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(54) **AAV-EXPRESSED M013 PROTEIN AS AN ANTI-INFLAMMATORY THERAPEUTIC FOR USE IN A METHOD OF TREATING INFLAMMATORY OCULAR DISEASE**

(57) Disclosed are methods and compositions for preventing, treating, and/or ameliorating one or more symptoms of inflammation in a mammal. In particular, viral vectors and medicaments containing them are disclosed, which are useful in the prophylaxis, therapy, or amelioration of symptoms of one or more inflammatory-mediated mammalian diseases, such as age-related macular degeneration (AMD), arthritis, Bechet's disease, Best macular dystrophy, corneal inflammation, diabetic retinopathy, drusen formation, dry AMD, dry eye, geographic atrophy, glaucomaocular neovascularization, Lupus erythematosus, macular degeneration, Mallat

Leventinese and Doyme honeycomb retinal dystrophy, nephritis, ocular hypertension, ocular inflammation, recurrent uveitis, Sorsby fundus dystrophy, vasculitis, vitreoretinopathy, wet AMD, or related disorders. In exemplary methods, administration of a pharmaceutical composition comprising a recombinant viral vector that delivers a secretable and cell-penetrating M013 protein or peptide to a subject in need thereof facilitated treatment of particular human disorders such as AMD, ocular neovascularization, uveitis, and related inflammatory ocular disease.

EP 4 050 020 A1

Description**BACKGROUND OF THE INVENTION****CROSS-REFERENCE TO RELATED APPLICATIONS**

[0001] The present application claims priority to United States Provisional Application No. 61/951,294; filed March 11, 2014 (pending; Atty. Dkt. No. 36689.343), the entire contents of which is incorporated herein in its entirety by express reference thereto.

STATEMENT REGARDING FEDERALLY SPONSORED RESEARCH OR DEVELOPMENT

[0002] This invention was made with government support under Grant Nos. R01-EY02025, RO1-EY021721, and R01-AI100987, each awarded by the National Institutes of Health. The government has certain rights in the invention.

NAMES OF THE PARTIES TO A JOINT RESEARCH AGREEMENT

[0003] Not Applicable.

FIELD OF THE INVENTION

[0004] The present invention relates generally to the fields of molecular biology and virology, and in particular, to the development of gene delivery vehicles. In this application, the use of a modified, cell-permeable version of the viral-specific M013 protein as an anti-inflammatory reagent is described. Methods are also provided for treating tissue-specific inflammation with the disclosed genetic constructs, using inflammatory eye disease as an exemplar disease target and AAV as the delivery vector to express the modified M013 protein in cells within the inflamed eye tissue.

DESCRIPTION OF RELATED ART

[0005] Dry age-related macular degeneration (AMD) has been associated with an increase in oxidative stress and inflammatory processes within the retina. Oxidized molecules, like 4-hydroxynonenal (4-HNE) and activated complement components, have been detected in the eyes of dry AMD patients, further evidencing the role of these processes in this disease.

[0006] What is lacking in the prior art are viral vectors that can be used to deliver therapeutic constructs to selected cell populations *in vitro* or *in vivo*. The development of such vectors, and compositions comprising them would provide a major advancement in human gene therapy, and particularly in the treatment of inflammatory-mediated diseases such as arthritis, diabetic retinopathy, dry AMD, vasculitis, recurrent uveitis, Bechet's disease, Lupus erythematosus, nephritis, and the like.

BRIEF SUMMARY OF THE INVENTION

[0007] The present invention overcomes these and other limitations inherent in the prior art by providing AAV vector constructs that are capable of, and optimized for, delivering anti-inflammatory peptides, such as those derived from the viral M013 protein, to selected mammalian cells and/or tissues. The invention also provides a robust methodology for the tissue-specific treatment and/or amelioration of one or more symptoms of oxidative stress and/or inflammation in a subject mammal. In particular, the invention provides novel and non-obvious AAV vector-based constructs that deliver a secretable, cell-penetrating, and virally-derived M013 gene product to selected mammalian cells. These genetic constructs encode M013 polypeptides that inhibit one or more proinflammatory cytokines [including, for example, IL-1 β and IL-18] in a variety of abnormal conditions in mammals, including, for example, ocular diseases such as age-related macular degeneration (AMD), arthritis, diabetic retinopathy, Bechet's disease, vasculitis, nephritis, lupus erythematosus, and recurrent uveitis.

[0008] The invention also provides new and useful methods for treating a variety of abnormal conditions, diseases, dysfunctions, and/or disorders that may result from, or be exacerbated by, one or more cellular oxidative stress pathways, and/or one or more steps of the inflammatory response process in mammals.

[0009] The present invention overcomes limitations in the prior art by providing new AAV-based gene therapy vectors that deliver a secretable, cell-penetrating peptide, such as viral M013 peptide or polypeptide, to one or more target host cells or tissues. When expressed in suitably-transformed mammalian host cells, the expressed gene product inhibits a variety of endogenous proinflammatory cytokines (including, without limitation, interleukins such as IL-1 β and IL-18). In

an exemplary embodiment, gene therapy constructs of the invention comprising M013-based peptides were useful in inhibiting the inflammatory response process in an established mammalian model of dry AMD.

[0010] The constructs described herein provide new and useful treatment strategies for a variety of mammalian diseases, conditions or disorders that are associated with, caused by, or exacerbated due to, one or more cellular oxidative stress responses or one or more steps in the inflammation process.

[0011] In exemplary embodiments, a viral-vectored tatM013 fusion peptide was exploited for treating tissue-based inflammatory disorders, using a method in which the vector construct was directly delivered to one or more selected target cell populations. Importantly, the viral-based expression constructs of the present invention (and in particular, those packaged in AAV-based virions) have been shown to reduce inflammation in a number of disease processes for which there are currently no effective therapies. The disclosed compositions and methods provide new therapeutic modalities for a host of inflammatory-mediated mammalian diseases, including, for example, those of the eye, the gut, and/or one or more internal organs or target tissues to which such viral vectors, such as rAAV-based expression cassettes can be safely, reliably, and suitably delivered.

[0012] Advantageously, the viral vectors and expression constructs of the present invention (as well as infectious virions and viral particles containing them) have an improved efficiency in transducing one or more of cells of a mammal, and in particular, one or more cells of a human eye, and facilitating the expression of M013 protein in cells transformed with the virus. In particular embodiments, the use of capsid-protein-modified viral particles to deliver the expression constructs of interest, has resulted in highly-efficient transduction of selected mammalian cells with populations of the therapeutic viral particles.

[0013] In an overall and general sense, the invention provides isolated and purified polynucleotides that encodes one or more of the disclosed rAAV vectors described herein, as well as pluralities of infectious adeno-associated viral virions that contain such a polynucleotide. Preferably, the vector constructs of the present invention include at least one nucleic acid segment that encodes at least one M013-derived peptide or protein operably linked to a promoter that is capable of expressing the nucleic acid segment in suitable mammalian cells that have been transformed with the vector construct.

[0014] In the practice of the invention, the disclosed rAAV vectors will preferably include at least one polynucleotide that comprises one or more promoters, one or more enhancers, one or more post-transcriptional regulatory sequences, one or more polyadenylation signals, or any combination thereof, operably linked to the nucleic acid segment that encodes the selected M013-derived therapeutic protein or peptide to permit expression of the construct in selected mammalian host cells.

[0015] In certain embodiments, such promoter(s) may include a homologous promoter, a heterologous promoter, an endogenous promoter, an exogenous promoter, a synthetic promoter, a hybrid promoter, a viral promoter, a cell-specific promoter, a tissue-specific promoter, or any combination thereof that is/are capable of expressing the M013-encoding nucleic acid segment in selected cells and/or tissues of a mammalian subject.

[0016] rAAV vectors prepared in accordance with the present invention may also optionally further include one or more additional nucleic acid sequences that are each operably linked to the therapeutic agent-encoding nucleic acid segment. Exemplary sequences include, but are not limited to, one or more selected from the group consisting of a viral-derived enhancer sequence, a mammalian-derived enhancer sequence, a synthetic enhancer, a cell- or tissue-specific enhancer sequence, one or more inverted terminal repeats, one or more multiple cloning sites, restriction cleavage sites, polyadenylation signals, post-transcriptional regulatory sequences, secretion signal sequences, or any combination thereof.

[0017] Exemplary post-transcriptional regulatory sequences include, but are not limited to, woodchuck hepatitis virus post-transcription regulatory elements, polyadenylation signal sequences, intron/exon junctions/splicing signals, synthetic elements, or any combination thereof.

[0018] The viral vectors of the present invention may also optionally further include an additional nucleic acid sequence region that comprises, consists essentially of, or consists of, one or more polylinkers, restriction sites, and/or multiple cloning region(s) to facilitate insertion (cloning) of one or more selected genetic elements, genes of interest, or therapeutic or diagnostic constructs into the vector at a pre-determined site.

[0019] In further aspects of the present invention, the exogenous polynucleotide(s) that may be delivered into suitable host cells by the disclosed viral vectors are preferably of mammalian origin, with polynucleotides encoding one or more polypeptides or peptides of human, non-human primate, porcine, bovine, ovine, feline, canine, equine, equine, caprine, or lupine origin also being preferred for certain uses of the disclosed vector compositions.

[0020] In certain embodiments, the disclosed viral vectors will further optionally include a second nucleic acid segment that expresses or encodes one or more diagnostic or therapeutic molecules, including, without limitation, one or more polypeptides, peptides, ribozymes, peptide nucleic acids, siRNA's, RNAi's, antisense oligonucleotides, antisense polynucleotides, antibodies, antigen binding fragments, or any combination thereof. Such additional therapeutic agents may include, without limitation, one or more agonists, antagonists, anti-apoptosis factors, inhibitors, receptors, cytokines, cytotoxins, erythropoietic agents, glycoproteins, growth factors, growth factor receptors, hormones, hormone receptors, interferons, interleukin inhibitors, interleukin receptors, nerve growth factors, neuroactive peptides, neuroactive peptide receptors, proteases, protease inhibitors, protein decarboxylases, protein kinases, protein kinase inhibitors, enzymes,

receptor binding proteins, transport proteins or one or more inhibitors thereof, serotonin receptors, or one or more uptake inhibitors thereof, serpins, serpin receptors, tumor suppressors, diagnostic molecules, molecular markers, chemotherapeutic agents, cytotoxins, or any combination thereof.

[0021] When combinational gene therapies are desired, two or more different molecules may be produced from a single rAAV expression system, or alternatively, a selected host cell may be transfected with two or more unique rAAV expression systems, each of which may comprise one or more distinct polynucleotides that encode one or more diagnostic and/or therapeutic agents.

[0022] The viral vectors of the present invention are preferably rAAV vectors, including, without limitation, those derived from, or comprised within a virion having a serotype that is selected from the group consisting of AAV serotype 1 (AAV1), AAV serotype 2 (AAV2), AAV serotype 3 (AAV3), AAV serotype 4 (AAV4), AAV serotype 5 (AAV5), AAV serotype 6 (AAV6), AAV serotype 7 (AAV7), AAV serotype 8 (AAV8), AAV serotype 9 (AAV9), AAV serotype 10 (AAV10), AAV serotype 11 (AAV11), AAV serotype 12 (AAV12), or any other serotype as known to one of ordinary skill in the viral arts.

[0023] In related embodiments, the invention further provides populations and pluralities of viral vectors, virions, infectious viral particles, or host cells that include one or more nucleic acid segments that encode a CPP, such as an M013-specific protein or peptide, operably linked to a selected promoter capable of expressing the encoded CPP in at least one mammalian cell of interest.

[0024] The invention further provides composition and formulations that include one or more of the proteins, nucleic acid segments, viral vectors, host cells, or viral particles of the present invention together with one or more pharmaceutically-acceptable buffers, diluents, or excipients. Such compositions may be included in one or more diagnostic or therapeutic kits, for diagnosing, preventing, treating or ameliorating one or more symptoms of a mammalian disease, injury, disorder, trauma or dysfunction, and in particular, for treatment of oxidative stress and/or inflammation in one or more human cells.

[0025] The invention further includes a method for providing a mammal in need thereof with a diagnostically- or therapeutically-effective amount of a selected M013-derived therapeutic agent, the method comprising providing to a cell, tissue or organ of a mammal in need thereof, an amount of one or more of the disclosed CPP-encoding rAAV vectors (such as those expressing M013 peptide; and for a time effective to provide the mammal with a diagnostically- or a therapeutically-effective amount of the encoded peptide.

[0026] In another embodiment, the invention also provides a method for diagnosing, preventing, treating, or ameliorating at least one or more symptoms of a disease, a disorder, a dysfunction, an injury, an abnormal condition, or trauma in a mammal. In an overall and general sense, the method includes at least the step of administering to a mammal in need thereof one or more of the disclosed M013-encoding rAAV genetic constructs, in an amount and for a time sufficient to diagnose, prevent, treat or ameliorate the one or more symptoms of the disease, disorder, dysfunction, injury, abnormal condition, or trauma in the mammal.

[0027] The invention also provides a method of transducing a population of mammalian cells. In an overall and general sense, the method includes at least the step of introducing into one or more cells of the population, a composition that comprises an effective amount of one or more of the M013-expressing rAAV vector constructs as disclosed herein.

[0028] In a further embodiment, the invention also provides isolated nucleic acid segments that encode one or more M013-specific proteins or peptides, as well as recombinant vectors, virus particles, infectious virions, and isolated host cells that comprise one or more of the modified M013-specific protein-encoding nucleic acids as described herein.

[0029] Additionally, the present invention provides compositions, as well as therapeutic and/or diagnostic kits that include one or more of the disclosed viral vector compositions, formulated with one or more additional ingredients, or prepared with one or more instructions for their use.

[0030] The invention also demonstrates methods for making, as well as methods of using the disclosed viral vectors in a variety of ways, including, for example, *ex situ*, *in vitro* and *in vivo* applications, methodologies, diagnostic procedures, and/or gene therapy methods. The disclosed vectors are particularly suited for viral vector-based human gene therapy regimens, and for delivering one or more genetic constructs, including, for example, an M013 peptide, to one or more populations of selected mammalian cells, either *in vivo* or *in vitro*.

[0031] In one aspect, the invention provides compositions comprising recombinant adeno-associated viral (AAV) vectors, virions, viral particles, and pharmaceutical formulations thereof, useful in methods for delivering genetic material encoding one or more beneficial or therapeutic product(s) to mammalian cells and tissues. In particular, the compositions and methods of the invention provide a significant advancement in the art through their use in the treatment, prevention, and/or amelioration of symptoms of one or more mammalian diseases associated with one or more oxidative stress or inflammatory responses.

[0032] In particular, the invention provides rAAV-based expression constructs that encode one or more mammalian therapeutic agent(s) (including, but not limited to, for example, M013-derived protein(s), polypeptide(s), peptide(s), enzyme(s), antibodies, antigen binding fragments, as well as variants, and/or active fragments thereof, for use in the treatment, prophylaxis, and/or amelioration of one or more symptoms of a mammalian disease, dysfunction, injury, and/or disorder.

[0033] In other embodiments, the invention also provides viral vectors that are comprised within an infectious viral particle or a virion, as well as pluralities of such virions or infectious particles. Such vectors and virions may be comprised within one or more diluents, buffers, physiological solutions, or pharmaceutical vehicles, or formulated for administration to a mammal in one or more diagnostic, therapeutic, and/or prophylactic regimens. The vectors, virus particles, virions, and pluralities thereof of the present invention may also be provided in excipient formulations that are acceptable for veterinary administration to selected livestock, exotics, domesticated animals, and companion animals (including pets and such like), as well as to non-human primates, zoological or otherwise captive specimens, and such like.

[0034] Compositions comprising one or more of the disclosed viral vectors, expression systems, infectious viral particles, or host cells also form part of the present invention, and particularly those compositions that further comprise at least a first pharmaceutically-acceptable excipient for use in therapy, and for use in the manufacture of medicaments for the treatment of one or more mammalian diseases, disorders, dysfunctions, or trauma, such as oxidative stress and/or inflammation, and/or related cellular processes.

[0035] Such pharmaceutical compositions may optionally further comprise one or more diluents, buffers, liposomes, a lipid, a lipid complex. Alternatively, the vectors of the present invention may be comprised within a plurality of microspheres, nanoparticles, liposomes, or a combination thereof. Pharmaceutical formulations suitable for administration to one or more cells, tissues, and/or organs of a human are particularly preferred.

[0036] Kits comprising one or more of the disclosed viral vectors (as well as one or more virions, viral particles, transformed host cells or pharmaceutical compositions comprising such vectors); and instructions for using such kits in one or more therapeutic, diagnostic, and/or prophylactic clinical embodiments are also provided by the present invention. Such kits may further comprise one or more reagents, restriction enzymes, peptides, therapeutics, pharmaceutical compounds, or means for delivery of the composition(s) to host cells, or to an animal (e.g., syringes, injectables, and the like). Exemplary kits include those for treating, preventing, or ameliorating the symptoms of a disease, deficiency, dysfunction, and/or injury, or may include components for the large-scale production of the viral vectors themselves, such as for commercial sale, or for use by others, including e.g., virologists, medical professionals, and the like.

[0037] Such kits may also further include one or more rAAV packaging components, such as a) a host cell that expresses at least one rep gene and/or at least one cap gene, b) a helper virus gene product; c) a helper virus, such as adenovirus or herpes virus, or d) any combination thereof to facilitate packaging of the expression vectors into suitable virions or infectious viral particles.

[0038] Another important aspect of the present invention concerns methods of use of the disclosed rAAV vectors, virions, expression systems, compositions, and host cells described herein in the preparation of medicaments for treating or ameliorating at least one symptom of a disease, a dysfunction, a disorder, an abnormal condition, a deficiency, injury, or trauma in an animal, and in particular, in one or more cells or tissues of a vertebrate mammal. Such methods generally involve direct administration to the mammal in need thereof, one or more of the disclosed CPP (and preferably M013 peptide)-expressing rAAV vectors, virions, viral particles, host cells, compositions, or pluralities thereof, in an amount and for a time sufficient to treat or ameliorate one or more symptoms of such a disease, dysfunction, disorder, abnormal condition, deficiency, injury, or trauma in the affected animal. The compositions of the invention may also be used for the diagnosis, and/or prophylaxis of one or more animals suspected of having such a condition, as well as for administration to one or more animals determined to be at risk for developing one or more such conditions.

BRIEF DESCRIPTION OF THE DRAWINGS

[0039] For promoting an understanding of the principles of the invention, reference will now be made to the embodiments, or examples, illustrated in the drawings and specific language will be used to describe the same. It will, nevertheless be understood that no limitation of the scope of the invention is thereby intended. Any alterations and further modifications in the described embodiments, and any further applications of the principles of the invention as described herein are contemplated as would normally occur to one of ordinary skill in the art to which the invention relates.

[0040] The following drawings form part of the present specification and are included to demonstrate certain aspects of the present invention. The application contains at least one drawing that is executed in color. Copies of this patent or patent application publication with color drawing(s) will be provided by the Patent and Trademark Office upon request and payment of the necessary fee. The invention may be better understood by reference to the following description taken in conjunction with the accompanying drawings, in which like reference numerals identify like elements, and in which:

FIG. 1A, FIG. 1B, FIG. 1C, and FIG. 1D show a myxoma virus M013 fusion protein construct can be used to inhibit interleukin (IL)-1 β secretion. **FIG. 1A:** The native M013 protein from myxoma virus has been demonstrated to inhibit both nuclear factor (NF)- κ B and NLRP3 inflammasome signaling pathways by interacting with key effector components. **FIG. 1B:** Map of lentiviral vector delivering the TatM013 fusion gene. The M013 and TatM013 fusion genes were cloned in-frame with the 2A peptide and puromycin resistance (puroR) sequence. **FIG. 1C:** Monocyte-derived THP-1 cells were transduced with a lentiviral vector delivering the M013-puroR fusion gene, the TatM013-puroR

fusion, or the puroR gene alone as a control. Stably transduced cells were incubated with IFN- γ (4 hr) and lipopolysaccharide (LPS) (18 hr). Secreted IL-1 β was quantified by ELISA. M013 protein was measured in cell extracts by Western blotting, and compared with tubulin control (inset) ($n = 3$, average $\pm$ SEM). FIG. 1D: ARPE-19 cells were transduced with either the TatM013-puroR or puroR lentiviral vector. Stably transduced cells were incubated with 30 μ M 4-hydroxynonenal (4-HNE, 18 hr). Secreted IL-1 β was quantified by ELISA ($n = 3$, average $\pm$ SEM). ASC, apoptosis-associated speck-like protein containing a caspase recruitment domain; AmpR, ampicillin resistance gene; cPPT, central polypurine tract; EF1, elongation factor-1; HIV LTR, human immunodeficiency virus long terminal repeat; I κ -B, inhibitor of NF- κ B; IKKK, I κ -B kinase kinase; n.s., not significant; ORI/Ori, origin of replication; RRE, Rev response element; RSV, Rous sarcoma virus; SV40, simian virus 40; TNF- α , tumor necrosis factor- α ; WPRE, woodchuck hepatitis virus posttranscriptional regulatory element. * $p < 0.05$; *** $p < 0.0001$;

FIG. 2A, FIG. 2B and FIG. 2C show the development of a secretable and cell-penetrating TatM013 fusion construct. **FIG. 2A:** Map of pCDH-EF1-sGFP-TatM013-T2A-Puro plasmid, an exemplary construct in accordance with one aspect of the present invention. In this exemplary construct, the TatM013 gene was fused to the secretable green fluorescent protein (sGFP) cDNA, separated by a furin cleavage-site sequence. The sGFP-TatM013 fusion gene was cloned in the lentiviral vector plasmid upstream and fused to the 2A signal peptide and puroR sequence. All these sequences were cloned in-frame, which, upon translation, generated the fused protein sGFP-TatM013 targeted to the secretory pathway. **FIG. 2B:** Expression of sGFP-TatM013 in lentivirus-transduced cells. HEK293T cells stably expressing GFP, a control peptide fused to GFP or sGFP-TatM013, were lysed in NP-40 lysis buffer (Lys). Proteins in the culture supernatant (Sup) were concentrated as described. A total of 30 μ g of protein from each sample was separated by SDS-PAGE, using a 12% gel. Expression of M013 was determined by Western blot using an anti-GFP antibody. **FIG. 2C:** sGFP-TatM013 protein exhibited a discrete perinuclear pattern of distribution in cells. Stably transduced HEK293T cells were generated by transfection with either unmodified GFP or sGFP-TatM013 lentiviral vector particles and further selection with puromycin. The expression of GFP was visualized by fluorescence microscopy. DAPI, 4',6-diamidino-2-phenylindole; Lys, lysate; Sup, supernatant;

FIG. 3A, FIG. 3B, FIG. 3C, and FIG. 3D show *in vitro* confirmation of the activity of the exemplary sGFP-TatM013-expressing AAV vector. **FIG. 3A:** Map of the pTR-smCBA-sGFP-TatM013 plasmid. The sGFP-TatM013 sequence was cloned in an AAV plasmid between AAV2 inverted terminal repeats (TRs). **FIG. 3B:** Cellular distribution of sGFP-TatM013. HEK293T cells were transfected with the pTR-smCBA-sGFP-TatM013 plasmid. GFP expression was validated at 48 hr, by fluorescence microscopy. As a control, HEK293T cells were transfected with pTR-smCBA-GFP plasmid, which delivers an unmodified GFP gene. **FIG. 3C:** Secretion of the sGFP-TatM013 fusion protein into the medium of transfected cells. Conditioned media from the cells imaged in **FIG. 3B** were concentrated. Protein samples from lysed cells (Lys) and conditioned media (Sup) were separated by SDS-PAGE, and GFP was detected with an anti-GFP antibody. **FIG. 3D:** The biological activity of secreted TatM013 was measured in human ARPE-19 cells (left). A total of 500 μ L of medium from cells transfected as in **FIG. 3B** (conditioned for 48 hr) was overlaid on ARPE-19 cells for 1 hr. Cells were incubated with or without 4-HNE (40 μ M) for 18 hr by adding this reagent to the wells without removing the conditioned medium, and secreted IL-1 β was quantified by ELISA. The biological activity of secreted TatM013 was measured in murine RAW 264.7 cells (right). Conditioned medium containing either GFP- or sGFP-TatM013-transfected cells was overlaid on RAW 264.7 cells. Cells were then incubated with or without LPS (18 hr), and secreted IL-1 β was quantified by ELISA ($n = 3$, average $\pm$ SEM). * $p \leq 0.05$; ** $p \leq 0.001$; *** $p \leq 0.0001$; and

FIG. 4A, FIG. 4B, FIG. 4C, and FIG. 4D show expression of an exemplary AAV-vectored TatM013 gene product in accordance with one aspect of the present invention inhibited the inflammatory response observed in an exemplary murine endotoxin-induced uveitis (EIU) model. **FIG. 4A:** Fundus of mice injected intravitreally with either AAV-GFP (left eye) or AAV-sGFP-TatM013 (right eye). Bright-field and fluorescence images of a noninjected eye (No Vector) are shown for comparison. **FIG. 4B:** One week later, three mice were injected intravitreally with 175 ng of LPS (both eyes), and their vitreous humor was harvested 24 hr later. The secreted IL-1 β concentration was quantified by ELISA (average $\pm$ SEM). **FIG. 4C:** Representative micrographs from eyes injected with either GFP or sGFP-TatM013 AAV vector after induction of uveitis (with 25 ng of LPS). Paraffin-embedded eyes were sectioned and stained (hematoxylin-eosin, H&E). Bright-field images were taken at original magnifications of $\times 5$ and $\times 10$ to observe infiltrating cells. **FIG. 4D:** Expression of the TatM013 gene product from AAV decreased inflammation in the EIU mouse model. Five mice were injected as described in **FIG. 4A**. Twenty-four hours after the injection of 25 ng of LPS mouse eyes were harvested and processed for paraffin sections. The eye sections were stained with H&E and the infiltrating cells were counted by a masked observer (average $\pm$ SEM). vg, vector genomes. * $p \leq 0.05$.

