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- **Debarbieux, Laurent**  
**92290 Chatenay-Malabry (FR)**

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(74) Representative: **Hoffmann Eitle**  
**Patent- und Rechtsanwälte PartmbB**  
**Arabellastraße 30**  
**81925 München (DE)**

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(56) References cited:  
**WO-A1-01/93904 WO-A1-2013/045863**  
**WO-A2-02/11549 WO-A2-2012/036580**

(83) **Declaration under Rule 32(1) EPC (expert solution)**

- **DOGAN BELGIN ET AL: "Multidrug resistance is common in Escherichia coli associated with ileal Crohn's disease.", INFLAMMATORY BOWEL DISEASES JAN 2013, vol. 19, no. 1, January 2013 (2013-01), pages 141-150, XP009172811, ISSN: 1536-4844**
- **SHENG HAIQING ET AL: "Application of bacteriophages to control intestinal Escherichia coli O157 : H7 levels in ruminants", APPLIED AND ENVIRONMENTAL MICROBIOLOGY, vol. 72, no. 8, August 2006 (2006-08), pages 5359-5366, XP009172812, ISSN: 0099-2240**

(62) Document number(s) of the earlier application(s) in accordance with Art. 76 EPC:  
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(73) Proprietors:

- **Ferring B.V.**  
**2132 JX Hoofddorp (NL)**
- **Institut Pasteur**  
**75015 Paris (FR)**

(72) Inventors:

- **Danglas, Pascal**  
**1162 Saint-Prex (CH)**

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- **WEGRZYN GRZEGORZ ET AL:** "Modulation of the susceptibility of intestinal bacteria to bacteriophages in response to Ag43 phase variation -- a hypothesis.", **MEDICAL SCIENCE MONITOR : INTERNATIONAL MEDICAL JOURNAL OF EXPERIMENTAL AND CLINICAL RESEARCH** JUN 2002, vol. 8, no. 6, June 2002 (2002-06), pages HY15-HY18, XP009172813, ISSN: 1234-1010
- **LUSIAK-SZELACHOWSKA M ET AL:** "Escherichia coli bacteriophages in human stool of patients with gastrointestinal tract diseases", **GASTROENTEROLOGIA POLSKA** 2008 PL, vol. 15, no. 2, 2008, pages 87-90, XP009172818, ISSN: 1232-9886
- **ROLHION NATHALIE ET AL:** "Adherent-invasive Escherichia coli in inflammatory bowel disease", **INFLAMMATORY BOWEL DISEASES, WILLIAMS AND WILKINS, HAGERSTOWN, MD, US**, vol. 13, no. 10, 1 October 2007 (2007-10-01), pages 1277-1283, XP009137421, ISSN: 1078-0998, DOI: 10.1002/IBD.20176 [retrieved on 2007-05-02]
- **MAURA DAMIEN ET AL:** "Intestinal colonization by enteroaggregative Escherichia coli supports long-term bacteriophage replication in mice", **ENVIRONMENTAL MICROBIOLOGY**, vol. 14, no. 8, Sp. Iss. SI, August 2012 (2012-08), pages 1844-1854, XP055047555,
- **MURUGANANTHAN ARAVINTH U ET AL:** "Clinical Risk Factors for Crohn's Disease Postoperative Recurrence are Reflected in Alterations in Mucosally Adherent Microbiota at Surgical Resection", **GASTROENTEROLOGY**, vol. 142, no. 5, Suppl. 1, May 2012 (2012-05), page S679, XP002727902, & **DIGESTIVE DISEASE WEEK (DDW); SAN DIEGO, CA, USA; MAY 19 -22, 2012**

**Description**FIELD OF THE INVENTION

5 **[0001]** The present invention lies in the field of bacteriophage therapy for use in the treatment of inflammatory bowel diseases, as further defined in the claims.

BACKGROUND

10 **[0002]** Bacteriophages are viruses that infect bacteria by specific interaction.

**[0003]** Crohn's disease (CD), also known as regional enteritis, is an inflammatory disease of the intestines that may affect any part of the gastrointestinal tract from mouth to anus, causing a wide variety of symptoms. It primarily causes abdominal pain, diarrhea, vomiting, or weight loss, but may also cause complications outside the gastrointestinal tract such as skin rashes, arthritis, inflammation of the eye, tiredness, and lack of concentration.

15 **[0004]** Although the exact cause of CD is still unknown, a combination of environmental factors and genetic predisposition seems to cause the disease. CD is thought to be an autoimmune disease, in which the body's immune system attacks the gastrointestinal tract, causing inflammation; it is classified as a type of inflammatory bowel disease (IBD).

**[0005]** In patients with CD, abnormal expression of carcinoembryonic antigen-related cell adhesion molecule 6 (CEACAM6) is observed at the apical surface of the ileal epithelium and CD ileal lesions are colonized by pathogenic adherent-invasive *Escherichia coli* (AIEC).

20 **[0006]** There is no known pharmaceutical or surgical cure for Crohn's disease. In particular, neither IBD in general nor CD in particular can be treated with antibiotics (aiming at combatting pathogenic *E. coli*). Treatment options are restricted to controlling symptoms, maintaining remission, and preventing relapse.

25 **[0007]** Maura et al, Environmental Microbiol, vol. 14, no. 8, Sp. Iss. SI, August 2012, pages 1844-1854 discloses isolation of the bacteriophage CLB\_P2.

**[0008]** Belgin et al, Inflammatory Bowel Diseases, Jan 2013, vol. 19, no.1, pages 141-150 discloses the treatment of Crohn's disease by antibiotics effective against AIEC.

SUMMARY OF THE INVENTION

30 **[0009]** The subject invention provides a pharmaceutical composition comprising:

a combination of two or more of the following strains and optionally a pharmaceutically acceptable carrier; for the treatment of inflammatory bowel disease (IBD), and

35 use thereof in a method of treating inflammatory bowel disease comprising administering to a subject in need thereof said combination of bacteriophage strains capable of producing a lytic infection in an adherent-invasive *Escherichia coli* strain thereby treating the subjects:

40 a bacteriophage strain P1 deposited with the French National Collection of Microorganisms at the Institut Pasteur under Accession Number CNCM 1-4694 or a variant thereof, wherein the variant has the same lytic activity and the same phenotypic characteristics as said bacteriophage strain;

45 a bacteriophage strain P2 deposited with the French National Collection of Microorganisms at the Institut Pasteur under Accession Number CNCM I-4695 or a variant thereof, wherein the variant has the same lytic activity and the same phenotypic characteristics as said bacteriophage strain;

a bacteriophage strain P3 deposited with the French National Collection of Microorganisms at the Institut Pasteur under Accession Number CNCM I-4696 or a variant thereof, wherein the variant has the same lytic activity and the same phenotypic characteristics as said bacteriophage strain;

50 a bacteriophage strain P4 deposited with the French National Collection of Microorganisms at the Institut Pasteur under Accession Number CNCM I-4697 or a variant thereof, wherein the variant has the same lytic activity and the same phenotypic characteristics as said bacteriophage strain;

55 a bacteriophage strain P5 deposited with the French National Collection of Microorganisms at the Institut Pasteur under Accession Number CNCM I-4698 or a variant thereof, wherein the variant has the same lytic activity and the same phenotypic characteristics as said bacteriophage strain;

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a bacteriophage strain P6 deposited with the French National Collection of Microorganisms at the Institut Pasteur under Accession Number CNCM I-4699 or a variant thereof, wherein the variant has the same lytic activity and the same phenotypic characteristics as said bacteriophage strain;

5 a bacteriophage strain P8 deposited with the French National Collection of Microorganisms at the Institut Pasteur under Accession Number CNCM I-4700 or a variant thereof, wherein the variant has the same lytic activity and the same phenotypic characteristics as said bacteriophage strain; and

10 bacteriophage strain CLB\_P2 deposited with the French National Collection of Microorganisms at the Institut Pasteur under Accession Number CNCM I-4675 or a variant thereof, wherein the variant has the same lytic activity and the same phenotypic characteristics as said bacteriophage strain.

[0010] For the purpose of the present invention, a variant of a bacteriophage strain is regarded as having the same lytic activity as said bacteriophage strain if it performs at least "+" against at least one of the AIEC strains LF82, 07081, 15 07082, 07076 and 06075 in the "*In vitro*" assay of the infectivity of bacteriophages in AIEC strains" described in Example 3 below. In a preferred embodiment, a variant is regarded as having the same lytic activity if it performs at least "+" against all five AIEC strains LF 82, LF 06075, LF 07076, LF 07081 and LF 07082 (AIEC strains LF 06075, LF 07076, LF 07081 and LF 07082 are also abbreviated herein as 06075, 07076, 07081 and 07082, respectively). These AIEC strains have been deposited by Université Lille 2 — Droit et Santé, 42 Rue Paul Duez, 59000 Lille (France) with the 20 French National Collection at Institut Pasteur under Accession Numbers CNCM 1-4712 (LF 82), CNCM 1-4713 (LF 06075), CNCM 1-4714 (LF 07076), CNCM 1-4715 (LF 07081) and CNCM 1-4716 (LF 07082).

[0011] For the purpose of the present invention, a variant of one of the bacteriophage strains P1 to P6, P8 and CLB\_P2 is regarded as having the same phenotypic characteristics as said bacteriophage strain if it has at least 80% sequence 25 identity on at least 70% of length, preferably at least 90% sequence identity on at least 80% of length and more preferably complete sequence identity on at least 90% of length (as determined by the BLAST algorithm) with the major capsid protein of bacteriophage wV8 (for variants of P1 to P6) or bacteriophage RB69 (for variants of P8) or bacteriophage JS98 (for variants of CLB\_P2), as described below in the section "Identification of Major Capsid Proteins".

[0012] A variant of a bacteriophage has the same lytic activity and the same phenotypic characteristics as the bacteriophage. 30

### DETAILED DESCRIPTION OF THE INVENTION

[0013] The subject invention provides a pharmaceutical composition comprising: the above combination and a pharmaceutically acceptable carrier; for the treatment of inflammatory bowel disease.

35 [0014] An "*adherent-invasive Escherichia coli* (AIEC) strain" as used herein should be understood as referring to an *E. coli* strain having a mean invasion potential of equal to or higher than 0.1% in a cell culture of the intestinal cell line I-407. In other words, an AIEC strain has the ability to invade an intestinal cell culture of I-407 with an invasion index equal or superior to 0.1% of the original inoculum (taken as 100%), when tested in accordance with the invasion assay described below in the section "Invasion Assay" (see also Darfeuille-Michaud et al. (2004), *Gastroenterology* 40 127:412-421).

[0015] Non-limiting examples of AIEC strains are LF82, LF82SK (deposited by Université d'Auvergne, 49 Boulevard François Mitterrand, 63001 Clermont-Ferrand (France) with the French National Collection at Institut Pasteur under Accession Number CNCM I-4723), those listed in Table 1 herein below and those listed in the following itemization (cf. 45 Darfeuille-Michaud et al. (2004), *Gastroenterology* 127:412-421, *especially page 417, Table 2*): LF31, LF71, LF123, LF138, LF9, LF15, LF28, LF50, LF65, LF119, LF128, LF130, LF73, LF100, LF110, LF134, LF105, LF49-2, LB11, and LF45-2. In one embodiment, the adherent-invasive *Escherichia coli* strain is LF82, 07081, 07082, 07076 or 06075, in particular LF82.

[0016] In one embodiment, the adherent-invasive *Escherichia coli* strain is present in the colon of the subject. In another embodiment, the adherent-invasive *Escherichia coli* strain is present in the ileum of the subject. In yet another 50 embodiment, the adherent-invasive *Escherichia coli* strain is present in one or more intestinal parts (small and/or large) of the subject.

[0017] In one embodiment, the bacteriophage strain is P1 which can be part of the combination and is deposited with the French National Collection of Microorganisms at the Institut Pasteur under Accession Number CNCM I-4694 or a variant thereof, wherein the variant has the same lytic activity and the same phenotypic characteristics as said bacteriophage strain. 55

[0018] In one embodiment, the bacteriophage strain is P2 which can be part of the combination and is deposited with the French National Collection of Microorganisms at the Institut Pasteur under Accession Number CNCM I-4695 or a variant thereof, wherein the variant has the same lytic activity and the same phenotypic characteristics as said bacteriophage strain.

ophage strain.

**[0019]** In one embodiment, the bacteriophage strain is P3 which can be part of the combination and is deposited with the French National Collection of Microorganisms at the Institut Pasteur under Accession

**[0020]** Number CNCM I-4696 or a variant thereof, wherein the variant has the same lytic activity and the same phenotypic characteristics as said bacteriophage strain.

**[0021]** In one embodiment, the bacteriophage strain is P4 which can be part of the combination and is deposited with the French National Collection of Microorganisms at the Institut Pasteur under Accession Number CNCM I-4697 or a variant thereof, wherein the variant has the same lytic activity and the same phenotypic characteristics as said bacteriophage strain.

**[0022]** In one embodiment, the bacteriophage strain is P5 which can be part of the combination and is deposited with the French National Collection of Microorganisms at the Institut Pasteur under Accession Number CNCM I-4698 or a variant thereof, wherein the variant has the same lytic activity and the same phenotypic characteristics as said bacteriophage strain.

**[0023]** In one embodiment, the bacteriophage strain is P6 which can be part of the combination and is deposited with the French National Collection of Microorganisms at the Institut Pasteur under Accession Number CNCM I-4699 or a variant thereof, wherein the variant has the same lytic activity and the same phenotypic characteristics as said bacteriophage strain.

**[0024]** In one embodiment, the bacteriophage strain is P8 which can be part of the combination and is deposited with the French National Collection of Microorganisms at the Institut Pasteur under Accession Number CNCM I-4700 or a variant thereof, wherein the variant has the same lytic activity and the same phenotypic characteristics as said bacteriophage strain.

**[0025]** In one embodiment, the bacteriophage strain is CLB\_P2 which can be part of the combination and is deposited with the French National Collection of Microorganisms at the Institut Pasteur under Accession Number CNCM I-4675 or a variant thereof, wherein the variant has the same lytic activity and the same phenotypic characteristics as said bacteriophage strain.

