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(54) **BETA-CORONAVIRUS ANTIGEN, PREPARATION METHOD THEREFOR AND USE THEREOF**

(57) The embodiments of the present disclosure relate to antigens of β -coronaviruses, preparation methods and uses thereof. The amino acid sequence of the antigen of the β -coronavirus includes an amino acid sequence arranged in a (A-B)-(A-B) pattern or an amino acid sequence arranged in a (A-B)-C-(A-B) pattern or an amino acid sequence arranged in a (A-B)-(A-B') pattern or an amino acid sequence arranged in a (A-B)-C-(A-B') pattern. The antigen of the β -coronavirus has a single-chain dimer structure. A single-chain dimer expressed according to examples of the present disclosure is stable in content and has excellent immunogenicity as an antigen of a β -coronavirus, and a vaccine prepared by using the single-chain dimer as an antigen of a β -coronavirus can elicit high-titer neutralizing antibodies in mice.

MERS-CoV-RBD dimer (E367-Y606)

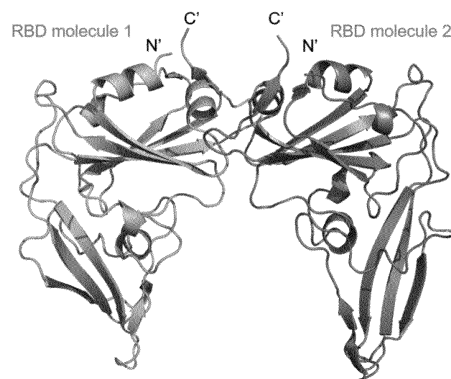


FIG. 8

Description**CROSS-REFERENCE**

[0001] This application claims priority to Chinese Patent Application No. CN202010085038.9 filed with China National Intellectual Property Administration, entitled "ANTIGENS OF β - CORONAVIRUSES, PREPARATION METHODS AND USES THEREOF", which is incorporated herein by reference in its entirety.

TECHNICAL FIELD

[0002] The present disclosure relates to the field of biomedical technology, and in particular, to antigens of β -coronaviruses, preparation methods and uses thereof.

BACKGROUND ART

[0003] Coronaviruses, belonging to the coronavirus genera of the family *Coronaviridae*, are positive-strand enveloped RNA viruses, the genome of which is the largest among all RNA viruses. Both animals and humans can be hosts of coronaviruses. Coronaviruses mainly infect the respiratory tracts and digestive tracts of mammals and birds, and seven kinds of coronaviruses are currently known to infect humans, including four (HCoV-229E, HCoV-NL63, OC43 and HKU1) that may cause a mild cold. Globally, there are three kinds of coronaviruses imposing the greatest threats to public health, namely severe acute respiratory syndrome coronavirus (SARS-CoV) that broke out in 2002-2003, Middle East respiratory syndrome coronavirus (MERS-CoV) that erupted in 2012 and persisted to date, and a novel coronavirus that broke out in 2019 (2019-nCoV), all of which are β -coronaviruses.

[0004] Middle East Respiratory Syndrome (MERS) is a disease caused by Middle East Respiratory Syndrome coronavirus (MERS-CoV) infection. In June 2012, the first MERS case was found in Saudi Arabia, and a novel coronavirus was isolated from a sputum sample of the case. This virus was subsequently named MERS-CoV by the coronavirus group of the International Committee on Taxonomy of Viruses. The virus spread in the Middle East and spread to Asia, Africa, Europe and North America. According to WHO statistics, as of October 6, 2015, there were 1,589 infections and 567 deaths worldwide, with a mortality rate of 35.6% (<http://www.who.int/entity/emergencies/mers-cov/en/>). In particular, the MERS epidemic imported from the Middle East to South Korea in May and June 2015 resulted in 186 infections and 36 deaths. Even one MERS case was imported into China. It brought a serious threat to the global public health system. MERS-CoV virus and SARS virus broke out in 2003 belong to β -*Coronavirus* subgenus, but they have a higher lethality rate than SARS-CoV. MERS-CoV may spread in the form of aerosol, and thus is difficult to prevent and control. Neutralizing antibodies to MERS-CoV can be detected in the serum of dromedary camels in many countries in the Middle East, suggesting that dromedary camel, which is an important vehicle in Middle East countries, is an intermediate host for MERS-CoV. Therefore, the sporadic MERS-CoV infection of humans in the Middle East has happened frequently since the discovery of MERS-CoV in 2012. As a result, with the increasing frequency of international communications, the risk of MERS spreading around the world has always existed. At present, there are still no vaccines and effective treatments in the world. Thus, it is urgent and important to develop a safe and effective vaccine against MERS-CoV.

[0005] In 2019, there was a case of pneumonia of unknown cause, which was identified as a coronavirus by using an electron microscope, and was temporarily named 2019 novel coronavirus (2019-nCoV), and later named SARS-CoV-2. The novel coronavirus can be transmitted from person to person through respiratory tracts and droplets, as well as through the air and digestive tracts. The source of infection is mainly patients infected with the novel coronavirus, but it is not ruled out that the asymptomatic cases are also the source of transmission. The disease may not occur immediately after infection of the virus, and the incubation period of the virus is relatively long, 1-14 days, which makes it difficult to prevent and control the disease. After entering a human body, the novel coronavirus, enters cells through angiotensin converting enzyme 2 (ACE 2) to infect the human body, causing the patient to have clinical symptoms such as fever, dry cough and muscle pain. Besides, a few of patients may have symptoms such as nasal obstruction, pharyngalgia and diarrhea and severe symptoms in some patients may rapidly progress to acute respiratory distress syndrome, septic shock, metabolic acidosis which is difficult to correct, and coagulation dysfunction, causing life danger. There is no specific drug or vaccine for the moment to prevent this virus, and only symptomatic support treatment is available.

[0006] In addition, some other coronaviruses also cause many serious animal diseases, especially posing a serious threat to agricultural livestock and pets. For instance, transmissible gastroenteritis virus (TGEV) can cause severe diarrhea in pigs with extremely high mortality, and its deletion mutant virus porcine respiratory coronavirus (PRCV) can cause severe respiratory diseases in pigs; feline infectious peritonitis virus (FIPV) can cause peritonitis and ascites aggregation in cats with high mortality; canine coronavirus (CCoV) can cause gastroenteritis symptoms in dogs to varying degrees, which spreads quickly and is difficult to control; and porcine epidemic diarrhea virus (PEDV) causes intestinal diseases such as porcine epidemic diarrhea, which is easy to spread in pigs with high mortality rate. There are also

murine, bovine and other coronaviruses. These coronaviruses pose a serious threat to human and animal health. Therefore, it is of great significance to develop vaccines against coronaviruses.

[0007] The surface spike protein (S protein) is the major neutralizing antigen of a coronavirus. The receptor binding domains (RBD) of the spike proteins (S proteins) of MERS-CoV, SARS-CoV and 2019-nCoV are considered as the most important antigen target domains to induce a body to produce neutralizing antibodies. The RBDs, as vaccines, can focus the neutralizing antibodies generated by body stimulation on the receptor binding of viruses, which can improve the immunogenicity and immune efficiency of the vaccines. MERS-CoV invades a cell by RBD binding to the host cell's receptor (CD26, also known as DPP4). In addition, both SARS-CoV and 2019-nCoV were found to enter a cell via their RBD binding to the host cell receptor hACE2.

[0008] The information disclosed herein is merely intended to provide a better understanding of the general background of the present disclosure and should not be construed as an acknowledgement or an implication in any form that the information constitutes the prior art that is already known to a person skilled in the art.

SUMMARY

OBJECTS OF THE DISCLOSURE

[0009] The present disclosure aims to provide antigens of β -coronaviruses, preparation methods and uses thereof. In examples of the present disclosure, based on the conclusion that MERS RBD-dimer protein could better elicit neutralizing antibodies than RBD-monomer protein, it was tried to link two nucleotide sequences encoding the identical or substantially identical RBD-monomer protein in tandem directly or via a linker fragment and to link the two expressed identical or substantially identical RBD-monomer proteins in tandem through the N-terminal and C-terminal flexible regions, and the results showed that the method could realize good expression of a single-chain RBD-dimer. Compared with a non-single-chain RBD-dimer protein formed by simply binding two RBD-monomers through cysteines therein with disulfide bonds, the single-chain RBD-dimer protein obtained in the examples of the present disclosure would not render the content of the RBD-dimer protein unstable in the production process due to unstable formation of the disulfide bonds. That is to say, the expression of most RBD-monomers and few RBD-dimers could be avoided, so that the dimeric RBD could be stably expressed and uniform in form with a greatly improved yield. Compared with the RBD-dimer protein formed by simply binding two RBD monomers through cysteines therein with disulfide bonds, the single-chain dimer expressed in the examples of the present disclosure had equivalent immunogenicity as an antigen of a β -coronavirus, and a vaccine prepared by using the single-chain dimer as the antigen of a β -coronavirus could elicit high-titer neutralizing antibodies in mice.

SOLUTION

[0010] In order to achieve the purpose of the present disclosure, examples of the present disclosure provide the following technical solution:

[0011] An antigen of a β -coronavirus, its amino acid sequence comprises an amino acid sequence arranged in a (A-B)-(A-B) pattern or an amino acid sequence arranged in a (A-B)-C-(A-B) pattern or an amino acid sequence arranged in a (A-B)-(A-B') pattern or an amino acid sequence arranged in a (A-B)-C-(A-B') pattern, where A-B represents a partial amino acid sequence or the entire amino acid sequence of a receptor binding domain of a surface spike protein of the β -coronavirus; C represents an amino acid linker sequence; and A-B' represents an amino acid sequence obtained by substitution, deletion or addition of one or more amino acids in the amino acid sequence of A-B. A protein encoded by A-B' has the identical or substantially identical immunogenicity as a protein encoded by A-B, and the antigen of the β -coronavirus has a single-chain dimer structure. Alternatively, the partial amino acid sequence of the receptor binding domain of the surface spike protein of the β -coronavirus is at least 50%, 60%, 70%, 80%, 90%, 95%, or 99% of the entire amino acid sequence of the receptor binding domain of the surface spike protein of the β -coronavirus.

[0012] In one possible embodiment of the above-mentioned antigen of a β -coronavirus, the β -coronavirus includes severe respiratory syndrome coronavirus, Middle East respiratory syndrome coronavirus, and 2019 novel coronavirus (also known as 2019-nCoV or SARS-CoV-2).

[0013] In one possible embodiment of the above-mentioned antigen of a β -coronavirus, the amino acid linker sequence includes a (GGG)_n linker sequence, where n represents the number of GGGs, which is an integer more than or equal to 1; alternatively, n is an integer selected from 1 to 10, and further, an integer selected from 1 to 5; and GGS represents amino acids G, G and S.

[0014] In one possible embodiment of the above-mentioned antigen of a β -coronavirus, when the β -coronavirus is the Middle East respiratory syndrome coronavirus, the partial or entire amino acid sequence of the receptor binding domain of the surface spike protein of the β -coronavirus is any one selected from the group consisting of the following amino acid sequences:

- (1) SEQ ID NO: 1, SEQ ID NO: 2, or SEQ ID NO: 3; and
 (2) an amino acid sequence obtained by substitution, deletion or addition of one or more amino acids in the amino acid sequence (1), where a protein encoded by the amino acid sequence has the identical or substantially identical immunogenicity as a protein encoded by the amino acid sequence (1).

[0015] Alternatively, the partial amino acid sequence of the receptor binding domain of the surface spike protein of the β -coronavirus includes SEQ ID NO: 2.

[0016] The sequences of SEQ ID NO: 1, SEQ ID NO: 2, and SEQ ID NO: 3 are all derived from a part of the MERS-CoV S protein (GenBank: AFS88936.1 on NCBI), which are E367-Y606 region, E367-N602 region, and V381-L588 region of the RBD of the MERS-CoV S protein, respectively.

[0017] In one possible embodiment of the above-mentioned antigen of a β -coronavirus, when the β -coronavirus is the Middle East respiratory syndrome coronavirus, the amino acid sequence of the antigen of the β -coronavirus includes any one selected from the group consisting of the following amino acid sequences:

- (1) two repeated amino acid sequences of SEQ ID NO: 1 linked in tandem by a GGSGGS linker sequence, namely E367-Y606-GGSGGS-E367-Y606;
- (2) two repeated amino acid sequences of SEQ ID NO: 1 linked in tandem by a GGS linker sequence, namely E367-Y606-GGS-E367-Y606;
- (3) two repeated amino acid sequences of SEQ ID NO: 1 linked directly in tandem, namely E367-Y606-E367-Y606.
- (4) two repeated amino acid sequences of SEQ ID NO: 2 linked in tandem by a GGS linker sequence, namely E367-N602-GGS-E367-N602;
- (5) two repeated amino acid sequences of SEQ ID NO: 2 linked directly in tandem by a GGS linker sequence, namely E367-N602-E367-N602;
- (6) two repeated amino acid sequences of SEQ ID NO: 3 linked by a GGSGGSGGSGGS linker sequence, namely V381-L588-GGSGGSGGSGGSGGSGGSGGS-V381-L588;
- (7) two repeated amino acid sequences of SEQ ID NO: 3 linked in tandem by a GGSGGSGGSGGS linker sequence, namely V381-L588-GGSGGSGGSGGSGGS-V381-L588;
- (8) two repeated amino acid sequences of SEQ ID NO: 3 linked in tandem by a GGSGGSGGS linker sequence, namely V381-L588-GGSGGSGGS-V381-L588;
- (9) two repeated amino acid sequences of SEQ ID NO: 3 linked in tandem by a GGS linker sequence, namely V381-L588-GGS-V381-L588; and
- (10) two repeated amino acid sequence of SEQ ID NO: 3 linked directly in tandem, namely V381-L588-V381-L588;

[0018] Alternatively, the amino acid sequence of the antigen of the β -coronavirus includes two repeated amino acid sequences of SEQ ID NO: 2 linked directly in tandem, namely E367-N602-E367-N602.

[0019] In one possible embodiment of the above-mentioned antigen of the β -coronavirus, when the β -coronavirus is the 2019 novel coronavirus, the partial or entire amino acid sequence of the receptor binding domain of the surface spike protein of the β -coronavirus is any one selected from the group consisting of the following amino acid sequences:

- (1) SEQ ID NO: 5, SEQ ID NO: 6, or SEQ ID NO: 7; and
- (2) an amino acid sequence obtained by substitution, deletion or addition of one or more amino acids to the amino acid sequence (1), where the protein encoded by the amino acid sequence has the identical or substantially identical immunogenicity as the protein encoded by (1).

[0020] Alternatively, the partial amino acid sequence of the receptor binding domain of the surface spike protein of the β -coronavirus includes SEQ ID NO: 6.

[0021] The sequences of SEQ ID NO: 5, SEQ ID NO: 6, and SEQ ID NO: 7 are all derived from a part of the S protein sequence of the WH01 strain of 2019-nCoV (GenBank on NCBI: QHR63250), which are R319-S530 region, R319-K537 region, and R319-F541 region of the RBD of the 2019-nCoV S protein, respectively.

[0022] In one possible embodiment of the above-mentioned antigen of a β -coronavirus, when the β -coronavirus is the 2019 novel coronavirus, the amino acid sequence of the antigen of β -coronavirus includes any one selected from the group consisting of the following amino acid sequences:

- two repeated amino acid sequences of SEQ ID NO: 5 linked directly in tandem, namely R319-S530-R319-S530;
- two repeated amino acid sequences of SEQ ID NO: 6 linked directly in tandem, namely R319-K537-R319-K537; and
- two repeated amino acid sequences of SEQ ID NO: 7 linked directly in tandem, namely R319-F541-R319-F541.