DESCRIPTION OF ILLUSTRATIVE EMBODIMENTS

[0041] Illustrative embodiments of the invention are described below. In the interest of clarity, not all features of an actual implementation are described in this specification. It will of course be appreciated that in the development of any

such actual embodiment, numerous implementation-specific decisions must be made to achieve the developers' specific goals, such as compliance with system-related and business-related constraints, which will vary from one implementation to another. Moreover, it will be appreciated that such a development effort might be complex and time-consuming, but would be a routine undertaking for those of ordinary skill in the art having the benefit of this disclosure.

[0042] Inflammation of the retina is a contributing factor in ocular diseases such as uveitis, diabetic retinopathy, and age-related macular degeneration (AMD). The M013 immunomodulatory protein from myxoma virus has been shown to interfere with the proinflammatory signaling pathways involving both the NLRP3 inflammasome and NF- κ B. In the present application, the inventors have developed and characterized an adeno-associated viral (AAV) vector that delivers a secretable and cell-penetrating form of the M013 protein (TatM013). The expressed TatM013 protein was secreted and blocked the endotoxin-induced secretion of interleukin (IL)-1 β in monocyte-derived cells and the reactive aldehyde-induced secretion of IL-1 β in retinal pigment epithelium cells. The local anti-inflammatory effects of AAV-delivered TatM013 were evaluated in an endotoxin-induced uveitis (EIU) mouse model after intravitreal injection of mice with an AAV2-based vector carrying either TatM013 fused to a secreted green fluorescent protein (GFP) tag (sGFP-TatM013) or to GFP itself. Expression of the sGFP-TatM013 transgene was demonstrated by fluorescence funduscopy in living mice. In EIU, the number of infiltrating cells and the concentration of IL-1 β in the vitreous body were significantly lower in the eyes injected with AAV-sGFP-TatM013 compared with the eyes injected with control AAV-GFP. These results suggested that a virus-derived inhibitor of the innate immune response, when delivered *via* AAV, provided an effective and generalized therapy for treatment of various inflammatory diseases of the eye.

[0043] Dry age-related macular degeneration (AMD) has been associated with an increase in oxidative stress and inflammatory processes within the retina. Oxidized molecules like 4-hydroxynonenal (4-HNE) and activated complement components have been detected in the eyes of dry AMD patients, supporting the role of these processes in the diseases.

[0044] Certain viral proteins encoded by large DNA viruses possess potent anti-inflammatory properties. In this application, the development of a modified cell-permeable version of one specific viral protein called M013 as an anti-inflammatory reagent to treat tissue-specific inflammation is disclosed. Using inflammatory eye disease as an exemplar disease target, AAV delivery vectors were developed that expressed a modified M013 protein in cells within the inflamed eye tissue.

[0045] Many drugs and reagents have been developed to treat pro-inflammatory diseases and some of these therapeutic compounds are derived from natural biological organisms, such as secreted anti-inflammatory proteins expressed by larger DNA viruses (reviewed in Lucas and McFadden, 2004). Unlike standard small molecule drugs and typical biological reagents such as monoclonal antibodies against single inflammatory targets, which tend to be monofunctional and act on one specific cellular target or pathway, viral proteins have evolved over millions of years to recognize multiple targets and pathways.

[0046] In the present invention, the M013 protein of myxoma virus [which is normally expressed as a small cytoplasmic protein that inhibits two distinct cellular inflammatory pathways: the inflammasome and the NF κ B signaling pathways, (Rahman, *et al.*, 2013; Rahman *et al.*, 2009)] has been exploited as a therapeutic for treating inflammatory-mediated conditions in mammals.

[0047] The M013 protein is relatively small (188 amino acids), contains a single pyrin domain, and physically interacts with two separate host proteins that regulate cellular inflammatory pathways: ASC-1 (a key scaffolding component of cellular inflammasomes) and NF κ B1/p105 (a key precursor utilized for NF κ B signaling). Thus, the expressed viral M013 protein shuts off two powerful cellular pro-inflammatory signaling cascades: inflammasomes that control the secretion of inflammatory cytokines like IL-1 β and IL-18, and NF κ B signaling that controls the secretion of another class of pro-inflammatory cytokines such as TNF, interferon, and IL-6.

[0048] Unlike secreted viral proteins that can be expressed as recombinant proteins and then used directly in patients as therapeutic anti-inflammatory protein-based drugs (Arsenault *et al.*, 2012), cytoplasmic proteins such as M013 must be delivered genetically- that is, an M013-encoding nucleic acid segment must be introduced into cells using a suitable vector, and that nucleic acid segment must be expressed inside the target cells to produce the encoded cytoplasmic protein. Thus, for proteins such as M013 (or for derivatives based on the M013 peptide) to be useful as a therapeutic agent in a mammal, suitable vectors (such as rAAV) must be engineered, and delivered to cells by an appropriate gene therapy protocol. For their use in the treatment of humans, such vectors must also be FDA-approved. The present invention provides enabling technology to meet these essential requirements.

[0049] Although a nucleic acid encoding the reporter molecule, GFP, has been utilized in the exemplary experimental results presented in the following Examples, the invention is not limit to constructs employing GFP, or for that matter, constructs employing one or more other "reporter" molecules or other gene products used primarily for visualization of the experimental results using, for example, microscopy. In fact, in many embodiments directed to therapeutic uses of the disclosed constructs, a variety of nucleic acid segments may be employed in place of the GFP reporter molecules exemplified herein. Such nucleic acids segments may include, without limitation, those encoding one or more proteins such as human serum albumin (including, without limitation, PMID:6171778 or PMID:6275391); opticin (including, without limitation, PMID:10636917 or PMID:22669977); a bacterial dihydrofolate reductase (or a destabilization domain thereof)

(such as the *E. coli* DHFR destabilization domain, PMID:20851347 or PMID:23029456, *etc.*), a FK506 binding protein (FKBP) destabilization domain (such as the domain from the FK506 binding protein that allows the stabilization of a fused protein only in the presence of its ligand Shield-1, *e.g.*, PMID:16959577, PMID:18836461); or one or more combinations thereof. Each of these proteins is known to those of ordinary skill in the art, and the amino acid sequences for each are on record in conventional sequence databases and archives (*e.g.*, SwissProt ID's NP_000468, CAB53459, AIW05158, and AAD40379, respectively). The constructs may also be engineered to further include one or more additional diagnostic and/or therapeutic-encoding nucleic acid segments where indicated.

RAAV VECTORS

[0050] Recombinant adeno-associated virus (AAV) vectors have been used successfully for *in vivo* gene transfer in numerous pre-clinical animal models of human disease, and have been used successfully for long-term expression of a wide variety of therapeutic genes (Daya and Berns, 2008; Niemeyer *et al.*, 2009; Owen *et al.*, 2002; Keen-Rhinehart *et al.*, 2005; Scallan *et al.*, 2003; Song *et al.*, 2004). AAV vectors have also generated long-term clinical benefit in humans when targeted to immune-privileged sites, *i.e.*, ocular delivery for Leber's congenital amaurosis (Bainbridge *et al.*, 2008; Maguire *et al.*, 2008; Cideciyan *et al.*, 2008). A major advantage of this vector is its comparatively low immune profile, eliciting only limited inflammatory responses and, in some cases, even directing immune tolerance to transgene products (LoDuca *et al.*, 2009).

EXEMPLARY DEFINITIONS

[0051] In accordance with the present invention, polynucleotides, nucleic acid segments, nucleic acid sequences, and the like, include, but are not limited to, DNAs (including and not limited to genomic or extragenomic DNAs), genes, peptide nucleic acids (PNAs) RNAs (including, but not limited to, rRNAs, mRNAs and tRNAs), nucleosides, and suitable nucleic acid segments either obtained from natural sources, chemically synthesized, modified, or otherwise prepared or synthesized in whole or in part by the hand of man.

[0052] Unless defined otherwise, all technical and scientific terms used herein have the same meaning as commonly understood by one of ordinary skill in the art to which this invention belongs. Although any methods and compositions similar or equivalent to those described herein can be used in the practice or testing of the present invention, the preferred methods, and compositions are described herein. For purposes of the present invention, the following terms are defined below for sake of clarity and ease of reference:

In accordance with long standing patent law convention, the words "a" and "an," when used in this application, including the claims, denote "one or more."

[0053] The terms "about" and "approximately" as used herein, are interchangeable, and should generally be understood to refer to a range of numbers around a given number, as well as to all numbers in a recited range of numbers (*e.g.*, "about 5 to 15" means "about 5 to about 15" unless otherwise stated). Moreover, all numerical ranges herein should be understood to include each whole integer within the range.

[0054] As used herein, the term "buffer" includes one or more compositions, or aqueous solutions thereof, that resist fluctuation in the pH when an acid or an alkali is added to the solution or composition that includes the buffer. This resistance to pH change is due to the buffering properties of such solutions, and may be a function of one or more specific compounds included in the composition. Thus, solutions or other compositions exhibiting buffering activity are referred to as buffers or buffer solutions. Buffers generally do not have an unlimited ability to maintain the pH of a solution or composition; rather, they are typically able to maintain the pH within certain ranges, for example from a pH of about 5 to 7.

[0055] As used herein, the term "carrier" is intended to include any solvent(s), dispersion medium, coating(s), diluent(s), buffer(s), isotonic agent(s), solution(s), suspension(s), colloid(s), inert (s), or such like, or a combination thereof that is pharmaceutically acceptable for administration to the relevant animal or acceptable for a therapeutic or diagnostic purpose, as applicable.

[0056] As used herein, the term "DNA segment" refers to a DNA molecule that has been isolated free of total genomic DNA of a particular species. Therefore, a DNA segment obtained from a biological sample using one of the compositions disclosed herein refers to one or more DNA segments that have been isolated away from, or purified free from, total genomic DNA of the particular species from which they are obtained. Included within the term "DNA segment," are DNA segments and smaller fragments of such segments, as well as recombinant vectors, including, for example, plasmids, cosmids, phage, viruses, and the like.

[0057] The terms "an effective amount" of an active agent refers to the amount of the active agent sufficient to elicit a desired biological response. As will be appreciated by those of ordinary skill in this art, the absolute amount of a particular agent that is effective may vary depending on such factors as the desired biological endpoint, the agent to be delivered, the target tissue, *etc.* Those of ordinary skill in the art will further understand that an "effective amount" may be administered in a single dose, or may be achieved by administration of multiple doses. For example, an effective

amount may be an amount sufficient to achieve one or more of the following: (i) prevent drusen formation; (ii) cause a reduction in drusen number and/or size (drusen regression); (iii) cause a reduction in or prevent lipofuscin deposits; (iv) prevent visual loss or slow the rate of visual loss; (v) prevent or slow the rate of choroidal neovascularization; (vi) cause a reduction in size and/or number of lesions characterized by choroidal neovascularization; (vii) improve visual acuity and/or contrast sensitivity; (viii) prevent or reduce the rate of photoreceptor or RPE cell atrophy or apoptosis; (ix) prevent or slow progression from the wet to the dry form of AMD.

[0058] As used herein, the terms "engineered" and "recombinant" cells are intended to refer to a cell into which an exogenous polynucleotide segment (such as DNA segment that leads to the transcription of a biologically active molecule) has been introduced. Therefore, engineered cells are distinguishable from naturally occurring cells, which do not contain a recombinantly introduced exogenous DNA segment. Engineered cells are, therefore, cells that comprise at least one or more heterologous polynucleotide segments introduced through the hand of man.

[0059] The phrases "isolated" or "biologically pure" refer to material that is substantially, or essentially, free from components that normally accompany the material as it is found in its native state. Thus, isolated polynucleotides in accordance with the invention preferably do not contain materials normally associated with those polynucleotides in their natural, or in situ, environment.

[0060] "Link" or "join" refers to any method known in the art for functionally connecting one or more proteins, peptides, nucleic acids, or polynucleotides, including, without limitation, recombinant fusion, covalent bonding, disulfide bonding, ionic bonding, hydrogen bonding, electrostatic bonding, and the like.

[0061] As used herein, the term "kit" may be used to describe variations of the portable, self-contained enclosure that includes at least one set of reagents, components, or pharmaceutically-formulated compositions to conduct one or more of the assay methods of the present invention. Optionally, such kit may include one or more sets of instructions for use of the enclosed reagents, such as, for example, in a laboratory or clinical application.

[0062] The terms "local administration" or "local delivery," in reference to delivery of a composition, formulation, or device of the invention, refer to delivery that does not rely upon transport of the agent to its intended target tissue *via* the vascular or lymphatic system from a site of administration that is remote from the intended target tissue. The agent is delivered directly to its intended target tissue or in the vicinity thereof, e.g. by injection or implantation. It will be appreciated that a small amount of the delivered agent may enter the vascular system and may ultimately reach the target tissue *via* the vascular system.

[0063] "Macular degeneration related condition" refers to any of a number of disorders and conditions in which the macula degenerates or loses functional activity. The degeneration or loss of functional activity can arise due to, for example, cell death, decreased cell proliferation, loss of normal biological function, or a combination of one or more of the foregoing. Macular degeneration can lead to and/or manifest as alterations in the structural integrity of the cells and/or extracellular matrix of the macula, alteration in normal cellular and/or extracellular matrix architecture, and/or the loss of function of macular cells. The cells can be any cell type normally present in or near the macula including RPE cells, photoreceptors, and capillary endothelial cells. AMD is the major macular degeneration related condition, but a number of others are known including, but not limited to, Best macular dystrophy, Sorsby fundus dystrophy, Mallat's Leventinese and Doyne honeycomb retinal dystrophy, and related disorders of the mammalian eye.

[0064] As used herein, the term "nucleic acid" includes one or more types of: polydeoxyribonucleotides (containing 2-deoxy-D-ribose), polyribonucleotides (containing D-ribose), and any other type of polynucleotide that is an N-glycoside of a purine or pyrimidine base, or modified purine or pyrimidine bases (including abasic sites). The term "nucleic acid," as used herein, also includes polymers of ribonucleosides or deoxyribonucleosides that are covalently bonded, typically by phosphodiester linkages between subunits, but in some cases by phosphorothioates, methylphosphonates, and the like. "Nucleic acids" include single- and double-stranded DNA, as well as single- and double-stranded RNA. Exemplary nucleic acids include, without limitation, gDNA; hnRNA; mRNA; rRNA, tRNA, micro RNA (miRNA), small interfering RNA (siRNA), small nucleolar RNA (snRNA), small nuclear RNA (snRNA), and small temporal RNA (stRNA), and the like, and any combination thereof.

[0065] The term "naturally-occurring" as used herein as applied to an object refers to the fact that an object can be found in nature. For example, a polypeptide or polynucleotide sequence that is present in an organism (including viruses) that can be isolated from a source in nature and which has not been intentionally modified by the hand of man in a laboratory is naturally-occurring. As used herein, laboratory strains of rodents that may have been selectively bred according to classical genetics are considered naturally-occurring animals.

[0066] "Ocular device" refers to a drug delivery device that has appropriate structure, dimensions, shape, and/or configuration and is made of appropriate materials so that it may be placed in or on the surface of the eye without causing unacceptable interference with the physiology or functioning of the eye. Preferably, placement of an ocular device does not significantly disrupt vision. An ocular device is typically a solid or semi-solid article of manufacture and is typically macroscopic, *i.e.*, visible with the naked eye.

[0067] "Ocular neovascularization" (ONV) is used herein to refer to choroidal neovascularization or retinal neovascularization, or both.

[0068] The term "operably linked," as used herein, refers to that the nucleic acid sequences being linked are typically contiguous, or substantially contiguous, and, where necessary to join two protein coding regions, contiguous and in reading frame. However, since enhancers generally function when separated from the promoter by several kilobases and intronic sequences may be of variable lengths, some polynucleotide elements may be operably linked but not contiguous.

[0069] As used herein, the term "patient" (also interchangeably referred to as "host" or "subject"), refers to any host that can serve as a recipient of one or more of the therapeutic or diagnostic formulations as discussed herein. In certain aspects, the patient is a vertebrate animal, which is intended to denote any animal species (and preferably, a mammalian species such as a human being). In certain embodiments, a patient may be any animal host, including but not limited to, human and non-human primates, avians, reptiles, amphibians, bovines, canines, caprines, canines, corvines, equines, felines, hircines, lapines, leporines, lupines, murines, ovines, porcines, racines, vulpines, and the like, including, without limitation, domesticated livestock, herding or migratory animals or birds, exotics or zoological specimens, as well as companion animals, pets, or any animal under the care of a veterinary or animal medical care practitioner.

[0070] The phrase "pharmaceutically acceptable" refers to molecular entities and compositions that preferably do not produce an allergic or similar untoward reaction when administered to a mammal, and in particular, when administered to a human. As used herein, "pharmaceutically acceptable salt" refers to a salt that preferably retains the desired biological activity of the parent compound and does not impart any undesired toxicological effects. Examples of such salts include, without limitation, acid addition salts formed with inorganic acids (e.g., hydrochloric acid, hydrobromic acid, sulfuric acid, phosphoric acid, nitric acid, and the like); and salts formed with organic acids including, without limitation, acetic acid, oxalic acid, tartaric acid, succinic acid, maleic acid, fumaric acid, gluconic acid, citric acid, malic acid, ascorbic acid, benzoic acid, tannic acid, pamoic (embonic) acid, alginate acid, naphthoic acid, polyglutamic acid, naphthalenesulfonic acids, naphthalenedisulfonic acids, polygalacturonic acid; salts with polyvalent metal cations such as zinc, calcium, bismuth, barium, magnesium, aluminum, copper, cobalt, nickel, cadmium, and the like; salts formed with an organic cation formed from N,N' dibenzylethylenediamine or ethylenediamine; and combinations thereof.

[0071] The term "pharmaceutically acceptable salt" as used herein refers to a compound of the present disclosure derived from pharmaceutically acceptable bases, inorganic or organic acids. Examples of suitable acids include, but are not limited to, hydrochloric, hydrobromic, sulfuric, nitric, perchloric, fumaric, maleic, phosphoric, glycolic, lactic, salicylic, succinic, toluene-p-sulfonic, tartaric, acetic, citric, methanesulfonic, formic, benzoic, malonic, naphthalene-2-sulfonic, trifluoroacetic and benzenesulfonic acids. Salts derived from appropriate bases include, but are not limited to, alkali such as sodium and ammonia.

[0072] As used herein, the term "polypeptide" is intended to encompass a singular "polypeptide" as well as plural "polypeptides," and includes any chain or chains of two or more amino acids. Thus, as used herein, terms including, but not limited to "peptide," "dipeptide," "tripeptide," "protein," "enzyme," "amino acid chain," and "contiguous amino acid sequence" are all encompassed within the definition of a "polypeptide," and the term "polypeptide" can be used instead of, or interchangeably with, any of these terms. The term further includes polypeptides that have undergone one or more post-translational modification(s), including for example, but not limited to, glycosylation, acetylation, phosphorylation, amidation, derivatization, proteolytic cleavage, post-translation processing, or modification by inclusion of one or more non-naturally occurring amino acids. Conventional nomenclature exists in the art for polynucleotide and polypeptide structures. For example, one-letter and three-letter abbreviations are widely employed to describe amino acids: Alanine (A; Ala), Arginine (R; Arg), Asparagine (N; Asn), Aspartic Acid (D; Asp), Cysteine (C; Cys), Glutamine (Q; Gln), Glutamic Acid (E; Glu), Glycine (G; Gly), Histidine (H; His), Isoleucine (I; Ile), Leucine (L; Leu), Methionine (M; Met), Phenylalanine (F; Phe), Proline (P; Pro), Serine (S; Ser), Threonine (T; Thr), Tryptophan (W; Trp), Tyrosine (Y; Tyr), Valine (V; Val), and Lysine (K; Lys). Amino acid residues described herein are preferred to be in the "L" isomeric form. However, residues in the "D" isomeric form may be substituted for any L-amino acid residue provided the desired properties of the polypeptide are retained.