**[0026]** In one aspect, it is envisaged that the pharmaceutical composition comprises more than one bacteriophage strain, also named "*a bacteriophage cocktail*". The bacteriophage cocktail of the present invention comprises any combination of two or more of P1, P2, P3, P4, P5, P6, P8 and CLB\_P2 and variants thereof having the same lytic activity, preferably the same lytic activity and the same phenotypic characteristics. Preferably, the bacteriophages in a bacteriophage cocktail intended for treatment of a specific subject or group of subjects will be selected on the basis of the AIEC strain or AIEC strains identified and selected for combatting.

**[0027]** Non-limiting examples of inflammatory bowel diseases are Crohn's disease (CD), ulcerative colitis (UC), chronic inflammatory bowel disease (chronic IBD) such as but not limited to microscopic colitis, celiac disease and vasculitis. In one embodiment, the IBD is CD or UC. In another embodiment, the inflammatory bowel disease is recurrence of ileal lesions after surgery (such as surgery for the removal of at least a part of the small intestine in CD patients). The recurrence can be measured by the Rutgeerts score.

**[0028]** In one embodiment, the IBD is not caused by a bacterial infection. This embodiment is based on the observation that IBD is an autoimmune disease which is not generally considered a bacterial disease. Instead, a bacterial infection may be concomitant to IBD, but is not necessarily the causative agent. This observation adds to the surprising finding of the present invention, namely applying bacteriophage therapy for the treatment of a disease which is not caused by bacteria.

**[0029]** For that reason, there can be — as an example — AIEC strains in family members of subjects suffering from an IBD, although these family members do not suffer from this disease. Likewise, AIEC strains can also be found in subjects neither suffering from IBD nor being related to subjects suffering from IBD, as can also be seen from Table 1 below.

**[0030]** "*Treating*" as used herein should be understood to encompass a decrease in one or more symptoms characteristic of the disease; a decrease in the rate of progression of the disease; recovery from the disease, cure from the disease, maintenance of remission and prophylaxis such as prevention of relapse.

**[0031]** A "*subject*" as used herein can be a male or a female subject. A subject can be a human being or any other mammal.

**[0032]** The dose and regimen of administration of a pharmaceutical composition of the invention will necessarily be dependent upon the therapeutic effect to be achieved (e.g. treatment of IBD) and may vary with the particular bacteriophage strains in the composition, the route of administration, and the age and condition of the individual subject to whom the medicament is to be administered.

**[0033]** A dosage for humans is likely to contain a dose of bacteriophage between  $10^4$  and  $10^{11}$  plaque forming units (pfu). The desired dose may be presented as one dose per day or as multiple sub-doses administered at appropriate intervals.

**[0034]** In the context of the present invention the term "*pharmaceutically acceptable carrier*" relates to pharmaceutically-

acceptable, non-toxic carriers, fillers or diluents, which are defined as vehicles commonly used to formulate pharmaceutical compositions for animal or human administration.

5 [0035] The pharmaceutical compositions of the present invention may further comprise pharmaceutically acceptable auxiliary agents, and optionally other therapeutic agents. Auxiliary agents, also named accessory ingredients, encompass those conventional in the art such as, but not limited to matrix-forming agents, thickeners, binders, lubricants, pH adjusting agents, protecting agents, viscosity enhancers, wicking agents, disintegrants, including non-effervescent and effervescent disintegrants, surfactants, anti-oxidants, wetting agents, colorants, flavoring agents, taste-masking agents, sweeteners, preservatives and so forth. In addition to being pharmaceutically acceptable, the auxiliary agents must be "acceptable" in the sense that they are compatible with the other ingredients of the composition, including the bacteriophage.

10 [0036] Pharmaceutical compositions and routes of administration include those suitable for or via oral (including buccal, sublingual and intraorbital), rectal, nasal, topical (including transdermal), ocular, otic, vaginal, bronchial, pulmonary or parenteral (including subcutaneous, intramuscular, intravenous, intradermal, intraperitoneal, intrapleural, intravesicular and intrathecal) administration or administration via an implant. The pharmaceutical composition or route of administration may be adapted to provide a targeted effect of bacteriophage strain of the invention. In a specific embodiment, a pharmaceutical composition of the invention is administered orally. The compositions may be prepared by any method well known in the art of pharmacy. Such methods include the step of bringing in association a bacteriophage strain of the invention with a pharmaceutically acceptable carrier and optionally one or more auxiliary agents.

15 [0037] Pharmaceutical compositions suitable for oral administration may be presented as discrete dosage units (dosage forms) such as pills, tablets, dragees or capsules, or as a powder or granules, or as a solution or suspension. The pharmaceutical composition may also be presented as a bolus or paste. The compositions can further be processed into a suppository or enema for rectal administration.

20 [0038] For parenteral administration, suitable compositions include aqueous and non-aqueous sterile injections. The compositions may be presented in unit-dose or multi-dose containers, for example sealed vials and ampoules, and may be stored in a freeze-dried (lyophilized) condition requiring only the addition of sterile liquid carrier, for example water, prior to use.

25 [0039] For transdermal administration, e.g., gels, patches or sprays can be contemplated.

[0040] Compositions or formulations suitable for pulmonary administration, e.g., by nasal inhalation, include fine dusts or mists which may be generated by means of metered dose pressurized aerosols, nebulizers or insufflators.

30 [0041] The specification includes a kit comprising a pharmaceutical composition of the invention and instructions for the use of the composition for a use as hereinbefore described, optionally together with packaging material.

[0042] The specification further provides a bacteriophage strain P1 deposited with the French National Collection of Microorganisms at the Institut Pasteur under Accession Number CNCM I-4694 or a variant thereof, wherein the variant has the same lytic activity, preferably the same lytic activity and the same phenotypic characteristics as said bacteriophage strain.

35 [0043] The specification further provides a bacteriophage strain P2 deposited with the French National Collection of Microorganisms at the Institut Pasteur under Accession Number CNCM I-4695 or a variant thereof, wherein the variant has the same lytic activity, preferably the same lytic activity and the same phenotypic characteristics as said bacteriophage strain.

40 [0044] The specification further provides a bacteriophage strain P3 deposited with the French National Collection of Microorganisms at the Institut Pasteur under Accession Number CNCM I-4696 or a variant thereof, wherein the variant has the same lytic activity, preferably the same lytic activity and the same phenotypic characteristics as said bacteriophage strain.

45 [0045] The specification further provides a bacteriophage strain P4 deposited with the French National Collection of Microorganisms at the Institut Pasteur under Accession Number CNCM I-4697 or a variant thereof, wherein the variant has the same lytic activity, preferably the same lytic activity and the same phenotypic characteristics as said bacteriophage strain.

50 [0046] The specification further provides a bacteriophage strain P5 deposited with the French National Collection of Microorganisms at the Institut Pasteur under Accession Number CNCM I-4698 or a variant thereof, wherein the variant has the same lytic activity, preferably the same lytic activity and the same phenotypic characteristics as said bacteriophage strain.

[0047] The specification further provides a bacteriophage strain P6 deposited with the French National Collection of Microorganisms at the Institut Pasteur under Accession Number CNCM I-4699 or a variant thereof, wherein the variant has the same lytic activity, preferably the same lytic activity and the same phenotypic characteristics as said bacteriophage strain.

55 [0048] The specification further provides a bacteriophage strain P8 deposited with the French National Collection of Microorganisms at the Institut Pasteur under Accession Number CNCM I-4700 or a variant thereof, wherein the variant has the same lytic activity and the same phenotypic characteristics as said bacteriophage strain.

EXAMPLES

**[0049]** The invention is further described in the following examples, which are not in any way intended to limit the scope of the invention as claimed.

METHODS

INVASION ASSAY

**[0050]** The Intestine-407 (I-407) cell line derived from human embryonic jejunum and ileum was used as a model of undifferentiated intestinal epithelial cells. It was purchased from Flow Laboratories (Flow Laboratories Inc., McLean, VA). **[0051]** Intestine-407 cells were seeded in 24-well tissue culture plates (Polylabo, Strasbourg, France) at a density of 4,105 cells/well and incubated for 20 hours. The cell monolayers were washed twice with PBS (pH 7.2). Bacterial invasion of epithelial cells was measured using the gentamicin protection assay (Falkow et al. (1987), Rev. Infect. Dis. 9 (Suppl. 5):S450-455). Each monolayer was inoculated in 1 mL of the cell culture medium lacking antibiotics with a multiplicity of infection of 10 bacteria per epithelial cell. After a 3-hour incubation period at 37°C with 5% CO<sub>2</sub>, the monolayers were washed 3 times with PBS. Fresh cell culture medium containing 100 µg/mL of gentamicin (Sigma, St. Louis, MO) was added for 1 hour to kill extracellular bacteria before lysis of the monolayers with 1% Triton X-100 (Sigma) in deionized water. This concentration of Triton X-100 had no effect on bacterial viability for at least 30 minutes. The samples were diluted and plated onto Mueller-Hinton agar plates to determine the number of colony-forming units. All results of *E. coli* invasive ability with Intestine-407 cell line were expressed as the percentage of intracellular bacteria compared with the initial inoculum, taken as 100%. All of the assays were performed at least 3 times in separate experiments.

IDENTIFICATION OF MAJOR CAPSID PROTEINS

**[0052]** Virion proteins were obtained by boiling 60 µl of a suspension of 10<sup>11</sup> pfu/ml of each bacteriophage for 10 min. 20 µl of the suspension were run on a precast 4—12% polyacrylamide gel. The gel was stained with Coomassie blue and the major bands were excised, subjected to trypsin digestion and analyzed by mass spectrometry at the Institut Pasteur microsequencing facility.

**[0053]** The peptide masses obtained were compared with the information in protein databases, allowing the identification of the closest known protein, i.e. wV8 for P1 to P6 and RB69 for P8 and JS98 for CLB\_P2 (see A. Villegas et al, Virology Journal 2009, 6:41 for characterization of wV8 and S. Zuber et al., Journal of Bacteriology 2007, 189:22, 8206 for characterization of RB69 and JS 98).

