassie. High level expression of all Fc binding proteins was

found under optimized conditions within the chosen period of time (data not shown). All expressed Fc binding proteins were soluble to more than 95 % according to SDS-PAGE.

Example 5: Purification of Fc binding proteins

[0100] Fc binding proteins were expressed in the soluble fraction of *E. coli* with a C-terminal StrepTagII (WSHPQFEK). The cells were lysed by two freeze/thaw cycles and the purification step was performed with Strep-Tactin[®]-resin according to the manufacturer's instructions (IBA, Goettingen, Germany). To avoid disulfide formation the buffers were supplemented with 1 mM DTT.

[0101] Alternatively, Fc binding proteins were expressed in the soluble fraction of *E. coli* with a C-terminal StrepTagII. The cells were resuspended in cell disruption buffer and lysed by a constant cell disruption system (Unit F8B, Holly Farm Business Park) at 1 kbar for two cycles. Purification step was performed with Strep-Tactin-resin (IBA, Goettingen, Germany) and additional gel filtration (Superdex 75 16/60; GE Healthcare) using an ÄKTExpress system (GE Healthcare) according to the manufacturer's instructions. To avoid disulfide formation buffers for Strep-Tactin-purification were supplemented with 1 mM DTT and citrate-buffer (20 mM Citrat, 150 mM NaCl, pH 6,0) was used as running buffer for gel filtration.

Example 6. The Fc binding proteins bind to IgG with high affinities (as determined by ELISA)

[0102] The affinities of the Fc binding proteins towards IgG₁ or IgG₂ or IgG₄ were determined using an Enzyme Linked Immunosorbent Assay (ELISA). IgG₁ or IgG₂ or IgG₄ containing antibodies (e.g. Cetuximab for IgG₁, Panitumumab for IgG₂, or Natalizumab for IgG₄) were immobilized on a 96 well Nunc MaxiSorb ELISA plate (2 µg/ml). After incubation for 16 h at 4 °C the wells were washed three times with PBST (PBS + 0.1 % Tween 20) and the wells were blocked with 3 % BSA in PBS (2 h at room temperature). The negative controls were wells blocked only with BSA. After blocking, the wells were washed three times with PBST and incubated for 1 h with the Fc binding protein (in PBST) at room temperature. After incubation the wells were washed three times with PBST and subsequently incubated with Strep-Tactin-HRP (1 :10000) (IBA, Goettingen, Germany) for 1 h at room temperature. Afterwards the wells were washed three times with PBST and three times with PBS. The activity of the horseradish peroxidase was visualized by adding TMB-Plus substrate. After 30 min the reaction was stopped by adding 0.2 M H₂SO₄ and the absorbance was measured at 450 nm. For example, as determined via ELISA, the K_D for human IgG₁ is 4.9 nM for IB14; 3.4 nM for domain Z; 3.1 nM for domain B; and 2.8 nM for domain C.

Example 7. The Fc binding proteins bind to IgG with high affinities (as determined with surface plasmon resonance experiments)

[0103] A CM5 sensor chip (GE Healthcare) was equilibrated with SPR running buffer. Surface-exposed carboxylic groups were activated by passing a mixture of EDC and NHS to yield reactive ester groups. 700-1500 RU *on*-ligand were immobilized on a flow cell, *off*-ligand was immobilized on another flow cell. Injection of ethanolamine after ligand immobilization removes non-covalently bound Fc binding protein. Upon ligand binding, protein analyte was accumulated on the surface increasing the refractive index. This change in the refractive index was measured in real time and plotted as response or resonance units (RU) versus time. The analytes were applied to the chip in serial dilutions with a suitable flow rate (µl/min). After each run, the chip surface was regenerated with regeneration buffer and equilibrated with running buffer. The control samples were applied to the matrix. Regeneration and re-equilibration were performed as previously mentioned. Binding studies were carried out by the use of the Biacore[®] 3000 (GE Healthcare) at 25 °C; data evaluation was operated via the BIAevaluation 3.0 software, provided by the manufacturer, by the use of the Langmuir 1:1 model (RI=0). Evaluated dissociation constants (K_D) were standardized against *off*-target and K_D values of different artificial Fc binding proteins for human IgG₁-Fc, Cetuximab (IgG₁), Natalizumab (IgG₄), or Panitumomab (IgG₂) are shown in Table 1.