[0023] Alternatively, the amino acid sequence of the antigen of the β -coronavirus includes two repeated amino acid

sequences of SEQ ID NO: 6 linked directly in tandem, namely R319-K537-R319-K537.

[0024] In one possible embodiment of the above-mentioned antigen of a β -coronavirus, when the β -coronavirus is the severe respiratory syndrome coronavirus, the partial or entire amino acid sequence of the receptor binding domain of the surface spike protein of the β -coronavirus is any one selected from the group consisting of the following amino acid sequences:

- (1) SEQ ID NO: 8; and
- (2) an amino acid sequence obtained by substitution, deletion or addition of one or more amino acids in the amino acid sequence (1), where a protein encoded by the amino acid sequence has the identical or substantially identical immunogenicity as a protein encoded by the amino acid sequence (1).

[0025] The sequence of SEQ ID NO: 8 is derived from a part of the S protein sequence of SARS-CoV (GenBank on NCBI: AAR07630), which is R306-Q523 region of the RBD of the SARS-CoV S protein.

[0026] In one possible embodiment of the above-mentioned antigen of a β -coronavirus, when the β -coronavirus is the severe respiratory syndrome coronavirus, the amino acid sequence of the antigen of the β -coronavirus includes two repeated amino acid sequences of SEQ ID NO: 8 linked directly in tandem, namely R306-Q523- R306-Q523.

[0027] In one possible embodiment of the above-mentioned antigen of a β -coronavirus, the nucleotide sequence encoding two repeated amino acid sequences of SEQ ID NO: 1 linked in tandem by the GGSGGS linker sequence is shown as SEQ ID NO: 9;

the nucleotide sequence encoding two repeated amino acid sequence of SEQ ID NO: 1 linked in tandem by the GGS linker sequence is shown as SEQ ID NO: 10;

the nucleotide sequence encoding two repeated amino acid sequence of SEQ ID NO: 1 linked directly in tandem is shown as SEQ ID NO: 11;

the nucleotide sequence encoding two repeated amino acid sequence of SEQ ID NO: 2 linked in tandem by the GGS linker sequence is shown as SEQ ID NO: 12;

the nucleotide sequence encoding two repeated amino acid sequence of SEQ ID NO: 2 linked directly in tandem is shown as SEQ ID NO: 13;

the nucleotide sequence encoding two repeated amino acid sequence of SEQ ID NO: 3 linked in tandem by the GGSGGSGGSGGSGGS linker sequence is shown as SEQ ID NO: 14;

the nucleotide sequence encoding two repeated amino acid sequence of SEQ ID NO: 3 linked in tandem by the GGSGGSGGSGGS linker sequence is shown as SEQ ID NO: 15;

the nucleotide sequence encoding two repeated amino acid sequence of SEQ ID NO: 3 linked in tandem by the GGSGGSGGS linker sequence is shown as SEQ ID NO: 16;

the nucleotide sequence encoding two repeated amino acid sequence of SEQ ID NO: 3 linked in tandem by the GGS linker sequence is shown as SEQ ID NO: 17;

the nucleotide sequence encoding two repeated amino acid sequence of SEQ ID NO: 3 linked directly in tandem is shown as SEQ ID NO: 18;

the nucleotide sequence encoding two repeated amino acid sequence of SEQ ID NO: 5 linked directly in tandem is shown as SEQ ID NO: 19;

the nucleotide sequence encoding two repeated amino acid sequence of SEQ ID NO: 6 linked directly in tandem is shown as SEQ ID NO: 20;

the nucleotide sequence encoding two repeated amino acid sequence of SEQ ID NO: 7 linked directly in tandem is shown as SEQ ID NO: 21; and

the nucleotide sequence encoding two repeated amino acid sequence of SEQ ID NO: 8 linked directly in tandem is shown as SEQ ID NO: 23.

[0028] The present disclosure also provides a method for preparing the above-mentioned antigen of a β -coronavirus, which includes the following steps: adding a sequence encoding a signal peptide to the 5'-terminal of a nucleotide sequence encoding the antigen of the β -coronavirus, adding a terminator codon to the 3'-terminal for cloning and expression, screening correct recombinants, transfecting the expression system cell for expression, collecting cell supernatants after expression, and purifying to obtain the antigen of the β -coronavirus.

[0029] In one possible embodiment of the above method, the cells of the expression system include mammalian cells, insect cells, yeast cells or bacterial cells. Alternatively, the mammalian cells include 293T cells or CHO cells, and the bacterial cells include *Escherichia coli* cells.

[0030] The present disclosure further provides a nucleotide sequence for encoding the above-mentioned antigen of a β -coronavirus, a recombinant vector including the nucleotide sequence, and an expression system cell including the recombinant vector.

[0031] The present disclosure further provides use of the above-mentioned antigen of a β -coronavirus, the nucleotide sequence encoding the antigen of the β -coronavirus, the recombinant vector including the nucleotide sequence, and the expression system cell including the recombinant vector in the preparation of a vaccine against the β -coronavirus.

[0032] The present disclosure further provides a vaccine against a β -coronavirus, which includes the above-mentioned antigen of a β -coronavirus and an adjuvant.

[0033] In one possible embodiment of the above-mentioned vaccine against a β -coronavirus, the adjuvant is selected from an aluminum adjuvant, an MF59 adjuvant or an MF59-like adjuvant. The present disclosure further provides a DNA vaccine against a β -coronavirus, which includes a recombinant vector including a DNA sequence encoding the above-mentioned antigen of a β -coronavirus.

The present disclosure further provides an mRNA vaccine against a β -coronavirus, which includes a recombinant vector including an mRNA sequence encoding the above-mentioned antigen of a β -coronavirus.

The present disclosure further provides a viral vector vaccine against a β -coronavirus, which includes a recombinant viral vector including a nucleotide sequence encoding the above-mentioned antigen of a β -coronavirus. Alternatively, the viral vector is one or more selected from the group consisting of an adenovirus vector, a poxvirus vector, an influenza virus vector and an adeno-associated virus vector.

Beneficial effects

[0034]

(1) In the antigen of a β -coronavirus of an example of the present disclosure, based on the conclusion that MERS RBD-dimer protein could better elicit neutralizing antibodies than RBD-monomer protein, it was found that the MERS RBD-dimer protein could form an end-to-end dimer by further analyzing the crystal structure of the MERS-CoV RBD-dimer protein. Therefore, the inventor tried to link two nucleotide sequences encoding the identical or substantially identical RBD-monomer proteins directly in tandem or via a linker fragment and to link two obtained identical or substantially identical RBD-monomer proteins in tandem through flexible regions at the N-terminal and C-terminal, and the results showed that the method could realize good expression of a single-chain dimer. Compared with a non-single-chain RBD-dimer protein formed by simply binding two RBD monomers through cysteines therein with disulfide bonds, the single-chain RBD-dimer protein obtained in the example of the present disclosure would not render the content of the RBD-dimer protein unstable in the production process due to unstable formation of the disulfide bonds. That is to say, the expression of most RBD-monomers and few RBD-dimers could be avoided, so that the RBD-dimer could be stably expressed and uniform in form with a greatly improved yield. Compared with the non-single-chain RBD-dimer protein formed by simply binding two RBD monomers through cysteines therein with disulfide bonds, the single-chain dimer expressed in the example of the present disclosure had equivalent immunogenicity as an antigen of a β -coronavirus, and a vaccine prepared by using the single-chain dimer as the antigen of a β -coronavirus could elicit high-titer neutralizing antibodies in mice.

(2) In the antigen of a β -coronavirus of an example of the present disclosure, based on the selection of amino acids in different regions of the contained RBD, the construct with the best expression was found from the first amino acid of START shown in FIG.14A to the amino acid before the last cysteine of STOP shown in FIG.14B, so that the influence of unpaired cysteines at the ends on the expression and the stability of the protein could be avoided to the greatest extent.

(3) In the antigen of a β -coronavirus of an example of the present disclosure, based on the selection of direct tandem connection or linker fragment-involved tandem connection of two nucleotide sequences encoding the identical or substantially identical RBD-monomer proteins, the highest expression level was found under the condition that no any exogenous linker sequence was introduced, i.e., two nucleotide sequences encoding the identical or substantially identical RBD-monomer proteins were linked directly in tandem, and the expression was also the safest because no exogenous sequence was added. Since various single-chain RBD-dimers obtained in the examples of the present disclosure had good immune effect as antigens of β -coronaviruses, the yield thereof would be crucial.

(4) In the antigen of a β -coronavirus of an example of the present disclosure, the involved end-to-end single-chain dimer structure is suitable for severe respiratory syndrome coronavirus, Middle East respiratory syndrome coronavirus and 2019 novel coronavirus.

BRIEF DESCRIPTION OF THE DRAWINGS

[0035]

FIG. 1 shows an ultraviolet absorption profile for the RBD protein obtained by means of a constructed vector pFastBac-SP-MERS-RBD (E367-Y606) in Example 1 subjected to molecular sieve chromatography using a Superdex 200

Hiload 16/60 column (GE), and an electrophoretogram for the protein subjected to SDS-PAGE under reduced conditions (+DTT) or non-reduced conditions (-DTT) obtained by collecting Dimer peaks and Monomer peaks in the ultraviolet absorption profile.

FIG. 2 is a schematic diagram of the immunization and MERS-CoV challenge strategies in Examples 2 to 7.

FIG. 3 shows the results of Example 3, i.e., the titers of MERS-CoV RBD specific IgG antibody in sera collected from mice according to the immunization strategy in FIG. 2 after the third immunization in Example 2, where Dimer indicates that MERS-CoV RBD-dimer was used as the immunogen; RBD-monomer indicates MERS-CoV RBD-monomer was used as the immunogen; AddaVax indicates the use of AddaVax adjuvant; Alum indicates the use of aluminum adjuvant; no indication of adjuvant means no use of adjuvant; and 3 μ g, 10 μ g, and 30 μ g indicate the amounts of the immunogen used per immunization. Significant difference analysis: ns, $P > 0.05$; *, $P < 0.05$; **, $P < 0.01$; ***, $p < 0.001$; ****, $P < 0.0001$.

FIG. 4 shows the results of Example 5, i.e., 90% neutralization titers antibodies against MERS-CoV pseudovirus in sera collected from mice according to the immunization strategy in FIG. 2 after the third immunization in Example 2, where Dimer indicates that MERS-CoV RBD dimer was used as the immunogen; RBD monomer indicates that MERS-CoV RBD monomer was used as the immunogen; AddaVax indicates the use of AddaVax adjuvant; Alum indicates the use of aluminum adjuvant; no indication of adjuvant means no use of adjuvant; and 3 μ g, 10 μ g, and 30 μ g indicate the amounts of the immunogen used per immunization. Significant difference analysis: ns, $P > 0.05$; ***, $p < 0.001$.

FIG. 5 shows the results of Example 6, i.e., 50% neutralization titers of antibodies against MERS-CoV euvirus (EMC strain) in sera collected from mice according to the immunization strategy in FIG. 2 after the third immunization in Example 2, where Dimer indicates that MERS-CoV RBD dimer was used as the immunogen; AddaVax indicates the use of AddaVax adjuvant; Alum indicates the use of aluminum adjuvant; no indication of adjuvant means no use of adjuvant; and 3 μ g, 10 μ g, and 30 μ g indicate the amounts of the immunogen used per immunization. Significant difference analysis: ns, $P > 0.05$; ***, $p < 0.001$; ****, $P < 0.0001$.

FIG. 6 shows the results of Example 7 in which mice after the third immunization were subjected to intranasal infection with adenovirus expressing hCD26 (hDPP4) according to the immunization strategy in FIG. 2, and were challenged with MERS-CoV 5 days later, followed by the detection of viral titers (TCID₅₀) on the tissue homogenates prepared from the lungs of the mice removed 3 days later. Dimer indicates that MERS-CoV RBD dimer was used as the immunogen; AddaVax indicates the use of AddaVax adjuvant; Alum indicates the use of aluminum adjuvant; no indication of adjuvant means no use of adjuvant; and 3 μ g, 10 μ g, and 30 μ g indicate the amounts of the immunogen used per immunization. Significant difference analysis: ns, $P > 0.05$; *, $P < 0.05$; **, $P < 0.01$; ***, $p < 0.001$; ****, $P < 0.0001$.

FIG. 7 shows the pathological results of the examination of the protective efficacy of the vaccine on the lung tissue of mice in Example 8 in which the lungs from the challenged mice in Example 7 after necroscopy were fixed in 4% paraformaldehyde, embedded in paraffin, stained with hematoxylin and eosin and sliced to obtain tissue sections for observation of pathological changes, where AddaVax indicates the use of AddaVax adjuvant; Alum indicates the use of aluminum adjuvant; and 3 μ g, 10 μ g, and 30 μ g indicate the amounts of the immunogen used per immunization. Slight, Mild and Severe indicate different grades of lung tissue lesions, respectively.

FIG. 8 shows the structure of MERS-CoV-RBD dimer (E367-Y606) analyzed in Example 9.

FIGS. 9A, 9B, and 9C show single-chain RBD dimers designed based on the MERS-CoV RBD-dimer structure in Example 10.

FIG. 10 shows the results of Western blot conducted on MERS-RBD-C1 to MERS-RBD-C10 single-chain dimers expressed in Example 10 under reduced conditions (+DTT) or non-reduced conditions (-DTT), where RBD Monomer is MERS-CoV RBD Monomer protein.

FIG. 11 shows an ultraviolet absorption profile for MERS-RBD-C5 single-chain dimer expressed in Example 11 subjected to molecular sieve chromatography using a Superdex 200 Hiload 16/60 column (GE), and the results of SDS-PAGE of the purified single-chain dimer under reduced conditions (+DTT) or non-reduced conditions (-DTT).

FIG. 12 shows the titers of MERS-CoV-RBD specific IgG antibody elicited by immunization of mice in Example 12 with single-chain MERS-CoV-RBD dimer and disulfide-linked non-single-chain dimer proteins, where sc-dimer is a single-chain dimer, and Dimer is a disulfide-linked non-single-chain dimer. Significant difference analysis: ns, $P > 0.05$; *, $P < 0.05$; ***, $p < 0.001$; ****, $P < 0.0001$.

FIG. 13 shows the 90% neutralization titers of antibodies against MERS-CoV pseudovirus elicited by immunization of mice with single-chain MERS-CoV-RBD dimer and disulfide-linked non-single-chain dimer proteins in Example 12, where sc-dimer is a single-chain dimer, and Dimer is a disulfide-linked non-single-chain dimer. Significant difference analysis: ns, $P > 0.05$; *, $P < 0.05$; ****, $P < 0.0001$.

FIGS. 14A and 14B are comparison diagrams of the receptor binding domains (RBDs) of β -coronaviruses in Example 13, where the sequences in the two figures were consecutive, and the following β -coronaviruses were aligned: MERS-CoV (AFS88936), SARS-CoV (AAS00003), SARS-CoV-2 (QHR63290), Bat-CoV_HKU5 (ABN10875),

Rousettus_bat-CoV (AOG30822), Bat-CoV_BM48-31 (ADK66841), Bat-CoV_HKU9 (ABN10911), Bat_Hp-betaCoV (AIL94216), SARS-related-CoV (APO40579), BtRs-Beta-CoV (QDF43825), Bat-SARS-like-CoV (AT098231), SARS-like-CoV_WIV16 (ALK02457), Bat-CoV (ARI44804), BtR1-Beta-CoV (QDF43815), HCoV_HKU1 (AZS52618), MCoV_MHV1 (ACN89742), BetaCoV_HKU24 (AJA91217), HCoV_OC43 (AAR01015), and BetaCoV_Erinaceus (AGX27810).