**[0054]** Alignment of the major capsid protein of bacteriophage wV8 with peptides obtained from mass spectrometry of the major capsid proteins of bacteriophages P1 to P6:

```
wV8 : MLTNSEKSRFFLADLTGEVQSIPTNTYGYISNLGLFRSAPITQTTFLMDLTDWDVSLLDVDRDSRKAE
P1 : FFLADLTGEVQSIPTNTYGYISNLGLFR
P2 : SRFFLADLTGEVQSIPTNTYGYISNLGLFRSAPITQTTFLMDLTDWDVSLLDVDR
P3 :
P4 :
P5 :
P6 :
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wV8 : TSAPERVRQISFPMMYFKEVESITPDEIQGVRQPGTANELTTEAVVRAKKLMKIRTKFDITREFLFMQ
P1 : QISFPMMYFKEVESITPDEIQGVRQPGTANELTTEAVVR TKFDITREFLFMQ
P2 : QISFPMMYFKEVESITPDEIQGVRQPGTANELTTEAVVR TKFDITREFLFMQ
P3 : QISFPMMYFKEVESITPDEIQGVRQPGTANELTTEAVVR TKFDITREFLFMQ
P4 : QISFPMMYFKEVESITPDEIQGVRQPGTANELTTEAVVR TKFDITREFLFMQ
P5 : QISFPMMYFKEVESITPDEIQGVRQPGTANELTTEAVVR TKFDITREFLFMQ
P6 : QISFPMMYFKEVESITPDEIQGVRQPGTANELTTEAVVR TKFDITREFLFMQ
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5  
wV8 : ALKGKVV DARGTLYADLYKQFDVEKKT VYFDL DNPADIDAAIEELRMHMEDEAKTGT VINGEEIHVV  
P1 : ALK GTLYADLYK KTVYFDL DNPADIDASIEELR TGT VINGEEIHVV  
P2 : ALK GTLYADLYK TGT VINGEEIHVV  
P3 : ALK GTLYADLYK TVYFDL DNPADIDASIEELR TGT VINGEEIHVV  
P4 : ALK GTLYADLYK TGT VINGEEIHVV  
P5 : ALK GTLYADLYK TIYFDL DNPADIDASIEELR TGT VINGEEIHVV  
P6 : ALK GTLYADLYKQFDVEK TIYFDL DNPADIDASIEELR TGT VINGEEIHVV

10  
wV8 : VDRLFFSKLVKHPKIRDAYLAQQTPLAWQQITGSLRTGGTDGVQAHMNTFYYGGVKFVQYNGKFKDKR  
P1 : VDR IRDAYLAQQTPLAWQQITGSLR FVQYNGK  
P2 : VDRLFFSK IRDAYLAQQTPLAWQQITGSLRTGGTDGVQAHMNTFYYGGVKFVQYNGK  
P3 : VDR DAYLAQQTPLAWQQITGSLR FVQYNGK  
P4 : VDR DAYLAQQTPLAWQQITGSLR FVQYNGK  
P5 : VDR DAYLAQQTPLAWQQITGSLRTGGADGVQAHMNTFYYGGVKFVQYNGK  
15  
P6 : DAYLAQQTPLAWQQITGSLRTGGADGVQAHMNTFYYGGVK

20  
wV8 : GKVHTLVSIDVAATVGVGHAFPNVSM LGEANNIFEVAYGPCPKMGYANTLGQELYVFEYEKDRDEGI  
P1 : MGYANTLGQELYVFEYEKDR  
P2 : MGYANTLGQELYVFEYEKDR  
P3 :  
P4 :  
P5 :  
P6 :

25  
wV8 : DFEAHSYMLPYCTRPQLLDVRS DAKPD  
P1 : PQLLDVDR  
P2 : PQLLDVDR  
P3 : PQLLDVDR  
P4 : PQLLDVDR  
30  
P5 : PQLLDVDR  
P6 : PQLLDVDR

[0055] Alignment of the major capsid protein of bacteriophage RB69 with peptides obtained from mass spectrometry of the major capsid protein of bacteriophage P8:

35  
RB69 : MTTIKTKAQLVDKWKELLEGEGLPEIANSKQAI IAKIFENQEKDFEVSPEYKDEKIAQAFGSFLTEAE  
P8 :

40  
RB69 : IGGDHGYN AQNIAAGQTS GAVTQIGPAVMGMVRRRAIPNLIAFDICGVQPMNSPTGQVFALRAVYKDP  
P8 :

45  
RB69 : IAAGAKEAFHPMYAPDAMFSGQGA AKKFPALAASTQTKVGD IYTHFFQETGT VYLQASAVTISSAD  
P8 : EAFHPMYAPDAMFSGQGA AK

50  
RB69 : DAAKLD AEI I KQMEAGALVEIAEGMATSIAELQEGFNGSTDNPNWEMGFRIDKQVIEAKSRQLKAAYS  
P8 : AAYS

50  
RB69 : IELAQDLRAVHGM DADAELS GILATEIMLEINREVV DWINYSAQVGKSGMTNIVGSKAGVDFDQDPID  
P8 : IELAQDLR EVVDWINYSAQVGK AGVDFDQDPID

55  
RB69 : IRGARWAGESFKALLFQIDKEAVEIARQTGRGEGNFIIASRN VVNLASVDTGISYAAQGLASGFNTD  
P8 : IR WAGESFK QTGRGEGNFIIASR

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RB69 : TTKSVFAGVLGGKYRVYIDQYAKQDYFTVGYKGANEMDAGIYYAPYVALTPLRGSDPKNFQPVMGFKT  
P8 : SVFAGVLGGKYRVYIDQYAKQDYFTVGYKGANEMDAGIYYAPYVALTPLR NFQPVMGFKT

5

RB69 : RYGIGVNPFAESSLQAPGARIQSGMPSILNSLGKNAYFRRVYVKGI  
P8 : RYGIGVNPFAESSLQAPGARIQSGMPSILNSLGK

10 **[0056]** Alignment of the major capsid protein of bacteriophage JS98 with peptides obtained from mass spectrometry of the major capsid protein of bacteriophage CLB\_P2.

JS98: MKKNALVQKWSALLENEALPEIVGASKQAIIAKIFENQEODILTAPEYRDEKISEAFGSFLTEAEI  
CLB\_P2:

15

JS98: GGDHGYDATNIAAGQTS GAVTQIGPAVMGMVRRRAIPHLIAFDICGVQPLNNPTGQVFALRAVYVKD  
CLB\_P2: AVYVKD

20

JS98: PIAAGAKEAFHPMYAPNAMFSGQAAETFEALAASKVLEVGKIYSHFFEATGSAHFQAVEAVTVDA  
CLB\_P2: PIAAGAK

25

JS98: GATDAAKLDAAVTALVEAGQLAEIAEGMATSI AELQEGFNGSTDNPNWEMGFRIDKQVIEAKSRQL  
CLB\_P2:

30

JS98: KASYSIELAQDLRAVHGMDADAELSGILATEIMLEINREVIDWINYSAQVGKSGMTNTVGAKAGVF  
CLB\_P2: ASYSIELAQDLR EVIDWINYSAQVGK AGVF

35

JS98: DFQDPIDIRGARWAGESFKALLFQIDKEAAE IARQTGRGAGNFIIASRNVVNVLAAVDTSVSYAAQ  
CLB\_P2: DFQDPIDIR WAGESFKALLFQIDKEAAE IAR GAGNFIIASR

40

JS98: GLGQGFNVDTTKAVFAGVLGGKYRVYIDQYARSDYFTIGYKGSNEMDAGIYYAPYVALTPLRGSDP  
CLB\_P2: AVFAGVLGGKYRVYIDQYAR GSNEMDAGIYYAPYVALTPLR

JS98: KNFQPVMGFKTRYGIGINPFADPAAQAPT KRIQNGMPDIVNSLGLNGYFRRVYVKGI  
CLB\_P2: NFQPVMGFKTRYGIGINPFADPAAQAPT KRIQNGMPDIVNSLGLNGYFR

### EXAMPLE 1

45

#### Isolation of AIEC strains

50 **[0057]** One hundred and sixty-six (166) adherent-invasive *Escherichia coli* (*E. coli*) strains, including *E. coli* strain LF82 (Table 1), were isolated as follows: The AIEC strains were isolated from fresh feces of CD patients, their family members and control subjects. The feces were diluted in tenfold dilutions up to -9. Each dilution was plated on different media. After incubation, colonies were sub-cultured, identified and the strains were tested for invasion capacity.

55 **[0058]** In detail, immediately after emission, fresh feces were introduced in a sterile container. The atmosphere was rendered anaerobic by addition of a moistened Anaerocult®. Samples were treated the day of sampling. About 1 g of feces were introduced in 9 mL of cysteinated ¼ strength Ringer solution in pre-weighed tubes; they were reweighed after introduction of the sample to determine its exact weight (first tenfold dilution). Eight further tenfold dilutions were made and 0.1 mL of each dilution was plated on different non-selective and selective media incubated in appropriated conditions: Columbia blood agar (CS) and CSH agar incubated for one week under anaerobic conditions, MRS medium incubated for 48h in an atmosphere enriched in CO<sub>2</sub>, McConkey and Cetrimide agar incubated for 48h in air. All incubations

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were done at 37°C. After incubation, colonies were counted, subcultured and identified by established phenotypic criteria.

**[0059]** A control subject was selected *vis-à-vis* a CD patient so that the control subject was of the same sex and age as the CD patient and had a similar family size as the CD patient (to take microflora variation within a family into consideration).

**[0060]** The protocol was approved by the local ethical committee in 2000. The patients were followed by the EPIMAD register, which is organized under an agreement between the Institut National de la Santé et de la Recherche Médicale (INSERM) and the Institut National de Veille Sanitaire (InVS) and is also supported by the François Aupetit Association, Lion's Club of Northwestern France, Ferring Laboratories, the Société Nationale Française de Gastroentérologie and Lille University Hospital.

Table 1 - AIEC strains

Number	Reference	Invasion I-407 Mean (%)	Invasion I-407 SEM (%)	Culture Medium <sup>1</sup>	Dilution	Level of <i>E. coli</i> (log UFC/g) <sup>2</sup>	Total Count (logUFC/g) <sup>3</sup>
	LF82	1.29	0.8	McC	-4	5.7	5.9
<i>AIEC isolated from CD patient</i>							
06259	C4-1	2.050	0.500	McC	-5	5.87	10.52
06254	C34-12	2.163	0.738	Cet	-2	2.91	10.14
06256	C34-2	0.550	0.170	McC	-7	7.91	10.14
06072	C39-1	0.2075	0.147	McC	-6	7.02	9.93
06073	C39-2	0.1374	0.097	McC	-6	7.02	9.93
06075	C39-4	0.2334	0.165	McC	-5	6.02	9.93
06076	C39-7	0.5900	0.417	Cet	-2	3.02	9.93
06087	C42-1	0.1095	0.055	McC	-5	5.82	9.58
06088	C42-2	0.1954	0.098	McC	-5	5.82	9.58
06089	C42-3	0.1930	0.097	Cet	-3	3.82	9.58
06398	C76-10	0.131	0.036	CS ana	-5	6.09	9.99
06011	C84-2	0.2580	0.129	McC	-6	6.96	9.23
06023	C97-1	0.1173	0.068	McC	-7	8.14	10.03
06024	C97-2	0.1303	0.075	McC	-6	7.14	10.03
06026	C98-1	0.1439	0.072	McC	-6	7.2	9.34
06027	C98-2	0.1122	0.065	McC	-6	7.2	9.34
06028	C98-4	0.2310	0.133	Cet	-2	3.20	9.34
06029	C99-1	1.0657	0.615	McC	-6	6.99	10.17
06030	C99-2	0.1613	0.081	McC	-6	6.99	10.17
06031	C99-3	0.2330	0.135	McC	-5	5.99	10.17
06033	C99-9	0.4667	0.269	Cet	-2	2.99	10.17
06150	C187-13	0.6675	0.472	CS ana	-7	7.93	9.97
06151	C187-14	1.0350	0.732	CS ana	-7	7.93	9.97
06152	C187-15	0.4375	0.253	CS ana	-7	7.93	9.97
06166	C190-1	0.2251	0.130	McC	-8	9.28	10.98
06167	C190-2	0.1247	0.072	McC	-8	9.28	10.98
06168	C190-3	0.1688	0.097	McC	-7	8.28	10.98
06169	C190-4	0.1373	0.079	McC	-6	7.28	10.98

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(continued)

<i>AIEC isolated from CD patient</i>								
5	06170	C190-6	0.7065	0.408	Cet	-3	4.28	10.98
	06171	C190-8	0.5827	0.336	Cet	-2	3.28	10.98
	06172	C190-7	0.5385	0.311	Cet	-2	3.28	10.98
	06173	C190-12	0.5182	0.299	CS ana	-9	10.28	10.98
10	06280	C203-7	0.185	0.087	Cet	-3	3.96	9.94
	06281	C203-9	0.393	0.023	Cet	-2	2.96	9.94
	06283	C204-4	0.253	0.092	McC	-6	7.00	9.78
15	06271	C205-2	0.153	0.052	McC	-6	6.93	9.97
	06278	C205-9	0.160	0.005	Cet	-2	2.93	9.97
	06351	C215-8	0.548	0.397	Cet	-5	5.93	9.91
	06352	C215-9	0.262	0.143	Cet	-5	5.93	9.91
20	06353	C215-12	1.960	1.340	Cet	-3	3.93	9.91
	06354	C215-13	1.339	1.281	Cet	-3	3.93	9.91
	06356	C215-10	2.260	1.540	Cet	-3	3.93	9.91
25	06357	C215-11	2.195	1.355	Cet	-3	3.93	9.91
	06358	C215-1	1.110	0.590	McC	-6	7.93	9.91
	06359	C215-2	1.523	0.928	McC	-6	7.93	9.91
	06360	C215-3	0.165	0.064	McC	-4	4.93	9.91
30	06361	C215-4	0.315	0.135	McC	-3	3.93	9.91
	06362	C215-5	0.980	0.720	McC	-3	3.93	9.91
	07074	C43-1	1.5825	1.3675	McC	-6	7.26	9.66
35	07075	C44-1	0.1822	0.0755	McC	-5	6.01	9.91
	07076	C44-2	0.5950	0.3350	McC	-5	6.01	9.91
	07077	C44-3	0.1432	0.0486	McC	-4	5.01	9.91
	07078	C44-4	0.3086	0.1764	McC	-4	5.01	9.91
40	07081	C44-9	0.5525	0.2675	Cet	-2	3.01	9.91
	07082	C45-1	0.4675	0.0925	McC	-5	5.94	9.46
	07086	C45-9	0.2110	0.0842	Cet	-2	2.94	9.46
45	07093	C50-2	1.3125	0.9375	McC	-5	5.99	7.69
	07035	C66-2	0.6475	0.3125	McC	-7	8.01	9.53
	07045	C71-1	0.2079	0.1226	McC	-5	5.95	10.58
	07046	C71-2	0.2030	0.0719	McC	-4	4.95	10.58
50	07048	C71-5	0.2388	0.1382	Cet	-2	2.95	10.58
	07051	C100-11A	0.6325	0.3425	MRS	-4	5.07	10.47
	07003	C112-4	0.2513	0.0861	McC	-5	6.14	10.88
55	07006	C112-10	0.8913	0.1863	Cet	-2	3.14	10.88
	07022	C121-8	0.1903	0.0448	Cet	-5	6.14	10.88
	07101	C55-1	0.678	0.022	McC	-6	6.95	10.38

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(continued)

<i>AIEC isolated from CD patient</i>								
5	07103	C55-3	9.175	2.775	McC	-5	5.95	10.38
	07107	C55-8A	4.425	0.075	Cet	-2	2.95	10.38
	07111	C60-1	0.232	0.028	McC	-6	6.84	8.20
	07113	C60-3	0.340	0.105	McC	-4	4.84	8.20
10	07126	C231-1	0.323	0.097	McC	-7	7.94	10.62
	07127	C231- 2	0.141	0.030	McC	-6	6.94	10.62
	07128	C231-5	0.365	0.095	Cet	-3	3.94	10.62
15	07134	C233-1	0.645	0.090	McC	-3	3.98	6.18
	07135	C233-3	1.510	0.390	McC	-2	2.98	6.18
	07136	C233-2	2.090	0.260	McC	-3	3.98	6.18
	07137	C233-11	1.108	0.168	CSH	-3	3.98	6.18
20	<i>AIEC isolated from family members of CD patients</i>							
	06066	C22-9	0.2710	0.192	CS ana	-7	7.94	10.12
	06381	C33-5	0.465	0.185	Cet	-2	2.90	9.46
25	06258	C35-5	0.873	0.428	McC	-6	7.64	10.14
	06086	C41-7	0.1242	0.072	Cet	-2	2.91	10.14
	06384	C47-2	0.180	0.010	McC	-6	7.07	9.62
	06386	C47-4	0.121	0.047	McC	-5	6.07	9.62
30	06097	C64-2	1.4550	1.029	McC	-5	6.03	9.80
	06099	C64-5	0.1175	0.068	Cet	-2	3.03	9.80
	06100	C64-6	1.1225	0.794	Cet	-2	3.03	9.8
35	06006	C81-1	0.2850	0.202	McC	-5	5.96	9.64
	06007	C81-2	0.3200	0.226	McC	-5	5.96	9.64
	06016	C85-1	0.7540	0.435	McC	-7	8.01	10.16
	06019	C85-5	1.4775	1.045	Cet	-2	3.01	10.16
40	06394	C87-7	0.130	0.030	MRS	-2	3.03	9.42
	06020	C92-1	0.1253	0.072	McC	-5	6.09	9.64
	06021	C92-2	0.1678	0.097	McC	-4	5.09	9.64
45	06022	C92-4	0.1229	0.071	McC	-2	3.09	9.64
	06396	C95-1	4.975	2.575	McC	-6	7.1	9.75
	06037	C102-1	0.6767	0.391	McC	-6	6.90	8.55
	06040	C102-7	0.2342	0.135	CS ana	-6	6.9	8.55
50	06080	C107-2	0.5050	0.357	McC	-6	7.1	9.55
	06042	C107-3	0.1900	0.110	McC	-6	7.1	9.55
	06043	C107-5	1.7600	1.245	McC	-6	7.1	9.55
55	06045	C107-10	0.1975	0.140	Cet	-2	3.1	9.55
	06046	C108-2	0.3925	0.278	McC	-6	7.12	10.54
	06049	C108-10	0.2425	0.171	CS ana	-7	8.12	10.54

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(continued)

<i>AIEC isolated from family members of CD patients</i>								
5	06057	C133-1	0.2475	0.175	McC	-6	6.98	10.49
	06101	C133-4	0.1809	0.090	Cet	-2	2.98	10.49
	06160	C189-2	1.3483	0.778	McC	-6	7.07	10.19
	06164	C189-16B	0.3295	0.190	CSH	-8	8.07	10.19
10	06176	C191-4	0.3975	0.281	McC	-5	5.96	10.34