Table 1. K_D values of Fc binding proteins for Ig

Fc binding protein	SEQ ID NO:	Kd [nM] IgG-Fc	Kd [nM] IgG ₁	Kd [nM] IgG ₄	Kd [nM] IgG ₂
IB14	78	1,28			
IB14 E43C	74	45,5			
IB14 G46C	75	72,9			
IB14 E47C	76	48,8			

(continued)

Fc binding protein	SEQ ID NO:	Kd [nM] IgG-Fc	Kd [nM] IgG ₁	Kd [nM] IgG ₄	Kd [nM] IgG ₂
IB14 K50C	77	42,8			
cs14	3	6,48	2.9	2,51	7,42
cs14 G46C	18		4,88	3,38	35,6
cs14 E43C	17		0,453	0,302	1,75
cs27	4		3,64	2,54	21,6
cs27 G46C	22		0,518	0,302	2,74
cs26	7		6,5	5,81	66,6
cs26 A46C	27		5,75	3,00	101
cs26 A46C (dimer)	32		2,59	1,86	20,5
cs26 A46C (trimer)	35		2,23	1,98	13,3
Domain C	84		4,91	3,95	58,1

Example 8. Fc binding proteins coupled to an epoxy-activated matrix (Sephacrose 6B)

[0104] Purified Fc binding proteins were coupled to epoxy-activated matrix (Sephacrose 6B, GE; Cat. No. 17-0480-01) according to the manufacturer's instructions (coupling conditions: pH 9.0 overnight, blocking for 5 h with ethanolamine). Cetuximab was used as IgG sample (5mg; 1 mg/ml matrix). Cetuximab was applied in saturated amounts to the matrix comprising immobilized Fc binding protein. The matrix was washed with 100 mM glycine buffer, pH 2.5 to elute Cetuximab that was bound to the immobilized IgG-binding protein. The concentration of the eluted IgG was measured by BLI (quantification with Protein A Octet-sensors and Cetuximab as standard) in order to determine the binding activity of the Fc binding proteins. **FIG. 2A** shows the coupling efficiency of IB14 (SEQ ID NO: 78) and IB14 variants with Cysteine in Positions 43, 46, 47, 50, or 58 (SEQ ID NOs: 74-77, respectively). The coupling efficiency of all IB14 variants with Cys in positions 43, 46, 47, 50, or 58 was higher than for IB14. **FIG. 2B** shows the coupling of cs14 46C (SEQ ID NO: 18) or cs14 43C (SEQ ID NO: 17) to Epoxy activated Sepharose 6B matrix compared to cs14 with an C-terminal Cys and compared to a commercially available Protein A. Coupling conditions were 450 μ M, 4500 nmol/ml, pH 9, 2 h or 18 h at 30°C, 1mM TCEP, 1 M (NH₄)₂SO₄. Only minor change in net coupling rate were observed between 2 h and 18 h. Coupling rates are shown in nmole (domain)/ml.

Example 9. Alkaline stability of Fc binding proteins coupled to an epoxy-activated matrix

[0105] Columns were incubated with 0.5 M NaOH for 0 h, 6 h, 18 h, 24 h, 36 h, or 72 h at room temperature (22 °C +/- 3 °C). The Ig binding activity of the immobilized proteins was analyzed before and after incubation with 0.5 M NaOH. The Ig binding activity of immobilized proteins before NaOH treatment was defined as 100 %. The remaining IgG binding activity after continuous 0.5 M NaOH treatment for 6 hours is shown in **FIG. 3A** for IB14 variants (SEQ ID NOs: 74-77). **FIG. 3B** shows caustic stability after continuous 0.5 M NaOH treatment for 0 h, 18 h, or 72 h for cs14 46C, cs14 43C, cs14, and for a commercially available Protein A. The proteins were immobilized to Epoxy Sepharose for 2 h at 30°C (4500 nmol/ml). The dynamic binding capacity DBC 10% was determined at 5 min residence time. Cs14 46C shows the highest DBC10% after 72 h of continuous 0.5 NaOH treatment for 19,5 mg/ml which is more than 20 % more than the value measured for the commercially available Protein A. **FIG. 4** shows the remaining binding capacity after continuous 0.5 M NaOH treatment for 6, 24, and 36 hours for cs26 46C (SEQ ID NO: 27) compared to cs26 (SEQ ID NO: 7) and to wildtype domain C (SEQ ID NO: 84). The binding capacity of cs26 A46C (monomer or dimer) remains > 20% after at least after 36 h 0.5 M NaOH incubation, see Table 2.