FIG. 15 is a mimic diagram showing the structure of SARS-CoV-RBD dimer or 2019-nCoV-RBD dimer in example 13 and the construct of the expression 2019-nCoV-RBD dimer, the 2019-nCoV-RBD monomer and the SARS-CoV-RBD dimer designed.

FIG. 16 shows the results of Western blot under reduced conditions (+DTT) or non-reduced conditions (-DTT) for several single-chain dimers of SARS-CoV-RBD and 2019-nCoV-RBD expressed in Example 13.

FIG. 17 shows the ultraviolet absorbance at 280 nm for 2019-nCoV-RBD-C2 antigen purified in Example 14, and the results of SDS-PAGE of the purified single-chain dimer under reduced conditions (+DTT) or non-reduced conditions (-DTT).

FIG. 18 shows the ultraviolet absorbance at 280 nm for SARS-CoV-RBD-C1 antigen purified in Example 14, and the results of SDS-PAGE of the purified single-chain dimer under reduced conditions (+DTT) or non-reduced conditions (-DTT).

FIG. 19 shows the titers of 2019-nCoV-RBD specific IgG antibody in sera collected from mice after three immunizations (19 days after the first immunization, 14 days after the second immunization, and 14 days after the third immunization) in Example 15, respectively, where sc-dimer indicates that single-chain nCoV-RBD dimer was used as the immunogen, and Monomer indicates that nCoV-RBD-monomer was used as the immunogen. Significant difference analysis: ****, $P < 0.0001$.

FIG. 20 shows the 90% neutralization titers of antibodies against 2019-nCoV pseudovirus in sera collected from mice after three immunizations (19 days after the first immunization, 14 days after the second immunization, and 14 days after the third immunization) in Example 15, where sc-dimer indicates that single-chain nCoV-RBD dimer was used as the immunogen, and Monomer indicates that nCoV-RBD-monomer was used as the immunogen. Significant difference analysis: ns, $P > 0.05$; **, $P < 0.01$; ****, $P < 0.0001$.

FIG. 21 shows the 50% neutralization titers of antibodies against 2019-nCoV euvirus (2020XN4276 strain) in sera collected after the second immunization (14 days after the second immunization) of mice in Example 15, where sc-dimer indicates that single-chain nCoV-RBD dimer was used as the immunogen, and Monomer indicates that nCoV-RBD-monomer was used as the immunogen.

FIG. 22 shows the titers of SARS-RBD-specific IgG antibody in sera collected after three immunizations of mice in Example 16 (19 days after the first immunization, 14 days after the second immunization, and 14 days after the third immunization), where sc-dimer indicates that single-chain SARS-CoV-RBD dimer was used as the immunogen, and Monomer indicates that SARS-CoV-RBD-monomer was used as the immunogen. Significant difference analysis: ns, $P > 0.05$; *, $P < 0.05$; **, $P < 0.01$; ****, $P < 0.0001$.

FIG. 23 shows the 90% neutralization titers of antibodies against SARS-CoV pseudovirus in sera collected from mice after three immunizations (19 days after the first immunization, 14 days after the second immunization, and 14 days after the third immunization) in Example 16, where sc-dimer indicates that single-chain SARS-CoV-RBD dimer was used as the immunogen, and Monomer indicates that SARS-RBD-monomer was used as the immunogen. Significant difference analysis: ns, $P > 0.05$; *, $P < 0.05$; **, $P < 0.01$; ***, $p < 0.001$; ****, $P < 0.0001$.

DETAILED DESCRIPTION OF THE PRESENT DISCLOSURE

[0036] In order to make the objects, technical solutions and advantages of the examples of the present disclosure clearer, the technical solutions in the examples of the present disclosure will be clearly and completely described below. Apparently, the described examples are some, but not all examples of the present disclosure. All other examples derived from the examples of the present disclosure by a person skilled in the art without creative work shall fall within the scope of protection of the present disclosure.

[0037] Furthermore, in the following detailed description, numerous specific details are set forth in order to provide a better understanding of the present disclosure. It will be understood by those skilled in the art that the present disclosure may be practiced without some of these specific details. In some examples, materials, elements, methods, procedures, and the like that are well known to those of skill in the art have not been described in detail so as not to obscure the present disclosure.

[0038] Throughout the specification and claims, unless expressly indicated otherwise, the terms "comprise" or "include", or variations such as "comprises" or "comprising", "includes" or "including" will be understood to imply the inclusion of a stated element or component but not the exclusion of any other element or component.

Explanation of Terms

[0039] Disulfide-linked non-single-chain RBD dimer and RBD monomer refer to those obtained by inserting a nucleotide sequence encoding RBD monomer into a vector, then transfecting cells of an expression system for expression, collecting cell supernatants after expression, and purifying, where two RBD monomers in a disulfide-linked non-single-chain RBD dimer are simply disulfide-bonded through cysteines therein. As used herein, disulfide-linked non-single-chain RBD dimer and non-single-chain RBD-dimer protein have the same meaning; and RBD monomer, monomeric RBD, and RBD-monomer protein all have the same meaning.

Single-chain RBD dimer is a recombinant protein obtained by linking two nucleotide sequences encoding identical or substantially identical RBD monomers in tandem directly or via a linker fragment, adding a sequence encoding a signal peptide to the 5'-terminal of the nucleotide sequence and a terminator codon to the 3'-terminal of the nucleotide sequence for cloning and expression, screening correct recombinants, transfecting cells of an expression system cell for expression, collecting cell supernatants after expression, and purifying, where the protein contains two RBD monomers which are identical or substantially identical and can be directly linked together with peptide bonds or linked together through a linker sequence (such as GGS, GGSGGS and the like). As used herein, single-chain RBD-dimer, single-chain RBD dimer, single-chain dimer, sc-RBD dimer, single-chain RBD dimer and the like all have the same meaning.

EXAMPLE 1: Preparation of Recombinant Baculovirus Expressing MERS-CoV Antigen, and Expression and Purification of RBD Protein

[0040] A nucleotide sequence (shown as SEQ ID NO: 24) encoding an amino acid RBD (E367-Y606) sequence (shown as SEQ ID NO: 1) in MERS-CoV S protein (having a sequence shown as GenBank: AFS88936.1) was cloned between EcoRI and XhoI restriction enzyme cutting sites of a pFastBac vector (pFastBac-SP, available from Invitrogen) containing gp67 signal peptide after the addition of a translation termination codon to the 3'-terminal thereof, so that the protein encoding region was subjected to fusion expression behind the signal peptide gp67 sequence for secretion of the protein of interest having 6 histidines at the C-terminal thereof, thereby obtaining a vector pFastBac-SP-MERS-RBD (E367-Y606). The vector was then transfected into the cells of the expression system for expression, and after expression, cell supernatants were collected and purified.

[0041] The obtained RBD protein was purified through molecular sieve chromatography using a Superdex 200 Hiload 16/60 column (GE), and a typical ultraviolet absorption profile for protein purification is shown in FIG. 1. There was one dimer peak and one monomer peak. SDS-PAGE was conducted on the elution peak of MERS-RBD protein in the vicinity of the elution volume of 78 mL. Under non-reduced conditions (without DTT), the size of the protein in the vicinity of the elution volume of 78 mL was approximately 60 Kd; whereas under reduced conditions (with DTT added), the size was approximately 30 Kd, which confirmed that the protein obtained in this peak was a dimer. SDS-PAGE was conducted on the elution peak in the vicinity of 90 mL of the elution volume, the size of the protein of interest was approximately 30 Kd under non-reduced conditions (without DTT) and reduced conditions, which confirmed that the peak was mainly RBD monomer. The dimer or monomer used in each of Examples 2 to 9 below was the disulfide-linked non-single-chain RBD dimer or RBD monomer obtained in this Example.

EXAMPLE 2 Experiment for Immunization of Mice with MERS-RBD Protein

[0042] MF59 (AddaVax used below was an MF59-like adjuvant) and aluminum adjuvant two commonly used adjuvants approved by SFDA, were used as vaccine components to provide more direct guidance for subsequent clinical trials. An in vitro neutralization experiment, as a classic method, was conducted to detect the protective efficacy of vaccines. Therefore, different doses of antigen were mixed with AddaVax adjuvant and Imject™ Alum adjuvant separately for immunization. The immunization groups, the types of RBD used in each group, the amount of RBD used in each immunization and the adjuvants are shown in Table 1 in which the blank space indicates "None".

[0043] MERS-RBD antigen (dimer or monomer) obtained in Example 1 was diluted with normal saline to a desired concentration and emulsified with adjuvants in groups. BALB/c mice aged 4-6 weeks (average weight 15-20 g, similarly hereinafter) were immunized in groups, with 6 mice in each group.

Table 1

Group	Immunogen	Forms	Dose	Adjuvant
1	RBD	Dimer	3 μ g	Alum
2	RBD	Dimer	10 μ g	Alum
3	RBD	Dimer	30 μ g	Alum

(continued)

Group	Immunogen	Forms	Dose	Adjuvant
4	RBD	Dimer	3 μ g	AddaVax
5	RBD	Dimer	10 μ g	AddaVax
6	RBD	Dimer	30 μ g	AddaVax
7	RBD	Monomer	3 μ g	Alum
8	RBD	Monomer	10 μ g	Alum
9	RBD	Monomer	30 μ g	Alum
10	RBD	Monomer	3 μ g	AddaVax
11	RBD	Monomer	10 μ g	AddaVax
12	RBD	Monomer	30 μ g	AddaVax
13	PBS	-		
14	PBS	-		Alum
15	PBS	-		Addavax

[0044] The immunization was conducted according to a strategy as shown in FIG. 2, i.e., by means of intramuscular injection into the thigh, each mouse received three immunizations with vaccine at day 0, day 21 and day 42, respectively, each time in a vaccination volume of 100 μ l. On the day 56 (namely, the 14th day after the third immunization), blood was collected from the tails of mice. Mouse sera were obtained by centrifugation at 3000 rpm for 10 minutes after standing, and stored at -20° C in a refrigerator for specific antibody titer assay and pseudovirus neutralization assay.

EXAMPLE 3 ELISA Assay of Vaccine-Elicited Specific Antibody Titer

[0045]

(1) The RBD-monomer protein of MERS-CoV was diluted to 3 μ g/ml with an ELISA coating solution (Solarbio, C1050), and 100 μ l of the resulting solution was added to each well of a 96-well ELISA plate (Corning, 3590) and placed at 4°C for 12 hours.

(2) The coating solution was removed, and then PBS was added to wash once. 5% skim milk prepared with PBS was added to a 96-well plate in an amount of 100 μ l per well as a blocking solution for blocking and placed at room temperature for 1 hour. After the completion of blocking, the plate was washed once with PBS solution.

(3) Mouse serum was diluted during blocking. Serum samples were also diluted with the blocking solution. Serum samples were diluted from 20-fold. Then 100 μ l of serum was added to each well of the ELISA plate, while the blocking solution was added for the negative control, incubated at 37°C for 2 hours, and then washed with PBST for 4 times.

(4) Goat anti-mouse IgG-HRP antibody (Abcam, ab6789) diluted 1:2000 with the blocking solution was added and incubated at 37°C for 1.5 hours, and then washed with PBST for 5-6 times. Plates were developed with TMB substrate, which was followed by stopping the reactions with 2 M hydrochloric acid for a proper time, and the absorbance was measured at 450 nm using a microplate reader. Antibody titer values were defined as the highest dilution of serum with a response value greater than 2.5 times the negative control value. The titer of a sample was defined as half of the lowest dilution (limit of detection) at which the response value was still less than 2.5-fold background value, namely, 1: 10.

[0046] As shown in FIG. 3, significantly different levels of antibodies were elicited for the RBD dimer group and monomer group at doses of 3 μ g and 10 μ g with the AddaVax adjuvant, and significantly different levels of antibodies were elicited for the two groups at doses of 3 μ g, 10 μ g and 30 μ g with the aluminum adjuvant, and the dimer group elicited higher levels of antibodies, indicating that the dimeric RBD antigen had a significantly higher ability to activate the antibody response in mice than the RBD monomer vaccine.

[0047] The RBD-monomer protein of MERS-CoV was used as the coating protein in all ELISA assays in the examples of the present disclosure.

EXAMPLE 4 Preparation of MERS-CoV Pseudovirus

PNL43-Luci Pseudovirus Packaging

[0048]

(1) Cell plating: on the day before transfection, 293T cells grown in logarithmic phase were harvested by trypsinization, counted, reseeded and cultured overnight in a 10 cm petri dish, and transfected (without antibiotics) when the confluence of the cells reached 70-90% over 18-24 hours.

(2) Plasmid co-transfection by a PEI method: a total of 20 μ g of plasmid (10 μ g of HIV pNL4-3.Luc.RE (Invitrogen) and 10 μ g of pCAGGS-MERS-S which was obtained by inserting a DNA sequence encoding MERS Spike protein (M1-H1352) into EcoRI and XhoI sites of pCAGGS vector) and 40 μ L of PEI (2 mg/ml) were dissolved in normal saline or HBS separately, to a final volume of 500 μ L, and mixed evenly. After standing for 5 minutes, the two solutions were mixed, followed by standing for 20 minutes. The mixture was then added dropwise to the cell culture dish, and 4-6 hours later, the cells were washed twice with PBS and provided with a fresh serum-free medium.

(3) Virus harvesting: after transfection for 48 hours, cells and supernatants were harvested, centrifuged slow at 1000 rpm for 10 minutes to remove cell debris, packed, and single use aliquots were stored at -80°C to avoid the decrease of virus titers caused by repeated freezing and thawing.

(4) Infection: on the first day, the cells were seeded and cultured overnight, and the cells reached 80-100% over 18-24 hours;

[0049] On the next day, the susceptible cells were washed with PBS to remove serum and infected with the collected viral supernatant, and the culture medium was changed to a serum-containing medium 4-6 hours later. According to the experimental requirements, Luciferase values could be measured at different time points, with reference to the Luciferase Assay System Protocol or the Dual Luciferase Reporter Assay System Protocol of Promega Company. The harvested virus solution was diluted 5-fold and added to Huh7 cells (human hepatoma cells) in a 96-well plate. After 4 hours of infection, the virus solution was discarded, and the cells were washed twice with PBS, and provided with DMEM complete medium containing 10% serum. The medium was discarded 48 hours later, and the cells were washed twice with PBS and added with a cell lysis solution. After freezing and thawing once at -80°C, 20 μ L of cell culture from each well was assayed for luciferase activity using a GloMax 96 Microplate Luminometer (Promega). TCID₅₀ was calculated by Reed-Muech method.

EXAMPLE 5 Pseudovirus Neutralization Assay of Immune Serum

[0050] The serum obtained in Example 2 was diluted in multiple ratios, mixed with 100 TCID₅₀ pseudovirus, and incubated for 30 minutes at 37°C. The mixture was then added to a 96-well plate completely covered with Huh7 cells. After incubation at 37°C for 4 hours, the virus solution was discarded, and the cells were washed twice with PBS, and provided with a complete medium DMEM containing 10% serum. After 48 hours, the culture medium was discarded, and the cells were washed twice with PBS and added with a cell lysis solution to assay the luciferase activity. Pseudovirus having spike protein on the surface infected cells to release DNA and express rather than replicate luciferase. If the pseudovirus could not infect the cells in the presence of neutralizing antibodies, the luciferase was not expressed. The neutralization titers of the serum were examined in this way.