	06177	C191-5	0.3185	0.225	McC	-5	5.96	10.34
	06293	C207-6	0.175	0.111	Cet	-2	2.93	10.13
15	06295	C208-6	0.116	0.047	Cet	-2	2.91	10.57
	06301	C211-1	0.285	0.242	McC	-2	2.87	10.07
	06329	C218-2	0.649	0.439	McC	-6	6.88	10.06
	06338	C218-13	0.208	0.047	Cet	-5	5.88	10.06
20	06341	C218-16	0.304	0.218	Cet	-4	4.88	10.06
	07064	C225-1	0.1280	0.0058	McC	-4	4.90	10.10
	07065	C225-2	0.8354	0.7146	McC	-4	4.90	10.10
25	07066	C225-5	0.9200	0.4150	McC	-6	6.87	9.49
	07067	C225-6	1.0792	0.5977	McC	-5	5.87	9.49
	07068	C226-1	0.1193	0.0334	McC	-5	6.06	10.58
	07073	C227-4	0.2164	0.1568	Cet	-2	2.88	10.06
30	07120	C228-2	0.228	0.013	McC	-2	3.17	9.72
	07121	C229-1	0.126	0.012	McC	-5	6.06	9.50
	07122	C229-2	0.117	0.034	McC	-4	5.06	9.50
35	07123	C229-7	0.190	0.045	Cet	-5	6.06	9.50
	07131	C232-5	0.190	0.122	Cet	-2	2.98	9.60
	07138	C235-1	0.658	0.193	McC	-5	6.02	9.42
<i>AIEC isolated from control subjects</i>								
40	06235	C174-6	2.833	2.468	Cet	-2	3.05	10.59
	06242	C177-1	0.251	0.175	McC	-6	6.99	9.95
	06103	C177-13	0.1461	0.073	CS ana	-7	7.99	9.95
45	06105	C177-2	0.1571	0.079	McC	-5	5.99	9.95
	06106	C178-23	0.4527	0.261	CSH	-5	6.03	10.24
	06108	C179-7	0.1103	0.064	Cet	-2	3.07	10.53
	06142	C181-5	1.1525	0.815	Cet	-3	4.01	9.96
50	06143	C183-12	0.2117	0.122	CSH	-7	8.2	9.95
	06121	C183-2	0.1867	0.108	McC	-4	5.2	9.95
	06122	C183-5	1.1025	0.780	Cet	-2	3.20	9.95
55	06145	C184-17	0.1095	0.077	CSH	-5	6.09	9.64
	06146	C185-22	0.3933	0.227	CSH	-5	5.88	9.88
	06126	C185-1	0.6050	0.428	McC	-4	4.88	9.88

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(continued)

<i>AIEC isolated from control subjects</i>								
5	06135	C185-2	0.4500	0.318	McC	-5	5.88	9.88
	06136	C185-3	0.4875	0.345	McC	-2	2.88	9.88
	06137	C185-6	0.3125	0.221	Cet	-2	2.88	9.88
	06127	C186-1	0.1063	0.053	McC	-6	6.86	10.04
10	06129	C186-4	0.4700	0.332	Cet	-3	3.86	10.04
	06158	C188-15	0.1052	0.061	CS ana	-6	6.94	9.60
	06196	C192-11	0.508	0.279	Cet	-2	2.92	9.44
15	06197	C195-1	0.206	0.157	McC	-5	5.95	9.72
	06198	C195-2	1.806	1.363	McC	-4	4.95	9.72
	06200	C195-6	2.498	1.482	Cet	-2	2.95	9.72
	06201	C196-1	0.218	0.105	McC	-7	7.87	9.61
20	06204	C196-4	0.307	0.163	McC	-5	5.87	9.61
	06212	C197-1	3.133	1.438	McC	-6	6.86	10.32
	06213	C197-2	0.445	0.042	McC	-6	6.86	10.32
25	06216	C197-6	0.886	0.782	Cet	-2	2.86	10.32
	06217	C198-1	0.143	0.112	McC	-6	7.07	10.03
	06218	C198-2	0.113	0.096	McC	-6	7.07	10.03
	06221	C199-3	5.367	3.132	McC	-4	5.06	9.79
30	06222	C199-4	1.353	0.942	McC	-3	4.06	9.79
	06223	C199-5	2.980	2.122	McC	-3	4.06	9.79
	06224	C199-6	5.398	2.837	McC	-3	4.06	9.79
35	06225	C200-1	0.538	0.277	McC	-3	4.06	9.79
	07032	C222-1	0.1038	0.0783	McC	-6	7.14	10.88
	07033	C222-2	1.2425	0.6575	McC	-6	7.14	10.88
40	07125	C230-1	0.300	0.055	McC	-5	6.0	10.2
	<sup>1</sup> McC = McConkey Agar (bioMérieux) Cet = Cetrimide Agar (bioMérieux) CS ana = anaerobic Columbia blood agar MRS = Man Rogosa Sharp Agar (Oxoid) CSH = Columbia SH Agar <sup>2</sup> The "level of <i>E. coli</i> " refers to the amount of the AIEC strain in the feces. <sup>3</sup> "Total Count" refers to all bacterial species in feces.							

**[0061]** CS ana culture medium has the following composition (per liter medium):

- 39 g of Columbia blood agar base (Oxoid)
- 5 g of glucose
- 0.3 g of cysteine chlorohydrate
- 5 g of agar
- pH 7.0 ± 0.2

**[0062]** The mixture is sterilized for 15 minutes at 121°C. Just before plating, 5% of horse blood is added.

**[0063]** CSH culture medium has the following composition (per liter medium):

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- 39 g of Columbia blood agar base (Oxoid)
- 3 g of cysteine chlorohydrate
- pH 6.8 ± 0.2

5 **[0064]** The mixture is sterilized for 15 minutes at 121°C. Just before plating, 2 ml of sterile ammonium citrate solution (0.25 g/10 ml water) are added. After incubation, bacteria using cysteine (and releasing sulfide) result in black colonies on this medium.

### 10 EXAMPLE 2

#### Phage isolation

15 **[0065]** Phages were isolated from sewage water as follows: sewage water was filtered at 0.2 µm and mixed with an equal volume of 2X Luria-Bertani (LB) medium. This mixture was inoculated with a fresh culture of LF82 strain and incubated on a shaker at 37°C overnight. Chloroform (1/10 volume) was added to the flask and placed on a shaker for one hour. The medium was centrifuged at 10,000 g for 10 min. 1 ml of the supernatant was collected and 1/10 vol. of chloroform was added. After a brief mix by vortex, the Eppendorf tube was centrifuged at 7,500 g for 5 min. To determine if phages were present in this extract, a drop (10 µl) of the supernatant was applied on an LB agar plate and allowed to dry. Using a platinum wire, the plate was streaked from the drop through the rest of the plate to isolate individual phages.

20 1 ml of a growing culture of LF82 strain was applied to cover the entire plate; the excess was removed and the plate was incubated at 37°C overnight. One or two plaques were picked up and resuspended in 200 µl of SM buffer (10 mM TrisHCl pH7, NaCl 200 mM, gelatin 0.03%). 20 µl of chloroform was added in each tube and tubes were briefly mixed by vortex and centrifuged at 7,500 g for 5 min. 10 µl of the supernatant was applied on a LB plate and allowed to dry and the previous procedure was repeated at least three times. Once the majority of isolated plaques were homogenous,

25 10 µl of the last resuspended plaque were added to 1 ml of growing culture of LF82 strain at OD 0.1 at 600 nm. This culture tube was incubated at 37°C for 2 to 4 hours until lysis occurred. After addition of 1/10 vol. of chloroform, the culture was transferred to an Eppendorf tube, centrifuged at 7,500 g for 5 min and cooled to 4°C, thereby obtaining the primary stock. Several dilutions of this stock were kept at 4°C and used to infect a larger volume of culture in order to prepare larger amounts of phages. Seven (7) phages were obtained as follows:

- vB\_EcoM\_LF82\_P1 (herein before and after P1) deposited with the French National Collection of Microorganisms at the Institut Pasteur under Accession Number CNCM I-4694;
- vB\_EcoM\_LF82\_P2 (herein before and after P2) deposited with the French National Collection of Microorganisms at the Institut Pasteur under Accession Number CNCM I-4695;
- 35 • vB\_EcoM\_LF82\_P3 (herein before and after P3) deposited with the French National Collection of Microorganisms at the Institut Pasteur under Accession Number CNCM I-4696;
- vB\_EcoM\_LF82\_P4 (herein before and after P4) deposited with the French National Collection of Microorganisms at the Institut Pasteur under Accession Number CNCM I-4697;
- vB\_EcoM\_LF82\_P5 (herein before and after P5) deposited with the French National Collection of Microorganisms at the Institut Pasteur under Accession Number CNCM I-4698;
- 40 • vB\_EcoM\_LF82\_P6 (herein before and after P6) deposited with the French National Collection of Microorganisms at the Institut Pasteur under Accession Number CNCM I-4699; and
- vB\_EcoM\_LF82\_P8 (herein before and after P8) deposited with the French National Collection of Microorganisms at the Institut Pasteur under Accession Number CNCM I-4700.

45 **[0066]** CLB\_P2, deposited with the French National Collection of Microorganisms at the Institut Pasteur under Accession Number CNCM I-4675, and its isolation is described in detail in Maura et al. Environmental Microbiology (2012) 14(8), 1844-1854.

**[0067]** P1 to P6 phages belong to the wV8 bacteriophage family.

50 **[0068]** P8 belongs to the RB69 bacteriophage family.

**[0069]** CLB\_P2 belongs to the JS98 bacteriophage family.

**[0070]** The classification into the wV8, RB69 and JS98 bacteriophage families was done based on the sequence of the major capsid protein.

55

EXAMPLE 3

**In vitro assays of the infectivity of bacteriophages in AIEC strains**

5 **[0071]** Plaque assay was carried out by contacting serial dilutions of bacteriophage solutions (from not diluted to 10<sup>-8</sup> dilution) with a Petri dish which surface was covered by one bacterium. After overnight incubation at 37°C plaques were counted. When the bacterium tested was the bacterial host (reference host) used to isolate bacteriophages it was considered that the plaque assay gave an efficiency of 100%. When the bacterium tested was not the original host, then the results were expressed by comparison to the reference host. A result greater than 80% (+++) means that the bacterium is a highly efficient host compared to the reference host, while a result between 0.1 and 80% (++) means that the bacterium is an efficient host, and a result below 0.1% (+) but above 0 means that the bacterium is a moderately efficient host, and finally 0 (-) means that the bacterium is totally resistant.

**Results**

15 **[0072]** Table 2 shows the result of the host spectrum of the 8 phages (as isolated/identified in Example 2) on 38 strains (out of the 166 strains isolated in Example 1, Table 1)

Table 2 Strains tested and effective efficiency of plating (EOP) obtained for each bacteriophage

Bacterial Strain	Bacteriophage							
	P1	P2	P3	P4	P5	P6	P8	CLB P2
LF82	+++	+++	+++	+++	+++	+++	+++	+++
06023	-	-	-	-	-	-	-	++
06030	-	-	-	-	-	-	+	+++
06033	++	++	++	++	++	++	+	+++
06066	-	-	-	-	-	-	+++	-
06072	-	-	-	-	-	-	+	++
06073	-	-	-	-	-	-	+	+++
06075	+++	+++	+++	+++	+++	+++	+	++
06088	++	++	++	++	++	++	-	-
06089	++	++	++	++	++	++	-	-
06122	++	++	++	++	++	++	-	-
06150	-	-	-	-	-	-	+	-
06351	++	++	++	++	++	++	-	++
06353	+	+	+	+	+	+	-	-
06354	-	-	-	-	-	-	-	-
06356	++	+	+	+	+	+	-	-
06357	+	+	+	+	+	+	-	-
06358	++	+	+	+	+	+	-	-
06359	++	+	+	+	+	+	-	-
06361	+	++	+	+	+	+	+	-
06362	-	-	-	-	-	-	-	-
07045	-	-	-	-	-	-	-	-
07046	-	-	-	-	-	-	-	++
07048	-	-	-	-	-	-	-	-
07051	-	-	-	-	-	-	-	-

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(continued)

Bacterial Strain	Bacteriophage							
	P1	P2	P3	P4	P5	P6	P8	CLB P2
07075	-	-	-	-	-	-	-	-
07076	++	+++	++	+	+	+	+++	++
07077	-	-	-	-	-	-	-	-
07078	++	+++	+	+	+	+	+	++
07081	++	+++	++	+	+	++	+++	++
07082	+++	+++	+++	+++	+++	+++	+++	++
07107	+++	++	+++	++	+++	+++	-	+++
07126	+++	+++	+++	+++	+++	+++	-	++
07127	+++	++	+++	+++	++	++	-	++
07128	-	-	-	-	-	-	-	++
07134	-	-	-	-	-	-	-	-
07135	-	-	-	-	-	-	-	++
07136	-	-	-	-	-	-	-	-
07137	-	-	-	-	-	-	++	-

Table 3 number of strains infected by phages:

Efficacy	P1	P2	P3	P4	P5	P6	P8	CLB P2
+	3	5	6	9	9	8	8	0
++	10	8	8	6	6	7	1	13
+++	5	6	5	4	4	4	4	5
<b>Total/38</b>	18	19	19	19	19	19	13	18
Numbers indicate the number of strains infected by one bacteriophage								

EXAMPLE 4

**In vivo replication of bacteriophages in the gut of mice**

[0073] *In vivo* replication of bacteriophages in the gut of mice was evaluated as follows:

First, the strain LF82 was engineered to carry two antibiotic resistance genes conferring respectively resistance to Streptomycin and Kanamycin. This new bacterial strain was named LF82SK and its invasive properties were verified as to be similar to the original LF82 strain.

[0074] Three (3) groups of two (2) mice each:

- Group 1: non-colonized mice + phages
- Group 2: LF82SK colonized mice
- Group 3: LF82SK colonized mice + phages

[0075] Streptomycin (5 g/L) was added to drinking water of all animals 3 days before day 0 and kept along the experiment.

[0076] At day 0, LF82SK was administered to Group 2 and 3 in order to allow the strain to colonize mice's gut.

[0077] At day 4, 200 µl of a cocktail of P2 + P6 bacteriophages was administered to Group 1 and 3 (gavage solution 10<sup>8</sup> pfu/ml) once in the morning and once in the afternoon. P2+P6 bacteriophages were also added to the drinking water

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( $10^8$  pfu/ml). At day 5 in the morning, mice were sacrificed to evaluate the number of bacteria and bacteriophages in the ileum and in the feces.

Results:

5

*Bacteria (E. coli):*

**[0078]**

10

Group 1: no bacteria;  