Table 2. Caustic stability of cs26 A46C compared to cs26 and to wildtype domain C

Fc binding protein	0 h	6 h	24 h	36 h	vs C @ 36 h
cs26 A46C	100	95	83	78	42%
cs26 A46C (dimer)	100	92	81	72	24.8 %
cs26	100	105	82	73	27.7 %

(continued)

Fc binding protein	0 h	6 h	24 h	36 h	vs C @ 36 h
domain C	100	95	76	62	

Example 10. Fc binding proteins coupled to agarose-based chromatography beads Praesto™ Pure45

[0106] Purified Fc binding proteins were coupled to agarose-based chromatography beads (Praesto™ Pure45, Purolite; Cat. No. PR01262-166) according to the manufacturer's instructions (coupling conditions: pH 9.5, 3 hours, 35 °C, 4.1 M NaSO₄, blocking overnight with ethanolamine). Polyclonal human IgG Gammanorm® (Ocatpharm) was used as IgG sample (conc. 2,2 mg/ml). Polyclonal hlgG sample was applied in saturated amounts to the matrix comprising immobilized Fc binding protein. The matrix was washed with 100 mM Citrate buffer, pH 2.0 to elute hlgG that was bound to the immobilized Fc binding protein. Dynamic binding capacity was determined for cs26 46C (monomer and dimer; (SEQ ID NOs: 27, 32) compared to recombinant wildtype Protein A by the mass of injected hlgG at 10% breakthrough at 6 min residence time. FIG. 5 shows that Cs26 46C has a 61.2 % higher DBC at 6 min residence time than recombinant Protein A.

Example 11. Elution of IgG from Fc binding proteins coupled to agarose-based chromatography beads Praesto™ Pure45 and/or Pure85

[0107] Purified Fc binding proteins (cs26 46C) were coupled to agarose-based chromatography beads (Praesto™ Pure45 or Pure 5) according to the manufacturer's instructions. Polyclonal human IgG Gammanorm® and monoclonal IgG₁, antibody Cetuximab was used as IgG sample (conc. 2.2 mg/ml), loading up to DBC10%. Polyclonal hlgG sample was applied in saturated amounts to the matrix comprising immobilized Fc binding protein. In a two-step process, the matrix was first washed with 100 mM Citrate buffer, pH 3.5 and then with 100 mM Citrate buffer, pH 2.0 to elute hlgG that was bound to the immobilized Fc binding protein. Table 3 shows that almost 100 % of the bound IgG was eluted from beads coupled with cs26 46C at pH 3.5.

Table 3. Elution of IgG from cs26 46C coupled to beads

Antibody	Eluted protein at 0.1 M citrate pH 3.5 (%)	Eluted protein at 0.1 M citrate pH 2.0 (%)	Recovery: Load/Elution (%)
Gammanorm	99.8	0.2	85
Cetuximab	100	0	86

Claims

1. An Fc binding protein comprising one or more domains, wherein the amino acid in position 50, the amino acid in position 51, or the amino acid in position 53 corresponding to SEQ ID NO: 2 is Cysteine, wherein said Fc binding protein is stable under alkaline conditions.
2. The Fc binding protein of claim 1, wherein at least one domain comprises an amino acid sequence of SEQ ID NO: 2 or an amino acid sequence with at least 89.5 % identity thereto wherein the amino acid in position 50, the amino acid in position 51, or the amino acid in position 53 corresponding to SEQ ID NO: 2 is Cysteine.
3. The Fc binding protein of claim 1, wherein at least one domain comprises an amino acid sequence of SEQ ID NOs: 3-16 or an amino acid sequence with at least 89.5 % identity thereto, respectively, wherein the amino acid in 50, the amino acid in position 51, or the amino acid in position 53 corresponding to SEQ ID NOs: 3-16 is Cysteine.
4. The Fc binding protein of claim 1, wherein at least one domain comprises an amino acid sequence of SEQ ID NOs: 7-8 or an amino acid sequence with at least 89.5 % identity thereto, respectively, wherein the amino acid in position 50, the amino acid in position 51, or the amino acid in position 53 corresponding to SEQ ID NOs: 7-8 is Cysteine.
5. The Fc binding protein of claim 1, wherein at least one domain comprises an amino acid sequence selected from the group of SEQ ID NOs: 77, and 90 - 95, or an amino acid sequence with at least 89.5 % identity thereto, respectively.