[0073] "Protein" is used herein interchangeably with "peptide" and "polypeptide," and includes both peptides and polypeptides produced synthetically, recombinantly, or in vitro and peptides and polypeptides expressed in vivo after nucleic acid sequences are administered into a host animal or human subject. The term "polypeptide" is preferably intended to refer to any amino acid chain length, including those of short peptides from about two to about 20 amino acid residues in length, oligopeptides from about 10 to about 100 amino acid residues in length, and longer polypeptides including from about 100 amino acid residues or more in length. Furthermore, the term is also intended to include enzymes, i.e., functional biomolecules including at least one amino acid polymer. Polypeptides and proteins of the present invention also include polypeptides and proteins that are or have been post-translationally modified, and include any sugar or other derivative(s) or conjugate(s) added to the backbone amino acid chain.

[0074] The term "promoter," as used herein refers to a region or regions of a nucleic acid sequence that regulates transcription.

[0075] "Purified," as used herein, means separated from many other compounds or entities. A compound or entity may be partially purified, substantially purified, or pure. A compound or entity is considered pure when it is removed

from substantially all other compounds or entities, *i.e.*, is preferably at least about 90%, more preferably at least about 91%, 92%, 93%, 94%, 95%, 96%, 97%, 98%, 99%, or greater than 99% pure. A partially or substantially purified compound or entity may be removed from at least 50%, at least 60%, at least 70%, or at least 80% of the material with which it is naturally found, *e.g.*, cellular material such as cellular proteins and/or nucleic acids.

[0076] The term "recombinant" indicates that the material (*e.g.*, a polynucleotide or a polypeptide) has been artificially or synthetically (non-naturally) altered by human intervention. The alteration can be performed on the material within or removed from, its natural environment, or native state. Specifically, *e.g.*, a promoter sequence is "recombinant" when it is produced by the expression of a nucleic acid segment engineered by the hand of man. For example, a "recombinant nucleic acid" is one that is made by recombining nucleic acids, *e.g.*, during cloning, DNA shuffling or other procedures, or by chemical or other mutagenesis; a "recombinant polypeptide" or "recombinant protein" is a polypeptide or protein which is produced by expression of a recombinant nucleic acid; and a "recombinant virus," *e.g.*, a recombinant AAV virus, is produced by the expression of a recombinant nucleic acid.

[0077] The term "regulatory element," as used herein, refers to a region or regions of a nucleic acid sequence that regulates transcription. Exemplary regulatory elements include, but are not limited to, enhancers, post-transcriptional elements, transcriptional control sequences, and such like.

[0078] The term "RNA segment" refers to an RNA molecule that has been isolated free of total cellular RNA of a particular species. Therefore, RNA segments can refer to one or more RNA segments (either of native or synthetic origin) that have been isolated away from, or purified free from, other RNAs. Included within the term "RNA segment," are RNA segments and smaller fragments of such segments.

[0079] "Retinal neovascularization" (RNV) refers to the abnormal development, proliferation, and/or growth of retinal blood vessels, *e.g.*, on the retinal surface.

[0080] The term "regulatory element," as used herein, refers to a region or regions of a nucleic acid sequence that regulates transcription. Exemplary regulatory elements include, but are not limited to, enhancers, post-transcriptional elements, transcriptional control sequences, and such like.

[0081] The term "substantially corresponds to," "substantially homologous," or "substantial identity," as used herein, denote a characteristic of a nucleic acid or an amino acid sequence, wherein a selected nucleic acid or amino acid sequence has at least about 70 or about 75 percent sequence identity as compared to a selected reference nucleic acid or amino acid sequence. More typically, the selected sequence and the reference sequence will have at least about 76, 77, 78, 79, 80, 81, 82, 83, 84 or even 85 percent sequence identity, and more preferably, at least about 86, 87, 88, 89, 90, 91, 92, 93, 94, or 95 percent sequence identity. More preferably still, highly homologous sequences often share greater than at least about 96, 97, 98, or 99 percent sequence identity between the selected sequence and the reference sequence to which it was compared.

[0082] The percentage of sequence identity may be calculated over the entire length of the sequences to be compared, or may be calculated by excluding small deletions or additions which total less than about 25 percent or so of the chosen reference sequence. The reference sequence may be a subset of a larger sequence, such as a portion of a gene or flanking sequence, or a repetitive portion of a chromosome. However, in the case of sequence homology of two or more polynucleotide sequences, the reference sequence will typically comprise at least about 18-25 nucleotides, more typically at least about 26 to 35 nucleotides, and even more typically at least about 40, 50, 60, 70, 80, 90, or even 100 or so nucleotides.

[0083] When highly-homologous fragments are desired, the extent of percent identity between the two sequences will be at least about 80%, preferably at least about 85%, and more preferably about 90% or 95% or higher, as readily determined by one or more of the sequence comparison algorithms well-known to those of skill in the art, such as *e.g.*, the FASTA program analysis described by Pearson and Lipman (1988).

[0084] The term "subject," as used herein, describes an organism, including mammals such as primates, to which treatment with the compositions according to the present invention can be provided. Mammalian species that can benefit from the disclosed methods of treatment include, but are not limited to, apes; chimpanzees; orangutans; humans; monkeys; domesticated animals such as dogs and cats; livestock such as horses, cattle, pigs, sheep, goats, and chickens; and other animals such as mice, rats, guinea pigs, and hamsters.

[0085] "Sequential administration" of two or more agents refers to administration of two or more agents to a subject such that the agents are not present together in the subject's body at greater than *de minimis* concentrations. Administration of the agents may, but need not, alternate. Each agent may be administered multiple times.

[0086] "Significant sequence homology" as applied to an amino acid sequence means that the sequence displays at least approximately 20% identical or conservatively replaced amino acids, preferably at least approximately 30%, at least approximately 40%, at least approximately 50%, at least approximately 60% identical or conservatively replaced amino acids, desirably at least approximately 70% identical or conservatively replaced amino acids, more desirably at least approximately 80% identical or conservatively replaced amino acids, and most desirably at least approximately 90% amino acid identical or conservatively replaced amino acids relative to a reference sequence. When two or more sequences are compared, any of them may be considered the reference sequence. % identity can be calculated using

a FASTA or BLASTP algorithm, using default parameters. A PAM250 or BLOSUM62 matrix may be used. For purposes of calculating % identical or conservatively replaced residues, a conservatively replaced residue is considered identical to the residue it replaces. Conservative replacements may be defined in accordance with Stryer, L, Biochemistry, 3rd ed., 1988, according to which amino acids in the following groups possess similar features with respect to side chain properties such as charge, hydrophobicity, aromaticity, etc. (1) Aliphatic side chains: G, A, V, L, I; (2) Aromatic side chains: F, Y, W; (3) Sulfur-containing side chains: C, M; (4) Aliphatic hydroxyl side chains: S, T; (5) Basic side chains: K, R, H; (6) Acidic amino acids: D, E, N, Q; and (7) Cyclic aliphatic side chain: P.

[0087] "Substantial sequence homology" as applied to a sequence means that the sequence displays at least approximately 60% identity, desirably at least approximately 70% identity, more desirably at least approximately 80% identity, and most desirably at least approximately 90% identity relative to a reference sequence. When two or more sequences are compared, any of them may be considered the reference sequence. % identity can be calculated using a FASTA, BLASTN, or BLASTP algorithm, depending on whether amino acid or nucleotide sequences are being compared. Default parameters may be used, and in exemplary embodiments, a PAM250 and/or BLOSUM62 matrix or such like may be employed in the practice of the invention.

[0088] As used herein, the term "structural gene" is intended to generally describe a polynucleotide, such as a gene, that is expressed to produce an encoded peptide, polypeptide, protein, ribozyme, catalytic RNA molecule, or antisense molecule.

[0089] A "sustained release formulation" is a composition of matter that comprises a therapeutic agent as one of its components and further comprises one or more additional components, elements, or structures effective to provide sustained release of the therapeutic agent, optionally in part because of the physical structure of the formulation. Sustained release is release or delivery that occurs either continuously or intermittently over an extended period, e.g., at least several days, at least several weeks, at least several months, at least several years, or even longer, depending upon the particular formulation employed.

[0090] The term "subject," as used herein, describes an organism, including mammals such as primates, to which treatment with the compositions according to the present invention can be provided. Mammalian species that can benefit from the disclosed methods of treatment include, but are not limited to, humans, non-human primates such as apes; chimpanzees; monkeys, and orangutans, domesticated animals, including dogs and cats, as well as livestock such as horses, cattle, pigs, sheep, and goats, or other mammalian species including, without limitation, mice, rats, guinea pigs, rabbits, hamsters, and the like.

[0091] "Transcriptional regulatory element" refers to a polynucleotide sequence that activates transcription alone or in combination with one or more other nucleic acid sequences. A transcriptional regulatory element can, for example, comprise one or more promoters, one or more response elements, one or more negative regulatory elements, and/or one or more enhancers.

[0092] As used herein, a "transcription factor recognition site" and a "transcription factor binding site" refer to a polynucleotide sequence(s) or sequence motif(s), which are identified as being sites for the sequence-specific interaction of one or more transcription factors, frequently taking the form of direct protein-DNA binding. Typically, transcription factor binding sites can be identified by DNA footprinting, gel mobility shift assays, and the like, and/or can be predicted based on known consensus sequence motifs, or by other methods known to those of ordinary skill in the art.

[0093] "Transcriptional unit" refers to a polynucleotide sequence that comprises at least a first structural gene operably linked to at least a first cis-acting promoter sequence and optionally linked operably to one or more other cis-acting nucleic acid sequences necessary for efficient transcription of the structural gene sequences, and at least a first distal regulatory element as may be required for the appropriate tissue-specific and developmental transcription of the structural gene sequence operably positioned under the control of the promoter and/or enhancer elements, as well as any additional cis sequences that are necessary for efficient transcription and translation (e.g., polyadenylation site(s), mRNA stability controlling sequence(s), etc).

[0094] As used herein, the term "transformed cell" is intended to mean a host cell whose nucleic acid complement has been altered by the introduction of one or more exogenous polynucleotides into that cell.

[0095] As used herein, the term "transformation" is intended to generally describe a process of introducing an exogenous polynucleotide sequence (e.g., a viral vector, a plasmid, or a recombinant DNA or RNA molecule) into a host cell or protoplast in which the exogenous polynucleotide is incorporated into at least a first chromosome or is capable of autonomous replication within the transformed host cell. Transfection, electroporation, and "naked" nucleic acid uptake all represent examples of techniques used to transform a host cell with one or more polynucleotides.

[0096] "Treating" or "treatment of" as used herein, refers to providing any type of medical or surgical management to a subject. Treating can include, but is not limited to, administering a composition comprising a therapeutic agent to a subject. "Treating" includes any administration or application of a compound or composition of the invention to a subject for purposes such as curing, reversing, alleviating, reducing the severity of, inhibiting the progression of, or reducing the likelihood of a disease, disorder, or condition or one or more symptoms or manifestations of a disease, disorder, or condition. A composition of this invention can be administered to a subject who has developed a macular degeneration

related condition risk of developing an infection relative to a member of the general population. A composition of this invention can be administered to a subject who has developed an eye disorder such as exudative or non-exudative AMD, diabetic retinopathy, uveitis, Bechet's disease, a localized or a generalized inflammation, or such like, or to a subject that is at increased risk of developing such a disorder (as compared to a member of the general population). In certain aspects, the inventors contemplate that the compositions of the present invention may also be administered prophylactically, *i.e.*, before development of any symptom or manifestation of the condition, where such prophylaxis is warranted. Typically, in such cases, the subject will be one that has been diagnosed for being "at risk" of developing such a disease or disorder, either as a result of familial history, medical record, or the completion of one or more diagnostic or prognostic tests indicative of a propensity for subsequently developing such a disease or disorder.

[0097] The term "a sequence essentially as set forth in SEQ ID NO:X" means that the sequence substantially corresponds to a portion of SEQ ID NO:X and has relatively few nucleotides (or amino acids in the case of polypeptide sequences) that are not identical to, or a biologically functional equivalent of, the nucleotides (or amino acids) of SEQ ID NO:X. The term "biologically functional equivalent" is well understood in the art, and is further defined in detail herein. Accordingly, sequences that have about 85% to about 90%; or more preferably, about 91% to about 95%; or even more preferably, about 96% to about 99%; of nucleotides that are identical or functionally equivalent to one or more of the nucleotide sequences provided herein are particularly contemplated to be useful in the practice of the invention.

[0098] Suitable standard hybridization conditions for the present invention include, for example, hybridization in 50% formamide, 5× Denhardt's solution, 5× SSC, 25 mM sodium phosphate, 0.1% SDS and 100 µg/ml of denatured salmon sperm DNA at 42°C for 16 hr followed by 1 hr sequential washes with 0.1× SSC, 0.1% SDS solution at 60°C to remove the desired amount of background signal. Lower stringency hybridization conditions for the present invention include, for example, hybridization in 35% formamide, 5× Denhardt's solution, 5× SSC, 25 mM sodium phosphate, 0.1% SDS and 100 µg/ml denatured salmon sperm DNA or *E. coli* DNA at 42°C for 16 h followed by sequential washes with 0.8× SSC, 0.1% SDS at 55°C. Those of skill in the art will recognize that conditions can be readily adjusted to obtain the desired level of stringency.

[0099] Naturally, the present invention also encompasses nucleic acid segments that are complementary, essentially complementary, and/or substantially complementary to at least one or more of the specific nucleotide sequences specifically set forth herein. Nucleic acid sequences that are "complementary" are those that are capable of base-pairing according to the standard Watson-Crick complementarity rules. As used herein, the term "complementary sequences" means nucleic acid sequences that are substantially complementary, as may be assessed by the same nucleotide comparison set forth above, or as defined as being capable of hybridizing to one or more of the specific nucleic acid segments disclosed herein under relatively stringent conditions such as those described immediately above.

[0100] As described above, the probes and primers of the present invention may be of any length. By assigning numeric values to a sequence, for example, the first residue is 1, the second residue is 2, *etc.*, an algorithm defining all probes or primers contained within a given sequence can be proposed:

n to $n + y$,

where n is an integer from 1 to the last number of the sequence, and y is the length of the probe or primer minus one, where $n + y$ does not exceed the last number of the sequence. Thus, for a 25-basepair probe or primer (*i.e.*, a "25 mer"), the collection of probes or primers correspond to bases 1 to 25, bases 2 to 26, bases 3 to 27, bases 4 to 28, and so on over the entire length of the sequence. Similarly, for a 35-basepair probe or primer (*i.e.*, a "35-mer"), exemplary primer or probe sequence include, without limitation, sequences corresponding to bases 1 to 35, bases 2 to 36, bases 3 to 37, bases 4 to 38, and so on over the entire length of the sequence. Likewise, for 40-mers, such probes or primers may correspond to the nucleotides from the first basepair to bp 40, from the second bp of the sequence to bp 41, from the third bp to bp 42, and so forth, while for 50-mers, such probes or primers may correspond to a nucleotide sequence extending from bp 1 to bp 50, from bp 2 to bp 51, from bp 3 to bp 52, from bp 4 to bp 53, and so forth.

[0101] In certain embodiments, it will be advantageous to employ one or more nucleic acid segments of the present invention in combination with an appropriate detectable marker (*i.e.*, a "label,"), such as in the case of employing labeled polynucleotide probes in determining the presence of a given target sequence in a hybridization assay. A wide variety of appropriate indicator compounds and compositions are known in the art for labeling oligonucleotide probes, including, without limitation, fluorescent, radioactive, enzymatic or other ligands, such as avidin/biotin, *etc.*, which are capable of being detected in a suitable assay. In particular embodiments, one may also employ one or more fluorescent labels or an enzyme tag such as urease, alkaline phosphatase or peroxidase, instead of radioactive or other environmentally less-desirable reagents. In the case of enzyme tags, colorimetric, chromogenic, or fluorogenic indicator substrates are known that can be employed to provide a method for detecting the sample that is visible to the human eye, or by analytical methods such as scintigraphy, fluorimetry, spectrophotometry, and the like, to identify specific hybridization with samples containing one or more complementary or substantially complementary nucleic acid sequences. In the case of so-called "multiplexing" assays, where two or more labeled probes are detected either simultaneously or sequentially, it may be desirable to label a first oligonucleotide probe with a first label having a first detection property or parameter (for example, an emission and/or excitation spectral maximum), which also labeled a second oligonucleotide probe with a second

label having a second detection property or parameter that is different (*i.e.*, discreet or discernable from the first label). The use of multiplexing assays, particularly in the context of genetic amplification/detection protocols are well-known to those of ordinary skill in the molecular genetic arts.

[0102] The term "vector," as used herein, refers to a nucleic acid molecule (typically comprised of DNA) capable of replication in a host cell and/or to which another nucleic acid segment can be operatively linked so as to bring about replication of the attached segment. A plasmid, cosmid, or a virus is an exemplary vector.

EXAMPLES

[0103] The following examples are included to demonstrate preferred embodiments of the invention. It should be appreciated by those of skill in the art that the techniques disclosed in the examples that follow represent techniques discovered by the inventor to function well in the practice of the invention, and thus can be considered to constitute preferred modes for its practice. However, those of skill in the art should, in light of the present disclosure, appreciate that many changes can be made in the specific embodiments which are disclosed and still obtain a like or similar result without departing from the spirit and scope of the invention.

EXAMPLE 1 - RAAV VECTOR CONSTRUCTS

[0104] The present example describes the construction of a novel recombinant variant of M013 that is engineered by fusion to a protein transduction domain peptide sequence (in this case, a specific protein domain of the tat protein from HIV has been used). This protein is expressed as a secreted fusion tatM013 protein from a recombinant AAV vector that can deliver the modified tatM013 gene into target tissues in order to inhibit inflammation locally within the transduced tissue. Thus, the therapeutic tatM013 fusion protein inhibits inflammatory signals not only from cells that have taken up the AAV-tatM013 virus vector (and thus express the therapeutic tatM013 protein directly), but also in neighboring cells that take up the secreted version of the expressed tatM013 protein itself. This "dual inhibition" then broadly inhibits multiple key pro-inflammatory pathways within various classes of cells within the AAV vector-transduced tissue, and this suppression will last as long as the AAV-M013 vector is active and expresses the therapeutic tatM013 fusion protein. The inflamed tissue highlighted in this application is designed to mimic various inflammatory diseases of the eye, but it is contemplated that similar anti-inflammatory effects may be forced in other tissues that are suitable for similar AAV-based gene delivery treatment regimens.

[0105] A variety of factors including oxidative stress, inflammation with a possible autoimmune component, genetic background (*e.g.*, mutations), and environmental or behavioral features such as smoking and diet may contribute to the pathogenesis of AMD in manners that are as yet poorly understood (Zarbin, 2004). Regardless of the underlying etiology, the clinical hallmark of AMD is the appearance of drusen, localized deposits of lipoproteinaceous material that accumulate in the space between the RPE and Bruch's membrane, which separates the RPE from the choroidal vessels (choriocapillaris). Drusen are typically the earliest clinical finding in AMD. The existence of macular drusen is a strong risk factor for the development of both wet and dry forms of AMD (Ambati *et al.*, 2003).

[0106] Ocular inflammation can affect a large number of eye structures including the conjunctiva, cornea, episclera, sclera, uveal tract, retina, vasculature, optic nerve, and orbit. Evidence of ocular inflammation can include the presence of inflammation-associated cells such as white blood cells (*e.g.*, neutrophils, macrophages) in the eye, the presence of endogenous inflammatory mediators known in the art, one or more symptoms such as eye pain, redness, light sensitivity, blurred vision and floaters, *etc.* Uveitis is a general term that refers to inflammation in the uvea of the eye, *e.g.*, in any of the structures of the uvea, including the iris, ciliary body or choroid. Specific types of uveitis include iritis, iridocyclitis, cyclitis, pars planitis, and choroiditis. Uveitis can arise from a number of different causes and is associated with a number of different diseases, including, but not limited to, rheumatic diseases such as rheumatic diseases (*e.g.*, ankylosing spondylitis and juvenile rheumatoid arthritis), certain infectious diseases such as tuberculosis and syphilis, other conditions such as sarcoidosis, systemic lupus erythematosus, chemical injury, trauma, surgery, *etc.* Keratitis refers to inflammation of the cornea. Keratitis has a diverse array of causes including bacterial, viral, or fungal infection, trauma, and allergic reaction. Amoebic infection of the cornea, *e.g.*, caused by *Acanthamoeba*, is a particular problem for contact lens wearers. Scleritis refers to inflammation of the sclera. Uveitis, keratitis, and scleritis, and methods for their diagnosis are well known in the art. Symptoms of the various inflammatory conditions that affect the eye can include, but are not limited to, eye pain, redness, light sensitivity, tearing, blurred vision, floaters, swelling, discomfort, or a combination of such symptoms. Ocular inflammation of various types is known to occur in association with a variety of local or systemic diseases, some of which are noted above. In some instances, however, the cause of the inflammation may remain unknown.

MATERIALS AND METHODS:

[0107] The myxoma virus M013 coding sequence was fused to the cell-penetrating peptide sequence derived from the HIV Tat protein using PCR, and called tatM013. The chimeric product was then fused either to a puromycin-resistance gene in a lentiviral vector plasmid, or to a secretable form of GFP through a furin cleavage site. The sequence of the TatM013 insert was verified by DNA sequencing and cloned in an AAV plasmid. The monocytic cell line THP-1 and the retina pigmented epithelium like cell line, ARPE-19, were transduced with lentiviral vectors to generate stable cell lines by selecting for puromycin resistant cells. The expression of the recombinant TatM013 protein was verified by western blotting, and the biological effects of this expressed protein on LPS- or 4-HNE-induced secretion of IL-1 β were measured by ELISA. The cellular localization of the secretable TatM013 protein was determined by fluorescent microscopy. Detection of secreted TatM013 protein was inferred by western blotting using cell conditioned media. Transfer of biological activity was determined using the ARPE-19 and Raw 264.7 cells. The *in vivo* anti-inflammatory effects of the TatM013 protein expressed from an AAV vector were determined by using the endotoxin-induced uveitis (EIU) mouse model.

[0108] In these *in vitro* experiments, the conditioned media of three independent cultures were analyzed. Animal studies were conducted in accordance with the guidelines of the Institutional Animal Care and Use Committee (IACUC) of the University of Florida (Gainesville, FL). These studies were conducted with the minimal number of mice needed to investigate endpoints that could not be addressed with *in vitro* experiments. Animal were properly anesthetized with ketamine and xylazine for in-life assessment by funduscopy and humanely killed, using carbon dioxide inhalation, for the analysis of tissue.