[0051] The results of immunogenicity assays after the third immunization are shown in FIG. 4. The result shown that the RBD dimer (E367-Y606) elicited neutralizing antibodies after three immunizations, regardless of the adavax adjuvant group or the aluminum adjuvant group (indicated by +Alum). Particularly, the mean value of the neutralizing antibodies NT₉₀ in the AddaVax adjuvant 10 μ g group could reach more than 1:1000 (as shown in FIG. 4); whereas the RBD-monomer (E379-E589) did not elicit neutralizing antibodies after three immunizations except low neutralizing antibody production in 2 mice (as shown in FIG. 4). Pseudovirus neutralization assay demonstrated that the neutralizing antibodies induced by the RBD-dimer was much higher than that induced by the monomeric RBD.

[0052] The RBD monomer (E379-E589) was obtained by the following method: a nucleic acid fragment (shown as SEQ ID NO: 25) encoding the amino acid (E379-E589) sequence (shown as SEQ ID NO: 4) in MERS-CoV S protein was inserted into EcoRI and XhoI restriction enzyme cutting sites of pFastBac-SP to allow fusion expression of the protein coding region behind the signal peptide gp67 sequence for secretion of the protein of interest having 6 histidines at the C-terminal thereof, thereby obtaining a vector pFastBac-SP-MERS-RBD (E379-E589).

EXAMPLE 6 Euvirus Neutralization of Immune Serum (EMC Strain)

[0053] Neutralization assay was conducted with serum after three immunizations for MERS-CoV euvirus (EMC strain,

disclosed in Yao Y, Bao L, Deng W, et al. An animal model of MERS produced by infection of rhesus macaques with MERS coronavirus. *J Infect Dis.* 2014;209(2):236-242. doi:10.1093/infdis/jit590, supplied by the institute of laboratory animals of Peking Union Medical College). The results are shown in FIG. 5. The results showed that both AddaVax adjuvant and aluminum adjuvant could elicit high neutralizing antibodies in mice. The highest group (Addavax adjuvant 10 μ g and RBD dimer) achieved an IC₅₀ greater than 1: 600. This result demonstrated that the dimeric RBD could elicit a higher level of neutralizing antibodies in mice by MERS-CoV euvirus neutralization assay.

EXAMPLE 7 Challenge Protection Experiment

[0054] Mice immunized three times in Example 2 were intranasally infected with adenovirus expressing hCD26 (hDPP4) on day 77, as shown in FIG. 2. This allowed transient expression of MERS-CoV receptor hCD26 in lung, making mice susceptible to MERS-CoV (see Chi H et al. DNA vaccine encoding Middle East responsive syndrome coronavirus S1 protein induces protective immune responses in mice [J]. *Vaccine*, 2017, 35 (16): 2069-2075). Five days later, experiments with MERS-CoV (EMC strain) challenge were conducted with a challenge dose of 5×10^5 pfu. Three days later after challenge, the lungs of mice were harvested, and tissue homogenate prepared therefrom was used to detect virus titers (TCID₅₀). The results are shown in FIG. 6. Compared with PBS control group, the viral load in lung tissue of mice in vaccine group decreased significantly. The viral load in the group with AddaVax adjuvant 3 μ g and RBD dimer decreased by nearly 1000 times compared with that of the PBS group, showing a good protective efficacy. These results showed that the RBD dimer, as a vaccine, had a markedly significant protective efficacy against MERS-CoV challenge.

EXAMPLE 8 Validation of Vaccine Protection for The Lung Tissue of Mice

[0055] The lung tissue of mice in the MERS-CoV challenge experiment in Example 7 was fixed in 4% paraformaldehyde, and then stained with hematoxylin and eosin, and tissue sections were used to observe the pathological changes of the lung, with results as shown in FIG. 7. Lung tissues of all control mice (namely, PBS group) exhibited severe interstitial pneumonia, pulmonary alveolitis, diffuse inflammatory cell infiltration, and necrosis of bronchial epithelial cells (as shown in FIG. 7). However, milder lesions were observed in the group of mice immunized with the RBD-dimer and the pulmonary alveolus was highly visible with lower infiltration of inflammatory cells because both AddaVax and Alum adjuvants could greatly alleviate the lung injury caused by virus challenge. The small histopathological changes in the lung likely resulted from a direct inoculation of high amount (5×10^5 pfu) of virus intranasally. Therefore, the RBD-dimer could substantially reduce the lung injury caused by MERS-CoV infection.

EXAMPLE 9 Crystallization and Structure Determination of MERS-RBD-Dimer

[0056] The RBD (E367-Y606) protein was expressed according to the method of Example 1. After purification, the dimer protein peaks were collected. The protein was concentrated to 10 mg/ml and mixed with the reservoir solution in a volume ratio of 1:1, and then protein crystal screening was carried out by mosquito[®] Protein Crystallization Screening Liquid Workstation (TTP LabTech). Diffraction-quality crystals of MERS-CoV RBD-dimer were obtained at 18°C. The crystals were collected at the Shanghai Synchrotron Radiation Facility (SSRF), and finally 2.8 Å diffraction data were obtained. The data were processed with HKL2000 software, and the structure was solved by the molecular replacement module, with the structure of MERS-CoV RBD (PDB: 4KQZ) as the search models. The results are shown in FIG. 8.

EXAMPLE 10 Structure Design of a Single-Chain RBD Dimer (Sc-RBD Dimer) Based on MERS-RBD Dimer

[0057] Based on the MERS-RBD crystal structure of FIG. 8, the N-terminal (N') and C-terminal (C') of the two subunits of RBD were arranged in an end-to-end form. The N-terminal and the C-terminal each had an invisible flexible sequence (as shown in FIG. 9A), which inspired us to link two subunits as a tandem repeat single chain, namely, single-chain RBD dimer (sc-RBD dimer).

[0058] The first design (as shown in FIG. 9A) was as follows:

- (1) two GGS linker sequences were added between two repeated tandem (E367-Y606) sequences to obtain MERS-RBD-C1 (abbreviated as C1), where the nucleotide sequence encoding the amino acid sequence was SEQ ID NO: 9;
- (2) one GGS linker sequence was added between two repeated tandem (E367-Y606) sequences to obtain MERS-RBD-C2 (abbreviated as C2), where the nucleotide sequence encoding the amino acid sequence is SEQ ID NO: 10;
- (3) two repeated (E367-Y606) sequences were directly linked in tandem to obtain MERS-RBD-C3 (abbreviated as C3), where the nucleotide sequence encoding the amino acid sequence is SEQ ID NO: 11.

[0059] The second design (as shown in FIG. 9B), in order to avoid the effects of cysteine residue (C603) at the position

603 of the C-terminal on expression, a truncated construct at C-terminal residue N602 was conducted, which was specifically as follows:

[0060] (4) one GGS linker sequence was added between two repeated tandem (E367-N602) sequences to obtain MERS-RBD-C4 (abbreviated as C4), where the nucleotide sequence encoding the amino acid sequence was SEQ ID NO: 12;

[0061] (5) two repeated (E367-N602) sequences were linked in tandem directly to obtain MERS-RBD-C5 (abbreviated as C5), where the nucleotide sequence encoding the amino acid sequence was SEQ ID NO: 13.

[0062] The third design in which (as shown in FIG. 9C) structurally visible sequences were directly expressed and linked by linker sequences of different lengths was specifically as follows:

[0063] (6) five GGS linker sequences were added between two repeated tandem (V381-L588) sequences to obtain MERS-RBD-C6 (abbreviated as C6), where the nucleotide sequence encoding the amino acid sequence was SEQ ID NO: 14;

[0064] (7) four GGS linker sequences were added between two repeated tandem (V381-L588) sequences to obtain MERS-RBD-C7 (abbreviated as C7), where the nucleotide sequence encoding the amino acid sequence was SEQ ID NO: 15;

[0065] (8) three GGS linker sequences were added between two repeated tandem (V381-L588) sequences to obtain MERS-RBD-C8 (abbreviated as C8), where the nucleotide sequence encoding the amino acid sequence was SEQ ID NO: 16;

[0066] (9) one GGS linker sequence was added between two repeated tandem (V381-L588) sequences to obtain MERS-RBD-C9 (abbreviated as C9), where the nucleotide sequence encoding the amino acid sequence was SEQ ID NO: 17;

[0067] (10) two repeated sequences (V381-L588) were directly linked in tandem to obtain MERS-RBD-C10 (abbreviated as C10), where the nucleotide sequence encoding the amino acid sequence was SEQ ID NO: 18.

[0068] The 5'-terminal of a nucleotide sequence encoding the above MERS-RBD-C1 to C10 was added with a nucleotide sequence encoding the MERS-S protein self-signal peptide (MIHSVFLLMFLLTPTES), while the 3'-terminal was added with a nucleotide sequence encoding six histidines. A terminator codon was then added to the 3'-terminal, and the obtained nucleotide sequence was inserted between the EcoRI and XhoI restriction enzyme cutting sites of a pCAGGS vector, and a Kozak sequence gccacc was contained upstream of an initiator codon. The above plasmid was transfected into 293T cells, and 48 hours later, the supernatant was collected, and the N-terminus of the protein of interest was provided with a signal peptide. Western blot method was used to detect the expression of the protein of interest, with the results as shown in FIG. 10. The results showed that all constructs were expressed except C2. Under both reduced (+DTT) and non-reduced (-DTT) conditions, the protein was about the size of the dimer (50-60 Kda). Among them, C4 and C5 were expressed at the highest levels. In view of that no any exogenous sequence was introduced and the sequence of the MERS-CoV itself was completely used, the C5 construct would be more advantageous and safer for clinical use. The efficacy of MERS-RBD-C5 as a vaccine would be further assessed.

EXAMPLE 11 Mammalian Expression of Single-Chain MERS-CoV RBD Dimer (Sc-RBD Dimer) and Protein Purification

[0069] MERS-RBD-C5 was expressed using mammalian 293T cells. After transfection of the plasmid into 293T cells, expression was conducted and the supernatant was harvested. The cell supernatant was filtered through a 0.22 μ m filtration membrane to remove cell debris. The supernatant of cell culture was purified by Ni affinity chromatography column (HisTrap) overnight at 4°C. The resin was washed with buffer A (20 mM Tris, 150 mM NaCl, pH 8.0) to remove non-specific binding proteins. Finally, the protein of interest was eluted from the resin with buffer B (20 mM Tris, 150 mM NaCl, pH 8.0, 300 mM imidazole), and the eluent was concentrated to be within 5 ml with a concentration tube of 10K MWCO. The protein of interest was further purified by molecular sieve chromatography using a Superdex 200 HiLoad 16/60 column (GE). The buffer for molecular sieve chromatography was 20 mM Tris and 150 mM NaCl, with pH 8.0. After the molecular sieve chromatography, there was only one main peak near the elution volume of 80 mL. Proteins were collected for SDS-PAGE analysis. As can be seen from the results of SDS-PAGE, MERS-RBD-C5 protein showed a distinct protein band between 55 and 72 kd, which was the size of RBD dimer. It was demonstrated that single-chain MERS-RBD dimer was obtained, as shown in FIG. 11. By using the method of Example 10, 293T cells were used to express and purify the non-single-chain MERS RBD dimer for comparison with the sc-RBD dimer.

EXAMPLE 12: Mice Immunized with Single-Chain MERS-CoV RBD Dimer (Sc-RBD Dimer) Protein

[0070] The single-chain MERS-RBD dimer antigen obtained in Example 11 was diluted in normal saline and emulsified with adjuvants in groups. Then BALB/c mice aged 4-6 weeks were immunized in groups, with 6 mice in each group. In addition, one group of mice was immunized with PBS as a negative control. A group of mice immunized with 293T cells

expressed a non-single-chain form of the dimer. Each mouse received three immunizations of vaccine by intramuscular injection into the thigh, at day 0, day 21 and day 42, respectively, at a vaccination volume of 100 μ l each time (containing 10 μ g of immunogen). Orbital blood was collected from mice 19 days later after the first immunization, 14 days later after the second immunization and 14 days later after the third immunization. Mouse serum was obtained by centrifugation at 3000 rpm for 10 minutes after standing, and stored in a refrigerator at -20°C for specific antibody detection and pseudovirus neutralization detection.

[0071] The serum specific antibody titer of the mice was detected by ELISA assay, using the method as shown in Example 3, with the results shown in FIG. 12. The RBD-sc-dimer group mice and disulfide-linked non-single-chain RBD-dimer group (indicated by Dimer) mice could be induced to produce antibody response. The titer mean value of the sc-dimer group was higher than that of the Dimer group, and the two groups had a significant difference after three immunizations (*, $P < 0.05$). The results showed that the sc-dimer had excellent immunogenicity as the disulfide-linked non-single-chain RBD-dimer.

[0072] The pseudovirus neutralization experiment was carried out with reference to Example 5, with the results shown in FIG. 13. The sc-dimer group mice and disulfide-linked non-single-chain RBD-dimer group (indicated by Dimer) mice could be induced to produce antibody response. The titer mean value of the sc-dimer group was higher than that of the Dimer group, and there was a significant difference between the two groups after the first immunization and the second immunization (FIG. 13). The mean values of the pseudovirus neutralization titers of sc-dimer group mice after three immunizations were already greater than 1: 1000. The results indicated that the vaccine developed by the sc-dimer had great clinical development potential.

EXAMPLE 13 Application of Single-Chain RBD Dimer Technology in Other Coronavirus Vaccines

[0073] To verify that this concept can be applied to vaccine design of all other coronaviruses, we compared the Receptor Binding Domains (RBDs) of the 19 common β coronaviruses, the result is shown in FIG. 14A and 14B. All β -coronavirus RBDs exhibited a conserved cysteine at position C603 of MERS-CoV, as shown in FIG. 14B. 2019-nCoV (hereinafter referred to as nCoV) and SARS-CoV were selected for verification. According to the structure of SARS-RBD (PDB: 3D0G), the crystal structure of SARS-RBD was molded into the crystal structure of MERS-RBD dimer at a resolution of 2.8 Å by using Pymol software. A simulated SARS-RBD dimer structure as shown in FIG. 15 was obtained. It was found that, like MERS-RBD dimer, SARS-RBD dimer also existed in the form of end-to-end (as shown in FIG. 15). Since the RBD region of 2019-nCoV shared more than 75 % homology with SARS-CoV, it was expected that the RBD dimer of 2019-nCoV would form this end-to-end arrangement. Considering that dimers in MERS-CoV could induce neutralizing antibodies with higher titers than monomers, it was considered that single-chain dimers (sc-dimers) were still used to design vaccines. Firstly, based on the S protein sequence of the 2019-nCoV WH01 strain, the construct of three single-chain dimers (sc-dimer) was designed, as shown in FIG. 15: (1) two R319-S530 were linked in tandem and named nCoV-RBD-C1 (the nucleotide sequence encoding the amino acid sequence was SEQ ID NO: 19); (2) two R319-K537 were linked in tandem and named nCoV-RBD-C2 (the nucleotide sequence encoding the amino acid sequence was SEQ ID NO: 20); (3) two R319-F541 were linked in tandem and named nCoV-RBD-C3 (the nucleotide sequence encoding the amino acid sequence was SEQ ID NO: 21); and (4) a further monomer was constructed as R319-F541, named nCoV-RBD-C4 (the nucleotide sequence encoding the amino acid sequence was SEQ ID NO: 22). In addition, a single-chain dimer of SARS-CoV was constructed, and two R306-Q523 were linked in tandem, as shown in FIGS. 14A and 14B, named SARS-CoV-RBD-C1 (the nucleotide sequence encoding this amino acid sequence was SEQ ID NO: 23).