6. The Fc binding protein of claim 1, wherein said domain has a deletion of 1, 2, or 3 amino acids within the first 4 amino acids of its N-terminus and/or a deletion of 1 or 2 amino acids within the first 2 amino acids of its C-terminus.
7. The Fc binding protein of claims 1-6, wherein the protein comprises 2, 3, 4, 5, 6, 7, or 8 domains linked to each other.
8. The Fc binding protein of claim 7, wherein the protein is a homo-multimer or a hetero-multimer.
9. The Fc binding protein of claim 8, wherein one or more domains are linked to each other directly or with one or more linkers, preferably peptide linkers.
10. The Fc binding protein of any one of claims 1-9, wherein the protein is conjugated to a solid support, preferably via the Cysteine in position 50, 51, or 53.
11. The Fc binding protein of any one of claims 1-10, wherein said protein binds to IgG₁, IgG₂, IgG₄, IgM, IgA, Ig fragments comprising the Fc region, fusion proteins comprising an Fc region of an Ig, and conjugates comprising an Fc region of an Ig.
12. An affinity separation matrix comprising the Fc binding protein of any one of claims 1 to 11.
13. Use of the Fc binding protein of any one of claims 1 to 11 or of the affinity separation matrix of claim 12 for affinity purification of any protein comprising an Fc sequence.
14. A method of affinity purification of a protein comprising an Fc sequence, the method comprising: (a) providing a liquid that contains protein comprising an Fc sequence; (b) providing an affinity separation matrix comprising at least one Fc binding protein of any one of claims 1 to 11 coupled to said affinity separation matrix; (c) contacting said affinity separation matrix with the liquid under conditions that permit binding of the at least one Fc binding protein of any one of claims 1 to 11 to a protein comprising an Fc sequence; and (d) eluting said protein comprising an Fc sequence from said affinity purification matrix.

Patentansprüche

1. Fc-bindendes Protein, das eine oder mehrere Domänen umfasst, wobei die Aminosäure in Position 50, die Aminosäure in Position 51 oder die Aminosäure in Position 53 entsprechend SEQ ID NO: 2 Cystein ist, wobei das Fc-bindende Protein unter alkalischen Bedingungen stabil ist.
2. Fc-bindendes Protein nach Anspruch 1, wobei mindestens eine Domäne eine Aminosäuresequenz der SEQ ID NO: 2 oder eine Aminosäuresequenz mit mindestens 89,5 % Übereinstimmung dazu umfasst, wobei die Aminosäure in Position 50, die Aminosäure in Position 51 oder die Aminosäure in Position 53 entsprechend SEQ ID NO: 2 Cystein ist.
3. Fc-bindendes Protein nach Anspruch 1, wobei mindestens eine Domäne jeweils eine Aminosäuresequenz der SEQ ID NOs: 3 bis 16 oder eine Aminosäuresequenz mit mindestens 89,5 % Übereinstimmung dazu umfasst, wobei die Aminosäure in Position 50, die Aminosäure in Position 51 oder die Aminosäure in Position 53 entsprechend SEQ ID NO: 3 bis 16 Cystein ist.
4. Fc-bindendes Protein nach Anspruch 1, wobei mindestens eine Domäne jeweils eine Aminosäuresequenz der SEQ ID NOs: 7 und 8 oder eine Aminosäuresequenz mit mindestens 89,5 % Übereinstimmung dazu umfasst, wobei die Aminosäure in Position 50, die Aminosäure in Position 51 oder die Aminosäure in Position 53 entsprechend SEQ ID NO: 7 und 8 Cystein ist.
5. Fc-bindendes Protein nach Anspruch 1, wobei mindestens eine Domäne jeweils eine Aminosäuresequenz umfasst, die ausgewählt ist aus der Gruppe SEQ ID NOs: 77 und 90 bis 95, oder eine Aminosäuresequenz mit mindestens 89,5 % Übereinstimmung dazu umfasst.
6. Fc-bindendes Protein nach Anspruch 1, wobei die Domäne eine Deletion von 1, 2 oder 3 Aminosäuren innerhalb der ersten 4 Aminosäuren ihres N-Terminus und/oder eine Deletion von 1 oder 2 Aminosäuren innerhalb der ersten 2 Aminosäuren ihres C-Terminus aufweist.