[0109] Cell Culture. Human HEK293T and mouse RAW 264.7 cell lines were grown in Dulbecco's modified Eagle's medium (DMEM) supplemented with 10% fetal bovine serum (FBS) and 1% penicillin-streptomycin (Pen-Strep) solution. The human ARPE-19 cell line was grown in DMEM/F12 (50/50) medium supplemented with 10% FBS and 1% Pen-Strep. The human THP-1 cell line was grown in RPMI 1640 medium supplemented with 10% FBS and 1% Pen-Strep. All the cell cultures were maintained in an incubator at 37°C with 5% CO₂. All stable cell lines generated by lentiviral vector transduction were grown in the corresponding medium supplemented with puromycin at a dose of 1 μ g/mL.

[0110] Concentration of Conditioned Medium. Conditioned M199 low-protein medium (Invitrogen Life Technologies, Grand Island, NY, USA) was harvested and placed in a 15-mL conical tube and centrifuged at 2500 $\times$ g for 5 min to remove any cellular debris. The conditioned medium was then passed through a 50-kDa molecular weight cut off (MWCO) Amicon-Ultra centrifugal filter (EMD Millipore, Billerica, MA, USA) by centrifugation at 14,000 $\times$ g for 15 min in a tabletop microcentrifuge. The flow-through was then harvested and passed through a 3-kDa Amicon-Ultra centrifugal filter (EMD Millipore) by centrifugation at 14,000 $\times$ g for 30 min in a tabletop microcentrifuge. The concentrated medium containing molecules with molecular weights between 50 and 3 kDa was collected by inversion and centrifugation of the filter at 1000 $\times$ g for 1 min.

[0111] Viral Vectors. All the lentiviral vectors were created with the pCDH-EF1-MCS-T2A-Puro plasmid (Systems Biosciences, Mountain View, CA, USA). The transgenes were cloned, using the *EcoRI* and the *NotI* restriction sites in the multiple cloning sites. Plasmids were propagated in DH5 α cells and sequenced by the dideoxy-chain-termination method. To generate viral particles, the plasmids were cotransfected with the pPACKH1 lentiviral vector packaging kit (Systems Biosciences) in HEK293T cells. The lentiviral vector-containing media were harvested 48 hr after the cotransfection and were centrifuged at 2500 $\times$ g for 5 min at 4°C. The vector-containing media were passed through a 0.22- μ m (pore size) syringe filter.

[0112] The sGFP-TatM013 vector plasmid was created by subcloning the TatM013 fusion construct downstream of Ig κ -GFP (IgG kappa light chain leader sequence fused to the GFP sequence) while maintaining the reading frame of these elements. This gene construct was then excised, using the *XbaI* (5') and *XhoI* (3') sites, and cloned in the pTR-smCBA AAV plasmid. The final pTR-smCBA-Ig κ -GFP- TatM013 plasmid was grown in SURE 2 cells (Agilent, Santa Clara, CA, USA) and purified by passage through a CsCl gradient. The DNA was packaged, purified, and titered by the Center for Vision Research Vector Core at the University of Florida according to previously published methods.

[0113] Isolation of Vitreous. Mouse vitreous was isolated using a protocol developed by Skeie and colleagues. Briefly, mice were humanely killed, using CO₂. A transverse incision was made in the cornea with a surgical scalpel to allow the aqueous humor to flow. Excess aqueous humor was absorbed with a paper towel. The retina, vitreous, and lens were eviscerated by squeezing the eye from the back. Tissues were collected in a sterile 1.5-mL tube and later transferred into an Amicon Ultra centrifugal filter (cutoff, 50 kDa; EMD Millipore). An additional 100 μ L of phosphate-buffered saline (PBS) supplemented with protease inhibitor cocktail (Thermo Fisher Scientific, Rockford, IL, USA) was added to the tube. Samples were centrifuged at 14,000 $\times$ g for 15 min, and vitreous flow-through was collected. Protein concentration of the vitreous was determined by DC protein assay (Bio-Rad, Hercules, CA, USA) according to the manufacturer's protocol and stored at -20°C until assayed.

[0114] ELISA. The ELISA kit for mouse IL-1 β was purchased from Peprotech (Peprotech, Rocky Hill, NJ, USA). The concentration of IL-1 β was determined according to the manufacturer's protocol. The concentration of human IL-1 β was determined with a RayBiotech human IL-1 β ELISA kit (RayBiotech, Norcross, GA, USA) according to the manufacturer's

protocol. In both cases, a total of 100 μ L of either tissue homogenate or cell culture medium was analyzed.

[0115] Western Blots. The cells corresponding to the conditioned media were lysed in NP-40 lysis buffer supplemented with protease inhibitor cocktail (Thermo Fisher Scientific) and 2 mM EDTA. The protein concentration of the samples was measured with the DC protein assay (Bio-Rad) according to the manufacturer's protocol. Protein lysates were diluted in Laemmli buffer containing 100 μ M dithiothreitol (DTT) and boiled for 5 min. Equal amounts of protein were separated by sodium dodecyl sulfate (SDS)-polyacrylamide gel electrophoresis and transferred onto a polyvinylidene difluoride (PVDF) membrane, using an iBlot system (Invitrogen Life Technologies) as recommended by the manufacturer. This membrane was blocked with a proprietary blocking buffer from Li-Cor Biosciences (Lincoln, NE, USA) for 1 hr at room temperature and incubated overnight with the designated primary antibody at 4°C. The membrane was then washed four times with PBS-0.1% Tween 20 and incubated with the corresponding IRDye secondary antibody (1:5000 dilution in blocking buffer; Li-Cor Biosciences). Finally, the membrane was washed as done previously and scanned with an Odyssey infrared imaging system (Li-Cor Biosciences).

[0116] Endotoxin-Induced Uveitis Mouse Model. Mice of the C57BL/6J strain were injected intravitreally with 3×10^9 vector genomes in each eye. Two weeks after injection the mice were inspected by spectral domain optical coherence tomography, using a BiopTigen high-resolution instrument, to exclude inflammatory infiltrates resulting from injection of vector. One month after injection, green fluorescent protein (GFP) expression was observed by fluorescence funduscopy. The next day mice were injected intravitreally in each eye with 25 ng of lipopolysaccharide (LPS). After 24 hr, these mice were killed, and their eyes were enucleated and placed in 4% paraformaldehyde at 4°C overnight. The eyes were placed in PBS after fixation and stored at 4°C until embedding. Eyes were dehydrated and embedded in paraffin in preparation for sectioning. Embedded eyes were sectioned through the cornea-optic nerve axis, at a thickness of 12 μ m. Eight step sections were collected in independent slides, with sections on the same slide having a difference of 80 μ m. Slides were stained with hematoxylin and eosin to visualize infiltrating cells. These cells were quantified in images of the sections by an individual who was ignorant of the treatment group.

[0117] Funduscopy. A Micron III digital fundus retinal imaging microscope (Phoenix Research Laboratories, Pleasanton, CA, USA) was used to monitor GFP expression in life. Conscious mice had their eyes dilated with 1% atropine and 2.5% phenylephrine. Mice were then anesthetized with a mixture of ketamine and xylazine in normal saline (0.9% sodium chloride solution USP, pH 5.0; Baxter, Deerfield, IL, USA). To avoid loss of moisture from the ocular surface during the procedure, mice received a drop of 2.5% hypromellose ophthalmic demulcent solution (Gonak; AKORN, Lake Forest, IL, USA). Bright-field fundus images were acquired using the same exposure times. GFP fluorescence was measured with the fluorescence filter, using the same exposure times for all the eyes.

[0118] Statistical Analysis. Data from *in vitro* experiments are reported as averages $\pm$ standard deviation. Values were compared by analysis of variance followed by a Student Newman-Keuls *t*-test to determine significant differences between groups. For animal studies, values are reported as averages $\pm$ standard error of the mean and statistical comparison was done by Mann-Whitney U test to avoid potential biases introduced by outliers. $P < 0.05$ was considered significant (* $p \leq 0.05$, ** $p \leq 0.01$, *** $p \leq 0.001$).

RESULTS AND DISCUSSION

[0119] AA V-mediated expression of myxoma virus gene M013 can inhibit the secretion of IL-1 β in vitro. On inflammasome activation by appropriate stimulatory ligands, IL-1 β precursor is cleaved and secreted, thus providing a cytokine "readout" for inflammasome signaling. To validate the inhibitory effect of fusion constructs of M013 on the secretion of IL-1 β , a lentiviral vector was first used to generate human myeloid THP-1 cells that stably express the following:

- (1) M013 fused at its C terminus to the puromycin resistance gene (puroR);
- (2) M013 with an N-terminal cell-penetrating peptide (TatM013) fused to puroR; or
- (3) the puroR gene alone (FIG. 1B).

[0120] Activation of the inflammasome was induced by incubating the transduced THP-1 cell lines with interferon (IPN)- γ in the presence or absence of lipopolysaccharide (LPS). The concentration of secreted IL-1 β in the conditioned media was increased in the presence of LPS only in cells expressing the control puromycin resistance gene alone (FIG. 1C). In the THP-1 cells expressing either the unmodified M013 protein (~ 19kDa) or the TatM013 protein (~20 kDa) fused to puroR, the secretion of IL-1 β was significantly inhibited. Interestingly, the level of detectable expression of the TatM013 protein was considerably less than that of the unmodified M013 (FIG. 1C, inset) in total cellular extracts prepared from these THP-1 cell clones. Nevertheless, both proteins prevented the IFN/LPS-induced secretion of IL-1 β .

[0121] In the retina, induced IL-1 β can be secreted from microglia, endothelial cells (Kohno *et al.*, 2013), and cells of the retinal pigmented epithelium (RPE) (Kauppinen *et al.*, 2012). The RPE-derived cell line ARPE-19 was stably transfected with the same lentiviral vectors as described previously to study the effect of TatM013 on IL-1 β secretion by a

biologically relevant ocular cell type. The modified ARPE-19 cells expressed the TatM013-puroR fusion or the control puromycin-resistance gene alone. Inflammasome activation was induced by incubation of these cells with 4-hydroxy-2-nonenal (4-HNE); a reactive aldehyde generated by oxidative stress in the retina and reported to activate the inflammasome in these cells (Kauppinen *et al.*, 2012). In ARPE-19 cells, 4-HNE led to a 3-fold induction of IL-1 β secretion, but this induction was blocked by expression of the TatM013 gene compared with cells expressing GFP only (FIG. 1D). Together, these results demonstrate that ectopic expression of M013 or TatM013 protein inhibited inflammasome-activated IL-1 β secretion *in vitro*. Furthermore, this fusion of the M013 gene with the cell-penetrating peptide Tat did not seem to compromise the efficiency of this inhibitory function of M013.

[0122] Development of an AAV vector that expresses secretable and cell-penetrating M013. The various cellular sources of IL-1 β in the eye make it difficult to prevent its secretion by a gene therapy strategy that blocks IL-1 β secretion only from cells transduced with the AAV vector. To permit the visualization of TatM013 protein, which has the potential to impact surrounding cells that may not be directly transduced by the gene therapy vector, the TatM013 coding sequence was fused to a sequence encoding a secretable version of GFP. This fusion construct was expressed as a single recombinant protein containing the N-terminal secretory signal from the Ig κ gene fused to GFP and linked at the C terminus to the TatM013 gene and separated by a furin cleavage site sequence (FIG. 2A). This secretion system has been demonstrated to deliver and secrete a small peptide derived from angiotensin (Verma *et al.*, 2012). To study the expression of the novel sGFP-TatM013 fusion construct, it was cloned in a lentiviral plasmid under the control of the elongation factor (EF)-1 α promoter and fused to the puromycin resistance (puroR) gene through a self-cleaving 2A peptide (FIG. 2A). Expression of the fused sGFP-TatM013 protein was demonstrated by Western blot, which revealed a band of the predicted size using either an antibody against the 2A peptide (not shown) or against GFP itself (FIG. 2B). By using fluorescence microscopy, it was observed that control GFP had a cytoplasmic distribution, whereas sGFP-TatM013 had a punctate pattern characteristic of proteins within the secretory pathway (Kakihana *et al.*, 2013; Stow and Murray, 2013; Reinhardt *et al.*, 2014) (FIG. 2C). This result suggested that the sGFP-TatM013 gene product could be expressed, and targeted for secretion.

[0123] For stable delivery to the eye, an AAV vector was designed to express sGFP-TatM013. AAV has proven to be safe and effective in multiple clinical trials intended to treat retinal disease (Hauswirth *et al.*, 2008; Maguire *et al.*, 2008; Simonelli *et al.*, 2010; Testa *et al.*, 2013; MacLaren *et al.*, 2014). This construct was subcloned in a plasmid containing the inverted terminal repeats (TRs) of AAV2 and a truncated chimeric cytomegalovirus (CMV) β -actin (smCBA) promoter, which is known to be ubiquitously active in the retina (Beltran *et al.*, 2010) (FIG. 3A). HEK293T cells were transfected with the AAV plasmid delivering either control GFP or sGFP-TatM013 to determine the effect of the secretion signal on the distribution of the fused protein. By using fluorescence microscopy, a similar punctate pattern of distribution was observed for GFP only among cells transfected with sGFP-TatM013 (FIG. 3B). To demonstrate that the sGFP-TatM013 protein was proteolyzed by furin and secreted, conditioned media was harvested from cells transfected with either control GFP or sGFP-TatM013. Medium was concentrated and a portion of it was fractionated on an SDS-polyacrylamide gel. In a Western blot (FIG. 3C), GFP was detected in the conditioned medium of sGFP-TatM013-expressing cells and was absent in the conditioned medium of GFP-expressing cells, suggesting that secretion and proteolysis at the furin cleavage site of the fusion protein did occur. The biological activity of the extracellular proteolyzed TatM013 was demonstrated by its ability to inhibit inflammasome signaling in cells incubated with this conditioned medium. Incubation of cells with TatM013-conditioned medium caused a significant decrease in the amount of IL-1 β produced by cells treated with an inflammatory agent (LPS or 4-HNE) when compared with cells that were incubated with control medium (FIG. 3D). Although IL-1 β release was repressed by the TatM013 conditioned medium, no cell death was observed after exposure to any of the conditioned media. These results demonstrated that secretable and cell-penetrating TatM013 could be expressed by AAV and that this protein retained its anti-inflammasome activity when exported into the conditioned medium.

[0124] AAV vector-mediated expression of sGFP-TatM013 protein is biologically active. With the goal of treating inflammatory retinal disease, delivery of therapeutic vector to the vitreous compartment of the eye may be clinically advantageous, because intravitreal injections are routinely performed in outpatient office visits. The sGFP-TatM013 plasmid was therefore packaged in an AAV-based vector containing four tyrosine-to-phenylalanine (Y $\rightarrow$ F) and one threonine-to-valine (T $\rightarrow$ V) mutation on its capsid surface: AAV2(quadY $\rightarrow$ F+T $\rightarrow$ V). Y $\rightarrow$ F mutations were at amino acid positions 272, 444, 500, and 730 and the T $\rightarrow$ V mutation was at amino acid position 491 of the AAV2 capsid protein. This pentuple-mutant variant was capable of infecting multiple cell types in the retina after injection into the vitreous compartment (Kay *et al.*, 2013). AAV vector was injected into the vitreous fluid of right eyes (3×10^9 vector genomes delivered) of C57B/6J mice. As a control, the left eyes were injected with the same capsid mutant driving only GFP. This vector was chosen to control for the impact of ocular injection on the release of neurotrophic factors such as basic fibroblast growth factor (bFGF) and ciliary neurotrophic factor (CNTF) as reported in the literature (Wen *et al.*, 1995; Qin and Rodrigues, 2010). Two weeks after injection all mice were inspected by spectral domain optical coherence tomography (SD-OCT) to rule out retinal detachments and inflammatory infiltrates in the vitreous. Importantly, neither the GFP-expressing virus nor the sGFP-TatM013-expressing virus led to ocular inflammation. Mice were evaluated by fluorescence

funduscopy 4 weeks after vector injection. Fluorescence imaging revealed that the AAV2(quadY-F+T-V)-smCBA-mediated GFP expression was robust and panretinal, with the most intense labeling surrounding retinal blood vessels. In contrast, AAV2(quadY-F+T-V)-mediated sGFP-TatM013 exhibited a diffuse pattern of fluorescence, suggesting secretion of the fused protein. This diffuse fluorescence, however, was not observed in noninjected control animals, indicating that the signal was caused by the expression of sGFP (FIG. 4A).

[0125] To test the biological activity of this AAV vector in a model of acute ocular inflammation, the well-characterized endotoxin-induced uveitis (EIU) mouse model was used (Rosenbaum *et al.*, 1980). Increases in IL-1 β and other cytokines have been reported in this model (Shen *et al.*, 2000), making it a suitable *in vivo* system for testing the efficacy of the viral vectors of the present invention. The biological activity of AAV-mediated sGFP-TatM013 was next evaluated by inducing an inflammatory response in the treated eyes by an injection of LPS 5 weeks after AAV injection. Levels of IL-1 β in the vitreous body were quantified by subjecting the harvested vitreous to an ELISA. The concentration of IL-1 β was significantly lower in the samples from eyes treated with sGFP-TatM013 than in the GFP-treated control eyes (FIG. 4B). To quantify the effect of this vector on the recruitment of infiltrating cells, the minimal dose of LPS (25 ng) was used that would induce uveitis, allowing these cells to be counted by light microscopy. To quantify the amount of infiltrating leukocytes in response to LPS challenge, eyes were harvested 24 hr after LPS injection, when inflammatory cell infiltration response peaks in the C57BL/6J strain (Shen *et al.*, 2000), and fixed and embedded in paraffin for sectioning and staining with hematoxylin and eosin (FIG. 4C). The average number of immune cells infiltrating the vitreous chamber in AAV-sGFP-TatM013-treated eyes was reduced by nearly 50% relative to eyes treated with control AAV-GFP (FIG. 4D). These results indicate that expression of the fusion gene sGFP-TatM013 significantly reduced the infiltrating inflammatory cell response in this widely used model of ocular inflammation. Furthermore, these results indicated that the TatM013 fusion gene had anti-inflammatory activities that could be measured both in cultured cells and in the eyes of test animals.

[0126] Inflammation contributes to the pathology of chronic diseases affecting many organ systems and tissues, including the CNS. In the brain, activation of glial cells is thought to play a role in the etiology of Alzheimer disease and amyotrophic lateral sclerosis (ALS) (Hall *et al.*, 1998; Chiu *et al.*, 2009). In the retina, chronic inflammation has been implicated in age-related macular degeneration (AMD), but the sources of that inflammation are a matter of conjecture and include such disparate molecules as oxidized lipids, bis-retinoids (Anderson *et al.*, 2013), double-stranded RNA (Tarallo *et al.*, 2012), and amyloid peptides (Cao *et al.*, 2013; Liu *et al.*, 2013). To suppress inflammation that may arise from more than one stimulus affecting a variety of cell types within complex tissues such as the eye, a generalized anti-inflammatory strategy perfected by poxviruses was exploited. The myxoma virus M013 gene is expressed as a small (~19 kDa) cytoplasmic PYD-containing protein that interacts with two important regulatory components of innate immunity: NP- κ B/p105 and the ASC of the inflammasomes. Expression of M013 protein thus blocks two important innate immune signaling cascades and reduces the secretion of inflammatory cytokines under their control: for example, IL-1 β and IL-18 under inflammasome control and TNF and IL-6 under NF- κ B control (Johnston *et al.*, 2005; Rahman, and McFadden, 2011; Rahman *et al.*, 2009). In infections of susceptible hosts (rabbits of the genus *Oryctolagus*), suppression of the immune response by myxoma is so successful that a single infectious virion is sufficient to kill an adult rabbit (Shope, 1932). Viruses deleted for the M013 gene, however, are severely attenuated because of the inability of the mutant virus to inhibit innate immune and inflammatory responses of the host (Johnston *et al.*, 2005; Rahman, and McFadden, 2011). Some of the immunomodulators expressed by this virus act only within this rabbit species, but others, such as M013, inhibit host cell targets in a wide variety of species (Liu *et al.*, 2010; Lucas and McFadden, 2004). Thus, the M013 protein inhibits both NF- κ B and inflammasome signaling in human and murine cells, as well as in rabbits (Rahman, and McFadden, 2011).

[0127] Although the retinal pigment epithelium (RPE) plays a major role in the secretion of inflammatory cytokines in diseases such as uveitis and AMD, other important sources of proinflammatory molecules include Müller glia, the major resident macroglial cells of the retina, retinal microglia, and vascular endothelial cells. Because there are multiple sources of induced inflammatory pathways, an AAV2-based capsid mutant was constructed that expresses a secretable and cell-penetrating fused version of M013. The N-terminal signal sequence from immunoglobulin κ led to efficient secretion of an sGFP-TatM013 fusion protein, with the GFP cleaved from TatM013 by furin during the secretion phase. The cell penetration signal derived from HIV tat permitted entry of the TatM013 fusion protein into intact nontransduced cells in the surrounding tissue. Cell culture experiments indicated that this fused TatM013 gene retained the immune suppressive properties of the parental M013 protein. The sGFP-TatM013 fusion protein inhibited the secretion of ligand-induced IL-1 β in relevant cell types, the mouse monocyte line RAW 246.7, human myeloid THP-1 cells, and the RPE-like human ARPE-19 line, even after incubation in media from various cells that had secreted M013. When injected in mouse vitreous this capsid mutant of AAV coexpressed recombinant M013 protein and GFP with a pattern of fluorescence consistent with a secreted protein. Furthermore, transduction with this AAV vector also led to a significant decrease in the recruitment of inflammatory cells in the vitreous, when eyes were challenged with LPS 1 month after vector delivery in the EIU mouse model.

[0128] The eye presents a particularly useful tissue to test immunomodulatory therapy against chronic inflammatory syndromes, using AAV gene therapy vectors armed with this secreted tat-fusion version of M013. Inflammation in the

eye is localized and, because the TatM013 protein is secreted from cells that take up and express the AAV-TatM013 vector, a low dose of vector delivered to the vitreous compartment should protect all layers of the retina from inflammatory signals that arise locally. Indeed, monoclonal antibodies that bind vascular endothelial growth factor (VEGF)-A are delivered directly to the vitreous in order to block neovascularization from the choroid, on the other side of the neural retina (Abedi *et al.*, 2014). Tseng and colleagues demonstrated that activation of the inflammasome could be associated with the low-grade inflammation associated with dry AMD (Tseng *et al.*, 2013). Activation of the NLRP3 inflammasome has also been reported in donor samples and in mouse models of AMD (Tarallo *et al.*, 2012; Kerur *et al.*, 2013). In addition to uveitis, the inventors contemplate that many other diseases of the eye may also be amenable to therapy using one or more of the AAV-TatM013-based vectors and expression systems disclosed herein. For example, a mouse model of geographic atrophy has been developed, based on oxidative stress in the RPE (Seo *et al.*, 2012). A secreted, cell-penetrating M013 peptide, delivered to suitable neighboring cells may be able to protect photoreceptor structure and function in the face of increased oxidative stress.