[0074] A nucleotide sequence encoding the above nCoV-RBD-C1 to C4 and a nucleotide sequence encoding SARS-CoV-RBD-C1 (SEQ ID NO: 23) were added with the nucleotide sequence encoding the MERS-S protein self-signal peptide (MIHSVFLLMFLLTPTES). After the nucleotide sequence encoding six histidines was added to the 3'-terminal, a terminator codon was added to the 3'-terminal, and inserted into the pCAGGS vector EcoRI and XhoI restriction enzyme cutting sites, and the Kozak sequence gccacc was contained upstream of the initiator codon thereof. The above plasmid was transfected into 293T cells. The supernatant was harvested 48 hours later and the expression of the protein of interest was detected by Western blot. The expression results are shown in FIG. 16. The results showed that the expression of nCoV-RBD-C2 was the highest among several antigen designs of 2019-nCoV. The SARS-CoV-RBD-C1 construct also had a high protein expression.

[0075] The above results indicated that in the design of a single-chain dimer of the β -coronavirus, the optimal construct is from the first amino acid of FIG. 14A (marked as Start) to an amino acid before the last cysteine of FIG. 14B (marked as Stop).

EXAMPLE 14 Expression and Purification of Single-Chain 2019-nCoV-RBD Dimer Antigen and Single-Chain SARS-CoV-RBD Dimer Antigen

[0076] Mammalian 293T cells were used to express nCoV-RBD-C2. After the plasmid was transfected into 293T cells,

the supernatant was harvested. The cell supernatant was filtered through a 0.22 μ m filtration membrane to remove cell debris. The supernatant of cell culture was purified by Ni affinity chromatography column (HisTrap) overnight at 4°C. The resin was washed with buffer A (20 mM Tris, 150 mM NaCl, pH 8.0) to remove non-specific binding proteins. Finally, the protein of interest was eluted from the resin with buffer B (20 mM Tris, 150 mM NaCl, pH 8.0, 300 mM imidazole), and the eluent was concentrated to be within 5 ml with a concentration tube of 10K MWCO. The protein of interest was further purified by molecular sieve chromatography with a Superdex 200 Hiload 16/60 column (GE). The buffer for molecular sieve chromatography was 20 mM Tris and 150 mM NaCl, with PH 8.0. After molecular sieve chromatography, there was only one main peak near the elution volume of 80 mL. Proteins were collected for SDS-PAGE analysis. As can be seen from the results of SDS-PAGE, nCoV-RBD-C2 protein showed a distinct protein band between 48-63 kd, which was the size of RBD-dimer. It was demonstrated that single-chain 2019-nCoV-RBD dimer was obtained, as shown in FIG. 17. The purity was more than 95%. The results showed that such construct could produce sufficient and high-purity single-chain 2019-nCoV dimer protein.

[0077] The monomeric RBD protein of 2019-nCoV (obtained by expression of nCoV-RBD-C4 construct), the monomeric RBD protein of SARS-CoV (SARS-CoV RBD R306-F527, having an amino acid sequence and a nucleotide sequence encoding the amino acid sequence as shown as SEQ ID NO: 26 and SEQ ID NO: 27) and the single-chain dimer protein of SARS-CoV (obtained by expression of SARS-CoV-RBD-C1 construct) were expressed and purified in the same way.

[0078] As shown in FIG. 18, the result of the single-chain dimer protein of SARS-CoV showed that after the molecular sieve chromatography, there was only one main peak near the elution volume of 80 mL. Proteins were collected for SDS-PAGE analysis. As can be seen from the results of SDS-PAGE, SARS-CoV-RBD-C1 protein of interest showed a distinct protein band between 55 and 72 kd, which was the size of RBD-dimer. It was demonstrated that single-chain SARS-RBD dimer was obtained, as shown in FIG. 18, and with high purity.

EXAMPLE 15 Mice Immunized with Single-Chain 2019-nCoV-RBD Dimer Protein

[0079] The single-chain 2019-nCoV-RBD dimer and 2019-nCoV-RBD monomer obtained in Example 14 was diluted in PBS solution and emulsified with AddaVax adjuvant in groups. Then BALB/c mice (average body weight 15-20g, the same applies below) aged 6-8 weeks were immunized in groups, with 8 mice in each group. Each mouse received three immunizations of vaccine by intramuscular injection into the thigh, at day 0, day 21 and day 42, respectively, at a vaccination volume of 100 μ l each time (containing 10 μ g of immunogen). Blood samples were collected from mice 19 days later after the first immunization, 14 days later after the second immunization and 14 days later after the third immunization. Mouse serum was obtained by centrifugation at 3000 rpm for 10 minutes after standing, and stored in a refrigerator at -20°C for specific antibody detection and pseudovirus neutralization detection.

[0080] The serum specific antibody titer of 2019-CoV RBD of the mice was detected by ELISA assay, using the method as shown in Example 3, with the results shown in FIG. 19. The single-chain dimeric RBD (indicated by sc-dimer) and monomeric RBD (indicated by Monomer) could induce mice to produce antibody response. The titer mean value of the single-chain dimeric RBD group was higher than that of the monomeric RBD group, and the two groups had a significant difference after three immunizations (FIG. 19). The single-chain dimeric RBD induced mice to produce antibodies at levels of up to approximately 1:10⁶ after three immunizations. The results showed that the immunogenicity of the single-chain dimeric RBD antigen was stronger than that of the single-chain dimeric RBD antigen, and it had great potential as a potential new coronavirus vaccine.

[0081] 2019-nCoV pseudovirus neutralization assay was carry out with reference to Example 5, with results as shown in FIG. 20. Neutralizing antibodies were induced in only the single-chain dimeric RBD (indicated by sc-dimer) group after the first immunization. Neutralizing antibodies were not detected in both the monomeric RBD (indicated by Monomer) and PBS groups, and there was a significant difference of neutralizing antibody titers between the single-chain dimeric RBD group and the monomeric RBD group (FIG. 20). After the second and third immunizations, both single-chain dimeric RBD and monomeric RBD could induce mice to produce neutralizing antibodies. After each immunization, the mean value of neutralizing antibody titers of the single-chain dimeric RBD group was higher than that of the monomeric RBD group (10-100 times higher), and there was a significant difference between the two groups after each immunization (FIG. 20). The single-chain dimeric RBD induced mice to produce antibodies at levels of up to approximately 1:10⁴ after three immunizations. The results showed that the single-chain dimeric RBD antigen could induce mice to produce higher neutralizing antibody level than the monomeric RBD antigen, and the single-chain dimeric RBD antigen had high advantages in use.

[0082] Neutralization assay was conducted with serum after the second immunization for 2019-nCoV euvirus (2020XN4276 strain, which was published in Lu J, du Plessis L, Liu Z, et al. Genomic Epidemiology of SARS-CoV-2 in Guangdong Province, China. Cell. 2020;181(5):997-1003.e9. doi:10.1016/j.cell.2020.04.023, provided by Guangdong Provincial Center for Disease Control and Prevention). The experimental results are shown in FIG. 21. The results showed that the RBD dimer could induce mice to produce high levels of neutralizing antibodies against the novel coronavirus. The highest neutralizing NT50 was greater than 4096, and the lowest NT50 was 512 in a mouse. However,

the neutralizing antibodies against the novel coronavirus were detected in only 2 of the 8 mice in the RBD monomer group, with lower NT50, which were 128 and 256, respectively. The results indicated that the dimeric RBD could induce mice to produce higher levels of neutralizing antibodies against the novel coronavirus.

EXAMPLE 16 Mice Immunized with Single-Chain SARS-RBD Dimer Protein

[0083] The single-chain SARS-RBD dimer and SARS-RBD monomer obtained in Example 14 were diluted in PBS solution and emulsified with AddaVax adjuvant in groups. Then BALB/c mice aged 6-8 weeks were immunized in groups, with 6 mice in each group. Each mouse received three immunizations of vaccine by intramuscular injection into the thigh, at day 0, day 2 and day 42, respectively, at a vaccination volume of 100 μ l each time (containing 10 μ g of immunogen). Blood samples were collected from mice 19 days later after the first immunization, 14 days later after the second immunization and 14 days later after the third immunization. Mouse serum was obtained by centrifugation at 3000 rpm for 10 minutes after standing, and stored in a refrigerator at -20°C for specific antibody detection and pseudovirus neutralization detection.

[0084] The serum specific antibody titer of SARS-RBD of the mice was detected by ELISA assay, using the method shown in Example 3, with the results shown in FIG. 22. The single-chain dimeric RBD (indicated by sc-dimer) and monomeric RBD (indicated by Monomer) could induce mice to produce antibody response. The titer mean value of the single-chain dimeric RBD group was higher than that of the monomeric RBD group, and the two groups had a significant difference after the second and the third immunizations (FIG.22). The dimeric RBD induced mice to produce antibodies at levels of up to approximately 1:10⁶ after three immunizations. The results showed that the immunogenicity of the dimeric RBD antigen was stronger than that of the monomeric RBD antigen.

[0085] The neutralization assay was conducted for SARS-CoV pseudovirus with reference to Example 5, with the results shown in FIG. 23. After the first and second immunizations, both dimeric RBD (indicated by sc-dimer) group and monomeric RBD (indicated by Monomer) group could induce mice to produce neutralizing antibodies. The mean value of neutralizing antibody titers of the dimeric RBD group was higher, and there was a significant difference between the two groups (FIG. 23). After the third immunization, the mean value of neutralizing antibody titers of the dimeric RBD group was still higher than that of the monomeric RBD group, and there was a significant difference (FIG. 23). The levels of neutralizing antibodies induced by dimeric RBD in mice after three immunizations were higher than 1:10³. The results showed that the dimeric RBD antigen could induce mice to produce higher neutralizing antibody level than the monomeric RBD antigen, and the dimeric RBD antigen had high advantages in use.

[0086] Finally, it should be noted that the above examples are only intended to illustrate rather than limit the technical solutions of the present disclosure. Although the present disclosure has been described in detail with reference to the foregoing examples, it will be understood by a person skilled in the art that the technical solutions described in the foregoing examples may still be modified, or some technical features may be equivalently replaced; and such modifications or substitutions do not depart from the spirit and scope of the corresponding technical solutions of the examples of the present disclosure.

INDUSTRIAL PRACTICAL APPLICABILITY

[0087] The examples of the present disclosure relate to antigens of β -coronaviruses, preparation methods and uses thereof. An antigen of a β -coronavirus, its amino acid sequence comprises an amino acid sequence arranged in a (A-B)-(A-B) pattern or an amino acid sequence arranged in a (A-B)-C-(A-B) pattern or an amino acid sequence arranged in a (A-B)-(A-B') pattern or an amino acid sequence arranged in a (A-B)-C-(A-B') pattern, where A-B represents a partial amino acid sequence or the entire amino acid sequence of a receptor binding domain of a surface spike protein of the β -coronavirus; C represents an amino acid linker sequence; A-B' represents an amino acid sequence obtained by substitution, deletion or addition of one or more amino acids in the amino acid sequence of A-B; a protein encoded by A-B' has the identical or substantially identical immunogenicity as a protein encoded by A-B; and the antigen of the β -coronavirus has a single-chain dimer structure. The single-chain dimer expressed according to the examples of the present disclosure is stable in content and has excellent immunogenicity as an antigen of a β -coronavirus, and the vaccine prepared by using the single-chain dimer as an antigen of a β -coronavirus can elicit high-titer neutralizing antibodies in mice.

SEQUENCE LISTING

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<120> ANTIGENS OF -CORONAVIRUSES, PREPARATION METHODS AND USES THEREOF

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<150> CN202010085038.9

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35 40 45

Leu Ser Leu Phe Ser Val Asn Asp Phe Thr Cys Ser Gln Ile Ser Pro
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Ala Ala Ile Ala Ser Asn Cys Tyr Ser Ser Leu Ile Leu Asp Tyr Phe
65 70 75 80

Ser Tyr Pro Leu Ser Met Lys Ser Asp Leu Ser Val Ser Ser Ala Gly
85 90 95

Pro Ile Ser Gln Phe Asn Tyr Lys Gln Ser Phe Ser Asn Pro Thr Cys
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 35 40 45

Leu Ser Leu Phe Ser Val Asn Asp Phe Thr Cys Ser Gln Ile Ser Pro
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55 Ala Ala Ile Ala Ser Asn Cys Tyr Ser Ser Leu Ile Leu Asp Tyr Phe
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Ser Tyr Pro Leu Ser Met Lys Ser Asp Leu Ser Val Ser Ser Ala Gly
 85 90 95

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55 Lys Leu Leu Ser Leu Phe Ser Val Asn Asp Phe Thr Cys Ser Gln Ile
35 40 45

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 Ala Gly Pro Ile Ser Gln Phe Asn Tyr Lys Gln Ser Phe Ser Asn Pro
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 Thr Cys Leu Ile Leu Ala Thr Val Pro His Asn Leu Thr Thr Ile Thr
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 Lys Pro Leu Lys Tyr Ser Tyr Ile Asn Lys Cys Ser Arg Leu Leu Ser
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 Asp Asp Arg Thr Glu Val Pro Gln Leu Val Asn Ala Asn Gln Tyr Ser
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 Pro Cys Val Ser Ile Val Pro Ser Thr Val Trp Glu Asp Gly Asp Tyr
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 Tyr Arg Lys Gln Leu Ser Pro Leu Glu Gly Gly Gly Trp Leu Val Ala
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 Leu Thr Lys Leu Leu Ser Leu Phe Ser Val Asn Asp Phe Thr Cys Ser

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15	Ser Ser Ala Gly Pro	Ile Ser Gln Phe	Asn Tyr Lys Gln Ser Phe Ser
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40	Asp Tyr Tyr Arg Lys	Gln Leu Ser Pro	Leu Glu Gly Gly Gly Trp Leu
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15

Tyr Ala Trp Asn Arg Lys Arg Ile Ser Asn Cys Val Ala Asp Tyr Ser
35 40 45

Val Leu Tyr Asn Ser Ala Ser Phe Ser Thr Phe Lys Cys Tyr Gly Val
50 55 60

20

Ser Pro Thr Lys Leu Asn Asp Leu Cys Phe Thr Asn Val Tyr Ala Asp
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25

Ser Phe Val Ile Arg Gly Asp Glu Val Arg Gln Ile Ala Pro Gly Gln
85 90 95

Thr Gly Lys Ile Ala Asp Tyr Asn Tyr Lys Leu Pro Asp Asp Phe Thr
100 105 110

30

Gly Cys Val Ile Ala Trp Asn Ser Asn Asn Leu Asp Ser Lys Val Gly
115 120 125

35

Gly Asn Tyr Asn Tyr Leu Tyr Arg Leu Phe Arg Lys Ser Asn Leu Lys
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45