EP 3 521 304 B9

7. Fc-bindendes Protein nach den Ansprüchen 1 bis 6, wobei das Protein 2, 3, 4, 5, 6, 7 oder 8 miteinander verknüpfte Domänen umfasst.
- 5 8. Fc-bindendes Protein nach Anspruch 7, wobei es sich bei dem Protein um ein Homomultimer oder ein Heteromultimer handelt
9. Fc-bindendes Protein nach Anspruch 8, wobei eine oder mehrere Domänen direkt oder mit einem oder mehreren Linkern miteinander verknüpft sind, vorzugsweise Peptidlinkern.
- 10 10. Fc-bindendes Protein nach einem der Ansprüche 1 bis 9, wobei das Fc-bindende Protein an einen festen Träger konjugiert ist, vorzugsweise über das Cystein in Position 50, 51 oder 53.
11. Fc-bindendes Protein nach einem der Ansprüche 1 bis 10, wobei das Protein an IgG₁, IgG₂, IgG₄, IgM, IgA, Ig-Fragmente umfassend die Fc-Region, Fusionsproteine umfassend eine Fc-Region eines Ig, und Konjugate umfassend eine Fc-Region eines Ig bindet.
- 15 12. Affinitätstrennungsmatrix, umfassend das Fc-bindende Protein nach einem der Ansprüche 1 bis 11.
13. Verwendung des Fc-bindenden Proteins nach einem der Ansprüche 1 bis 11 oder der Affinitätstrennungsmatrix nach Anspruch 12 zur Affinitätsreinigung eines beliebigen Proteins, das eine Fc-Sequenz umfasst.
- 20 14. Verfahren zur Affinitätsreinigung eines Proteins, das eine Fc-Sequenz umfasst, wobei das Verfahren Folgendes umfasst: (a) Bereitstellen einer Flüssigkeit, die ein eine Fc-Sequenz umfassendes Protein enthält; (b) Bereitstellen einer Affinitätstrennungsmatrix, die mindestens ein mit der Affinitätstrennungsmatrix gekoppeltes Fc-bindendes Protein nach einem der Ansprüche 1 bis 11 umfasst; (c) In-Kontakt-Bringen der Affinitätstrennungsmatrix mit der Flüssigkeit unter Bedingungen, die eine Bindung des mindestens einen Fc-bindenden Proteins nach einem der Ansprüche 1 bis 11 an ein eine Fc-Sequenz umfassendes Protein erlauben; und (d) Eluieren des eine Fc-Sequenz umfassenden Proteins aus der Affinitätsreinigungsmatrix.
- 25

30

Revendications

1. Protéine de liaison à Fc comprenant un ou plusieurs domaines, dans laquelle l'acide aminé en position 50, l'acide aminé en position 51 ou l'acide aminé en position 53 correspondant à la SEQ ID NO : 2 est la cystéine, ladite protéine de liaison à Fc étant stable dans des conditions alcalines.
- 35 2. Protéine de liaison à Fc selon la revendication 1, dans laquelle au moins un domaine comprend une séquence d'acides aminés de la SEQ ID NO : 2 ou une séquence d'acides aminés présentant au moins 89,5 % d'identité avec celle-ci ; l'acide aminé en position 50, l'acide aminé en position 51 ou l'acide aminé en position 53 correspondant à la SEQ ID NO : 2 étant la cystéine.
- 40 3. Protéine de liaison à Fc selon la revendication 1, dans laquelle au moins un domaine comprend respectivement une séquence d'acides aminés des SEQ ID NO : 3 à 16 ou une séquence d'acides aminés présentant au moins 89,5 % d'identité avec celles-ci ; l'acide aminé en position 50, l'acide aminé en position 51 ou l'acide aminé en position 53 correspondant aux SEQ ID NO : 3 à 16 étant la cystéine.
- 45 4. Protéine de liaison à Fc selon la revendication 1, dans laquelle au moins un domaine comprend respectivement une séquence d'acides aminés des SEQ ID NO : 7 et 8 ou une séquence d'acides aminés présentant au moins 89,5 % d'identité avec celles-ci ; l'acide aminé en position 50, l'acide aminé en position 51 ou l'acide aminé en position 53 correspondant aux SEQ ID NO : 7 et 8 étant la cystéine.
- 50 5. Protéine de liaison à Fc selon la revendication 1, dans laquelle au moins un domaine comprend respectivement une séquence d'acides aminés choisie dans le groupe des SEQ ID NO : 77, et 90 à 95, ou une séquence d'acides aminés présentant au moins 89,5 % d'identité avec celles-ci.
- 55 6. Protéine de liaison à Fc selon la revendication 1, dans laquelle ledit domaine a une délétion de 1, 2 ou 3 acides aminés dans les 4 premiers acides aminés de son extrémité N-terminale et/ou une délétion de 1 ou 2 acides aminés dans les 2 premiers acides aminés de son extrémité C-terminale.