Group 2: in ileum -  $10^6$  cfu/g organ; in feces -  $10^8$  cfu/organ;  
Group 3: in ileum and feces: bacteria all lysed by phages.

*Phages:*

15

**[0079]**

20

Group 1: in ileum -  $10^6$  pfu/g organ; in feces -  $10^7$  pfu/organ;  
Group 2: no phages  
Group 3: in ileum -  $10^6$  pfu/g organ; in feces -  $10^{10}$  pfu/organ;

**[0080]** In the feces, there were 100 times more phages in Group 3 than in Group 1 showing the multiplication of the phages *in vivo*.

25

EXAMPLE 5

***In vivo* replication of bacteriophages in the gut of mice**

30

**[0081]** *In vivo* replication of bacteriophages in the gut of mice was evaluated as follows:

12 mice were dispatched into three (3) groups of four (4) mice each:

Group 1: non-colonized mice + phages  
Group 2: LF82SK-colonized mice  
Group 3: LF82SK-colonized mice + phages

35

**[0082]** Streptomycin (5 g/L) was added to drinking water of all animals 3 days before day 0 and kept along the experiment.

**[0083]** At day 0, LF82SK was given to mice of Group 2 and 3 in order to allow the strain to colonize mice's gut.

40

**[0084]** At day 4, bacteriophages (cocktail of P2+P6+P8 at  $10^8$  pfu/mL each) were added in the drinking water of Group 1 and 3.

**[0085]** At day 5, mice were sacrificed to evaluate the number of bacteria and bacteriophages in the ileum and in the feces. 100  $\mu$ l of ileal homogenates from the three groups were taken to extract whole DNA using Maxwell<sup>®</sup> 16 Tissue DNA purification kit from Promega.

45

Results:

*Bacteria (E. coli):*

**[0086]**

50

Group 1: no bacteria;  
Group 2: in ileum -  $3.2 \cdot 10^6$  cfu/g of organ; in feces -  $1.2 \cdot 10^9$  cfu/g of feces;  
Group 3: in ileum and feces: bacteria all lysed by phages.

55

*Phages:*

**[0087]**

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Group 1: in ileum -  $1.4 \cdot 10^6$  pfu/g of organ ; in feces -  $5.2 \cdot 10^6$  pfu/g of feces;

Group 2: no phages

Group 3: in ileum -  $2.6 \cdot 10^6$  pfu/g of organ; in feces -  $1.0 \cdot 10^9$  pfu/g of feces;

5 **[0088]** In the feces, there were 200 times more phages in Group 3 than in Group 1 showing the multiplication of the phages *in vivo*.

**[0089]** DNA extracted from ileal sections was used to run quantitative PCR using two sets of primers. One set of primers (SEQ ID NO: 30-31) served to amplify DNA from "all bacteria" present in the sample while the second set (SEQ ID NO: 32-33) was used to amplify specifically DNA from "*E. coli*" bacteria. After normalization, results were expressed as the ratio of *E. coli* versus all bacteria.

10 **[0090]** Group 1: qPCR amplifications were successful with all bacteria primers but not with *E. coli* primers. The ratio could not be calculated.

**[0091]** Group 2: qPCR amplifications were successful with both set of primers. The average ratio was 0.6 (60% of total bacteria were *E. coli* bacteria)

15 **[0092]** Group 3: qPCR amplifications were successful with both set of primers. The average ratio was 0.1 (10% of total bacteria were *E. coli* bacteria). Note that one mouse displayed a ratio of 0.4 while the three others displayed much lower values (0.06; 0.0002; 0.002).

**[0093]** In consequence bacteriophages were able to reduce the level of ileal colonization of LF82 bacteria by at least one order of magnitude in three mice out of four.

20

### EXAMPLE 6

#### ***In vivo* assay of the infectivity of bacteriophages**

25 **[0094]** Two cocktails of phages are selected for testing in wild-type (WT) mice and in CEACAM6 mice infected with the LF82 *E. coli* strain isolated from the CD patients. In both WT mice and in CEACAM6 mice infected with the LF82 *E. coli* strain isolated from the CD patients, bacteriophages are administered to the mice by oral gavage in CMC. This kind of administration has many advantages: known quantity of bacteriophage administration and immediate gastric acidity neutralization. Phages are daily administered to the mice during the entire study.

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- Mice are sacrificed at 5 days after LF82 administration.
- Main criteria: quantification of LF82 in ileal and colonic adherent flora of the mice.
- Minor criteria:

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- Evaluation of weight
- Stool consistency.
- Presence of fecal blood (macro and bio)

40

- Luminal flora (conventional flora + LF82 + phages)

- At sacrifice: Macroscopic and histologic examinations, adherent ileal and colonic flora + LF82 + phages,
- At sacrifice: Macroscopic and histologic examinations, adherent ileal and colonic flora + LF82 + phages,

45 **[0095]** Biological parameters of inflammation are monitored, and bacteriophage translocation in the mesenteric lymph nodes (MLN), liver and spleen is searched for.

**[0096]** Inflammation markers (MPO, pro-inflammatory cytokines IL-6, IL-12 and antiinflammatory cytokines IL-10) are monitored. Bacteriophage and AIEC translocation in MLN, liver and spleen is searched for.

**[0097]** Follow-up of bacteriophage elimination takes place in stools of mice receiving the bacteriophage cocktail without the LF82 strain.

50

### EXAMPLE 7

#### ***In vivo* assay of a cocktail of phages on the LF82 strain.**

55 **[0098]** *In vivo* replication of bacteriophages (cocktail of P2+P6+P8+CLB P2) in the gut of mice was evaluated as follows: 20 mice were dispatched into two (2) groups of ten (10) mice each:

Group 1: LF82SK-colonized mice

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Group 2: LF82SK-colonized mice + phages

[0099] Streptomycin (5 g/L) was added to drinking water of all animals 3 days before day 0 and kept along the experiment.

[0100] At day 0, LF82SK was given to mice of both groups in order to allow the strain to colonize mice's gut.

[0101] At day 3, bacteriophages (cocktail of P2+P6+P8+CLB\_P2 at  $10^8$  pfu/mL each) were given to mice of Group 2 by gavage.

[0102] At day 4 and 7, 5 mice of each group were sacrificed to evaluate the number of bacteria and bacteriophages in the ileum, in the colon and in the feces. 100  $\mu$ l of ileal and colonic homogenates from the two groups were taken to extract whole DNA using Maxwell<sup>®</sup> 16 Tissue DNA purification kit from Promega.

Results:

Level of LF82 in stools:

[0103] At day 4 and 7 levels of LF82 were:

in group 1:  $7 \cdot 10^9$  ;  $1 \cdot 10^9$  cfu/g

in group 2:  $8 \cdot 10^7$  ;  $5 \cdot 10^8$  cfu/g

Level of Phages in stools:

[0104] At day 4 and 7 levels of Phages were:

in group 1: none

in group 2:  $5 \cdot 10^9$ ;  $6 \cdot 10^9$  pfu/g

[0105] In the presence of the phage cocktail the level of LF82 in stools was significantly lower than in their absence showing that the phage cocktail was able to infect LF82 inside mice's gut.

Level of LF82 in organs:

[0106] at day 4 levels of LF82 were:

in ileum of group 1: 100% of bacteria are E. coli (LF82)

in ileum of group 2: 20% of bacteria are E. coli (LF82)

in colon of group 1: 40% of bacteria are E. coli (LF82)

in colon of group 2: 2% of bacteria are E. coli (LF82)

[0107] at day 7 levels of LF82 were:

in ileum of group 1: 100% of bacteria are E. coli (LF82)

in ileum of group 2: 50% of bacteria are E. coli (LF82)

in colon of group 1: 25% of bacteria are E. coli (LF82)

in colon of group 2: 10% of bacteria are E. coli (LF82)

Level of Phages in organs:

[0108] at day 4 levels of Phages were:

in ileum of group 1: none

in ileum of group 2:  $7 \cdot 10^8$  pfu/g

in colon of group 1: none

in colon of group 2:  $5 \cdot 10^{10}$  pfu/g

[0109] at day 7 levels of LF82 were:

in ileum of group 1: none

in ileum of group 2:  $7 \times 10^8$  pfu/g  
in colon of group 1: none  
in colon of group 4:  $2 \times 10^8$  pfu/g

5 **[0110]** At day 2 and 5 the level of LF82 was reduced in both ileum and colon in the group treated by phages. This shows that phages infect LF82 in gut sections and not only in stools. Concomitantly, the level of phages at day 7 stays as high as at day 2 showing that phage can last several days in the gut after a unique initial administration.

10 EXAMPLE 8

***In vivo* assay of the infectivity of bacteriophages**

**[0111]** *In vivo* assay of the infectivity of bacteriophages (cocktail of P2+P6+P8) in CEACAM6 mice infected with LF82SK was evaluated as follows:

15 48 mice were dispatched into three (4) groups as follows:

- Group 1: non-colonized mice (8 mice)
- Group 2: non-colonized mice + phages (12 mice)
- Group 3: LF82SK-colonized mice (16 mice)
- 20 Group 4: LF82SK-colonized mice + phages (12 mice)

**[0112]** DSS (dextran sulfate) 0.25% was introduced in the drinking water 3 days before day 0 and kept along the experiment.

**[0113]** Streptomycin (5mg) was administrated by oral gavage to all animals 1 day before day 0.

25 **[0114]** At day 0, LF82SK was administered to mice of Group 3 and 4 in order to allow the strain to colonize mice's gut.

**[0115]** At day 1, phages (cocktail of P2+P6+P8 at  $10^7$  pfu/mL each) were administered once to each mouse of Group 2 and 4 by oral gavage in CMC. This kind of administration has many advantages: known quantity of bacteriophage administration and immediate gastric acidity neutralization.

30 **[0116]** At day 1, 4 mice from Group 3 were sacrificed to evaluate the number of bacteria in the ileum, in the colon and in the feces before the administration of phages.

**[0117]** At day 2, respectively 4, 6, 6 and 6 mice from Groups 1, 2, 3 and 4 were sacrificed to evaluate the number of bacteria and bacteriophages in the ileum, in the colon and in the feces.

**[0118]** At day 5, respectively 4, 6, 6 and 6 mice from Groups 1, 2, 3 and 4 were sacrificed to evaluate the number of bacteria and bacteriophages in the ileum, in the colon and in the feces.

35 **[0119]** 100  $\mu$ l of ileal, colon and feces homogenates from the four groups were taken to extract whole DNA using Maxwell<sup>®</sup> 16 Tissue DNA purification kit from Promega. Weight, stool consistency and presence of fecal blood were monitored daily.

**[0120]** DNA extracted from ileal sections was used to run quantitative PCR using one set of primers (SEQ ID NO: 44-45) to amplify a specific gene (pMT1) from LF82. Results were expressed as the number of copies of this gene per gram of tissues.

40

LF82 pMT1 F (SEQ ID NO:44) CCATTCATGCAGCAGCTCTTT  
LF82 pMT1 R (SEQ ID NO:45) ATCGGACAACATTAGCGGTGT

45 Results:

**[0121]** Values represent the median values obtained for each group of mice.

**[0122]** In group 1, neither LF82 nor Phages were detected along the experiment.

50 Level of LF82 in stools:

**[0123]**

At day 1: the level of LF82 in Groups 3 and 4 were  $5 \times 10^9$  and  $6 \times 10^9$  cfu/g resp.

55 At day 2, 3 and 5 levels of LF82 were:

in group 3:  $3 \times 10^9$  ;  $5 \times 10^8$  ;  $5 \times 10^7$  cfu/g  
in group 4:  $5 \times 10^5$  ;  $5 \times 10^5$ ;  $5 \times 10^3$  cfu/g

Level of Phages in stools:

**[0124]** At day 2, 3 and 5 levels of Phages were:

5 in group 2:  $5 \times 10^5$  pfu/g; not detected; not detected  
in group 4:  $1 \times 10^9$ ;  $1 \times 10^7$ ;  $5 \times 10^6$  pfu/g

**[0125]** In the presence of phages the level of LF82 in stools was significantly lower than in their absence. Concomitantly, the level of phages was significantly higher in mice colonised by LF82 than in LF82-free mice. Both data confirmed that phages can infect LF82 in the gut.

Level of LF82 in organs:

**[0126]**

15 at day 2 levels of LF82 were:

in ileum of group 3:  $2 \times 10^6$  copies of pMT1/g  
in ileum of group 4:  $8 \times 10^4$  copies of pMT1/g  
20 in colon of group 3:  $2 \times 10^7$  copies of pMT1/g  
in colon of group 4:  $1 \times 10^5$  copies of pMT1/g

at day 5 levels of LF82 were:

25 in ileum of group 3:  $5 \times 10^4$  copies of pMT1/g  
in ileum of group 4:  $8 \times 10^4$  copies of pMT1/g  
in colon of group 3:  $6 \times 10^6$  copies of pMT1/g  