[0129] The expression of the TatM013 fusion protein significantly inhibited the lipopolysaccharide (LPS) induced secretion of IL-1 β from THP-1 cells. Stable ARPE-19 cells transduced with either empty lentivirus vector control or TatM013 lentivirus vector were stimulated with 4-HNE at 30 μ M for 24 hrs. The levels of secreted IL-1 β in the conditioned media were lower in cells expressing the TatM013 construct than in the empty lentivirus, vector control cells. The cellular distribution of sGFP-TatM013 in transfected cells was punctate in contrast to the diffuse cytoplasmic distribution of GFP alone. The fused sGFP-TatM013 construct was detected in transfected cell lysates, whereas the cleaved GFP was detected in the corresponding cell conditioned media. Incubation of either ARPE-19 or Raw 264.7 cells with this conditioned medium significantly decreased both the LPS and the 4-HNE induced secretion of IL-1 β . The *in vivo* effects were determined by intravitreally injecting the sGFP-TatM013 or GFP control vector in C57B/6 mice and by inducing intraocular inflammation one month later through an injection of LPS. The sGFP-TatM013 injected eyes had a significantly lower number of infiltrating inflammatory cells when compared to control GFP injected eyes.

[0130] The expression of TatM013 inhibited LPS- and 4-HNE-induced secretion of IL-1 β . The viral constructs described herein offer a secretable form of the TatM013 protein that is useful in blocking retinal and RPE inflammation, as demonstrated by its effect in the EIU mouse model.

[0131] These data provide the first evidence of a virus-derived inhibitor of innate immunity expressed from a gene-therapy vector that demonstrates usefulness for protecting neighboring, nontransduced cells against inflammatory insults, such as those emanating from a specific target tissue.

EXAMPLE 2 -USE OF VECTOR CONSTRUCTS FOR NON-OCULAR INDICATIONS

[0132] In addition to the use of the constructs in treatment of ocular disorders, the disclosed rAAV-tatM013 vectors may also be used to prevent, reduce, treat, and/or ameliorate one or more symptoms of inflammation in mammalian tissues other than those of the eye.

[0133] Such conditions include, for example, but not limited to, inflammation of one or more tissues or organs in the mammalian body such as the intestines, the liver, the pancreas, as well as one or more delocalized or diffuse sites within the body (including, for example, the peritoneal cavity) that are amenable to gene transduction by one or more of the disclosed AAV-based genetic constructs.

EXAMPLE 3 -SYSTEMIC ADMINISTRATION OF VECTOR CONSTRUCTS IN IMMUNOSUPPRESSIVE THERAPY MODALITIES

[0134] Systemic treatment with rAAV-vectored TatM013 therapeutic constructs, such as those described herein may be facilitated by regulating production of the fusion protein with one or more exogenous inducers.

[0135] Even without specific regulation of the TatM013 gene, localized AAV-mediated expression of the secreted M013 may also be of significant value for treating one or more chronic autoimmune disorders or other organ-specific inflammatory diseases.

EXAMPLE 4 -EXEMPLARY SECRETION SIGNALS FOR RETINAL EXPRESSION*

[0136] In the practice of the present invention, the inventors contemplate the use of vectors comprising a nucleic acid segment that encodes one or more of a number of secretion signals for the preparation of therapeutic expression systems. In addition to the exemplary secretion signal utilized in the previous examples, one or more of the following secretion signal sequences may be employed in particular aspects of the invention wherein expression in the retina is desirable. Such secretion signals include, without limitation:

TABLE 1
SECRETION SIGNAL SEQUENCES FOR RETINAL EXPRESSION

	PROTEIN	UNIPROT NO.	SIGNAL SEQUENCE	SEQ ID NO:
5	CNTF	P26992	MAAPVPWACCAVLAAAAAWYA	1
	PEDF	P36955	MQALVLLLCIGALLGHSSC	2
	FGF10	015520	MWKWILTHCASAFPHLPGCCCCCFLLFLVSSVPVTC	3
	PDGF-A	P04085	MRTLACLLLLGCGYLAHVLA	4
10	Gas6	Q14393	MAPSLSPGPAALRRAPQLLLLLLAAECALA	5
	TIMP3	P35625	MTPWLGLIVLLGSWSLGDWGAEA	6
	VEGF-A	P15692	MNFLLSWVFIWSLALLLYLHHAKWSQA	7
	TGF- β 1	P01137	MPPSGLRLLLLLLPLLWLLVLTGRPAAG	8
	CFH	P08603	MRLAKIICLMLWAICVA	9
15	IL-8	P10145	MTSKLAVALLAAFLISAALC	10
	MCP-1	P13500	MKVSAALLCCLLIAATFIPQGLA	11
	GNDF	P39905	MKLWDVVAVCLVLLHTASA	12

20 **EXAMPLE 5 -EXEMPLARY CELL PENETRATING PEPTIDES**

[0137] In the practice of the present invention, the inventors contemplate the use of vectors comprising a nucleic acid segment that encodes one or more of a number of cell penetrating peptides (CPP) for the preparation of particular therapeutic expression systems. In addition to the use of M013 peptides as shown in the previous examples, one or more of the following amino acid sequences may also find particular usefulness in certain aspects of the invention.

[0138] In those embodiments, a vector may be prepared in which the nucleotide sequence of the M013 peptide shown above is replaced with a nucleic acid sequence that encodes one or more of the CPP amino acid sequences including, without limitation:

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TABLE 2
EXEMPLARY CELL PENETRATING PEPTIDES USEFUL IN THE PRACTICE OF THE PRESENT INVENTION

Peptide Amino Acid Sequence	SEQ ID NO:	Origin
<i>From Heparan Binding Proteins:</i>		
RKKRRRESRKKRRRES	13	DPV3
GRPRESGKKRKRLKP	14	DPV6
GKRKKGKLGKKRDP	15	DPV7
GKRKKGKLGKKRPRS	16	DPV7b
RKKRRRESRRARRSPRHL	17	DPV3/10
SRRARRSPRESGKKRKRR	18	DPV10/6
VKRGKLRHVRPRVTRMDV	19	DPV1047
SRRARRSPRHLGSG	20	DPV10
LRERQSRLRRERQSR	21	DPV15
GAYDLRRERQSLRRERQSR	22	DPV15b
<i>From RNA Binding Proteins:</i>		
RKKRRQRRR	23	HIV-1 Tat
RRRNRTRRNRRRVR	24	FHV coat
TRQARNRRRRWRERQR	25	HIV-1 Rev
TRRQTRRARNR	26	HTLV-II Rex
KMTRAQRRAAARNRWTA	27	BMV Gag
NAKTRRHERRKLAIER	28	P22N
MDAQTRRRERRAEKQAWKAAN	29	λ N(1-22)
TAKTRYKARRAELIAERR	30	ϕ 21N(12-29)
TRNKRNRIQEQLNRK	31	Yeast PrP6
<i>From DNA Binding Proteins:</i>		
PRRRSSRPVRRRRPRVSRRRRRGGRRRR	32	Protamine 1
RIKAERKMRNRIAASKRKLEIAR	33	Human cJun
KRIRRERNKMAAAKSRNRRRELTD	34	Human cFos
KRARNTEAARRSRARKLQRMKQ	35	Yeast GCN4
RQIKIWFQNRMRMKWK	36	Penetratin
RVIRVWFQNKCKDKK	37	Islet-1
SKRTRQTYTRYQTLEKEHFENRYITRRRRIDIANALSLSERQIKIWFQNRMRMKSKKDR	38	Fushi-tarazu
SQIKIWFQNKRAKIKK	39	Engrailed-2
RQVTIWFQNRVRKEKK	40	HoxA-13
KQINNWFQNRKRHWK	41	Knotted-1

(continued)

EXEMPLARY CELL PENETRATING PEPTIDES USEFUL IN THE PRACTICE OF THE PRESENT INVENTION

Peptide Amino Acid Sequence	SEQ ID NO:	Origin
RHIKIWFQNRMRMKWKK	42	PDX-1
<i>From Signal Peptides:</i>		
MGLGLHLLVLAALQGAKKKRKV	43	Ig(v)
MVKSIGSWILVLFVAMWSDVGLCKKRPKP	44	BPrPp(1-30)
MANLGYWLLALFVTMTDVGGLCKKRPKP	45	MPrPp(1-28)
AAVLLPVLLAAPVQRKQKLP	46	K-FGF+NLS
AAVLLPVLLAAP	47	K-FGF + NLS
<i>From Antimicrobial Peptides:</i>		
RRIRPRPPRLPRPRPLPFPRPG	48	Bac7
VDKGSYLP RPTPPRIYNRN	49	Pyrrhocoricin
KCFQWQRNMRKVRGPPVSCI R	50	Lactoferrin 19-40
TRSSRAGLQWPVGRVHRLLRK	51	Buforin 2
GIGAVLKVLTG L PALISWIKRKRQQ	52	Melittin
GIGKWLHSAKKFGKAFVGEIMNS	53	Magainin 2
LLGDFFRKSKEKIGKEFRIVQRIKDFLRNLVPRTE SC	54	LL-37
RGGRLSY SRRRFSTSTGR	55	SynB1
YKQCHKKGKKGSG	56	Crotamine
ALWKTLLKKVLKAPKKKRKV	57	S4(13)-PV(rev)
HARIKPTFRRLKWKYKGKEW	58	L-2
<i>From Viral Proteins:</i>		
TKRRI TP KDVIDVRSVTTEINT	59	E(ms)
RQGAARVTSWLGRQLRIAGKRLEGRSK	60	VP22
NAATATRG RSAASRPTQRPRAPARSASRPRRPVQ	61	HIV-1 VPR 77-92
RHSRIGIIQQRRTRNG	62	Ribotoxin2 L31oop
KLIKGRTPIKFGKADCDRPPKHSQNGMGK	63	PreS2-TLM
PLSSIFSRI GDP	64	VT5
DPKGD PKGVTVTVTVTGKGDPKPD	65	
<i>From Natural Protein:</i>		
RRIPNRRPRR	66	HRSV
RLRWR	67	AIP6
MVRRFLVTLRIRACGPPPRVRV	68	ARF(1-22)
MVTVLFRRLRIRACGPPPRVRV	69	M918

(continued)

EXEMPLARY CELL PENETRATING PEPTIDES USEFUL IN THE PRACTICE OF THE PRESENT INVENTION

Peptide Amino Acid Sequence	SEQ ID NO:	Origin
LLIILRRRIKQAHASHK	70	pVEC
LSTAADMQQGWTGDMASG	71	Azurin p18
LSTAADMQQGWTGDMASGLDKDYLPDD	72	Azurin p28
KFHTFPQTAIGVGAP	73	hCT18-32
VPTLK (PMLKE, VPALR, VSALK, IPALK)	74	Bip
PFVYLI	75	C105Y
PIEVCMYREP	76	FGF12
From Peptide Libraries:		
R8, R9, R10, R12 Polyarginine		
KETWWETWTEWSQPKKRKV	77	Pep-1
GLAFLGLGAAGSTMGAWSQPKKRKV	78	MPG
GWTLSAGYLLGKINLKALAALAKKIL	79	Transportan
AGYLLGHINLHHLAHLAibHHIL	80	TH
KLALKALKALKAAKLKLA	81	MAP
RRWWRRWRR	82	W/R
GLWRALWRLRLSLWRLWRA	83	CADY
LIRLWSHLIHWFNRLKWKKK	84	EB-1
WEAALAEALAEALAEHLAEALAEALAA	85	GALA
LKTLTETLKELTKLTTEL	86	MAP12
QLALQLALQALQAALQLA	87	MAP17
(PPR)3, (PPR)4, (PPR)5, (PPR)6		(PPR)n
(PPR)3, (PPR)4, (PPR)5, (PPR)6		(PPR)n
GPSQPTYPGDDAPVRDLIRFYRDLQRYLNWTRHRY	88	aPP4R1
GPSQPTYPGDDAPVRDLIRFYRDLRRYLNWTRHRY	89	aPP5R1
GPSQPTYPGDDAPVRDLRRFYRDLRRYLNWTRHRY	90	aPP6R1
G(PLXX)NP1		PoliProline-based
VRLPPPVRLPPPVRLLPPP	91	SAP
VELPPPVELPPPVELPPP	92	SAP(E)
FKYDKKVRTRVVKH	93	SVM1
RASKRDGSWVKKLHRIE	94	SVM2
KGTYKKKLMRIPLKGT	95	SVM3
LYKKGPAKKGRPPLRGWFH	96	SVM4

EXEMPLARY CELL PENETRATING PEPTIDES USEFUL IN THE PRACTICE OF THE PRESENT INVENTION

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[0139] (Adapted from a review by Francesca Milleti, doi:10.1016/j.drudis.2012.03.002; 2012).

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[0192] It should be understood that the examples and embodiments described herein are for illustrative purposes only and that various modifications or changes in light thereof will be suggested to persons skilled in the art and are to be included within the spirit and purview of this application and the scope of the appended claims. All references, including publications, patent applications and patents, cited herein are hereby incorporated by reference to the same extent as if each reference was individually and specifically indicated to be incorporated by reference and was set forth in its entirety herein. Recitation of ranges of values herein are merely intended to serve as a shorthand method of referring individually to each separate value falling within the range, unless otherwise indicated herein, and each separate value is incorporated into the specification as if it were individually recited herein.

[0193] The description herein of any aspect or embodiment of the invention using terms such as "comprising," "having,"

"including," or "containing," with reference to an element or elements is intended to provide support for a similar aspect or embodiment of the invention that "consists of," "consists essentially of," or "substantially comprises" that particular element or elements, unless otherwise stated or clearly contradicted by context (e.g., a composition described herein as comprising a particular element should be understood as also describing a composition consisting of that element, unless otherwise stated or clearly contradicted by context).

[0194] All of the compositions and methods disclosed and claimed herein can be made and executed without undue experimentation in light of the present disclosure. While the compositions and methods of this invention have been described in terms of preferred embodiments, it will be apparent to those of skill in the art that variations may be applied to the compositions and methods and in the steps or in the sequence of steps of the method described herein without departing from the concept, spirit and scope of the invention. More specifically, it will be apparent that certain agents that are chemically- and/or physiologically-related may be substituted for the agents described herein while the same or similar results would be achieved. All such similar substitutes and modifications apparent to those skilled in the art are deemed to be within the spirit, scope, and concept of the invention as defined by the appended claims.

[0195] The present invention will now be described by way of reference to the following clauses:

1. A viral vector comprising a polynucleotide that comprises a first isolated nucleic acid segment that encodes a modified virally-derived, M013 polypeptide, a second isolated nucleic acid segment encoding a secretion signal peptide, each operably linked to a promoter sequence that expresses the encoded fusion polypeptide in one or more selected mammalian host cells transformed with a viral particle comprising the vector.

2. The viral vector according to clause 1, wherein the vector is a retroviral vector, a lentiviral vector, an adenoviral vector, a Herpes viral vector, a Hepatitis viral vector, an SV40 vector, an EBV vector, or an adeno-associated virus (AAV) vector.

3. The viral vector according to clause 1 or clause 2, wherein the vector is packaged in an viral particle that comprises a viral capsid protein, in which at least one surface- exposed amino acid is substituted by a non-native residue, wherein the viral particle has an increased transduction efficiency, an altered cellular specificity, or a combination thereof, as compared to a viral particle comprising the wild-type capsid protein, when the particle is introduced into a selected mammalian host cell.

4. The viral vector according to any preceding clause, wherein the vector is a self-complementary rAAV (scAAV) vector.

5. The viral vector according to any preceding clause, a) characterized as an AAV serotype 1 vector (AAV1), an AAV serotype 2 vector (AAV2), an AAV serotype 3 vector (AAV3), an AAV serotype 4 vector (AAV4), an AAV serotype 5 vector (AAV5), an AAV serotype 6 vector (AAV6), an AAV serotype 7 vector (AAV7), an AAV serotype 9 vector (AAV9), an AAV serotype 10 vector (AAV10), an AAV serotype 11 vector (AAV11), or an AAV serotype 12 vector; or b) packaged within a virion particle comprising an AAV1 capsid protein, an AAV2 capsid protein, an AAV3 capsid protein, an AAV4 capsid protein, an AAV5 capsid protein, an AAV6 capsid protein, an AAV7 capsid protein, an AAV8 capsid protein, an AAV10 capsid protein, an AAV11 capsid protein, or an AAV12 capsid protein.

6. The viral vector according to any preceding clause, wherein the polynucleotide further comprises an enhancer, a post-transcriptional regulatory sequence, a polyadenylation signal, an inverted terminal repeat, a multiple cloning site, or any combination thereof, operably linked to the first and the second nucleic acid segments.

7. The viral vector according to any preceding clause, wherein the promoter is a homologous promoter, a heterologous promoter, an endogenous promoter, an exogenous promoter, a viral-derived promoter, a mammalian-specific promoter, a tissue-specific promoter, a cell-specific promoter, a constitutive promoter, an inducible promoter, a human cell-specific promoter, or any combination thereof.

8. The viral vector according to any preceding clause, wherein the polynucleotide further comprises a sequence region that expresses or encodes a first therapeutic molecule, a first diagnostic molecule, or a combination thereof.

9. The viral vector according to any preceding clause, wherein the first therapeutic molecule or the first diagnostic molecule is selected from the group consisting of a polypeptide, a peptide, a ribozyme, a peptide nucleic acid, an siRNA, an RNAi, an antisense oligonucleotide, an antisense polynucleotide, an antibody, an antigen binding fragment, and any combination thereof.

10. The viral vector according to any preceding clause, wherein the modified, virally-derived M013 polypeptide comprises a TatM013 peptide or an M013 peptide, fused to one or more protein transduction domains that facilitate or aid in transport of the resulting fusion protein across a mammalian cell membrane, or facilitates secretion of the resulting fusion protein from one or more mammalian cells comprising the vector.

11. The viral vector according to any preceding clause, further comprising a cell penetrating peptide selected from any one of the amino acid sequences set forth in SEQ ID NO : 13 to SEQ ID NO : 111.

12. The viral vector according to any preceding clause, wherein the one or more protein transduction domains comprise an N-terminal mammalian secretion signal peptide.

13. The viral vector according to any preceding clause, wherein the one or more protein transduction domains comprise an N-terminal mammalian secretion signal peptide selected from the group consisting of an IgK peptide, a glucagon-like peptide, a CNTF peptide, a PEDF peptide, a FGF10 peptide, a PDGF-A peptide, a Gas6 peptide, a CFH peptide, a GDNF peptide, an IL-8 peptide, an MCP-1 peptide, a TIMP3 peptide, a synthetic peptide, a peptide signal sequence as set forth in any one of SEQ ID NO: 1 to 12, and combinations thereof.

14. The viral vector according to any preceding clause, comprised within a viral particle or an infectious virion.

15. A plurality of virions or infectious viral particles comprising a viral vector in accordance with any preceding clause.

16. An isolated non-human mammalian host cell comprising a viral vector in accordance with any preceding clause.

17. A composition comprising: a) a viral vector in accordance with any one of clauses 1 to 14 or the plurality of virions or infectious viral particles in accordance with clause 15; and b) a pharmaceutically-acceptable buffer, diluent, or excipient.

18. The composition according to clause 17, formulated for administration to a human.

19. The composition according to clause 17 or clause 18, comprised within a kit for diagnosing, preventing, treating or ameliorating one or more symptoms of a mammalian disease, defect, disorder, dysfunction, injury, or trauma.

20. The composition according to any one of clauses 17 to 19, for use in therapy or prophylaxis.

21. The composition according to any one of clauses 17 to 20, for use in the therapy of inflammation in a mammal.

22. Use of a composition in accordance with any one of clauses 17 to 21, in the manufacture of a medicament for diagnosing, preventing, treating or ameliorating one or more symptoms of inflammation in a mammal.

23. Use according to clause 22, in the manufacture of a medicament for treating or ameliorating one or more symptoms of arthritis, diabetic retinopathy, dry AMD, vasculitis, recurrent uveitis, Bechet's disease, Lupus erythematosus, or nephritis.

24. A kit comprising:

(1) a component selected from the group consisting of:

(a) the viral vector in accordance with any one of clauses 1 to 14; or

(b) the virion or plurality of infectious viral particle in accordance with clause 15; and

(2) instructions for using the component in the diagnosis, prevention, treatment, or amelioration of one or more symptoms of inflammation in a human.

25. The kit according to clause 24, further comprising an rAAV packaging component, such as a) a host cell that expresses at least one rep gene and/or at least one cap gene, b) a helper virus gene product; or c) a helper virus selected from adenovirus or herpes virus that facilitates expression of the rAAV viral vector.

26. A method for the treatment or the amelioration of one or more symptoms of a disease or an abnormal condition

that is associated with, or that results from, oxidative stress or inflammation in one or more cells of a mammalian subject, comprising administering to the mammalian subject an amount of a composition in accordance with any one of clauses 17 to 21, and for a time effective to treat or ameliorate the one or more symptoms of the disease or the abnormal condition.

27. The method according to clause 26, wherein the inflammation is associated with arthritis, diabetic retinopathy, macular degeneration, vasculitis, recurrent uveitis, Bechet's disease, Lupus erythematosus, nephritis, or any combination thereof.

28. The method according to clause 26 or clause 27, wherein the disease or abnormal condition is selected from the group consisting of age-related macular degeneration (AMD), wet AMD, dry AMD, geographic atrophy, drusen formation, dry eye, diabetic retinopathy, vitreoretinopathy, corneal inflammation, Best macular dystrophy, Sorsby fundus dystrophy, Mallatia Leventinese and Doyme honeycomb retinal dystrophy, uveitis, ocular hypertension, glaucoma, and combinations thereof.

29. The method according to any one of clauses 26 to 28, wherein the composition is locally administered to a first tissue within the mammalian subject.

30. The method according to any one of clauses 26 to 29, wherein the composition is directly administered to one or both eyes of the mammalian subject.

31. The method according to any one of clauses 26 to 30, wherein the composition is directly administered to one or both eyes of the mammalian subject via intravitreal injection, subretinal injection, injection into the anterior chamber of the eye, injection or application locally to the cornea, subconjunctival injection, subtenon injection, or by topical application.

32. The method according to any one of clauses 26 to 31, wherein the mammalian subject is suspected of having, has been diagnosed with, or is at risk for developing, a disease or abnormal condition that is associated with, or caused by, oxidative stress or inflammation in one or more cells of the subject.

33. The method according to any one of clauses 26 to 32, wherein the step of administering results in decreased secretion of IL-1 β in one or more cells of the mammalian subject.