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20 25 30

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35 40 45

Val Leu Tyr Asn Ser Ala Ser Phe Ser Thr Phe Lys Cys Tyr Gly Val
50 55 60

Ser Pro Thr Lys Leu Asn Asp Leu Cys Phe Thr Asn Val Tyr Ala Asp
65 70 75 80

Ser Phe Val Ile Arg Gly Asp Glu Val Arg Gln Ile Ala Pro Gly Gln
85 90 95

Thr Gly Lys Ile Ala Asp Tyr Asn Tyr Lys Leu Pro Asp Asp Phe Thr
100 105 110

Gly Cys Val Ile Ala Trp Asn Ser Asn Asn Leu Asp Ser Lys Val Gly
115 120 125

Gly Asn Tyr Asn Tyr Leu Tyr Arg Leu Phe Arg Lys Ser Asn Leu Lys
130 135 140

Pro Phe Glu Arg Asp Ile Ser Thr Glu Ile Tyr Gln Ala Gly Ser Thr
145 150 155 160

Pro Cys Asn Gly Val Glu Gly Phe Asn Cys Tyr Phe Pro Leu Gln Ser
165 170 175

Tyr Gly Phe Gln Pro Thr Asn Gly Val Gly Tyr Gln Pro Tyr Arg Val
180 185 190

Val Val Leu Ser Phe Glu Leu Leu His Ala Pro Ala Thr Val Cys Gly
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20 25 30

Tyr Ala Trp Glu Arg Lys Lys Ile Ser Asn Cys Val Ala Asp Tyr Ser
35 40 45

Val Leu Tyr Asn Ser Thr Phe Phe Ser Thr Phe Lys Cys Tyr Gly Val
50 55 60

Ser Ala Thr Lys Leu Asn Asp Leu Cys Phe Ser Asn Val Tyr Ala Asp
65 70 75 80

Ser Phe Val Val Lys Gly Asp Asp Val Arg Gln Ile Ala Pro Gly Gln
85 90 95

Thr Gly Val Ile Ala Asp Tyr Asn Tyr Lys Leu Pro Asp Asp Phe Met
100 105 110

Gly Cys Val Leu Ala Trp Asn Thr Arg Asn Ile Asp Ala Thr Ser Thr
115 120 125

Gly Asn Tyr Asn Tyr Lys Tyr Arg Tyr Leu Arg His Gly Lys Leu Arg
130 135 140

Pro Phe Glu Arg Asp Ile Ser Asn Val Pro Phe Ser Pro Asp Gly Lys
145 150 155 160

Pro Cys Thr Pro Pro Ala Leu Asn Cys Tyr Trp Pro Leu Asn Asp Tyr
165 170 175

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Gly Phe Tyr Thr Thr Thr Gly Ile Gly Tyr Gln Pro Tyr Arg Val Val
180 185 190

5 Val Leu Ser Phe Glu Leu Leu Asn Ala Pro Ala Thr Val Cys Gly Pro
195 200 205

10 Lys Leu Ser Thr Asp Leu Ile Lys Asn Gln
210 215

<210> 9

<211> 1458

<212> DNA

15 <213> Artificial Sequence

<220>

<221> CDS

<222> (1) .. (1458)

20 <223> A nucleotide sequence encoding two repeated amino acid sequences
of SEQ ID NO: 1 linked in tandem by the GSGGS linker sequence

<400> 9

25	gaagcaaaac cttctggctc agttgtggaa caggctgaag gtgttgaatg tgatttttca	60
	cctcttctgt ctggcacacc tcctcagggt tataatttca agcgttttgt tttaccaat	120
	tgcaattata atcttaccaa attgctttca ctttttctg tgaatgattt tacttgtagt	180
30	caaatatctc cagcagcaat tgctagcaac tgttattctt cactgatttt ggattacttt	240
	tcatacccac ttagtatgaa atccgatctc agtggttagtt ctgctgggtcc aatatccag	300
	tttaattata aacagtcctt ttctaataccc acatgtttga ttttagcgac tgttcctcat	360
35	aaccttacta ctattactaa gcctcttaag tacagctata ttaacaagtg ctctcgtctt	420
	ctttctgatg atcgactga agtacctcag ttagtgaacg ctaatcaata ctcaccctgt	480
	gtatccattg tcccatccac tgtgtgggaa gacggtgatt attataggaa acaactatct	540
40	ccacttgaag gtgggtggctg gcttgttgct agtggctcaa ctggtgccat gactgagcaa	600
	ttacagatgg gctttggtat tacagttcaa tatggtacag acaccaatag tgtttgcccc	660
	aagcttgaat ttgctaata caaaaaatt gcctctcaat taggcaattg cgtggaatat	720
45	ggcggctcag gcggtcaga agcaaaacct tctggctcag ttgtggaaca ggctgaaggt	780
	gttgaatgtg atttttcacc tcttctgtct ggcacacctc ctcaggttta taatttcaag	840
50	cgtttggttt ttaccaattg caattataat cttaccaaata tgctttcact ttttctgtg	900
	aatgatttta cttgtagtca aatatctcca gcagcaattg ctagcaactg ttattcttca	960
	ctgatttttg attacttttc ataccactt agtatgaaat ccgatctcag tgtagttct	1020
55	gctgggtcaa tatccagtt taattataaa cagtcctttt ctaatccac atgtttgatt	1080
	ttagcgactg ttcctcataa cttactact attactaagc ctcttaagta cagctatatt	1140

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aacaagtgct ctcgtcttct ttctgatgat cgtactgaag tacctcagtt agtgaacgct 1200
 aatcaatact caccctgtgt atccattgtc ccatccactg tgtgggaaga cggtgattat 1260
 5 tataggaaac aactatctcc acttgaaggt ggtggctggc ttgttgctag tggctcaact 1320
 gttgccatga ctgagcaatt acagatgggc tttggtatta cagttcaata tggtagacac 1380
 accaatagtg tttgccccaa gcttgaatth gctaatagaca caaaaattgc ctctcaatta 1440
 10 ggcaattgag tggaatat 1458

<210> 10
 <211> 1449
 <212> DNA
 15 <213> Artificial Sequence

<220>
 <221> CDS
 20 <222> (1)..(1449)
 <223> A nucleotide sequence encoding two repeated amino acid sequences
 of SEQ ID NO: 1 linked in tandem by a GGS linker sequence

<400> 10
 25 gaagcaaaac cttctggctc agttgtggaa caggctgaag gtgttgaatg tgatttttca 60
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 tgcaattata atcttaccac attgctttca cttttttctg tgaatgattt tacttgtagt 180
 30 caaatatctc cagcagcaat tgctagcaac tgttattctt cactgatttt ggattacttt 240
 tcataccacac ttagtatgaa atccgatctc agtgttagtt ctgctgggtcc aatatcccag 300
 ttttaattata aacagtcctt ttctaataccc acatgtttga ttttagcgac tgttcctcat 360
 35 aaccttacta ctattactaa gcctcttaag tacagctata ttaacaagtg ctctcgtctt 420
 ctttctgatg atcgtactga agtacctcag ttagtgaacg ctaatacaata ctacccctgt 480
 gtatccattg tcccatccac tgtgtgggaa gacggtgatt attataggaa acaactatct 540
 40 ccacttgaag gtggtggctg gcttgttgct agtggtcaa ctggtgcat gactgagcaa 600
 ttacagatgg gctttggtat tacagttcaa tatggtacag acaccaatag tgtttgcccc 660
 aagcttgaat ttgctaata caaaaaatt gcctctcaat taggcaattg cgtggaatat 720
 45 ggcggctcag aagcaaaacc ttctggctca gttgtggaac aggctgaagg tgttgaatgt 780
 gattttttcac ctcttctgtc tggcacacct cctcaggttt ataatttcaa gcgtttggtt 840
 tttaccaatt gcaattataa tcttaccaaa ttgctttcac ttttttctgt gaatgatttt 900
 50 actttagatc aaatatctcc agcagcaatt gctagcaact gttattcttc actgattttg 960
 gattactttt cataccact tagtatgaaa tccgatctca gtgttagttc tgctgggtcca 1020
 55 atatcccagt ttaattataa acagtccttt tctaatacca catgtttgat ttttagcgact 1080
 gttcctcata accttactac tattactaag cctcttaagt acagctatat taacaagtgc 1140

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	tctcgtcttc tttctgatga tcgtactgaa gtacctcagt tagtgaacgc taatcaatac	1200
	tcaccctgtg tatccattgt cccatccact gtgtgggaag acggtgatta ttataggaaa	1260
5	caactatctc cacttgaagg tgggtggctgg cttgttgcta gtggctcaac tgttgccatg	1320
	actgagcaat tacagatggg ctttgggtatt acagttcaat atggtacaga caccaatagt	1380
	gtttgcccc aagcttgaatt tgctaatac acaaaaattg cctctcaatt aggcaattgc	1440
10	gtggaatat	1449
	<210> 11	
	<211> 1440	
15	<212> DNA	
	<213> Artificial Sequence	
	<220>	
20	<221> CDS	
	<222> (1) .. (1440)	
	<223> Nucleotide sequence encoding two repeated amino acid sequences of SEQ ID NO: 1 directly linked in tandem	
	<400> 11	
25	gaagcaaaac cttctggctc agttgtggaa caggctgaag gtgttgaatg tgatttttca	60
	cctcttctgt ctggcacacc tcctcagggt tataatattca agcgtttgggt ttttaccaat	120
	tgcaattata atcttaccaa attgctttca cttttttctg tgaatgattt tacttgtagt	180
30	caaatatctc cagcagcaat tgctagcaac tgttattctt cactgatttt ggattacttt	240
	tcatacccac ttagtatgaa atccgatctc agtggttagtt ctgctgggtcc aatatccag	300
	tttaattata aacagtcctt ttctaataccc acatgtttga ttttagcgac tgttcctcat	360
35	aaccttacta ctattactaa gcctcttaag tacagctata ttaacaagtg ctctcgtctt	420
	ctttctgatg atcgtactga agtacctcag ttagtgaacg ctaatcaata ctaccctgt	480
40	gtatccattg tcccatccac tgtgtggga gacggtgatt attataggaa acaactatct	540
	ccacttgaag gtgggtggctg gcttgttgct agtggctcaa ctgttgccat gactgagcaa	600
	ttacagatgg gctttgggtat tacagttcaa tatggtacag acaccaatag tgtttgcccc	660
45	aagcttgaat ttgctaata cacaataatt gcctctcaat taggcaattg cgtggaatat	720
	gaagcaaaac cttctggctc agttgtggaa caggctgaag gtgttgaatg tgatttttca	780
	cctcttctgt ctggcacacc tcctcagggt tataatattca agcgtttgggt ttttaccaat	840
50	tgcaattata atcttaccaa attgctttca cttttttctg tgaatgattt tacttgtagt	900
	caaatatctc cagcagcaat tgctagcaac tgttattctt cactgatttt ggattacttt	960
55	tcatacccac ttagtatgaa atccgatctc agtggttagtt ctgctgggtcc aatatccag	1020
	tttaattata aacagtcctt ttctaataccc acatgtttga ttttagcgac tgttcctcat	1080

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aaccttacta ctattactaa gcctcttaag tacagctata ttaacaagtg ctctcgtctt 1140
 ctttctgatg atcgtactga agtacctcag ttagtgaacg ctaatcaata ctcaccctgt 1200
 5 gtatccattg tcccatccac tgtgtgggaa gacggtgatt attataggaa acaactatct 1260
 ccacttgaag gtggtggctg gcttgttgct agtggctcaa ctggtgccat gactgagcaa 1320
 ttacagatgg gctttggtat tacagttcaa tatggtacag acaccaatag tgtttgcccc 1380
 10 aagcttgaat ttgctaata caaaaaatt gcctctcaat taggcaattg cgtggaatat 1440

 <210> 12
 <211> 1425
 <212> DNA
 15 <213> Artificial Sequence

 <220>
 <221> CDS
 20 <222> (1)..(1425)
 <223> A nucleotide sequence encoding two repeated amino acid sequences
 of SEQ ID NO: 2 linked in tandem by a GGS linker sequence

 <400> 12
 25 gaagcaaaac cttctggctc agttgtggaa caggctgaag gtggtgaatg tgatttttca 60
 cctcttctgt ctggcacacc tcctcagggt tataatttca agcgtttggt ttttaccaat 120
 tgcaattata atcttaccac attgctttca ctttttctg tgaatgattt tactttagt 180
 30 caaatatctc cagcagcaat tgctagcaac tgttattctt cactgatttt ggattacttt 240
 tcataccac ttagtatgaa atccgatctc agtggttagt ctgctggtcc aatatcccag 300
 ttaattata aacagtcctt ttctaattcc acatgtttga ttttagcgac tgttcctcat 360
 35 aaccttacta ctattactaa gcctcttaag tacagctata ttaacaagtg ctctcgtctt 420
 ctttctgatg atcgtactga agtacctcag ttagtgaacg ctaatcaata ctcaccctgt 480
 gtatccattg tcccatccac tgtgtgggaa gacggtgatt attataggaa acaactatct 540
 40 ccacttgaag gtggtggctg gcttgttgct agtggctcaa ctggtgccat gactgagcaa 600
 ttacagatgg gctttggtat tacagttcaa tatggtacag acaccaatag tgtttgcccc 660
 aagcttgaat ttgctaata caaaaaatt gcctctcaat taggcaatgg cggctcagaa 720
 45 gcaaaacctt ctggctcagt tgtggaacag gctgaagggt ttgaatgtga tttttcacct 780
 cttctgtctg gcacacctcc tcagggttat aatttcaagc gtttggtttt taccaattgc 840
 aattataatc ttaccaaatt gctttcactt ttttctgtga atgattttac ttgtagtcaa 900
 50 atatctccag cagcaattgc tagcaactgt tattcttcac tgattttgga ttacttttca 960
 taccactta gtatgaaatc cgatctcagt gttagttctg ctggtccaat atcccagttt 1020
 55 aattataaac agtccttttc taatcccaca tgtttgattt tagcgactgt tcctcataac 1080
 cttactacta ttactaagcc tcttaagtac agctatatta acaagtgtc tcgtcttctt 1140

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tctgatgatc gtactgaagt acctcagtta gtgaacgcta atcaatactc accctgtgta 1200

tccattgtcc catccactgt gtgggaagac ggtgattatt ataggaaaca actatctcca 1260

5 cttgaagggtg gtggctggct tgttgctagt ggctcaactg ttgccatgac tgagcaatta 1320

cagatgggct ttgggtattac agttcaatat ggtacagaca ccaatagtgt ttgccccaaag 1380

cttgaatttg ctaatgacac aaaaattgcc tctcaattag gcaat 1425

10

<210> 13

<211> 1416

<212> DNA

<213> Artificial Sequence

15

<220>

<221> CDS

<222> (1) .. (1416)

20 <223> Nucleotide sequence encoding two repeated amino acid sequences of
SEQ ID NO: 2 directly linked in tandem