in colon of group 4:  $2 \times 10^5$  copies of pMT1/g

30 Level of Phages in organs:

**[0127]**

35 at day 2 levels of Phages were:

in ileum of group 2: not detected  
in ileum of group 4:  $8 \times 10^5$  pfu/g  
in colon of group 2:  $5 \times 10^4$  pfu/g  
40 in colon of group 4:  $5 \times 10^6$  pfu/g

at day 5 levels of LF82 were:

45 in ileum of group 2: not detected  
in ileum of group 4: not detected  
in colon of group 2: not detected  
in colon of group 4:  $2 \times 10^4$  pfu/g

**[0128]** At day 2, the level of LF82 was reduced in both ileum and colon in the group treated by phages. This shows that phages infected LF82 in gut sections and not only in stools. Concomitantly, the level of phages was significantly higher in mice colonised by LF82 than in LF82-free mice.

**[0129]** At day 5, the level of LF82 in ileum was too weak to see a difference between the two groups while in colon samples the level of LF82 was still reduced in the group that received phages compared to the groups that did not. Concomitantly, we could only detect phages in colon of mice colonised by LF82. This shows that the effect of phages in reducing LF82 can last several days after the initial administration.

55 **[0130]** Despite high colonisation level of LF82 observed in this experiment, no sign of colitis was observed in any of the groups.

SEQUENCE LISTING

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	Met	Leu	Thr	Asn	Ser	Glu	Lys	Ser	Arg	Phe	Phe	Leu	Ala	Asp	Leu	Thr
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			35					40					45			
	Leu	Thr	Asp	Trp	Asp	Val	Ser	Leu	Leu	Asp	Ala	Val	Asp	Arg	Asp	Ser
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			115					120					125			
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40	Asn	Thr	Leu	Gly	Gln	Glu	Leu	Tyr	Val	Phe	Glu	Tyr	Glu	Lys	Asp	Arg
					325					330					335	
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			340						345					350		
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 50 55

25

<210> 4  
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 <213> Artificial Sequence

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<220>  
 <223> Peptide from bacteriophage strain P1-P6 aligning with position 77-115 of the major capsid protein of bacteriophage wV8 (SEQ ID NO: 1)

35

<400> 4

Gln Ile Ser Phe Pro Met Met Tyr Phe Lys Glu Val Glu Ser Ile Thr  
 1 5 10 15  
 Pro Asp Glu Ile Gln Gly Val Arg Gln Pro Gly Thr Ala Asn Glu Leu  
 20 25 30  
 Thr Thr Glu Ala Val Val Arg  
 35

40

<210> 5  
 <211> 16  
 <212> PRT  
 <213> Artificial Sequence

45

<220>  
 <223> Peptide from bacteriophage strain P1-P6 aligning with position 124-139 of the major capsid protein of bacteriophage wV8 (SEQ ID NO: 1)

50

<400> 5

55

Thr Lys Phe Asp Ile Thr Arg Glu Phe Leu Phe Met Gln Ala Leu Lys  
 1 5 10 15

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<210> 6  
<211> 9  
<212> PRT  
<213> Artificial Sequence  
5  
<220>  
<223> Peptide from bacteriophage strain P1-P5 aligning with position 147-155 of the major capsid protein of bacteriophage wV8 (SEQ ID NO: 1)  
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<400> 6  
  
Gly Thr Leu Tyr Ala Asp Leu Tyr Lys  
1 5  
15  
<210> 7  
<211> 15  
<212> PRT  
<213> Artificial Sequence  
20  
<220>  
<223> Peptide from bacteriophage strain P6 aligning with position 147-161 of the major capsid protein of bacteriophage wV8 (SEQ ID NO: 1)  
25  
<400> 7  
  
Gly Thr Leu Tyr Ala Asp Leu Tyr Lys Gln Phe Asp Val Glu Lys  
1 5 10 15  
30  
<210> 8  
<211> 22  
<212> PRT  
<213> Artificial Sequence  
35  
<220>  
<223> Peptide from bacteriophage strain P1 aligning with position 162-183 of the major capsid protein of bacteriophage wV8 (SEQ ID NO: 1)  
40  
<400> 8  
  
Lys Thr Val Tyr Phe Asp Leu Asp Asn Pro Asn Ala Asp Ile Asp Ala  
1 5 10 15  
45 Ser Ile Glu Glu Leu Arg  
20  
<210> 9  
<211> 21  
<212> PRT  
<213> Artificial Sequence  
50  
<220>  
<223> Peptide from bacteriophage strain P3 aligning with position 163-183 of the major capsid protein of bacteriophage wV8 (SEQ ID NO: 1)  
55  
<400> 9

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Thr Val Tyr Phe Asp Leu Asp Asn Pro Asn Ala Asp Ile Asp Ala Ser  
1 5 10 15  
Ile Glu Glu Leu Arg  
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5

<210> 10  
<211> 21  
<212> PRT  
<213> Artificial Sequence

10

<220>  
<223> Peptide from bacteriophage strain P5-P6 aligning with position 163-183 of the major capsid protein of bacteriophage wV8 (SEQ ID NO: 1)

15

<400> 10

Thr Ile Tyr Phe Asp Leu Asp Asn Pro Asn Ala Asp Ile Asp Ala Ser  
1 5 10 15  
Ile Glu Glu Leu Arg  
20

20

<210> 11  
<211> 16  
<212> PRT  
<213> Artificial Sequence

25

<220>  
<223> Peptide from bacteriophage strain P1 and P3-P6 aligning with position 192-207 of the major capsid protein of bacteriophage wV8 (SEQ ID NO: 1)

30

<400> 11

Thr Gly Thr Val Ile Asn Gly Glu Glu Ile His Val Val Val Asp Arg  
1 5 10 15

35

<210> 12  
<211> 21  
<212> PRT  
<213> Artificial Sequence

40

<220>  
<223> Peptide from bacteriophage strain P2 aligning with position 192-212 of the major capsid protein of bacteriophage wV8 (SEQ ID NO: 1)

45

<400> 12

Thr Gly Thr Val Ile Asn Gly Glu Glu Ile His Val Val Val Asp Arg  
1 5 10 15  
Leu Phe Phe Ser Lys  
20

50

<210> 13  
<211> 22  
<212> PRT  
<213> Artificial Sequence

55

<220>

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<223> Peptide from bacteriophage strain P1 aligning with position 219-240 of the major capsid protein of bacteriophage wV8 (SEQ ID NO: 1)

<400> 13

5

Ile Arg Asp Ala Tyr Leu Ala Gln Gln Thr Pro Leu Ala Trp Gln Gln  
1 5 10 15  
Ile Thr Gly Ser Leu Arg  
20

10

<210> 14

<211> 49

<212> PRT

<213> Artificial Sequence

15

<220>

<223> Peptide from bacteriophage strain P2 aligning with position 219-267 of the major capsid protein of bacteriophage wV8 (SEQ ID NO: 1)

20

<400> 14

Ile Arg Asp Ala Tyr Leu Ala Gln Gln Thr Pro Leu Ala Trp Gln Gln  
1 5 10 15  
Ile Thr Gly Ser Leu Arg Thr Gly Gly Thr Asp Gly Val Gln Ala His  
20 25 30  
Met Asn Thr Phe Tyr Tyr Gly Gly Val Lys Phe Val Gln Tyr Asn Gly  
35 40 45  
Lys

25

30

<210> 15

<211> 20

<212> PRT

<213> Artificial Sequence

35

<220>

<223> Peptide from bacteriophage strain P3-P4 aligning with position 221-240 of the major capsid protein of bacteriophage wV8 (SEQ ID NO: 1)

40

<400> 15

Asp Ala Tyr Leu Ala Gln Gln Thr Pro Leu Ala Trp Gln Gln Ile Thr  
1 5 10 15  
Gly Ser Leu Arg  
20

45

<210> 16

<211> 47

<212> PRT

<213> Artificial Sequence

50

<220>

<223> Peptide from bacteriophage strain P5 aligning with position 221-267 of the major capsid protein of bacteriophage wV8 (SEQ ID NO: 1)

55

<400> 16

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5  
 Asp Ala Tyr Leu Ala Gln Gln Thr Pro Leu Ala Trp Gln Gln Ile Thr  
 1 5 10 15  
 Gly Ser Leu Arg Thr Gly Gly Ala Asp Gly Val Gln Ala His Met Asn  
 20 25 30  
 Thr Phe Tyr Tyr Gly Gly Val Lys Phe Val Gln Tyr Asn Gly Lys

35 40 45

10  
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 <211> 40  
 <212> PRT  
 <213> Artificial Sequence  
 15  
 <220>  
 <223> Peptide from bacteriophage strain P6 aligning with position 221-260 of the major capsid protein of bacteriophage wV8 (SEQ ID NO: 1)

20  
 <400> 17  
 Asp Ala Tyr Leu Ala Gln Gln Thr Pro Leu Ala Trp Gln Gln Ile Thr  
 1 5 10 15  
 Gly Ser Leu Arg Thr Gly Gly Ala Asp Gly Val Gln Ala His Met Asn  
 25 20 25 30  
 Thr Phe Tyr Tyr Gly Gly Val Lys  
 35 40

30  
 <210> 18  
 <211> 7  
 <212> PRT  
 <213> Artificial Sequence  
 35  
 <220>  
 <223> Peptide from bacteriophage strain P1, P3 and P4 aligning with position 261-267 of the major capsid protein of bacteriophage wV8 (SEQ ID NO: 1)

<400> 18  
 40  
 Phe Val Gln Tyr Asn Gly Lys  
 1 5

45  
 <210> 19  
 <211> 20  
 <212> PRT  
 <213> Artificial Sequence  
 50  
 <220>  
 <223> Peptide from bacteriophage strain P1-P2 aligning with position 317-336 of the major capsid protein of bacteriophage wV8 (SEQ ID NO: 1)

<400> 19  
 55  
 Met Gly Tyr Ala Asn Thr Leu Gly Gln Glu Leu Tyr Val Phe Glu Tyr  
 1 5 10 15  
 Glu Lys Asp Arg  
 20

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<210> 20  
<211> 8  
<212> PRT  
<213> Artificial Sequence

5

<220>  
<223> Peptide from bacteriophage strain P1-P6 aligning with position 355-362 of the major capsid protein of bacteriophage wV8 (SEQ ID NO: 1)

10

<400> 20

Pro Gln Leu Leu Val Asp Val Arg  
1 5

15

<210> 21  
<211> 522  
<212> PRT  
<213> Bacteriophage RB69

20

<400> 21

25

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35

40

45

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55

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Met Thr Thr Ile Lys Thr Lys Ala Gln Leu Val Asp Lys Trp Lys Glu  
1 5 10  
Leu Leu Glu Gly Glu Gly Leu Pro Glu Ile Ala Asn Ser Lys Gln Ala  
20 25 30  
5 Ile Ile Ala Lys Ile Phe Glu Asn Gln Glu Lys Asp Phe Glu Val Ser  
35 40 45  
Pro Glu Tyr Lys Asp Glu Lys Ile Ala Gln Ala Phe Gly Ser Phe Leu  
50 55 60  
10 Thr Glu Ala Glu Ile Gly Gly Asp His Gly Tyr Asn Ala Gln Asn Ile  
65 70 75 80  
Ala Ala Gly Gln Thr Ser Gly Ala Val Thr Gln Ile Gly Pro Ala Val  
85 90 95  
Met Gly Met Val Arg Arg Ala Ile Pro Asn Leu Ile Ala Phe Asp Ile  
100 105 110  
15 Cys Gly Val Gln Pro Met Asn Ser Pro Thr Gly Gln Val Phe Ala Leu  
115 120 125  
Arg Ala Val Tyr Gly Lys Asp Pro Ile Ala Ala Gly Ala Lys Glu Ala  
130 135 140  
Phe His Pro Met Tyr Ala Pro Asp Ala Met Phe Ser Gly Gln Gly Ala  
145 150 155 160  
20 Ala Lys Lys Phe Pro Ala Leu Ala Ala Ser Thr Gln Thr Lys Val Gly  
165 170 175  
Asp Ile Tyr Thr His Phe Phe Gln Glu Thr Gly Thr Val Tyr Leu Gln  
180 185 190  
25 Ala Ser Ala Gln Val Thr Ile Ser Ser Ser Ala Asp Asp Ala Ala Lys  
195 200 205  
Leu Asp Ala Glu Ile Ile Lys Gln Met Glu Ala Gly Ala Leu Val Glu  
210 215 220  
Ile Ala Glu Gly Met Ala Thr Ser Ile Ala Glu Leu Gln Glu Gly Phe  
225 230 235 240  
30 Asn Gly Ser Thr Asp Asn Pro Trp Asn Glu Met Gly Phe Arg Ile Asp  
245 250 255  
Lys Gln Val Ile Glu Ala Lys Ser Arg Gln Leu Lys Ala Ala Tyr Ser  
260 265 270  
Ile Glu Leu Ala Gln Asp Leu Arg Ala Val His Gly Met Asp Ala Asp  
275 280 285  
35 Ala Glu Leu Ser Gly Ile Leu Ala Thr Glu Ile Met Leu Glu Ile Asn  
290 295 300  
Arg Glu Val Val Asp Trp Ile Asn Tyr Ser Ala Gln Val Gly Lys Ser  
305 310 315 320  
Gly Met Thr Asn Ile Val Gly Ser Lys Ala Gly Val Phe Asp Phe Gln  
325 330 335  
40 Asp Pro Ile Asp Ile Arg Gly Ala Arg Trp Ala Gly Glu Ser Phe Lys  
340 345 350  
Ala Leu Leu Phe Gln Ile Asp Lys Glu Ala Val Glu Ile Ala Arg Gln  
355 360 365  
45 Thr Gly Arg Gly Glu Gly Asn Phe Ile Ile Ala Ser Arg Asn Val Val  
370 375 380  
Asn Val Leu Ala Ser Val Asp Thr Gly Ile Ser Tyr Ala Ala Gln Gly  
385 390 395 400  
Leu Ala Ser Gly Phe Asn Thr Asp Thr Thr Lys Ser Val Phe Ala Gly  
405 410 415  
50 Val Leu Gly Gly Lys Tyr Arg Val Tyr Ile Asp Gln Tyr Ala Lys Gln  
420 425 430  
Asp Tyr Phe Thr Val Gly Tyr Lys Gly Ala Asn Glu Met Asp Ala Gly  
435 440 445

55

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Ile Tyr Tyr Ala Pro Tyr Val Ala Leu Thr Pro Leu Arg Gly Ser Asp  
 450 455 460  
 Pro Lys Asn Phe Gln Pro Val Met Gly Phe Lys Thr Arg Tyr Gly Ile  
 465 470 475 480  
 5 Gly Val Asn Pro Phe Ala Glu Ser Ser Leu Gln Ala Pro Gly Ala Arg  
 485 490 495  
 Ile Gln Ser Gly Met Pro Ser Ile Leu Asn Ser Leu Gly Lys Asn Ala  
 500 505 510  
 10 Tyr Phe Arg Arg Val Tyr Val Lys Gly Ile  
 515 520

<210> 22

<211> 20

<212> PRT

15 <213> Artificial Sequence

<220>

<223> Peptide from bacteriophage strain P8 aligning with position 143-162 of the major capsid protein of bacteriophage RB69 (SEQ ID NO: 21)

20

<400> 22

Glu Ala Phe His Pro Met Tyr Ala Pro Asp Ala Met Phe Ser Gly Gln  
 1 5 10 15  
 25 Gly Ala Ala Lys  
 20

<210> 23

<211> 12

30 <212> PRT

<213> Artificial Sequence

<220>

<223> Peptide from bacteriophage strain P8 aligning with position 269-280 of the major capsid protein of bacteriophage RB69 (SEQ ID NO: 21)