34. A method of transducing a population of mammalian cells, comprising introducing into one or more cells of the population, a composition that comprises an effective amount of a viral vector in accordance with any one of clauses 1 to 17.

35. A method for treating or ameliorating one or more symptoms of oxidative stress or inflammation in a mammal, the method comprising administering to the mammal in need thereof, a viral vector in accordance with any one of clauses 1 to 17, in an amount and for a time sufficient to treat or ameliorate the one or more symptoms of oxidative stress or inflammation in the mammal.

36. The method according to clause 35, wherein the mammal is a human that has been diagnosed with macular degeneration, age-related macular degeneration (AMD), geographic atrophy, wet AMD, dry AMD, drusen formation, dry eye, diabetic retinopathy, vitreoretinopathy, Best macular dystrophy, Sorsby fundus dystrophy, Mallatia Leventinese and Doyme honeycomb retinal dystrophy, corneal inflammation, uveitis, ocular hypertension, glaucoma, or any combination thereof.

37. The method according to clause 35 or clause 36, wherein expression of the encoded therapeutic molecule in ocular cells a) treats or ameliorates one or more symptom of inflammation; b) reduces one or more adverse effects of oxidative stress; c) preserves one or more photoreceptor cells, d) restores one or more rod- and/or cone-mediated functions, e) restores visual behavior in one or both eyes of the mammal, or f) any combination thereof.

38. The method according to any one of clauses 35 to 37, wherein expression of the encoded therapeutic molecule persists in ocular cells for a period of at least three months, and preferably for at least six months or more, following administration of the viral vector to the one or both eyes of the mammal.

39. The method according to any one of clauses 35 to 38, wherein the viral vector is provided to the one or both

eyes by intravitreal injection, subretinal injection, injection into the anterior chamber of the eye, injection or application locally to the cornea, subconjunctival injection, subtenon injection, by topical application, or by any combination thereof.

5 40. The method according to any one of clauses 35 to 39, wherein the administering occurs as a single injection, or wherein the administering includes sequential injection of the composition that comprises the viral vector to the one or both eyes over a pre- determined treatment period of several weeks to several months.

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SEQUENCE LISTING

5 <110> University of Florida Research Foundation, Inc.
 McFadden, Grant
 Lewin, Alfred S.
 Lucas, Alexandra R.
 Ildefonso, Cristhian J.
 Rahman, Mohammed M.

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15 <140> PCT/US15/020001
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5 Val Thr Gly Lys Gly Asp Pro Lys Pro Asp
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Arg Arg Ile Pro Asn Arg Arg Pro Arg Arg
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EP 4 050 020 A1

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Leu Trp Arg Ala
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EP 4 050 020 A1

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Arg His Arg Tyr
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EP 4 050 020 A1

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Pro Pro

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Pro Pro

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<211> 12

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1 5 10

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<220>

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Pro Leu Ile Tyr Leu Arg Leu Leu Arg Gly Gln Phe
1 5 10

<210> 111

<211> 15

<212> PRT
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<400> 111

Lys Leu Trp Met Arg Trp Tyr Ser Pro Thr Thr Arg Arg Tyr Gly
 1 5 10 15

Claims

1. An adeno-associated virus (AAV) vector for use in treating one or more symptoms of inflammation in an eye of a mammal, wherein the viral vector comprises a polynucleotide that comprises a nucleic acid segment that encodes a fusion polypeptide comprising a virally-derived M013 polypeptide, a cell penetrating peptide, and a secretion signal peptide, wherein the nucleic acid segment is operably linked to a promoter that expresses the encoded fusion polypeptide in one or more mammalian host cells, and wherein the AAV vector is administered to the eye of the mammal via intravitreal injection.
2. The AAV vector for the use of claim 1,
 - (a) characterised as an AAV serotype 1 vector (AAV1), an AAV serotype 2 vector (AAV2), an AAV serotype 3 vector (AAV3), an AAV serotype 4 vector (AAV4), an AAV serotype 5 vector (AAV5), an AAV serotype 6 vector (AAV6), an AAV serotype 7 vector (AAV7), an AAV serotype 8 vector (AAV8), an AAV serotype 9 vector (AAV9), an AAV serotype 10 vector (AAV10), an AAV serotype 11 vector (AAV11), an AAV serotype 12 vector (AAV12), or a modified AAV serotype 2 vector (AAV2 quadY→F+T→V); or
 - (b) packaged within a virion particle comprising an AAV1 capsid protein, an AAV2 capsid protein, an AAV3 capsid protein, an AAV4 capsid protein, an AAV5 capsid protein, an AAV6 capsid protein, an AAV7 capsid protein, an AAV8 capsid protein, an AAV9 capsid protein, an AAV10 capsid protein, an AAV11 capsid protein, an AAV12 capsid protein, or an AAV2 quadY→F+T→V capsid protein.
3. The AAV vector for the use of any one of the preceding claims, wherein the polynucleotide further comprises an enhancer, a post-transcriptional regulatory sequence, a polyadenylation signal, an inverted terminal repeat, a multiple cloning site, or any combination thereof, operably linked to the nucleic acid segment.
4. The AAV vector for the use of any one of the preceding claims, wherein the fusion polypeptide comprises a TatM013 protein.
5. A plurality of virions or infectious viral particles comprising the AAV vector for the use in accordance with any one of the preceding claims.
6. A composition comprising the plurality of virions or infectious viral particles of claim 5 and a pharmaceutically-acceptable buffer, diluent, or excipient.
7. The composition of claim 7, which is formulated for intravitreal administration to a human.
8. The AAV vector for the use of any one of claims 1-4, the plurality of virions or infectious viral particles of claim 5, or the composition of claim 6 or 7, wherein the mammal has been diagnosed with macular degeneration, age-related macular degeneration (AMD), geographic atrophy, wet AMD, dry AMD, drusen formation, dry eye, diabetic retinopathy, vitreoretinopathy, corneal inflammation, uveitis, ocular hypertension or glaucoma.
9. An AAV vector for use in treating symptoms of an inflammatory-mediated disease selected from macular degeneration, age-related macular degeneration (AMD), geographic atrophy, wet AMD, dry AMD, drusen formation, dry eye, diabetic retinopathy, vitreoretinopathy, corneal inflammation, uveitis, ocular hypertension or glaucoma by transducing a population of mammalian cells by introducing a composition comprising an AAV vector packaged within a virion particle into one or more cells of the population via intravitreal injection, wherein the AAV vector comprises a

polynucleotide comprising a nucleic acid segment that encodes a fusion polypeptide that comprises a virally-derived M013 polypeptide, a cell penetrating peptide, and a secretion signal peptide, wherein the nucleic acid segment is operably linked to a promoter that expresses the encoded fusion polypeptide in one or more transduced mammalian host cells.

10. The AAV vector for the use of claim 9,

(a) characterized as an AAV serotype 1 vector (AAV1), an AAV serotype 2 vector (AAV2), an AAV serotype 3 vector (AAV3), an AAV serotype 4 vector (AAV4), an AAV serotype 5 vector (AAV5), an AAV serotype 6 vector (AAV6), an AAV serotype 7 vector (AAV7), an AAV serotype 8 vector (AAV8), an AAV serotype 9 vector (AAV9), an AAV serotype 10 vector (AAV10), an AAV serotype 11 vector (AAV11), an AAV serotype 12 vector (AAV12), or a modified AAV serotype 2 vector (AAV2 quadY→F+T→V); or

(b) packaged within a virion particle comprising an AAV1 capsid protein, an AAV2 capsid protein, an AAV3 capsid protein, an AAV4 capsid protein, an AAV5 capsid protein, an AAV6 capsid protein, an AAV7 capsid protein, an AAV8 capsid protein, an AAV9 capsid protein, an AAV10 capsid protein, an AAV11 capsid protein, an AAV12 capsid protein, or an AAV2 quadY→F+T→V capsid protein.

11. The AAV vector for the use of claim 9 or 10, wherein the fusion polypeptide comprises a TatM013 protein.

12. The AAV vector for the use of any one of claims 1-4 and 9-11, the plurality of virions or infectious viral particles of claim 5, or the composition of claim 6 or 7, wherein the promoter is an endogenous promoter, an exogenous promoter, a viral-derived promoter, a mammalian-specific promoter, a tissue-specific promoter, a cell-specific promoter, a constitutive promoter, an inducible promoter, or any combination thereof.

13. The AAV vector for the use of any one of claims 1-4 and 9-12, the plurality of virions or infectious viral particles of claim 5 or 12, or the composition of any one of claims 6, 7, and 12, wherein the secretion signal peptide is selected from the group consisting of: an Igκ peptide, a glucagon-like peptide, a CNTF peptide, a PEDF peptide, a FGF10 peptide, a PDGF-A peptide, a Gas6 peptide, a CFH peptide, a GDNF peptide, an IL-8 peptide, an MCP-1 peptide, and a TIMP peptide.

14. The AAV vector for the use of any one of claims 1-4 and 9-13, the plurality of virions or infectious viral particles of any one of claim 5, 12 and 13, or the composition of any one of claims 6, 7, 12 and 13, wherein the secretion signal peptide comprises any of the amino acid sequences as set forth in SEQ ID NOs: 1-12, or any combination thereof.

15. The AAV vector for the use of any one of claims 1-4 and 9-14, the plurality of virions or infectious viral particles of any one of claims 5 and 12-14, or the composition of any one of claims 6, 7, and 12-14, wherein the cell penetrating peptide comprises an amino acid sequence as set forth in any one of SEQ ID NOs: 13-111.

16. The AAV vector for the use of claim 2 or claim 10,

(a) characterised as a modified AAV serotype 2 vector (AAV2 quadY→F+T→V); or

(b) packaged within a virion particle comprising an AAV2 quadY→F+T→V capsid protein.

17. The AAV vector for the use of any one of claims 1-4 and 9-15, the plurality of virions or infectious viral particles of any one of claims 5 and 12-15, or the composition of any one of claims 6, 7, and 12-15, wherein the AAV vector is a self-complementary rAAV (scAAV) vector.