EP 3 521 304 B9

7. Protéine de liaison à Fc selon les revendications 1 à 6, ladite protéine comprenant 2, 3, 4, 5, 6, 7 ou 8 domaines liés entre eux.
- 5 8. Protéine de liaison à Fc selon la revendication 7, ladite protéine étant un homomultimère ou un hétéromultimère.
9. Protéine de liaison à Fc selon la revendication 8, dans laquelle un ou plusieurs domaines sont liés entre eux directement ou par un ou plusieurs lieurs, de préférence des lieurs peptidiques.
- 10 10. Protéine de liaison à Fc selon l'une quelconque des revendications 1 à 9, ladite protéine étant conjuguée à un support solide, de préférence par l'intermédiaire de la cystéine en position 50, 51 ou 53.
11. Protéine de liaison à Fc selon l'une quelconque des revendications 1 à 10, ladite protéine se liant aux fragments d'IgG₁, IgG₂, IgG₄, IgM, IgA, Ig comprenant la région Fc, aux protéines de fusion comprenant une région Fc d'une Ig et aux conjugués comprenant une région Fc d'une Ig.
- 15 12. Matrice de séparation par affinité comprenant la protéine de liaison à Fc selon l'une quelconque des revendications 1 à 11.
- 20 13. Utilisation de la protéine de liaison à Fc selon l'une quelconque des revendications 1 à 11 ou de la matrice de séparation par affinité selon la revendication 12 pour la purification par affinité de toute protéine comprenant une séquence Fc.
- 25 14. Procédé de purification par affinité d'une protéine comprenant une séquence Fc, le procédé comprenant : (a) la mise en place d'un liquide contenant une protéine comprenant une séquence Fc ; (b) la mise en place d'une matrice de séparation par affinité comprenant au moins une protéine de liaison à Fc selon l'une quelconque des revendications 1 à 11 couplée à ladite matrice de séparation par affinité ; (c) la mise en contact de ladite matrice de séparation par affinité avec ledit liquide dans des conditions permettant une liaison de l'au moins une protéine de liaison à Fc selon l'une quelconque des revendications 1 à 11 à une protéine comprenant une séquence Fc ; et (d) l'élution de ladite protéine comprenant une séquence Fc à partir de ladite matrice de purification par affinité.
- 30
- 35
- 40
- 45
- 50
- 55

FIG. 2

FIG. 2A Coupling efficiency of IB14 variants

Y-axis: coupled amount of protein in nmol/ml.