35

<400> 23

Ala Ala Tyr Ser Ile Glu Leu Ala Gln Asp Leu Arg  
 1 5 10

<210> 24

<211> 14

45 <212> PRT

<213> Artificial Sequence

<220>

<223> Peptide from bacteriophage strain P8 aligning with position 306-319 of the major capsid protein of bacteriophage RB69 (SEQ ID NO: 21)

50

<400> 24

Glu Val Val Asp Trp Ile Asn Tyr Ser Ala Gln Val Gly Lys  
 1 5 10

<210> 25

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<211> 13  
<212> PRT  
<213> Artificial Sequence

5 <220>  
<223> Peptide from bacteriophage strain P8 aligning with position 330-342 of the major capsid protein of bacteriophage RB69 (SEQ ID NO: 21)

10 <400> 25

Ala Gly Val Phe Asp Phe Gln Asp Pro Ile Asp Ile Arg  
1 5 10

15 <210> 26  
<211> 7  
<212> PRT  
<213> Artificial Sequence

20 <220>  
<223> Peptide from bacteriophage strain P8 aligning with position 346-352 of the major capsid protein of bacteriophage RB69 (SEQ ID NO: 21)

25 <400> 26

Trp Ala Gly Glu Ser Phe Lys  
1 5

30 <210> 27  
<211> 14  
<212> PRT  
<213> Artificial Sequence

35 <220>  
<223> Peptide from bacteriophage strain P8 aligning with position 368-381 of the major capsid protein of bacteriophage RB69 (SEQ ID NO: 21)

40 <400> 27

Gln Thr Gly Arg Gly Glu Gly Asn Phe Ile Ile Ala Ser Arg  
1 5 10

45 <210> 28  
<211> 50  
<212> PRT  
<213> Artificial Sequence

50 <220>  
<223> Peptide from bacteriophage strain P8 aligning with position 412-461 of the major capsid protein of bacteriophage RB69 (SEQ ID NO: 21)

55 <400> 28

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Ser Val Phe Ala Gly Val Leu Gly Gly Lys Tyr Arg Val Tyr Ile Asp  
 1 5 10 15  
 Gln Tyr Ala Lys Gln Asp Tyr Phe Thr Val Gly Tyr Lys Gly Ala Asn  
 20 25 30  
 5 Glu Met Asp Ala Gly Ile Tyr Tyr Ala Pro Tyr Val Ala Leu Thr Pro  
 35 40 45  
 Leu Arg  
 50

10 <210> 29  
 <211> 44  
 <212> PRT  
 <213> Artificial Sequence

15 <220>  
 <223> Peptide from bacteriophage strain P8 aligning with position 467-510 of the major capsid protein of bacteriophage RB69 (SEQ ID NO: 21)

20 <400> 29

Asn Phe Gln Pro Val Met Gly Phe Lys Thr Arg Tyr Gly Ile Gly Val  
 1 5 10 15  
 Asn Pro Phe Ala Glu Ser Ser Leu Gln Ala Pro Gly Ala Arg Ile Gln  
 20 25 30  
 25 Ser Gly Met Pro Ser Ile Leu Asn Ser Leu Gly Lys  
 35 40

30 <210> 30  
 <211> 18  
 <212> DNA  
 <213> Artificial Sequence

35 <220>  
 <223> All bacteria 16S gene forward primer

<400> 30  
 cggtgaatac gttcccg 18

40 <210> 31  
 <211> 22  
 <212> DNA  
 <213> Artificial Sequence

45 <220>  
 <223> All bacteria 16S gene reverse primer

<400> 31  
 tacggctacc ttgttagc tt 22

50 <210> 32  
 <211> 20  
 <212> DNA  
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55 <220>  
 <223> E. coli 16S gene forward primer

<400> 32

catgccgcgt gatatgaagaa 20

<210> 33

<211> 21

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<212> DNA

<213> Artificial Sequence

<220>

<223> E. coli 16S gene reverse primer

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<400> 33

cgggtaacgt caatgagcaa a 21

<210> 34

15

<211> 519

<212> PRT

<213> Bacteriophage JS98

<400> 34

20

25

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45

50

55

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	1			5						10					15	
	Glu	Ala	Leu	Pro	Glu	Ile	Val	Gly	Ala	Ser	Lys	Gln	Ala	Ile	Ile	Ala
				20					25					30		
5	Lys	Ile	Phe	Glu	Asn	Gln	Glu	Gln	Asp	Ile	Leu	Thr	Ala	Pro	Glu	Tyr
			35					40					45			
	Arg	Asp	Glu	Lys	Ile	Ser	Glu	Ala	Phe	Gly	Ser	Phe	Leu	Thr	Glu	Ala
		50					55					60				
	Glu	Ile	Gly	Gly	Asp	His	Gly	Tyr	Asp	Ala	Thr	Asn	Ile	Ala	Ala	Gly
	65				70						75					80
10	Gln	Thr	Ser	Gly	Ala	Val	Thr	Gln	Ile	Gly	Pro	Ala	Val	Met	Gly	Met
				85						90					95	
	Val	Arg	Arg	Ala	Ile	Pro	His	Leu	Ile	Ala	Phe	Asp	Ile	Cys	Gly	Val
				100					105					110		
	Gln	Pro	Leu	Asn	Asn	Pro	Thr	Gly	Gln	Val	Phe	Ala	Leu	Arg	Ala	Val
				115				120					125			
15	Tyr	Gly	Lys	Asp	Pro	Ile	Ala	Ala	Gly	Ala	Lys	Glu	Ala	Phe	His	Pro
		130					135					140				
	Met	Tyr	Ala	Pro	Asn	Ala	Met	Phe	Ser	Gly	Gln	Gly	Ala	Ala	Glu	Thr
	145					150						155				160
	Phe	Glu	Ala	Leu	Ala	Ala	Ser	Lys	Val	Leu	Glu	Val	Gly	Lys	Ile	Tyr
20				165						170					175	
	Ser	His	Phe	Phe	Glu	Ala	Thr	Gly	Ser	Ala	His	Phe	Gln	Ala	Val	Glu
				180					185					190		
	Ala	Val	Thr	Val	Asp	Ala	Gly	Ala	Thr	Asp	Ala	Ala	Lys	Leu	Asp	Ala
				195				200					205			
	Ala	Val	Thr	Ala	Leu	Val	Glu	Ala	Gly	Gln	Leu	Ala	Glu	Ile	Ala	Glu
25				210			215					220				
	Gly	Met	Ala	Thr	Ser	Ile	Ala	Glu	Leu	Gln	Glu	Gly	Phe	Asn	Gly	Ser
	225					230					235					240
	Thr	Asp	Asn	Pro	Trp	Asn	Glu	Met	Gly	Phe	Arg	Ile	Asp	Lys	Gln	Val
				245						250					255	
30	Ile	Glu	Ala	Lys	Ser	Arg	Gln	Leu	Lys	Ala	Ser	Tyr	Ser	Ile	Glu	Leu
				260					265					270		
	Ala	Gln	Asp	Leu	Arg	Ala	Val	His	Gly	Met	Asp	Ala	Asp	Ala	Glu	Leu
				275				280					285			
	Ser	Gly	Ile	Leu	Ala	Thr	Glu	Ile	Met	Leu	Glu	Ile	Asn	Arg	Glu	Val
		290					295					300				
35	Ile	Asp	Trp	Ile	Asn	Tyr	Ser	Ala	Gln	Val	Gly	Lys	Ser	Gly	Met	Thr
	305					310					315					320
	Asn	Thr	Val	Gly	Ala	Lys	Ala	Gly	Val	Phe	Asp	Phe	Gln	Asp	Pro	Ile
				325						330					335	
	Asp	Ile	Arg	Gly	Ala	Arg	Trp	Ala	Gly	Glu	Ser	Phe	Lys	Ala	Leu	Leu
				340					345					350		
40	Phe	Gln	Ile	Asp	Lys	Glu	Ala	Ala	Glu	Ile	Ala	Arg	Gln	Thr	Gly	Arg
				355				360					365			
	Gly	Ala	Gly	Asn	Phe	Ile	Ile	Ala	Ser	Arg	Asn	Val	Val	Asn	Val	Leu
				370			375					380				
	Ala	Ala	Val	Asp	Thr	Ser	Val	Ser	Tyr	Ala	Ala	Gln	Gly	Leu	Gly	Gln
45				385			390				395					400
	Gly	Phe	Asn	Val	Asp	Thr	Thr	Lys	Ala	Val	Phe	Ala	Gly	Val	Leu	Gly
				405						410					415	
	Gly	Lys	Tyr	Arg	Val	Tyr	Ile	Asp	Gln	Tyr	Ala	Arg	Ser	Asp	Tyr	Phe
				420				425						430		
	Thr	Ile	Gly	Tyr	Lys	Gly	Ser	Asn	Glu	Met	Asp	Ala	Gly	Ile	Tyr	Tyr
50				435				440					445			
	Ala	Pro	Tyr	Val	Ala	Leu	Thr	Pro	Leu	Arg	Gly	Ser	Asp	Pro	Lys	Asn
				450			455					460				
	Phe	Gln	Pro	Val	Met	Gly	Phe	Lys	Thr	Arg	Tyr	Gly	Ile	Gly	Ile	Asn
	465					470					475					480
	Pro	Phe	Ala	Asp	Pro	Ala	Ala	Gln	Ala	Pro	Thr	Lys	Arg	Ile	Gln	Asn
55				485						490					495	
	Gly	Met	Pro	Asp	Ile	Val	Asn	Ser	Leu	Gly	Leu	Asn	Gly	Tyr	Phe	Arg

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Arg Val Tyr Val Lys Gly Ile  
 500 505 510  
 515

5 <210> 35  
 <211> 13  
 <212> PRT  
 <213> Artificial Sequence

10 <220>  
 <223> Peptide from bacteriophage strain CLB\_P2 aligning with position 127-139 of the major capsid protein of bacteriophage JS98 (SEQ ID NO: 34)

15 <400> 35

Ala Val Tyr Gly Lys Asp Pro Ile Ala Ala Gly Ala Lys  
 1 5 10

20 <210> 36  
 <211> 12  
 <212> PRT  
 <213> Artificial Sequence

25 <220>  
 <223> Peptide from bacteriophage strain CLB\_P2 aligning with position 266-277 of the major capsid protein of bacteriophage JS98 (SEQ ID NO: 34)

30 <400> 36

Ala Ser Tyr Ser Ile Glu Leu Ala Gln Asp Leu Arg  
 1 5 10

35 <210> 37  
 <211> 14  
 <212> PRT  
 <213> Artificial Sequence

40 <220>  
 <223> Peptide from bacteriophage strain CLB\_P2 aligning with position 303-316 of the major capsid protein of bacteriophage JS98 (SEQ ID NO: 34)

45 <400> 37

Glu Val Ile Asp Trp Ile Asn Tyr Ser Ala Gln Val Gly Lys  
 1 5 10

50 <210> 38  
 <211> 13  
 <212> PRT  
 <213> Artificial Sequence

55 <220>  
 <223> Peptide from bacteriophage strain CLB\_P2 aligning with position 327-339 of the major capsid protein of bacteriophage JS98 (SEQ ID NO: 34)

<400> 38

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Ala Gly Val Phe Asp Phe Gln Asp Pro Ile Asp Ile Arg  
1 5 10

5 <210> 39  
<211> 22  
<212> PRT  
<213> Artificial Sequence

10 <220>  
<223> Peptide from bacteriophage strain CLB\_P2 aligning with position 343-364 of the major capsid protein of bacteriophage JS98 (SEQ ID NO: 34)

15 <400> 39

Trp Ala Gly Glu Ser Phe Lys Ala Leu Leu Phe Gln Ile Asp Lys Glu  
1 5 10 15  
Ala Ala Glu Ile Ala Arg  
20

20 <210> 40  
<211> 10  
<212> PRT  
<213> Artificial Sequence

25 <220>  
<223> Peptide from bacteriophage strain CLB\_P2 aligning with position 369-378 of the major capsid protein of bacteriophage JS98 (SEQ ID NO: 34)

30 <400> 40

Gly Ala Gly Asn Phe Ile Ile Ala Ser Arg  
1 5 10

35 <210> 41  
<211> 20  
<212> PRT  
<213> Artificial Sequence

40 <220>  
<223> Peptide from bacteriophage strain CLB\_P2 aligning with position 409-428 of the major capsid protein of bacteriophage JS98 (SEQ ID NO: 34)

45 <400> 41

Ala Val Phe Ala Gly Val Leu Gly Gly Lys Tyr Arg Val Tyr Ile Asp  
1 5 10 15  
Gln Tyr Ala Arg  
20

50 <210> 42  
<211> 21  
<212> PRT  
<213> Artificial Sequence

55 <220>  
<223> Peptide from bacteriophage strain CLB\_P2 aligning with position 438-458 of the major capsid protein of

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bacteriophage JS98 (SEQ ID NO: 34)

<400> 42

5 Gly Ser Asn Glu Met Asp Ala Gly Ile Tyr Tyr Ala Pro Tyr Val Ala  
1 5 10 15  
Leu Thr Pro Leu Arg  
20

10 <210> 43  
<211> 49  
<212> PRT  
<213> Artificial Sequence

15 <220>  
<223> Peptide from bacteriophage strain CLB\_P2 aligning with position 464-512 of the major capsid protein of bacteriophage JS98 (SEQ ID NO: 34)

20 <400> 43

Asn Phe Gln Pro Val Met Gly Phe Lys Thr Arg Tyr Gly Ile Gly Ile  
1 5 10 15  
Asn Pro Phe Ala Asp Pro Ala Ala Gln Ala Pro Thr Lys Arg Ile Gln  
20 25 30  
25 Asn Gly Met Pro Asp Ile Val Asn Ser Leu Gly Leu Asn Gly Tyr Phe  
35 40 45  
Arg

30 <210> 44  
<211> 21  
<212> DNA  
<213> Artificial Sequence

35 <220>  
<223> forward primer to amplify a specific gene (pMT1) from LF82

<400> 44  
ccattcatgc agcagctctt t 21

40 <210> 45  
<211> 21  
<212> DNA  
<213> Artificial Sequence

45 <220>  
<223> reverse primer to amplify a specific gene (pMT1) from LF82

50 <400> 45  
atcgacaac attagcgtg t 21

Claims

55 1. A pharmaceutical composition comprising a combination of two or more of  
— the bacteriophage strain P1 deposited with the French National Collection of Microorganisms at the Institut Pasteur under Accession Number CNCM I-4694 or a variant thereof, wherein the variant has the same lytic

activity and the same phenotypic characteristics as said bacteriophage strain P1;  
 — the bacteriophage strain P2 deposited with the French National Collection of Microorganisms at the Institut Pasteur under Accession Number CNCM I-4695 or a variant thereof, wherein the variant has the same lytic activity and the same phenotypic characteristics as said bacteriophage strain P2;  
 5 — the bacteriophage strain P3 deposited with the French National Collection of Microorganisms at the Institut Pasteur under Accession Number CNCM I-4696 or a variant thereof, wherein the variant has the same lytic activity and the same phenotypic characteristics as said bacteriophage strain P3;  
 — the bacteriophage strain P4 deposited with the French National Collection of Microorganisms at the Institut Pasteur under Accession Number CNCM I-4697 or a variant thereof, wherein the variant has the same lytic activity and the same phenotypic characteristics as said bacteriophage strain P4;  
 10 — the bacteriophage strain P5 deposited with the French National Collection of Microorganisms at the Institut Pasteur under Accession Number CNCM I-4698 or a variant thereof, wherein the variant has the same lytic activity and the same phenotypic characteristics as said bacteriophage strain P5;  
 — the bacteriophage strain P6 deposited with the French National Collection of Microorganisms at the Institut Pasteur under Accession Number CNCM I-4699 or a variant thereof, wherein the variant has the same lytic activity and the same phenotypic characteristics as said bacteriophage strain P6;  
 15 — the bacteriophage strain P8 deposited with the French National Collection of Microorganisms at the Institut Pasteur under Accession Number CNCM I-4700 or a variant thereof, wherein the variant has the same lytic activity and the same phenotypic characteristics as said bacteriophage strain P8;  