FIG. 1A

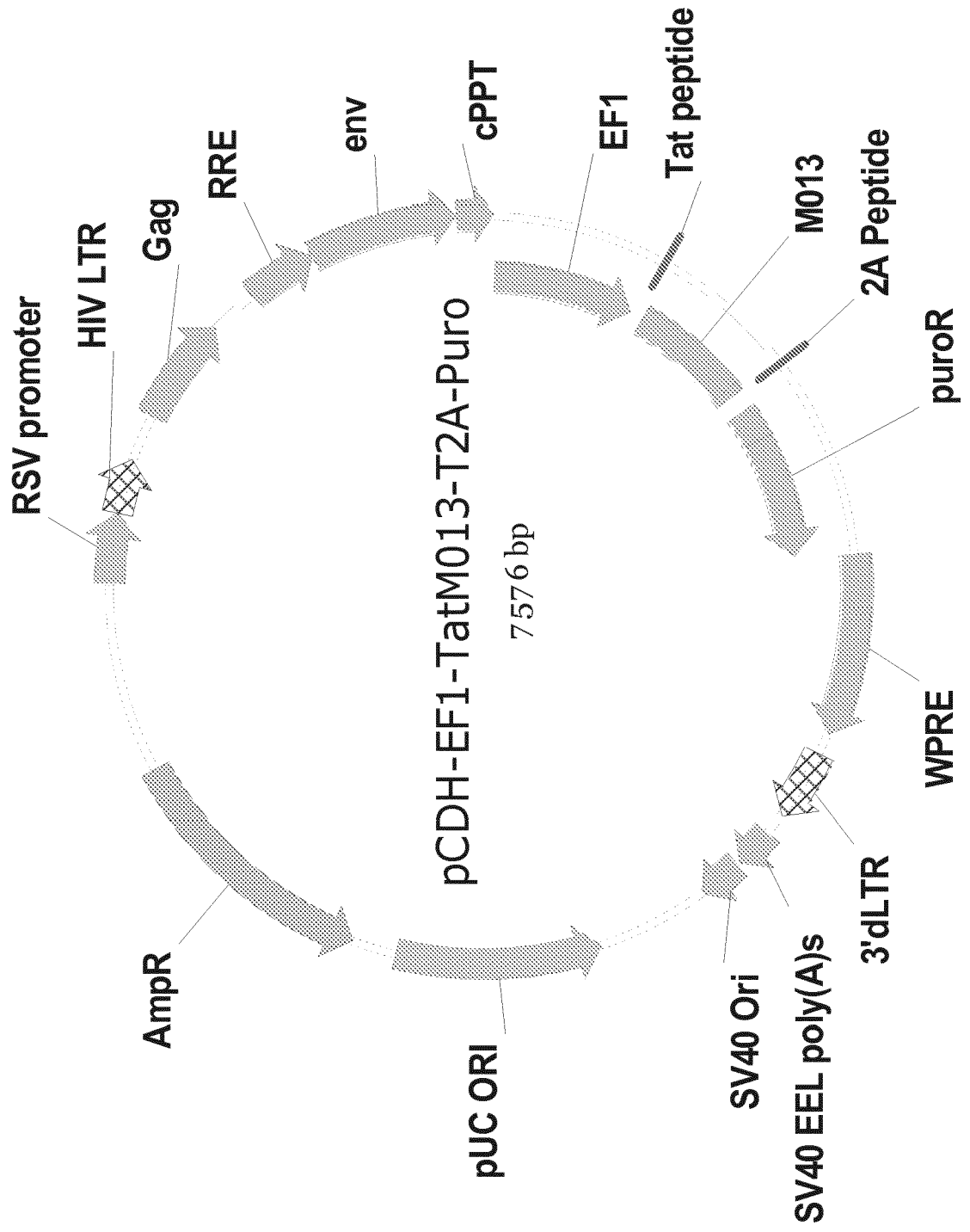


FIG. 1B

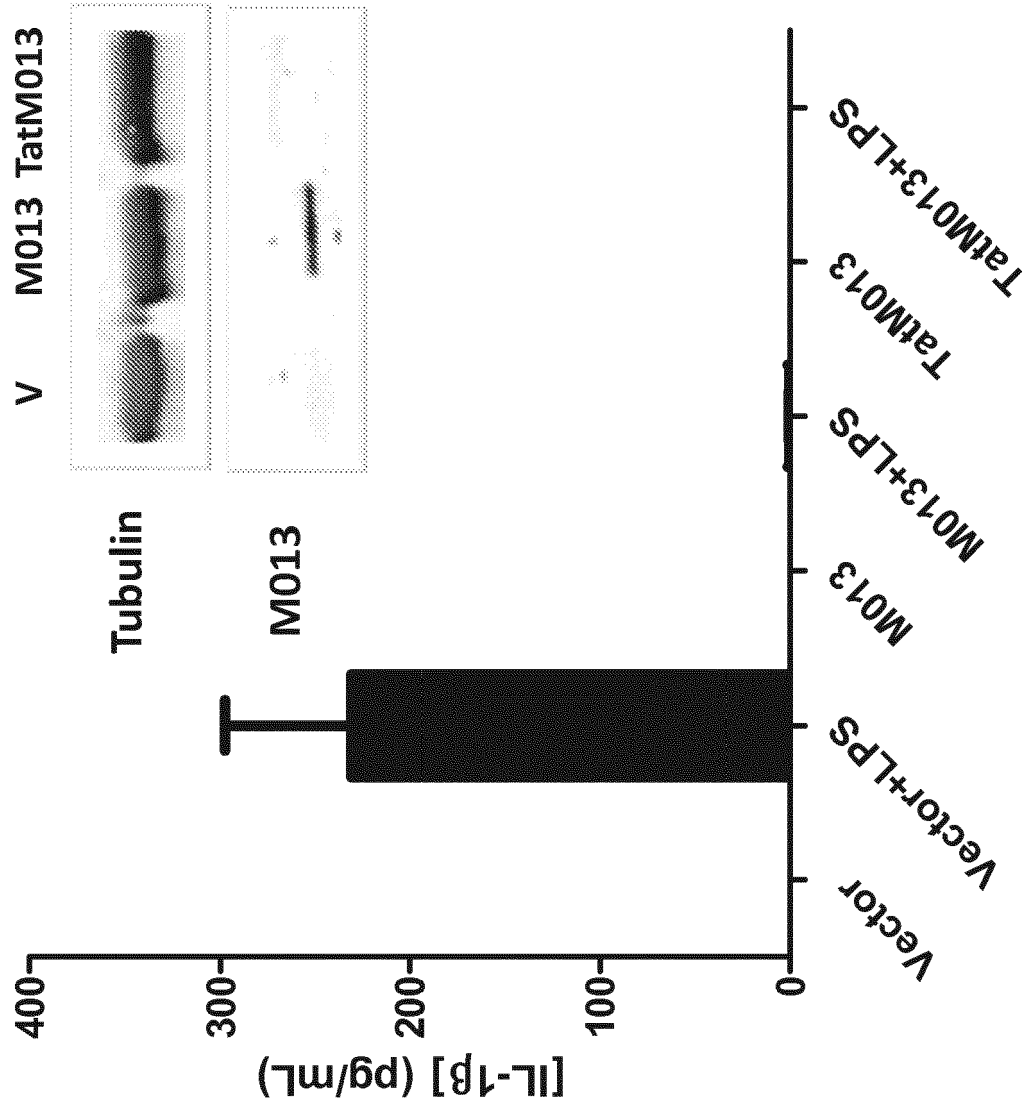


FIG. 1C

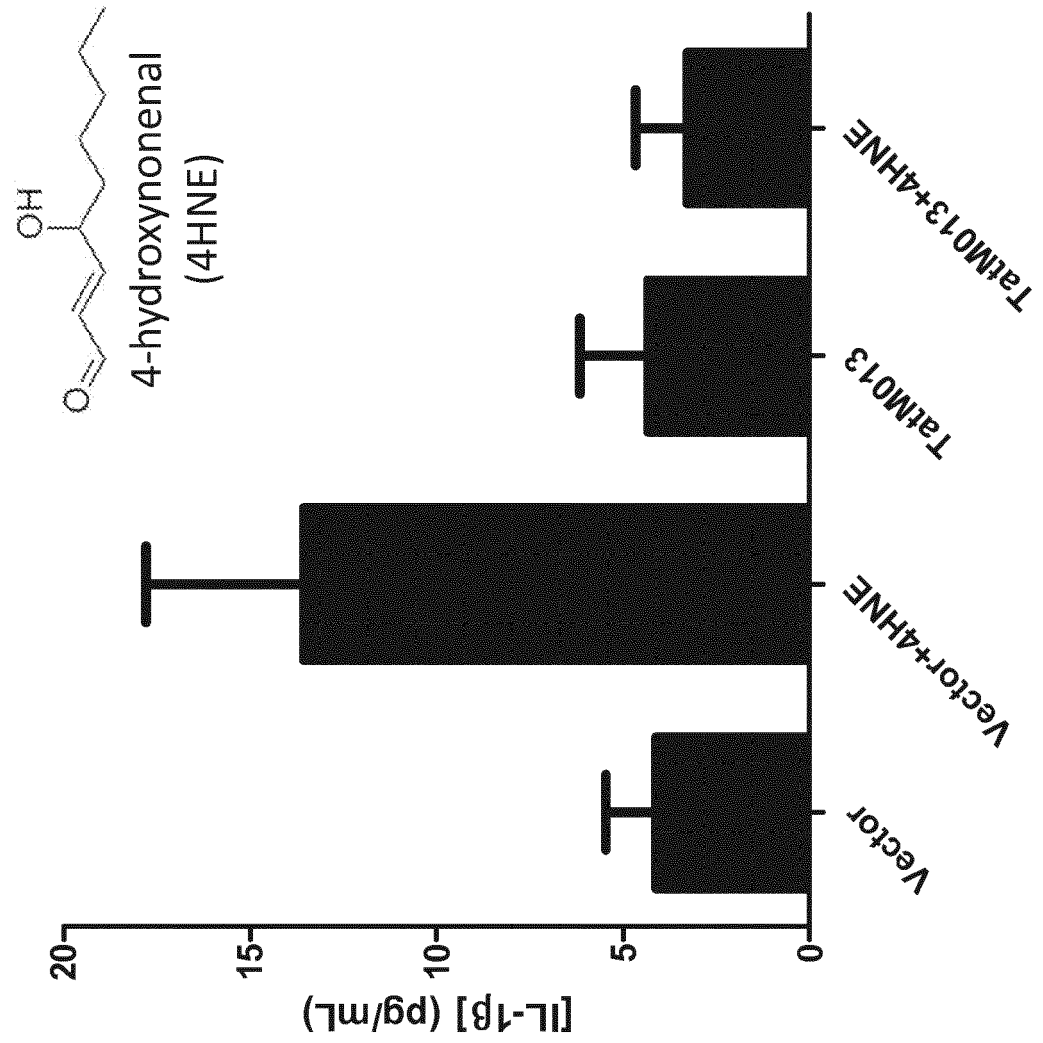


FIG. 2A

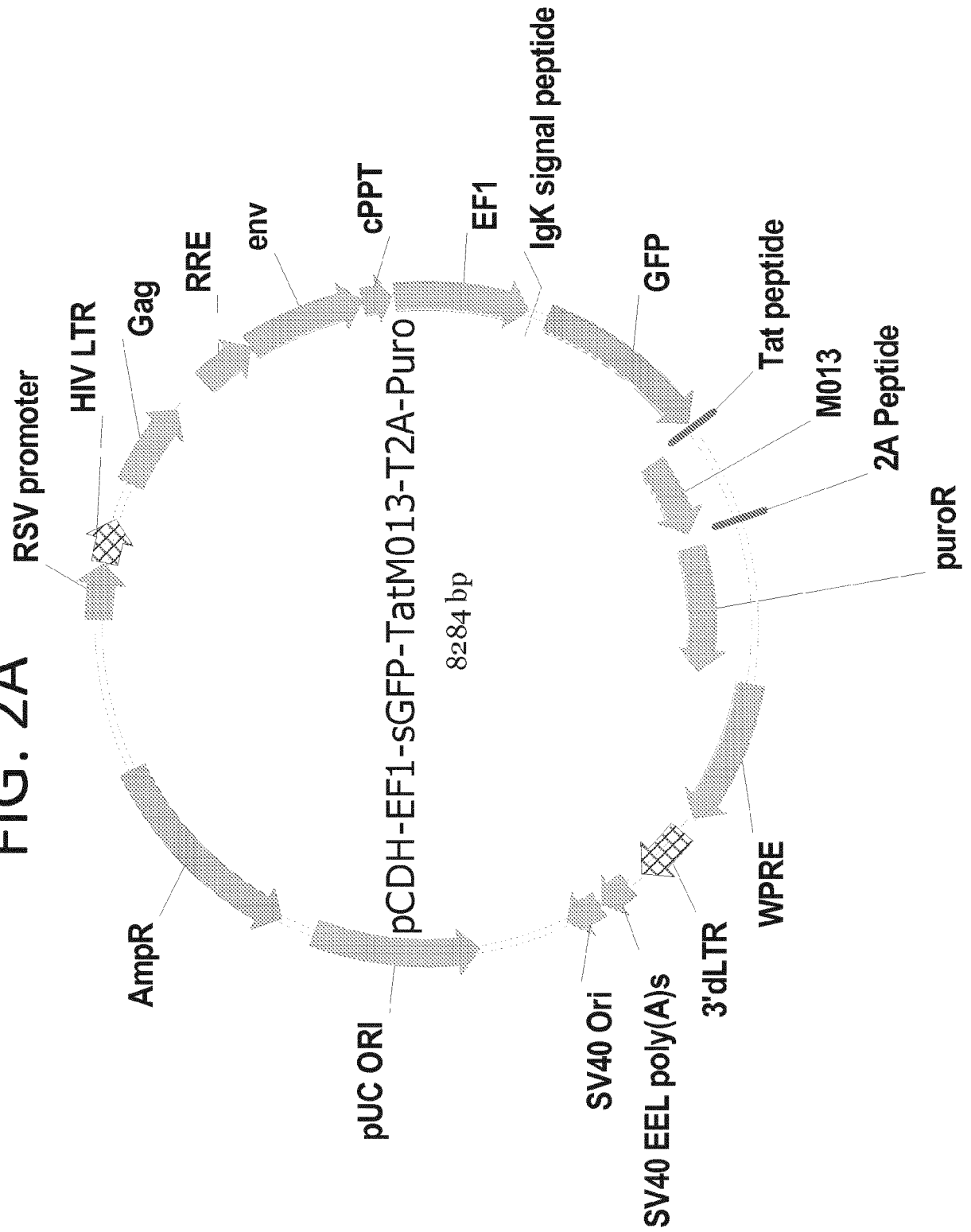


FIG. 2B

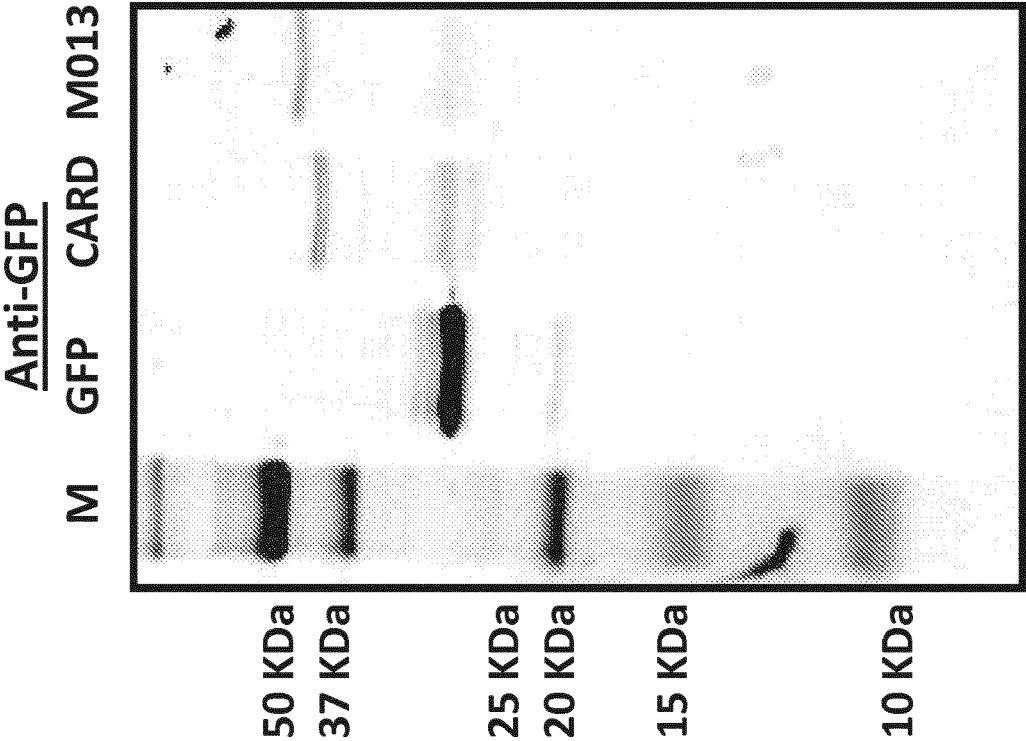


FIG. 2C

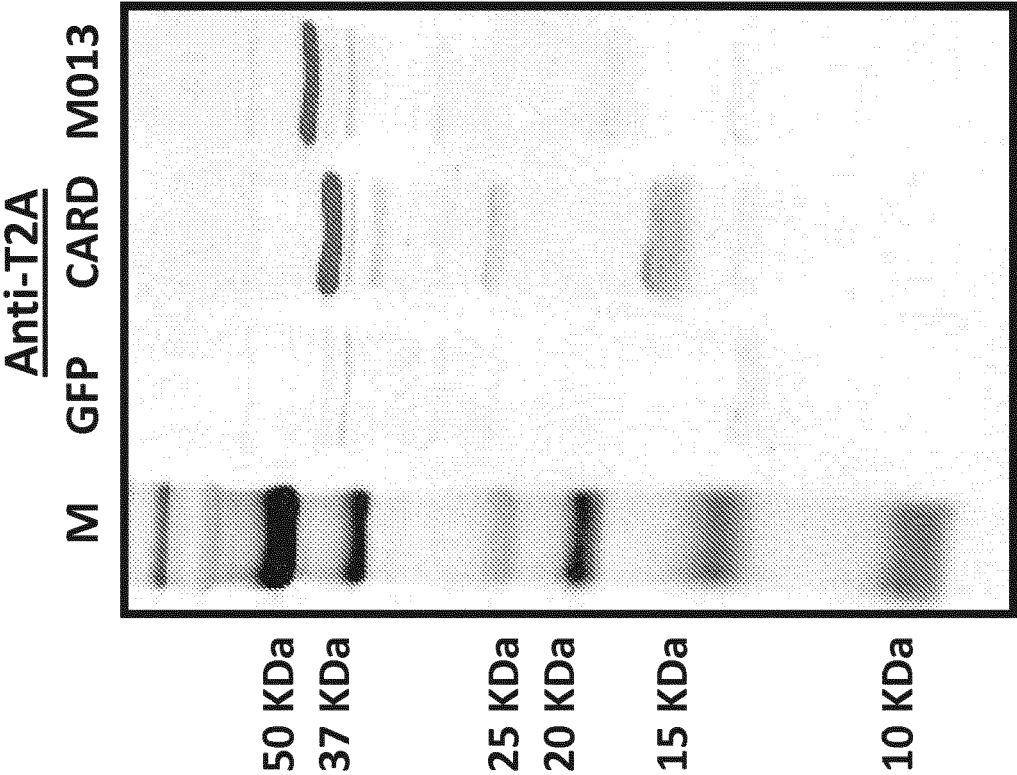


FIG. 2D

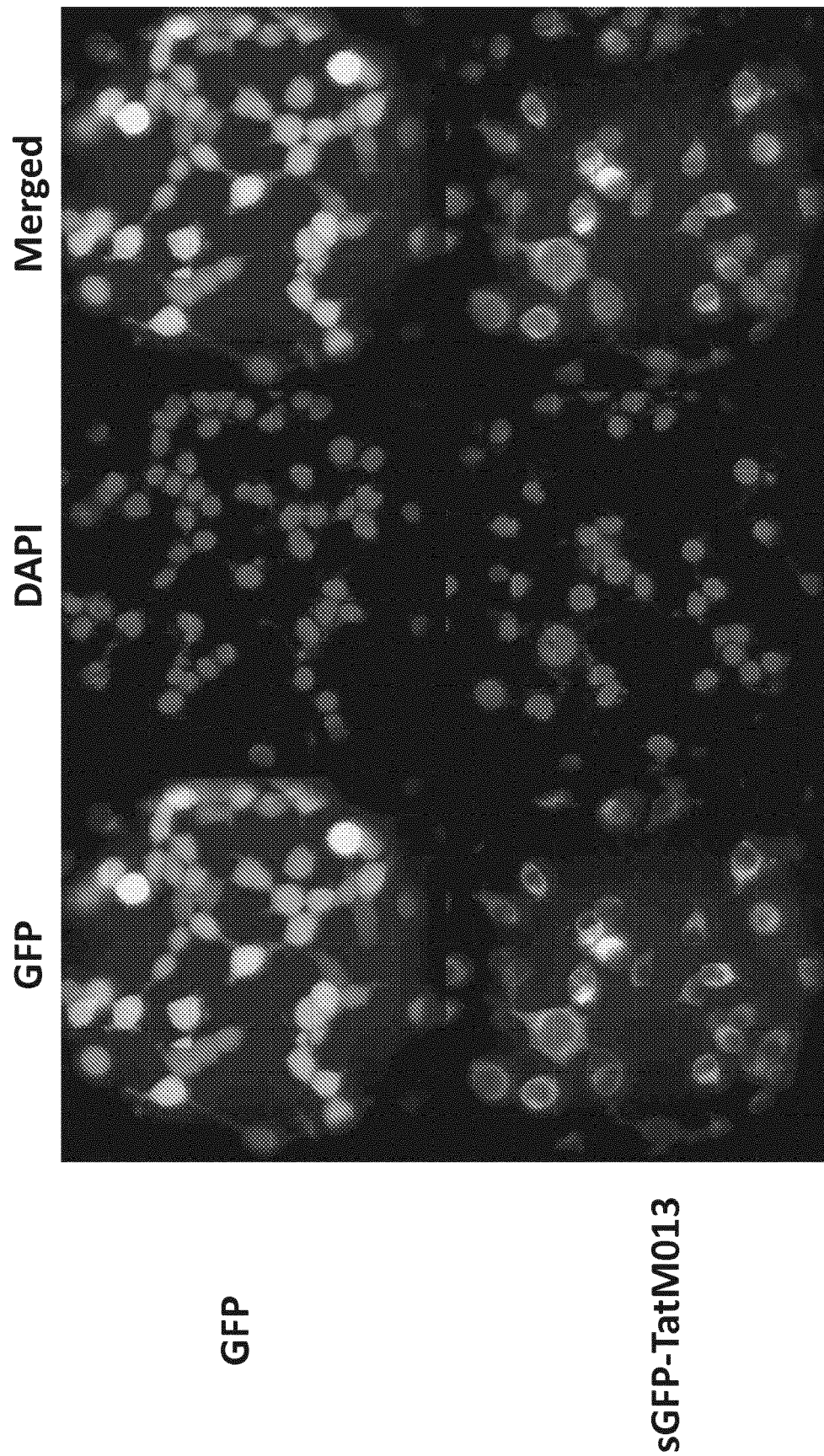


FIG. 3A

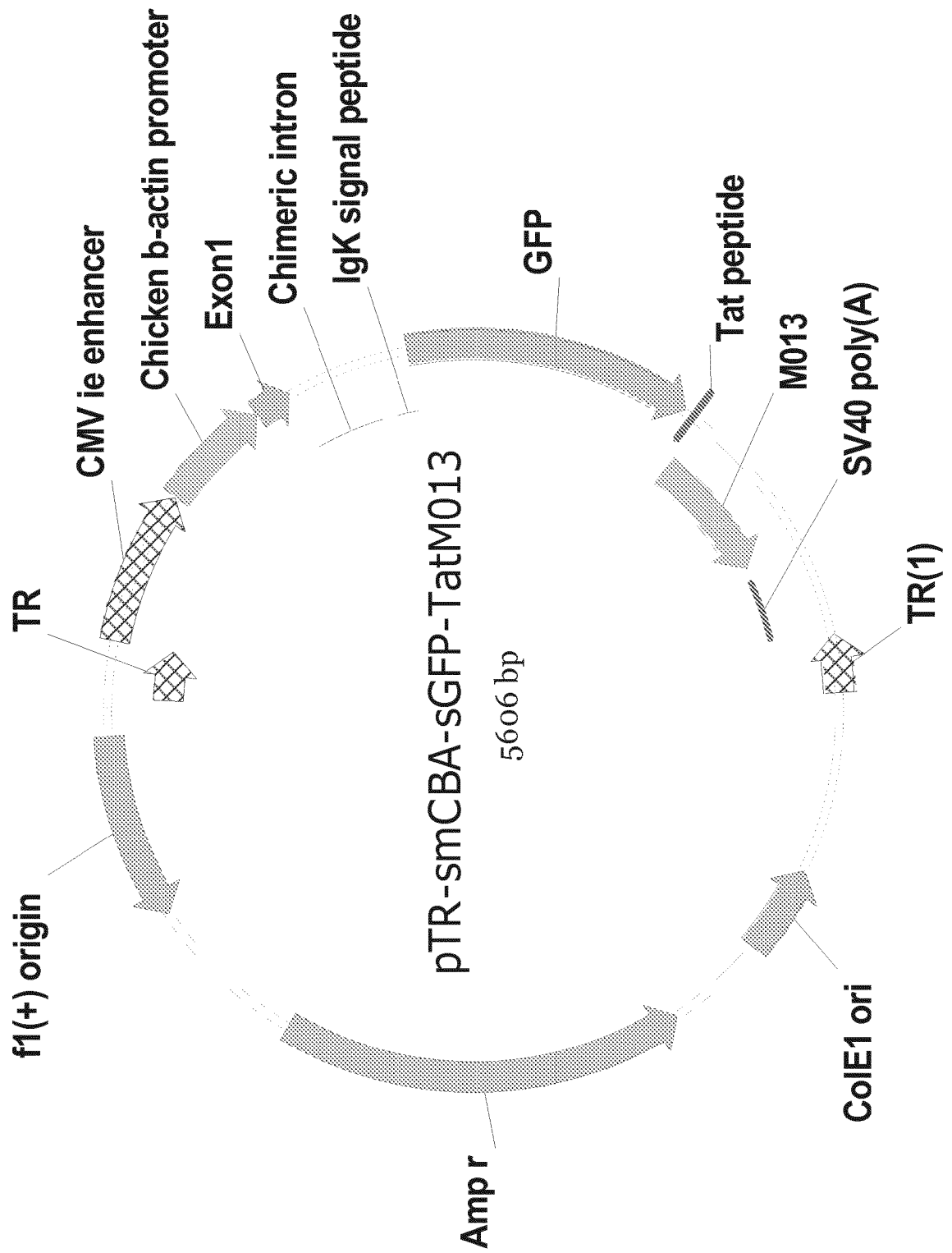


FIG. 3B

GFP

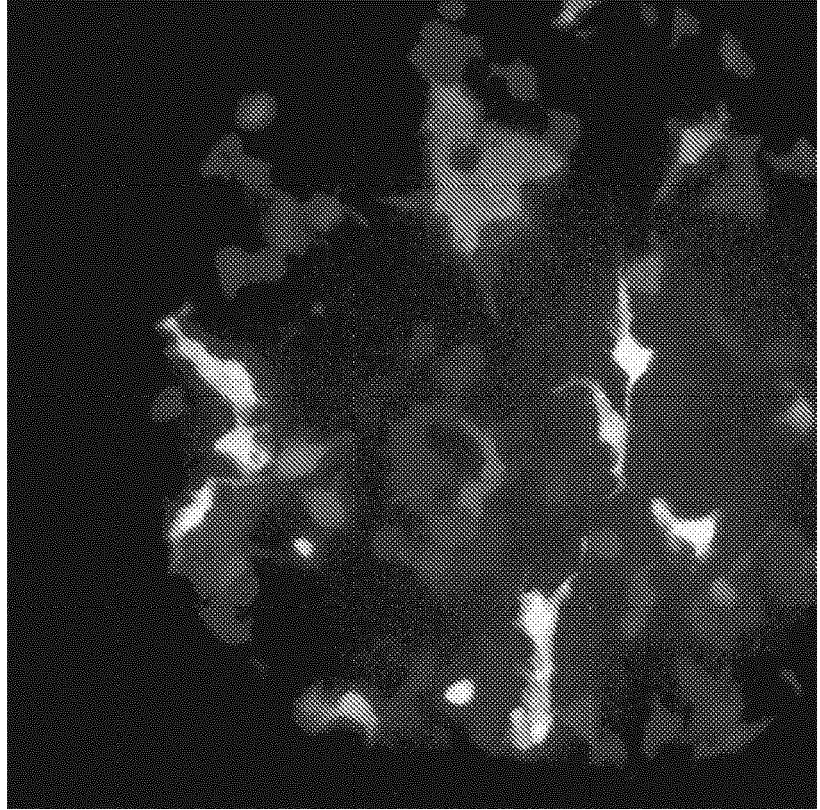


FIG. 3C

sGFP-FCS-TatM013

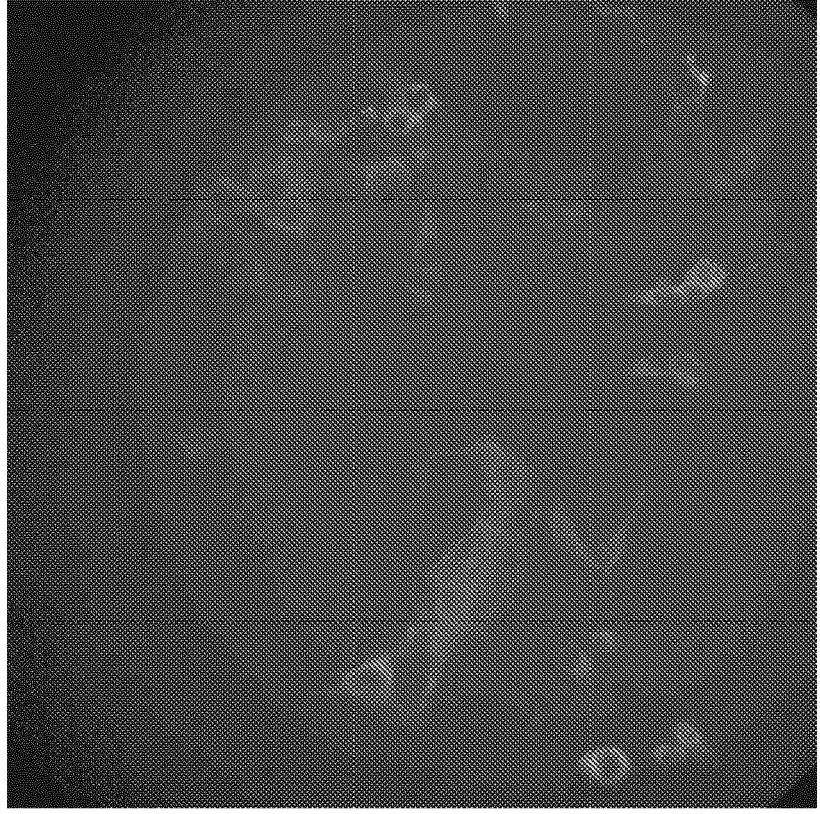


FIG. 3D

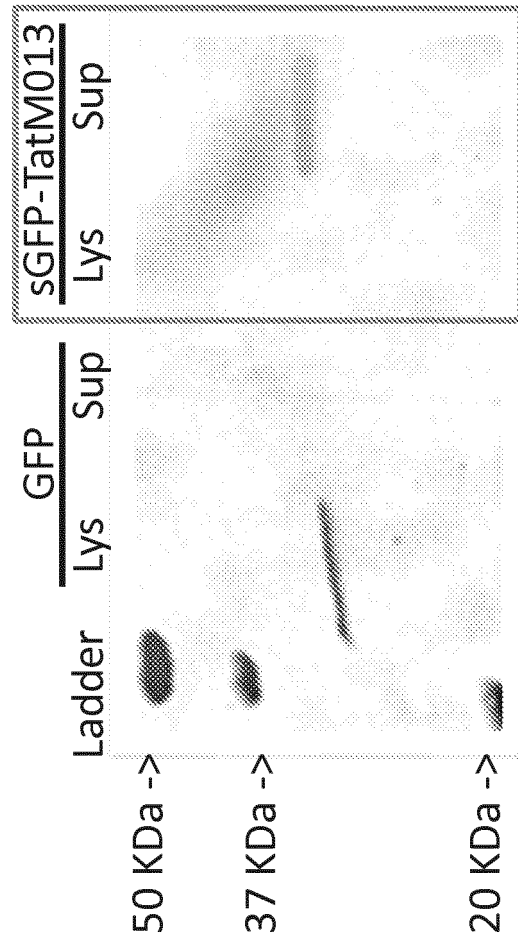


FIG. 3E

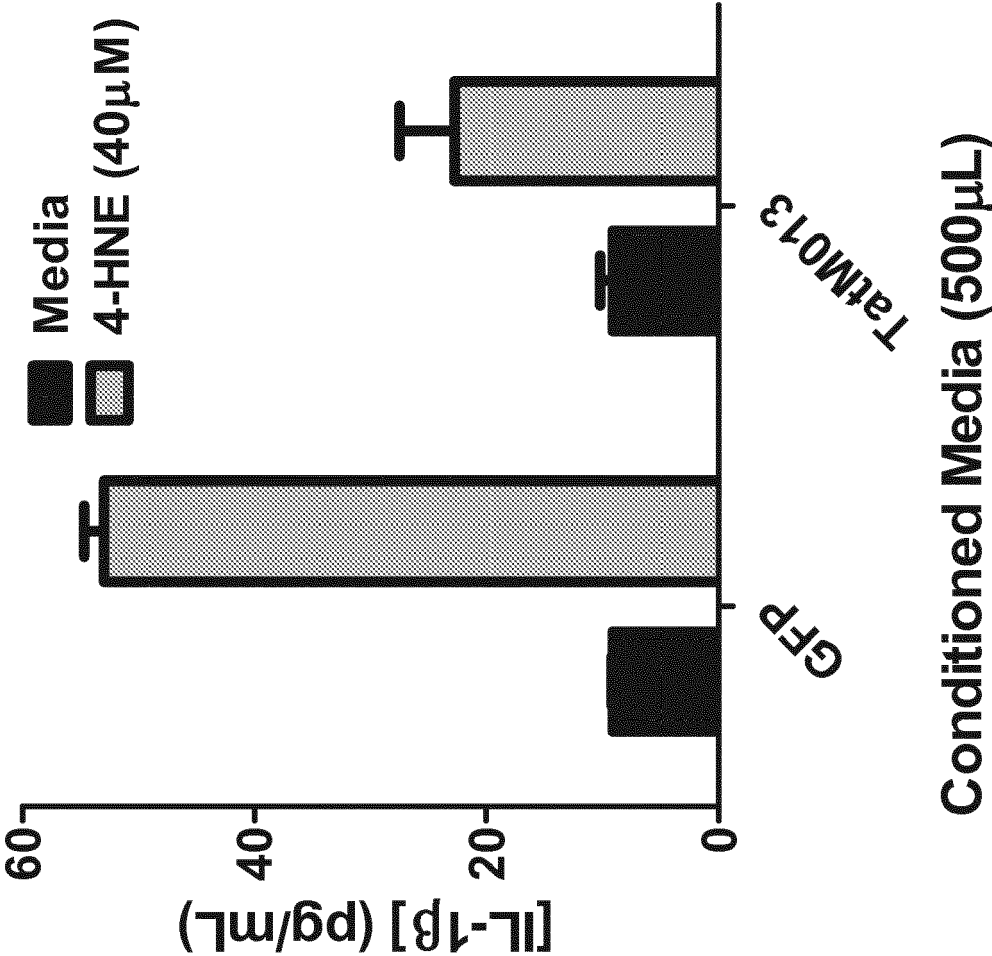
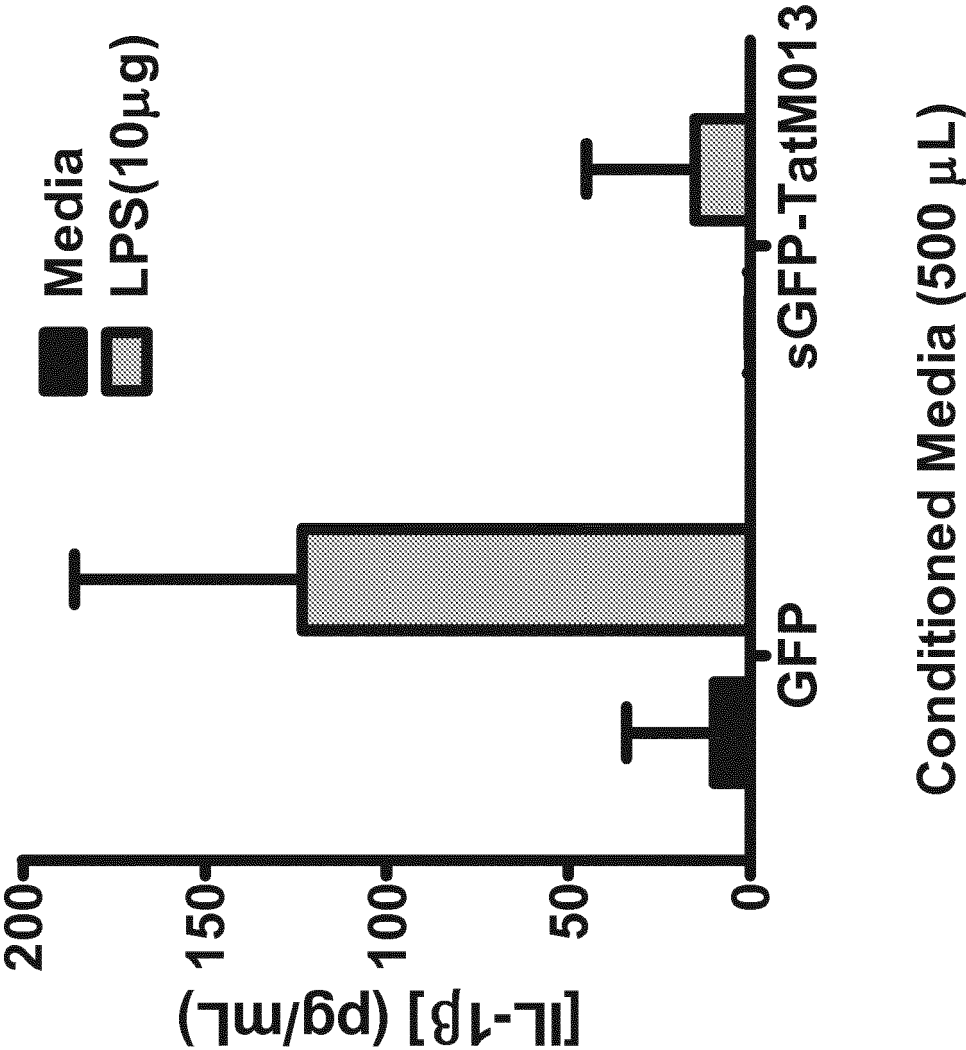


FIG. 3F



Conditioned Media (500 μL)

FIG. 4A

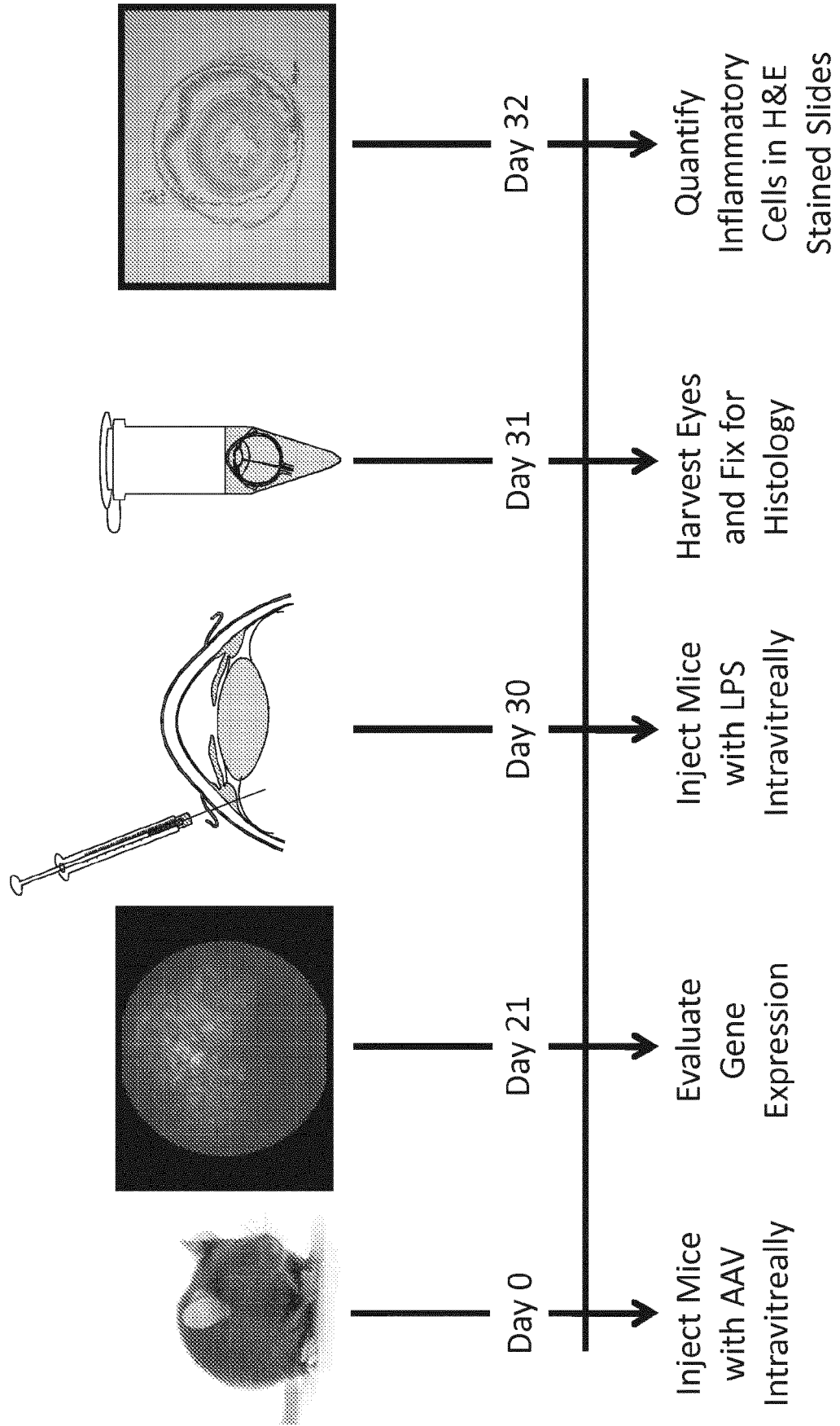


FIG. 4B

AAV2QUAD-GFP

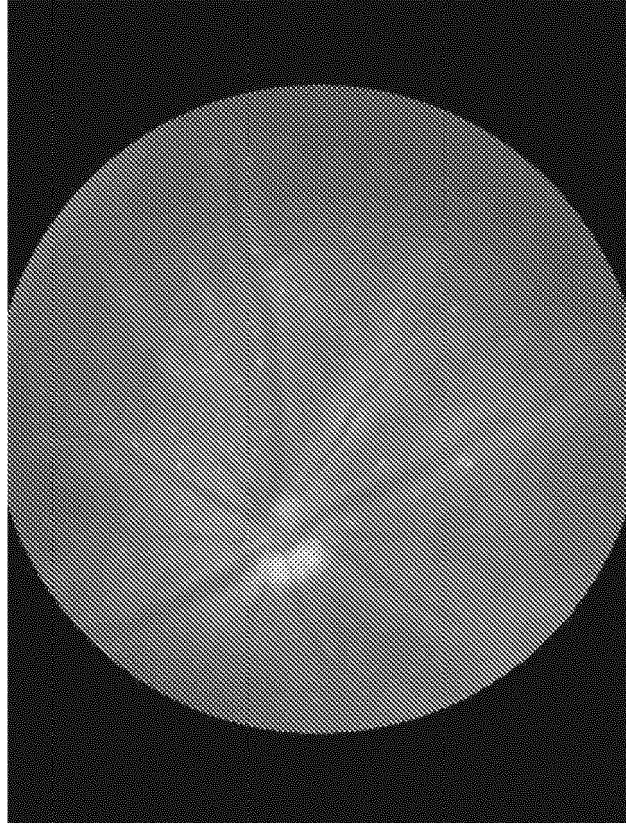


FIG. 4C

AAV2QUAD-sGFP-TatM013

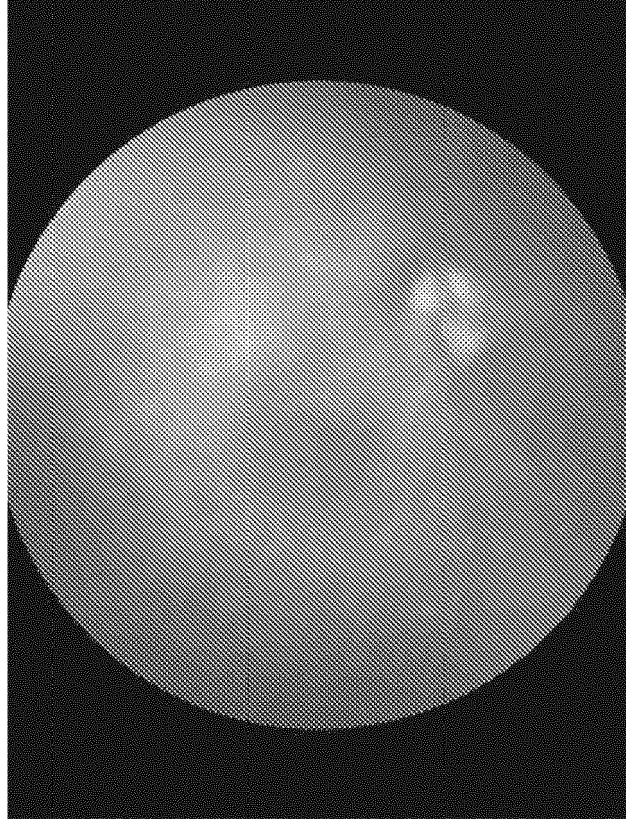


FIG. 4D

AAV2QUAD-GFP

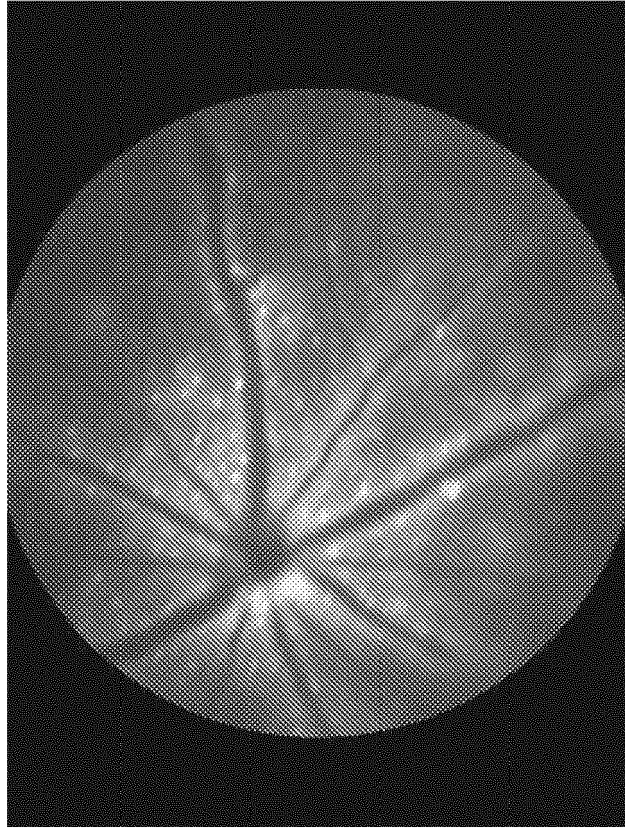


FIG. 4E

AAV2QUAD-sGFP-TatM013

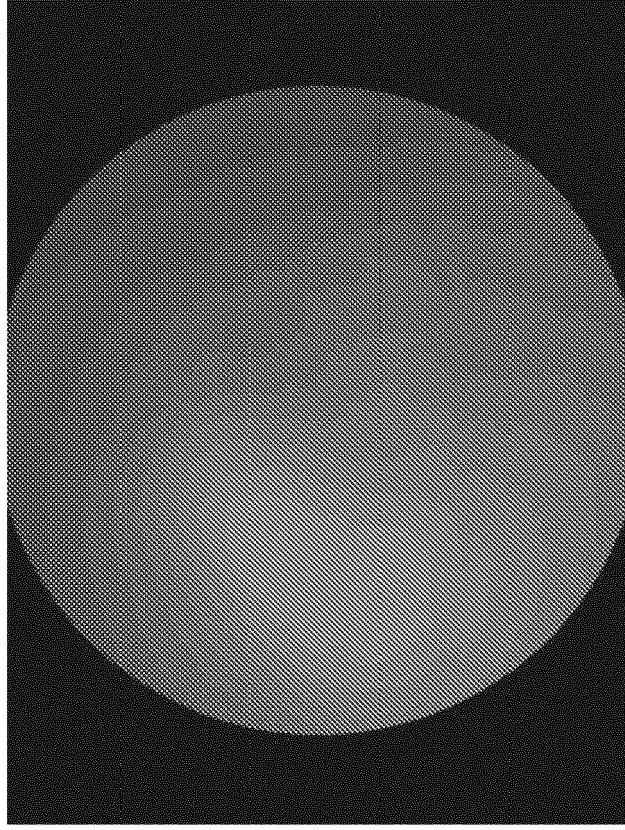


FIG. 4F

AAV2QUAD-GFP (5X)



FIG. 4G

AAV2QUAD-GFP (10X)

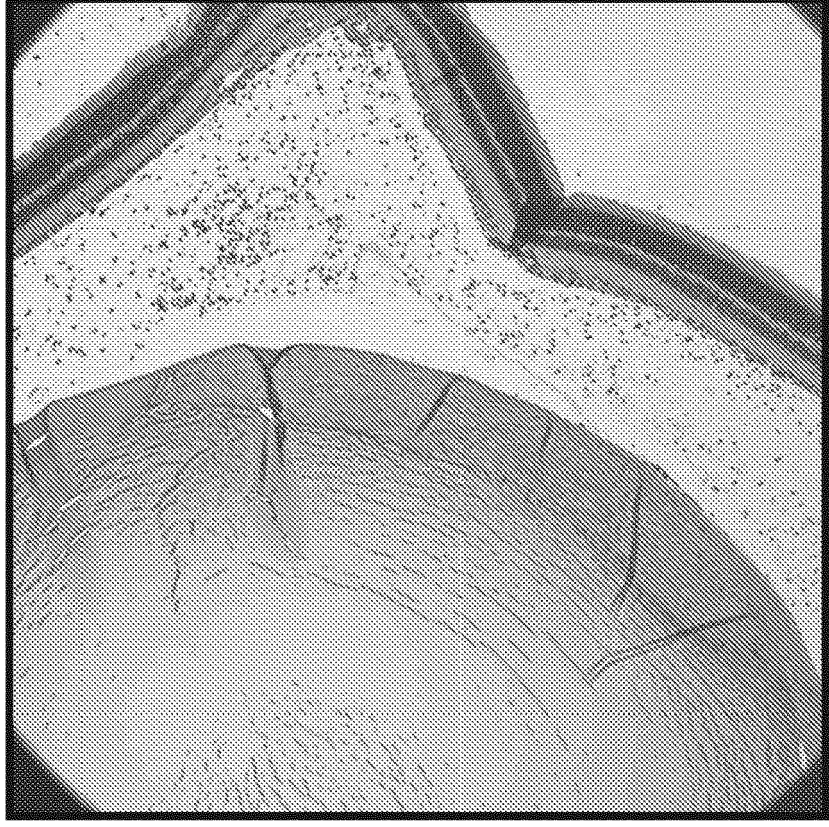


FIG.4H

AAV2QUAD-sGFP-TatM013 (5X)

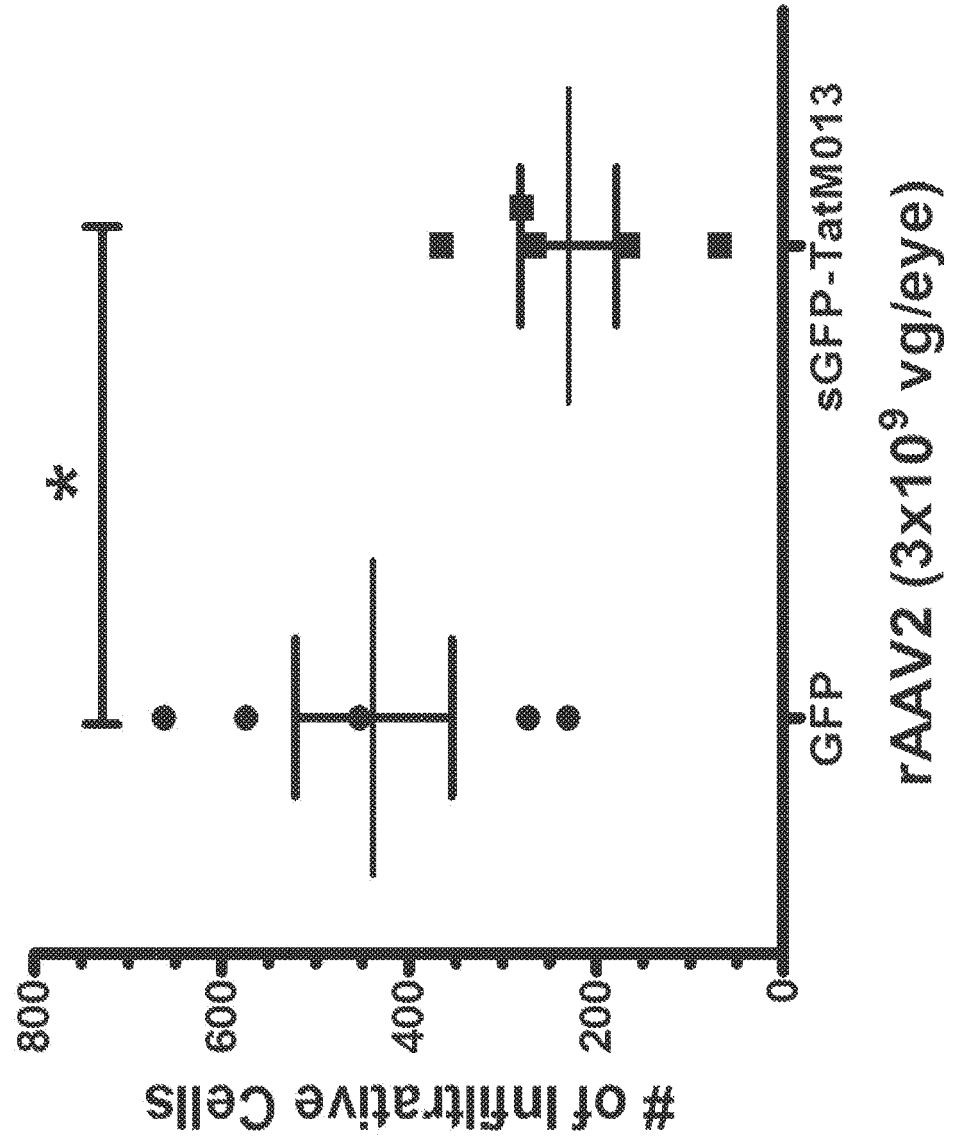


FIG.4I

AAV2QUAD-sGFP-TatM013 (10X)



FIG. 4J





EUROPEAN SEARCH REPORT

Application Number

EP 22 15 2431

DOCUMENTS CONSIDERED TO BE RELEVANT

Category	Citation of document with indication, where appropriate, of relevant passages	Relevant to claim	CLASSIFICATION OF THE APPLICATION (IPC)