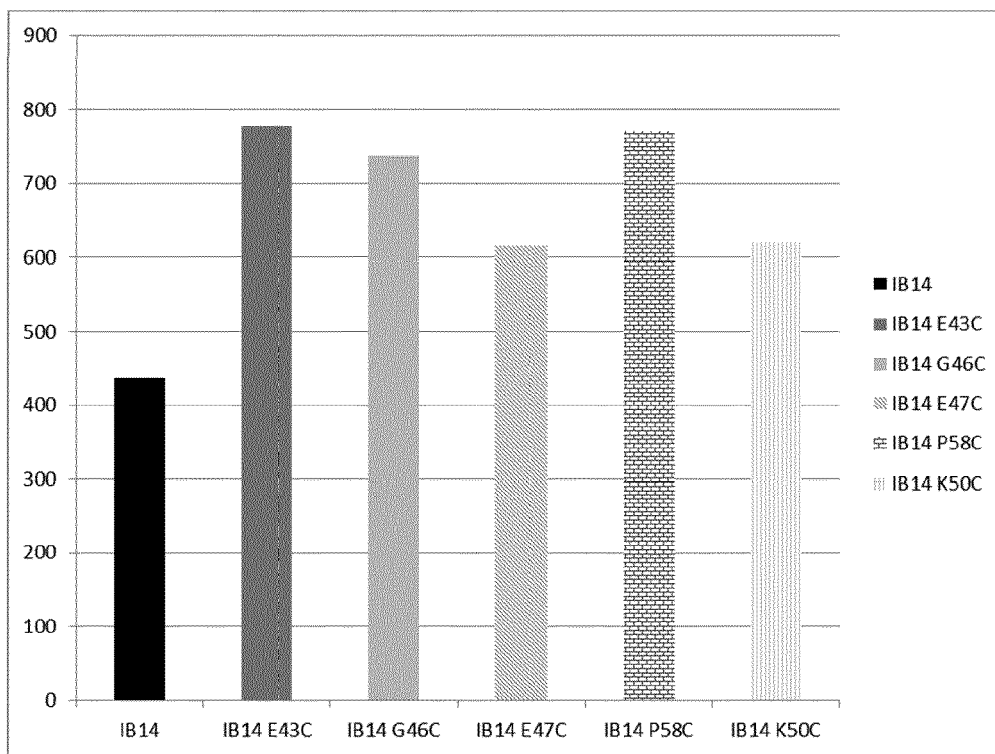


FIG. 2B Coupling of cs14 43C and cs14 46C to Epoxy-activated Sepharose 6B matrix

Y-axis: n(protein per ml resin) in nmol

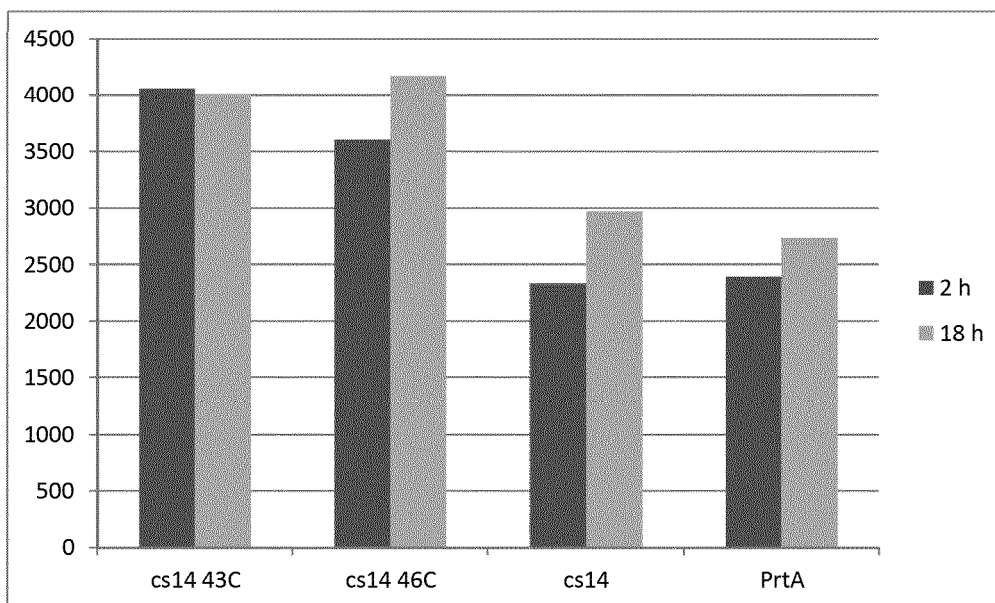


FIG 3.

FIG. 3A. Caustic stability of IB14 variants

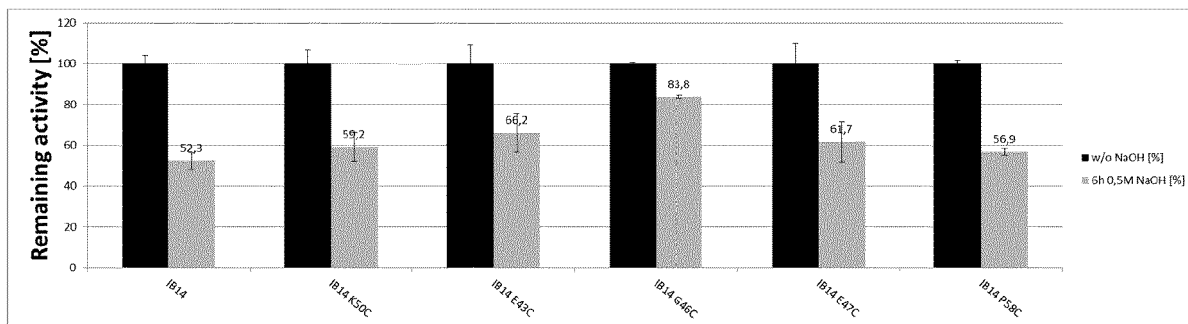


FIG. 3B. Caustic stability of cs14 variants

y-axis: DBC 10 % (mg/ml)