<400> 13

gaagcaaaac cttctggctc agttgtggaa caggctgaag gtgttgaatg tgatttttca 60

25 cctcttctgt ctggcacacc tcctcagggt tataatttca agcgtttggt ttttaccat 120

tgcaattata atcttaccat attgctttca ctttttctg tgaatgattt tactttagt 180

caaatatctc cagcagcaat tgctagcaac tgttattctt cactgatttt ggattacttt 240

30 tcatacccac ttagtatgaa atccgatctc agtggttagt ctgctgggtcc aatatcccag 300

tttaattata aacagtcctt ttctaatacc acatgtttga ttttagcgac tgttcctcat 360

aaccttacta ctattactaa gcctcttaag tacagctata ttaacaagtg ctctcgtctt 420

35 ctttctgatg atcgtactga agtacctcag ttagtgaacg ctaatcaata ctcaccctgt 480

gtatccattg tcccatccac tgtgtgggaa gacggtgatt attataggaa acaactatct 540

40 ccacttgaag gtgggtggctg gcttgttgct agtggtcaa ctgttgccat gactgagcaa 600

ttacagatgg gctttggtat tacagttcaa tatggtacag acaccaatag tgtttgcccc 660

aagcttgaat ttgctaata cacaataatt gcctctcaat taggcaatga agcaaacct 720

45 tctggctcag ttgtggaaca ggctgaagggt gttgaatgtg atttttcacc tcttctgtct 780

ggcacacctc ctcagggtta taatttcaag cgtttggttt ttaccaattg caattataat 840

cttaccaaaat tgctttcact ttttctgtg aatgatttta cttgtagtca aatatctcca 900

50 gcagcaattg ctagcaactg ttattcttca ctgattttgg attacttttc ataccactt 960

agtatgaaat ccgatctcag tgttagttct gctgggtccaa tatcccagtt taattataaa 1020

55 cagtcctttt ctaatcccac atgtttgatt ttagcgactg ttctcataa cttactact 1080

attactaagc ctcttaagta cagctatatt aacaagtgt ctcgtcttct ttctgatgat 1140

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cgtactgaag tacctcagtt agtgaacgct aatcaatact caccctgtgt atccattgtc 1200
 ccatccactg tgtgggaaga cggtgattat tataggaaac aactatctcc acttgaaggt 1260
 5 ggtggctggc ttgttgctag tggctcaact gttgccatga ctgagcaatt acagatgggc 1320
 tttggtatta cagttcaata tggtagacac accaatagtg tttgccccaa gcttgaattt 1380
 gctaattgaca caaaaattgc ctctcaatta ggcaat 1416
 10
 <210> 14
 <211> 1293
 <212> DNA
 <213> Artificial Sequence
 15
 <220>
 <221> CDS
 <222> (1)..(1293)
 <223> A nucleotide sequence encoding two repeated amino acid sequences
 20 of SEQ ID NO: 3 linked in tandem by the GSGSGSGSGSGSGS linker
 sequence
 <400> 14
 gttgaatgtg atttttcacc tcttctgtct ggcacacctc ctgaggttta taatttcaag 60
 25 cgtttggttt ttaccaattg caattataat cttaccaaatt tgctttcact ttttctgtg 120
 aatgatttta cttgtagtca aatatctcca gcagcaattg ctagcaactg ttattcttca 180
 ctgatttttg attacttttc ataccactt agtatgaaat ccgatctcag tgtagttct 240
 30 gctgggtcaa tatcccagtt taattataaa cagtcctttt ctaatccac atgtttgatt 300
 ttagcgactg ttcctcataa cttactact attactaagc ctcttaagta cagctatatt 360
 35 aacaagtgtc ctctgttctt ttctgatgat cgtactgaag tacctcagtt agtgaacgct 420
 aatcaatact caccctgtgt atccattgtc ccatccactg tgtgggaaga cggtgattat 480
 tataggaaac aactatctcc acttgaaggt ggtggctggc ttgttgctag tggctcaact 540
 40 gttgccatga ctgagcaatt acagatgggc tttggtatta cagttcaata tggtagacac 600
 accaatagtg tttgccccaa gcttggcggc tcaggcgggt caggcgggtc aggcgggtca 660
 ggcgggtcag ttgaatgtga tttttcacct cttctgtctg gcacacctcc tcaggtttat 720
 45 aatttcaagc gtttggtttt taccaattgc aattataatc ttaccaaatt gctttcactt 780
 ttttctgtga atgattttac ttgtagtcaa atatctccag cagcaattgc tagcaactgt 840
 tattcttcac tgatttttga ttacttttca taccactta gtatgaaatc cgatctcagt 900
 50 gtttagttctg ctggtccaat atcccagttt aattataaac agtccttttc taatcccaca 960
 tgtttgattt tagcgactgt tcctcataac cttactacta ttactaagcc tcttaagtac 1020
 55 agctatatta acaagtgtc tcgtcttctt tctgatgatc gtactgaagt acctcagtta 1080
 gtgaacgcta atcaatactc accctgtgta tccattgtcc catccactgt gtgggaagac 1140

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ggtgattatt ataggaaaca actatctcca cttgaagggtg gtggctggct tgttgctagt 1200
 ggctcaactg ttgccatgac tgagcaatta cagatgggct ttggtattac agttcaatat 1260
 5 ggtacagaca ccaatagtgt ttgcccgaag ctt 1293

 <210> 15
 <211> 1284
 <212> DNA
 10 <213> Artificial Sequence

 <220>
 <221> CDS
 15 <222> (1)..(1284)
 <223> A nucleotide sequence encoding two repeated amino acid sequences
 of SEQ ID NO: 3 linked in tandem by the GSGSGSGSGGS linker
 sequence

 <400> 15
 20 gttgaatgtg atttttcacc tcttctgtct ggcacacctc ctcaggttta taatttcaag 60
 cgtttggttt ttaccaattg caattataat cttaccaaata tgctttcact ttttctgtg 120
 aatgatttta cttgtagtca aatatctcca gcagcaattg ctagcaactg ttattcttca 180
 25 ctgatttttg attacttttc ataccactt agtatgaaat ccgatctcag tgtagttct 240
 gctggtccaa tatcccagtt taattataaa cagtcctttt ctaatccac atgtttgatt 300
 ttagcgactg ttcctcataa cttactact attactaagc ctcttaagta cagctatatt 360
 30 aacaagtgtc ctcgtcttct ttctgatgat cgtactgaag tacctcagtt agtgaacgct 420
 aatcaatact caccctgtgt atccattgtc ccatccactg tgtgggaaga cggtgattat 480
 tataggaaac aactatctcc acttgaagggt ggtggctggc ttgttgctag tggctcaact 540
 gttgccatga ctgagcaatt acagatgggc tttggtatta cagttcaata tggtagacac 600
 accaatagtg tttgcccga gcttgccggc tcaggcggct caggcggctc aggcggctca 660
 40 gttgaatgtg atttttcacc tcttctgtct ggcacacctc ctcaggttta taatttcaag 720
 cgtttggttt ttaccaattg caattataat cttaccaaata tgctttcact ttttctgtg 780
 aatgatttta cttgtagtca aatatctcca gcagcaattg ctagcaactg ttattcttca 840
 45 ctgatttttg attacttttc ataccactt agtatgaaat ccgatctcag tgtagttct 900
 gctggtccaa tatcccagtt taattataaa cagtcctttt ctaatccac atgtttgatt 960
 ttagcgactg ttcctcataa cttactact attactaagc ctcttaagta cagctatatt 1020
 50 aacaagtgtc ctcgtcttct ttctgatgat cgtactgaag tacctcagtt agtgaacgct 1080
 aatcaatact caccctgtgt atccattgtc ccatccactg tgtgggaaga cggtgattat 1140
 tataggaaac aactatctcc acttgaagggt ggtggctggc ttgttgctag tggctcaact 1200
 55 gttgccatga ctgagcaatt acagatgggc tttggtatta cagttcaata tggtagacac 1260

accaatagtg tttgccccaa gctt

1284

<210> 16
 <211> 1275
 <212> DNA
 <213> Artificial Sequence

<220>
 <221> CDS
 <222> (1)..(1275)
 <223> A nucleotide sequence encoding two repeated amino acid sequences
 of SEQ ID NO: 3 linked in tandem by the GGSGGSGGS linker sequence

<400> 16
 gttgaatgtg atttttcacc tcttctgtct ggcacacctc ctcaggttta taatttcaag 60
 cgtttggttt ttaccaattg caattataat cttaccaaatt tgctttcact tttttctgtg 120
 aatgatttta ctgtagtca aatatctcca gcagcaattg ctagcaactg ttattcttca 180
 ctgatttttg attacttttc ataccactt agtatgaaat ccgatctcag tgtagttct 240
 gctggtccaa tatcccagtt taattataaa cagtcctttt ctaatccac atgtttgatt 300
 ttagcgactg ttcctcataa cttactact attactaagc ctcttaagta cagctatatt 360
 aacaagtgtc ctctgttctt ttctgatgat cgtactgaag tacctcagtt agtgaacgct 420
 aatcaatact caccctgtgt atccattgtc ccatccactg tgtgggaaga cggtgattat 480
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 gttgccatga ctgagcaatt acagatgggc tttgggtatta cagttcaata tggtagacagac 600
 accaatagtg tttgccccaa gcttggcggc tcaggcggct caggcggctc agttgaatgt 660
 gatttttcac ctcttctgtc tggcacacct cctcaggttt ataatttcaa gcgtttggtt 720
 tttaccaatt gcaattataa tcttaccaa ttgctttcac ttttttctgt gaatgatttt 780
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 actgagcaat tacagatggg ctttgggtatt acagttcaat atggtacaga caccaatagt 1260
 gtttgcccca agctt 1275

<210> 17
 <211> 1257

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<212> DNA
<213> Artificial Sequence

5 <220>
<221> CDS
<222> (1) .. (1257)
<223> A nucleotide sequence encoding two repeated amino acid sequences
of SEQ ID NO: 3 linked in tandem by a GGS linker sequence

10 <400> 17
gttgaatgtg atttttcacc tcttctgtct ggcacacctc ctcaggttta taatttcaag 60
cgtttggttt ttaccaattg caattataat cttaccaaatt tgctttcact tttttctgtg 120
15 aatgatttta cttgtagtca aatatctcca gcagcaattg ctagcaactg ttattcttca 180
ctgatttttg attacttttc ataccactt agtatgaaat ccgatctcag tgtagttct 240
gctggtccaa tatcccagtt taattataaa cagtcctttt ctaatccac atgtttgatt 300
20 tttagcgactg ttcctcataa cttactact attactaagc ctcttaagta cagctatatt 360
aacaagtgtc ctctgttctt ttctgatgat cgtactgaag tacctcagtt agtgaacgct 420
aatcaatact caccctgtgt atccattgtc ccatccactg tgtgggaaga cggtgattat 480
25 tataggaaac aactatctcc acttgaaggt ggtggctggc ttgttgctag tggtcaact 540
gttgccatga ctgagcaatt acagatgggc tttggatta cagttcaata tggtagacag 600
accaatagtg tttgccccaa gcttggcggc tcagttgaat gtgatttttc acctcttctg 660
30 tctggcacac ctctcaggt ttataatttc aagcgttttg tttttaccaa ttgcaattat 720
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40 gatcgtagtg aagtacctca gtagtgaac gctaataaat actcaccctg tgtatccatt 1080
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45 ggtggtggct ggcttgttgc tagtggctca actgttgcca tgactgagca attacagatg 1200
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50 <210> 18
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55 <220>
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<223> Nucleotide sequence encoding two repeated amino acid sequences of
SEQ ID NO: 3 directly linked in tandem

<400> 18

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  cgtttggttt ttaccaattg caattataat cttaccaaatt tgctttcact tttttctgtg      120
  aatgatttta cttgtagtca aatatctcca gcagcaattg ctagcaactg ttattcttca      180
10 ctgatttttg attacttttc ataccactt agtatgaaat cggatctcag tgtagttct      240
  gctggtccaa tatcccagtt taattataaa cagtcctttt ctaatccac atgtttgatt      300
  ttgacgactg ttcctcataa cttactact attactaagc ctcttaagta cagctatatt      360
15 aacaagtgct ctogtcttct ttctgatgat cgtactgaag tacctcagtt agtgaacgct      420
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20 gttgccatga ctgagcaatt acagatgggc tttggtatta cagttcaata tggtagacagac      600
  accaatagtg tttgccccaa gcttggtgaa tgtgattttt cacctcttct gtctggcaca      660
  cctcctcagg ttataaattt caagcgtttg gtttttacca attgcaatta taatcttacc      720
25 aaattgcttt cacttttttc tgtgaatgat ttacttgta gtcaaataatc tccagcagca      780
  attgctagca actgttattc ttactgatt ttggattact tttcataccc acttagtatg      840
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  aagcctctta agtacagcta tattaacaag tgctctcgtc ttctttctga tgatcgctact     1020
35 gaagtacctc agttagttaa cgctaataca tactcaccct gtgtatccat tgtcccatcc     1080
  actgtgtggg aagacggtga ttattatagg aaacaactat ctccacttga aggtggtggc     1140
  tggcttggtg ctagtggctc aactgttgcc atgactgagc aattacagat gggctttggt     1200
40 attacagttc aatatggtac agacaccaat agtgtttgcc ccaagctt     1248

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<210> 19

<211> 1272

<212> DNA

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<220>

<221> CDS

<222> (1) .. (1272)

<223> Nucleotide sequence encoding two repeated amino acid sequences of
SEQ ID NO: 5 directly linked in tandem

<400> 19

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55 agagtgaac ctacagaatc aatcgtgaga tttcctaaca tcacaaacct ttgccctttc      60
  ggcgaggtgt ttaacgcaac aagatttga tcagtgtacg catggaacag aaagcgtata      120

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	tcaaactgcg tggcagatta ctcagtgtct tacaactcag catcattcag tacgtttaaa	180
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5	tcattttgtga tcagaggaga tgaagtgaga caaatcgac ctggacaaac aggaaagatt	300
	gccgattaca actacaaact tcctgatgat ttcaccggct gcgtgatcgc atggaactca	360
	aacaaccttg attcaaagg aggtggtaat tataattatt tgtataggct ctttcgtaag	420
10	agcaacttaa agccatttga gcgagatata tcaacagaaa tctaccaagc aggatcaaca	480
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	cctacaaacg gagtgggata ccaaccttac agagtgggtg tgctttcatt tgaacttctt	600
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25	gtgagacaaa tcgcacctgg acaaacagga aagattgccg attacaacta caaacttctt	960
	gatgatttca ccggctgcgt gatcgcagtg aactcaaaca accttgattc aaaggtaggt	1020
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30	gatattctcaa cagaaatcta ccaagcagga tcaacacctt gcaacggagt ggaaggattt	1140
	aactgctact ttcctcttca atcatcacgga tttcaacctt caaacggagt gggataccaa	1200
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	ggcgagggtgt ttaacgcaac aagatttgca tcagtgtacg catggaacag aaagcgtata	120
55	tcaaactgcg tggcagatta ctcagtgtct tacaactcag catcattcag tacgtttaaa	180
	tgctacggag tgtcacctac aaagctaaat gatctttgct ttacaaacgt gtacgcagat	240