Y	WO 02/26780 A2 (BURNHAM INST [US]; REED JOHN C [US]; GODZIK ADAM [US]; CHU ZHI LIANG []) 4 April 2002 (2002-04-04) * page 12, paragraph 3; sequence 14 * * page 49, paragraph 2 * * page 99, paragraph 1 * * page 100, line 12 - line 14 * -----	1-17	INV. C07K14/065 A61P37/00 A61P29/00 A61K38/16 A61K48/00 C07K14/005 C12N7/00 C12N15/86 A61K9/00
Y	US 2007/031410 A1 (HARTON JONATHAN A [US] ET AL) 8 February 2007 (2007-02-08) * paragraphs [0009], [0022], [0032], [0046]; sequence 22 * -----	1-17	
Y	DEBRA J. TAXMAN ET AL: "Inflammasome Inhibition as a Pathogenic Stealth Mechanism", CELL HOST & MICROBE, vol. 8, no. 1, 1 July 2010 (2010-07-01), pages 7-11, XP055390243, NL ISSN: 1931-3128, DOI: 10.1016/j.chom.2010.06.005 * page 8, column 1, paragraph 1; figure 1 * -----	1-17	TECHNICAL FIELDS SEARCHED (IPC) C07K A61P C12N A61K
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The present search report has been drawn up for all claims			
Place of search Munich		Date of completion of the search 12 July 2022	Examiner Zellner, Eveline
CATEGORY OF CITED DOCUMENTS X : particularly relevant if taken alone Y : particularly relevant if combined with another document of the same category A : technological background O : non-written disclosure P : intermediate document		T : theory or principle underlying the invention E : earlier patent document, but published on, or after the filing date D : document cited in the application L : document cited for other reasons ----- & : member of the same patent family, corresponding document	

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EUROPEAN SEARCH REPORT

Application Number

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Munich		12 July 2022	Zellner, Eveline
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