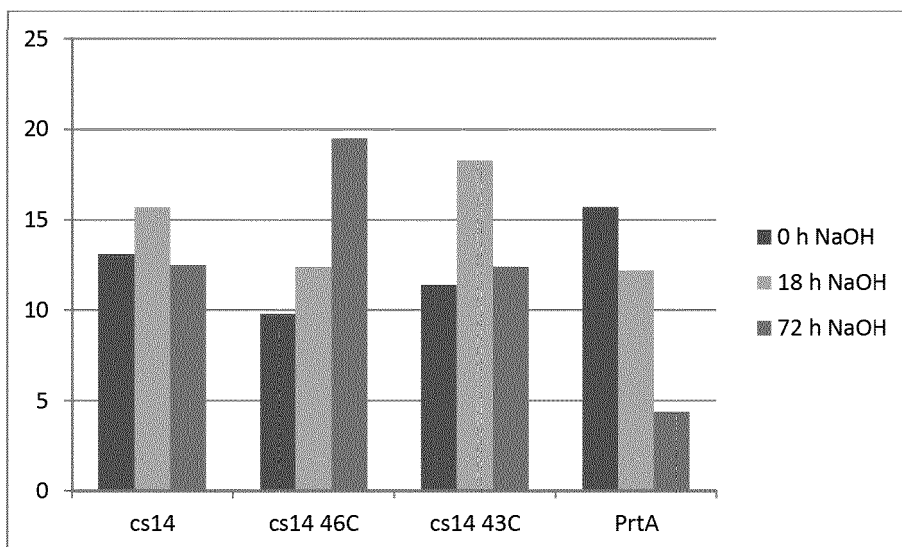


FIG. 4. Caustic stability of cs26 46C

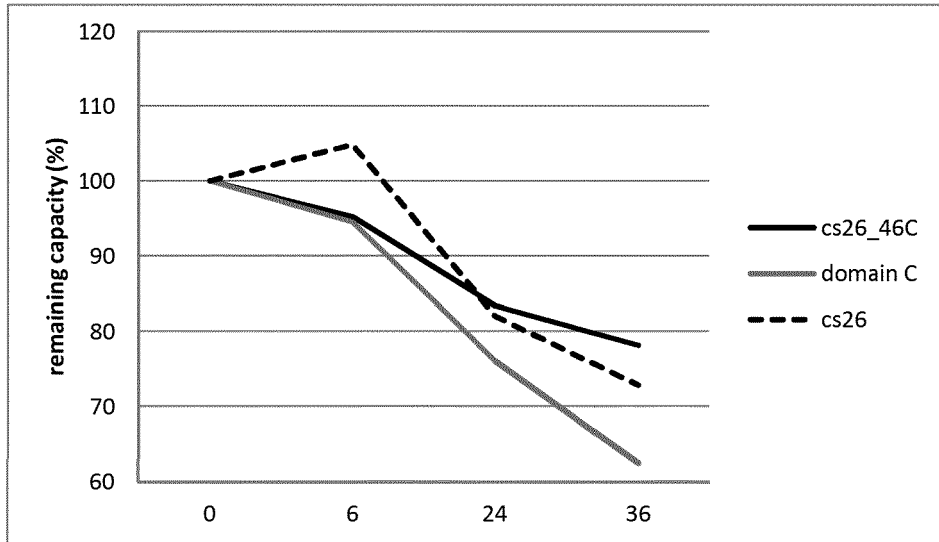
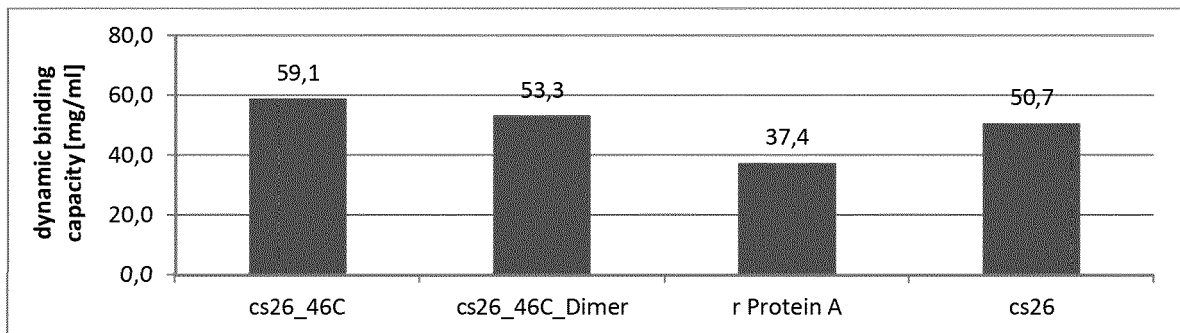


FIG. 5. DBC of cs26 46C



REFERENCES CITED IN THE DESCRIPTION

This list of references cited by the applicant is for the reader's convenience only. It does not form part of the European patent document. Even though great care has been taken in compiling the references, errors or omissions cannot be excluded and the EPO disclaims all liability in this regard.

Non-patent literature cited in the description

- **HUSE et al.** *J. Biochem. Biophys. Methods*, 2002, vol. 51, 217-231 **[0002]**
- A multilingual glossary of biotechnological terms: (IUPAC Recommendations). *Helvetica Chimica Acta*, 1995 **[0012]**
- **XIAOQUIN HUANG ; WEBB MILLER.** *Advances in Applied Mathematics*, 1991, vol. 12, 337-357 **[0032]**
- **THOMPSON et al.** *Nucleic Acids Res.*, 1994, vol. 22 (22), 4673-4680 **[0032]**