 20 — the bacteriophage strain CLB\_P2 deposited with the French National Collection of Microorganisms at the Institut Pasteur under Accession Number CNCM I-4675 or a variant thereof, wherein the variant has the same lytic activity and the same phenotypic characteristics as said bacteriophage strain CLB\_P2.

2. The pharmaceutical composition according to claim 1 comprising:

25 — the bacteriophage strain P2 deposited with the French National Collection of Microorganisms at the Institut Pasteur under Accession Number CNCM I-4695 or a variant thereof, wherein the variant has the same lytic activity and the same phenotypic characteristics as said bacteriophage strain P2;  
 — the bacteriophage strain P8 deposited with the French National Collection of Microorganisms at the Institut Pasteur under Accession Number CNCM I-4700 or a variant thereof, wherein the variant has the same lytic activity and the same phenotypic characteristics as said bacteriophage strain P8; and  
 30 — the bacteriophage strain CLB\_P2 deposited with the French National Collection of Microorganisms at the Institut Pasteur under Accession Number CNCM I-4675 or a variant thereof, wherein the variant has the same lytic activity and the same phenotypic characteristics as said bacteriophage strain CLB\_P2.

3. The pharmaceutical composition according to any one of claims 1 or 2, wherein said pharmaceutical composition is for use in the treatment of inflammatory bowel disease.

4. The pharmaceutical composition according to any one of claims 1 or 2 or the pharmaceutical composition for use according to claim 3, said pharmaceutical composition comprising a pharmaceutically acceptable carrier.

5. The pharmaceutical composition for use according to any one of claims 3 or 4, wherein the inflammatory bowel disease is Crohn's disease.

6. The pharmaceutical composition for use according to any one of claims 3 to 5, wherein the pharmaceutical composition is for oral administration.

**Patentansprüche**

1. Pharmazeutische Zusammensetzung, umfassend eine Kombination von zwei oder mehreren von

— dem Bakteriophagenstamm P1, hinterlegt bei der French National Collection of Microorganisms am Institut Pasteur unter der Hinterlegungsnummer CNCM I-4694, oder einer Variante davon, wobei die Variante die gleiche lytische Aktivität und die gleichen phänotypischen Charakteristika aufweist wie der Bakteriophagenstamm P1;  
 55 — dem Bakteriophagenstamm P2, hinterlegt bei der French National Collection of Microorganisms am Institut Pasteur unter der Hinterlegungsnummer CNCM I-4695, oder einer Variante davon, wobei die Variante die

gleiche lytische Aktivität und die gleichen phänotypischen Charakteristika aufweist wie der Bakteriophagenstamm P2;

— dem Bakteriophagenstamm P3, hinterlegt bei der French National Collection of Microorganisms am Institut Pasteur unter der Hinterlegungsnummer CNCM I-4696, oder einer Variante davon, wobei die Variante die gleiche lytische Aktivität und die gleichen phänotypischen Charakteristika aufweist wie der Bakteriophagenstamm P3;

— dem Bakteriophagenstamm P4, hinterlegt bei der French National Collection of Microorganisms am Institut Pasteur unter der Hinterlegungsnummer CNCM I-4697, oder einer Variante davon, wobei die Variante die gleiche lytische Aktivität und die gleichen phänotypischen Charakteristika aufweist wie der Bakteriophagenstamm P4;

— dem Bakteriophagenstamm P5, hinterlegt bei der French National Collection of Microorganisms am Institut Pasteur unter der Hinterlegungsnummer CNCM I-4698, oder einer Variante davon, wobei die Variante die gleiche lytische Aktivität und die gleichen phänotypischen Charakteristika aufweist wie der Bakteriophagenstamm P5;

— dem Bakteriophagenstamm P6, hinterlegt bei der French National Collection of Microorganisms am Institut Pasteur unter der Hinterlegungsnummer CNCM I-4699, oder einer Variante davon, wobei die Variante die gleiche lytische Aktivität und die gleichen phänotypischen Charakteristika aufweist wie der Bakteriophagenstamm P6;

— dem Bakteriophagenstamm P8, hinterlegt bei der French National Collection of Microorganisms am Institut Pasteur unter der Hinterlegungsnummer CNCM I-4700, oder einer Variante davon, wobei die Variante die gleiche lytische Aktivität und die gleichen phänotypischen Charakteristika aufweist wie der Bakteriophagenstamm P8;

— dem Bakteriophagenstamm CLB\_P2, hinterlegt bei der French National Collection of Microorganisms am Institut Pasteur unter der Hinterlegungsnummer CNCM I-4675, oder einer Variante davon, wobei die Variante die gleiche lytische Aktivität und die gleichen phänotypischen Charakteristika aufweist wie der Bakteriophagenstamm CLB\_P2.

2. Pharmazeutische Zusammensetzung gemäß Anspruch 1, umfassend:

— den Bakteriophagenstamm P2, hinterlegt bei der French National Collection of Microorganisms am Institut Pasteur unter der Hinterlegungsnummer CNCM I-4695, oder eine Variante davon, wobei die Variante die gleiche lytische Aktivität und die gleichen phänotypischen Charakteristika aufweist wie der Bakteriophagenstamm P2;

— den Bakteriophagenstamm P8, hinterlegt bei der French National Collection of Microorganisms am Institut Pasteur unter der Hinterlegungsnummer CNCM I-4700, oder eine Variante davon, wobei die Variante die gleiche lytische Aktivität und die gleichen phänotypischen Charakteristika aufweist wie der Bakteriophagenstamm P8; und

— den Bakteriophagenstamm CLB\_P2, hinterlegt bei der French National Collection of Microorganisms am Institut Pasteur unter der Hinterlegungsnummer CNCM I-4675, oder eine Variante davon, wobei die Variante die gleiche lytische Aktivität und die gleichen phänotypischen Charakteristika aufweist wie der Bakteriophagenstamm CLB\_2.

3. Pharmazeutische Zusammensetzung gemäß irgendeinem der Ansprüche 1 oder 2, wobei die pharmazeutische Zusammensetzung zur Verwendung bei der Behandlung von entzündlicher Darmerkrankung ist.

4. Pharmazeutische Zusammensetzung gemäß irgendeinem der Ansprüche 1 oder 2 oder pharmazeutische Zusammensetzung zur Verwendung gemäß Anspruch 3, wobei die pharmazeutische Zusammensetzung einen pharmazeutisch akzeptablen Träger umfasst.

5. Pharmazeutische Zusammensetzung zur Verwendung gemäß irgendeinem der Ansprüche 3 oder 4, wobei die entzündliche Darmerkrankung Morbus Crohn ist.

6. Pharmazeutische Zusammensetzung zur Verwendung gemäß irgendeinem der Ansprüche 3 bis 5, wobei die pharmazeutische Zusammensetzung für die orale Verabreichung ist.

**Revendications**

1. Composition pharmaceutique comprenant une combinaison de deux ou plus parmi

5 - la souche de bactériophage P1 déposée à la Collection Nationale Française de Microorganismes de l'Institut Pasteur sous le numéro d'accèsion CNCM I-4694 ou une variante de celle-ci, dans laquelle la variante présente la même activité lytique et les mêmes caractéristiques phénotypiques que ladite souche de bactériophage P1 ;  
- la souche de bactériophage P2 déposée à la Collection Nationale Française de Microorganismes de l'Institut Pasteur sous le numéro d'accèsion CNCM I-4695 ou une variante de celle-ci, dans laquelle la variante présente la même activité lytique et les mêmes caractéristiques phénotypiques que ladite souche de bactériophage P2 ;  
10 - la souche de bactériophage P3 déposée à la Collection Nationale Française de Microorganismes de l'Institut Pasteur sous le numéro d'accèsion CNCM I-4696 ou une variante de celle-ci, dans laquelle la variante présente la même activité lytique et les mêmes caractéristiques phénotypiques que ladite souche de bactériophage P3 ;  
- la souche de bactériophage P4 déposée à la Collection Nationale Française de Microorganismes de l'Institut Pasteur sous le numéro d'accèsion CNCM I-4697 ou une variante de celle-ci, dans laquelle la variante présente la même activité lytique et les mêmes caractéristiques phénotypiques que ladite souche de bactériophage P4 ;  
15 - la souche de bactériophage P5 déposée à la Collection Nationale Française de Microorganismes de l'Institut Pasteur sous le numéro d'accèsion CNCM I-4698 ou une variante de celle-ci, dans laquelle la variante présente la même activité lytique et les mêmes caractéristiques phénotypiques que ladite souche de bactériophage P5 ;  
- la souche de bactériophage P6 déposée à la Collection Nationale Française de Microorganismes de l'Institut Pasteur sous le numéro d'accèsion CNCM I-4699 ou une variante de celle-ci, dans laquelle la variante présente la même activité lytique et les mêmes caractéristiques phénotypiques que ladite souche de bactériophage P6 ;  
20 - la souche de bactériophage P8 déposée à la Collection Nationale Française de Microorganismes de l'Institut Pasteur sous le numéro d'accèsion CNCM I-4700 ou une variante de celle-ci, dans laquelle la variante présente la même activité lytique et les mêmes caractéristiques phénotypiques que ladite souche de bactériophage P8 ;  
- la souche de bactériophage CLB\_P2 déposée à la Collection Nationale Française de Microorganismes de l'Institut Pasteur sous le numéro d'accèsion CNCM I-4675 ou une variante de celle-ci, dans laquelle la variante présente la même activité lytique et les mêmes caractéristiques phénotypiques que ladite souche de bactériophage CLB\_P2.  
25

2. Composition pharmaceutique selon la revendication 1, comprenant :

30 - la souche de bactériophage P2 déposée à la Collection Nationale Française de Microorganismes de l'Institut Pasteur sous le numéro d'accèsion CNCM I-4695 ou une variante de celle-ci, dans laquelle la variante présente la même activité lytique et les mêmes caractéristiques phénotypiques que ladite souche de bactériophage P2 ;  
- la souche de bactériophage P8 déposée à la Collection Nationale Française de Microorganismes de l'Institut Pasteur sous le numéro d'accèsion CNCM I-4700 ou une variante de celle-ci, dans laquelle la variante présente la même activité lytique et les mêmes caractéristiques phénotypiques que ladite souche de bactériophage P8 ; et  
35 - la souche de bactériophage CLB\_P2 déposée à la Collection Nationale Française de Microorganismes de l'Institut Pasteur sous le numéro d'accèsion CNCM I-4675 ou une variante de celle-ci, dans laquelle la variante présente la même activité lytique et les mêmes caractéristiques phénotypiques que ladite souche de bactériophage CLB\_P2.

40 3. Composition pharmaceutique selon l'une quelconque des revendications 1 ou 2, dans laquelle ladite composition pharmaceutique est destinée à une utilisation dans le traitement d'une maladie inflammatoire de l'intestin.

45 4. Composition pharmaceutique selon l'une quelconque des revendications 1 ou 2 ou composition pharmaceutique pour une utilisation selon la revendication 3, ladite composition pharmaceutique comprenant un véhicule pharmaceutiquement acceptable.

5. Composition pharmaceutique pour une utilisation selon l'une quelconque des revendications 3 ou 4, dans laquelle la maladie inflammatoire de l'intestin est la maladie de Crohn.

50 6. Composition pharmaceutique pour une utilisation selon l'une quelconque des revendications 3 à 5, dans laquelle la composition pharmaceutique est destinée à une administration orale.

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## REFERENCES CITED IN THE DESCRIPTION

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### Patent documents cited in the description

- EP 13305568 [0131]

### Non-patent literature cited in the description

- **MAURA et al.** *Environmental Microbiol*, August 2012, vol. 14 (8), 1844-1854 [0007]
- **BELGIN et al.** *Inflammatory Bowel Diseases*, January 2013, vol. 19 (1), 141-150 [0008]
- **DARFEUILLE-MICHAUD et al.** *Gastroenterology*, 2004, vol. 127, 412-421 [0014] [0015]
- **FALKOW et al.** *Rev. Infect. Dis.*, 1987, vol. 9 (5), 450-455 [0051]
- **A. VILLEGAS et al.** *Virology Journal*, 2009, vol. 6, 41 [0053]
- **S. ZUBER et al.** *Journal of Bacteriology*, 2007, vol. 189 (22), 8206 [0053]
- **MAURA et al.** *Environmental Microbiology*, 2012, vol. 14 (8), 1844-1854 [0066]