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	tcattttgtga tcagaggaga tgaagtgaga caaatcgcac ctggacaaac aggaaagatt	300
	gccgattaca actacaaact tcctgatgat ttcaccggct gcgtgatcgc atggaactca	360
5	aacaaccttg attcaaaggt aggtggtaat tataattatt tgtataggct ctttcgtaag	420
	agcaacttaa agccatttga gcgagatatc tcaacagaaa tctaccaagc aggatcaaca	480
	ccttgcaacg gagtggaagg atttaactgc tactttcctc ttcaatcata cggatttcaa	540
10	cctacaaacg gagtgggata ccaaccttac agagtgggtg tgctttcatt tgaacttctt	600
	cacgcacctg caacagtgtg cggacctaa gagagcacga accttgtgaa gaataagaga	660
	gtgcaaccta cagaatcaat cgtgagattt cctaacaatca caaacctttg ccctttcggc	720
15	gaggtgttta acgcaacaag atttgcata gtgtacgcat ggaacagaaa gcgtatatca	780
	aactgcgtgg cagattactc agtgctttac aactcagcat cattcagtac gtttaaagtc	840
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	gattacaact acaaacttcc tgatgatttc accggctgcg tgatcgcatg gaactcaaac	1020
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25	aacttaaagc catttgagcg agatatctca acagaaatct accaagcagg atcaaacacct	1140
	tgcaacggag tggaaggatt taactgctac tttcctcttc aatcatagcg atttcaacct	1200
30	acaaacggag tgggatacca accttacaga gtggtggtgc tttcatttga acttcttcac	1260
	gcacctgcaa cagtgtgagg acctaagaag agcacgaacc ttgtgaagaa taag	1314
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40	<220> <221> CDS <222> (1)..(1338) <223> Nucleotide sequence encoding two repeated amino acid sequences of SEQ ID NO: 7 directly linked in tandem	
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50	tcaaactgcg tggcagatta ctacgtgctt tacaactcag catcattcag tacgttttaa	180
	tgctacggag tgtcacctac aaagctaaat gatctttgct ttacaaacgt gtacgcagat	240
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	aacaaccttg attcaaaggt aggtggtaat tataattatt tgtataggct ctttcgtaag	420

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agcaacttaa agccatttga gcgagatatc tcaacagaaa tctaccaagc aggatcaaca 480
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 5 cctacaaacg gagtgggata ccaaccttac agagtgggtg tgctttcatt tgaacttctt 600
 cacgcacctg caacagtgtg cggacctaag aagagcacga accttgtgaa gaataagtgc 660
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 aataagtgcg tgaacttt 1338
 30
 <210> 22
 <211> 669
 <212> DNA
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 35
 <220>
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 <222> (1)..(669)
 40 <223> The nucleotide sequence encoding R319-F541 amino acid sequence in
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 55 aacaaccttg attcaaagg aggtggtaat tataattatt tgtataggct ctttcgtaag 420
 agcaacttaa agccatttga gcgagatatc tcaacagaaa tctaccaagc aggatcaaca 480

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	ccttgcaacg gagtgggaagg atttaactgc tactttcctc ttcaatcata cggatttcaa	540
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25	agcaattgtg ttgcggacta ttccgtacta tataattoga ctttcttcag caccttcaag	180
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<220>
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ttacagatgg gctttggtat tacagttcaa tatggtacag acaccaatag tgtttgcccc 660
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<210> 25
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<212> DNA
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<220>
<221> CDS
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tcttcactga ttttgatta cttttcatat ccacttagta tgaaatccga tctcagtgtt      240
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5  ttgatttttag cgactgttcc tcataacctt actactatta ctaagcctct taagtacagc      360
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tcaactgttg ccatgactga gcaattacag atgggctttg gtattacagt tcaatatggt      600
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<210> 26
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<212> PRT
20 <213> SARS-CoV

<220>
<221> DOMAIN
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25 <223> R306-F527 amino acid sequence in the RBD of the S protein
sequence of SARS-CoV (GenBank on NCBI: AAR07630)

<400> 26

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1 5 10 15

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Leu Cys Pro Phe Gly Glu Val Phe Asn Ala Thr Lys Phe Pro Ser Val
20 25 30
35

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Tyr Ala Trp Glu Arg Lys Lys Ile Ser Asn Cys Val Ala Asp Tyr Ser
35 40 45

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40 Val Leu Tyr Asn Ser Thr Phe Phe Ser Thr Phe Lys Cys Tyr Gly Val
50 55 60

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45 Ser Ala Thr Lys Leu Asn Asp Leu Cys Phe Ser Asn Val Tyr Ala Asp
65 70 75 80

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Ser Phe Val Val Lys Gly Asp Asp Val Arg Gln Ile Ala Pro Gly Gln
85 90 95
50

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Thr Gly Val Ile Ala Asp Tyr Asn Tyr Lys Leu Pro Asp Asp Phe Met
100 105 110

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55 Gly Cys Val Leu Ala Trp Asn Thr Arg Asn Ile Asp Ala Thr Ser Thr
115 120 125

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Gly Asn Tyr Asn Tyr Lys Tyr Arg Tyr Leu Arg His Gly Lys Leu Arg
130 135 140

5 Pro Phe Glu Arg Asp Ile Ser Asn Val Pro Phe Ser Pro Asp Gly Lys
145 150 155 160

10 Pro Cys Thr Pro Pro Ala Leu Asn Cys Tyr Trp Pro Leu Asn Asp Tyr
165 170 175

Gly Phe Tyr Thr Thr Thr Gly Ile Gly Tyr Gln Pro Tyr Arg Val Val
180 185 190

15 Val Leu Ser Phe Glu Leu Leu Asn Ala Pro Ala Thr Val Cys Gly Pro
195 200 205

20 Lys Leu Ser Thr Asp Leu Ile Lys Asn Gln Cys Val Asn Phe
210 215 220

25 <210> 27
<211> 666
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30 <220>
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<222> (1) .. (666)
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the RBD encoding the S protein sequence of SARS-CoV (GenBank on
NCBI: AAR07630)

35 <400> 27
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40 tcaaattgtg ttgccgacta ttccgtccta tataatagca cggtcttctc gacgttcaag 180
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50 ccgtgcactc caccggcttt aaattgttat tggccggttaa acgactatgg cttttataca 540
acgacgggaa tagggtagca accttacaga gtagtagtac taagtttcga gctattaaat 600
55 gcgccggcca ccgtatgtgg gcccaagcta tcgacggacc taatcaagaa tcagtgtgtt 660
aatttc 666

Claims

1. An antigen of a β -coronavirus, its amino acid comprising an amino acid sequence arranged in a (A-B)-(A-B) pattern or an amino acid sequence arranged in a (A-B)-C-(A-B) pattern or an amino acid sequence arranged in a (A-B)-(A-B')
- 5 (A-B)-C-(A-B')
- 10 pattern, wherein A-B represents a partial amino acid sequence or an entire amino acid sequence of a receptor binding domain of a surface spike protein of the β -coronavirus; C represents an amino acid linker sequence; A-B' represents an amino acid sequence obtained by substitution, deletion or addition of one or more amino acids in the amino acid sequence of A-B; a protein encoded by A-B' has an identical or a substantially identical immunogenicity as a protein encoded by A-B; and the antigen of the β -coronavirus has a single-chain dimer structure.
2. The antigen of the β -coronavirus according to claim 1, wherein the β -coronavirus is selected from a group consisting of severe respiratory syndrome coronavirus, Middle East respiratory syndrome coronavirus, and 2019 novel coronavirus.
- 15 3. The antigen of the β -coronavirus according to claim 1, wherein the amino acid linker sequence comprises a (GGS)_n linker sequence, wherein n represents the number of GGSs, n is an integer more than or equal to 1; preferably, n is an integer selected from 1 to 10, and further preferably, n is an integer selected from 1 to 5.
- 20 4. The antigen of the β -coronavirus according to claim 1, wherein the partial amino acid sequence of the receptor binding domain of the surface spike protein of the β -coronavirus is at least 50%, 60%, 70%, 80%, 90%, 95%, or 99% of the entire amino acid sequence of the receptor binding domain of the surface spike protein of the β -coronavirus.
- 25 5. The antigen of the β -coronavirus according to claim 1, wherein:
when the β -coronavirus is the Middle East respiratory syndrome coronavirus, the partial or the entire amino acid sequence of the receptor binding domain of the surface spike protein thereof is any one selected from a group consisting of following amino acid sequences:
 - (1) SEQ ID NO: 1, SEQ ID NO: 2, or SEQ ID NO: 3;
 - 30 (2) an amino acid sequence obtained by substitution, deletion or addition of one or more amino acids in the amino acid sequence (1), wherein a protein encoded by the amino acid sequence has an identical or substantially identical immunogenicity as a protein encoded by the amino acid sequence (1);
 - alternatively, the partial amino acid sequence of the receptor binding domain of the surface spike protein thereof comprises SEQ ID NO: 2;
 - 35 when the β -coronavirus is the 2019 novel coronavirus, the partial or the entire amino acid sequence of the receptor binding domain of the surface spike protein thereof is any one selected from a group consisting of following amino acid sequences:
 - (3) SEQ ID NO: 5, SEQ ID NO: 6, or SEQ ID NO: 7;
 - 40 (4) an amino acid sequence obtained by substitution, deletion or addition of one or more amino acids in the amino acid sequence (3), wherein a protein encoded by the amino acid sequence has an identical or substantially identical immunogenicity as a protein encoded by the amino acid sequence (3);
 - alternatively, the partial amino acid sequence of the receptor binding domain of the surface spike protein comprises SEQ ID NO: 6; and
 - 45 when the β -coronavirus is the severe respiratory syndrome coronavirus, the partial or the entire amino acid sequence of the receptor binding domain of the surface spike protein thereof is any one selected from a group consisting of following amino acid sequences:
 - 50 (5) SEQ ID NO: 8;
 - (6) an amino acid sequence obtained by substitution, deletion or addition of one or more amino acids in the amino acid sequence (5), wherein a protein encoded by the amino acid sequence has an identical or substantially identical immunogenicity as a protein encoded by the amino acid sequence (5);
 - 55
6. The antigen of the β -coronavirus according to claim 5, wherein:
when the β -coronavirus is the Middle East respiratory syndrome coronavirus, the amino acid sequence of the antigen of the β -coronavirus comprises any one selected from a group consisting of following amino acid sequences:

- (1) two repeated amino acid sequences of SEQ ID NO: 1 linked in tandem by a GGSGGS linker sequence;
- (2) two repeated amino acid sequences of SEQ ID NO: 1 linked in tandem by a GGS linker sequence;
- (3) two repeated amino acid sequences of SEQ ID NO: 1 linked directly in tandem;
- (4) two repeated amino acid sequences of SEQ ID NO: 2 linked in tandem by a GGS linker sequence;
- (5) two repeated amino acid sequences of SEQ ID NO: 2 linked directly in tandem;
- (6) two repeated amino acid sequences of SEQ ID NO: 3 linked by a GGSGGSGGSGS linker sequence;
- (7) two repeated amino acid sequences of SEQ ID NO: 3 linked in tandem by a GGSGGSGGSGS linker sequence;
- (8) two repeated amino acid sequences of SEQ ID NO: 3 linked in tandem by a GGSGGSGGSGS linker sequence;
- (9) two repeated amino acid sequences of SEQ ID NO: 3 linked in tandem by a GGS linker sequence; and
- (10) two repeated amino acid sequences of SEQ ID NO: 3 linked directly in tandem;

alternatively, the amino acid sequence of the antigen of the β -coronavirus comprises two repeated amino acid sequences of SEQ ID NO: 2 directly linked in tandem;

when the β -coronavirus is the 2019 novel coronavirus, the amino acid sequence of the antigen of the β -coronavirus comprises any one selected from a group consisting of following amino acid sequences:

- (1) two repeated amino acid sequences of SEQ ID NO: 5 linked directly in tandem;
- (2) two repeated amino acid sequences of SEQ ID NO: 6 linked directly in tandem; and
- (3) two repeated amino acid sequences of SEQ ID NO: 7 linked directly in tandem;

alternatively, the amino acid sequence of the antigen of the β -coronavirus comprises two repeated amino acid sequences of SEQ ID NO: 6 directly linked in tandem; and

when the β -coronavirus is the severe respiratory syndrome coronavirus, the amino acid sequence of the antigen of the β -coronavirus comprises two repeated amino acid sequences of SEQ ID NO: 8 linked directly in tandem.

7. The antigen of the β -coronavirus according to claim 6, wherein:

- a nucleotide sequence encoding the two repeated amino acid sequences of SEQ ID NO: 1 linked in tandem by the GGSGGS linker sequence is shown as SEQ ID NO: 9;
- a nucleotide sequence encoding the two repeated amino acid sequences of SEQ ID NO: 1 linked in tandem by the GGS linker sequence is shown as SEQ ID NO: 10;
- a nucleotide sequence encoding the two repeated amino acid sequences of SEQ ID NO: 1 linked directly in tandem is shown as SEQ ID NO: 11;
- a nucleotide sequence encoding the two repeated amino acid sequences of SEQ ID NO: 2 linked in tandem by the GGS linker sequence is shown as SEQ ID NO: 12;
- a nucleotide sequence encoding the two repeated amino acid sequences of SEQ ID NO: 2 linked directly in tandem is shown as SEQ ID NO: 13;
- a nucleotide sequence encoding the two repeated amino acid sequences of SEQ ID NO: 3 linked in tandem by the GGSGGSGGSGGSGS linker sequence is shown as SEQ ID NO: 14;
- a nucleotide sequence encoding the two repeated amino acid sequences of SEQ ID NO: 3 linked in tandem by the GGSGGSGGSGGSGS linker sequence is shown as SEQ ID NO: 15;
- a nucleotide sequence encoding the two repeated amino acid sequences of SEQ ID NO: 3 linked in tandem by the GGSGGSGGSGS linker sequence is shown as SEQ ID NO: 16;
- a nucleotide sequence encoding the two repeated amino acid sequences of SEQ ID NO: 3 linked in tandem by the GGS linker sequence is shown as SEQ ID NO: 17;
- a nucleotide sequence encoding the two repeated amino acid sequences of SEQ ID NO: 3 linked directly in tandem is shown as SEQ ID NO: 18;
- a nucleotide sequence encoding the two repeated amino acid sequences of SEQ ID NO: 5 linked directly in tandem is shown as SEQ ID NO: 19;
- a nucleotide sequence encoding the two repeated amino acid sequences of SEQ ID NO: 6 linked directly in tandem is shown as SEQ ID NO: 20;
- a nucleotide sequence encoding the two repeated amino acid sequences of SEQ ID NO: 7 linked directly in tandem is shown as SEQ ID NO: 21; and
- a nucleotide sequence encoding the two repeated amino acid sequences of SEQ ID NO: 8 linked directly in tandem is shown as SEQ ID NO: 23.

- 5 8. A method for preparing the antigen of the β -coronavirus according to any one of claims 1-7, comprising following steps: adding a sequence encoding a signal peptide to a 5'-terminal of a nucleotide sequence encoding the antigen of the β -coronavirus according to any one of claims 1-7, adding a terminator codon to a 3'-terminal for cloning and expression, screening a correct recombinant, transfecting an expression system cell for expression, collecting a cell supernatant after expression, and purifying to obtain the antigen of the β -coronavirus.
- 10 9. The method according to claim 8, wherein the expression system cell is selected from a group consisting of mammalian cell, insect cell, yeast cell, and bacterial cell; preferably, the mammalian cell is 293T cell or CHO cell, and the bacterial cell is *Escherichia coli* cell.
11. A nucleotide sequence encoding the antigen of the β -coronavirus according to any one of claims 1-7.
11. A recombinant vector comprising the nucleotide sequence according to claim 10.
- 15 12. An expression system cell comprising the recombinant vector according to claim 11.
13. Use of the antigen of the β -coronavirus according to any one of claims 1-7, the nucleotide sequence according to claim 10, the recombinant vector according to claim 11, and the expression system cell according to claim 12 in the preparation of a vaccine against the β -coronavirus.
- 20 14. A β -coronavirus vaccine, comprising the antigen of the β -coronavirus according to any one of claims 1-7 and an adjuvant.
15. The β -coronavirus vaccine according to claim 14, wherein the adjuvant is selected from a group consisting of an aluminum adjuvant, an MF59 adjuvant, and an MF59-like adjuvant.
- 25 16. A β -coronavirus DNA vaccine, comprising a recombinant vector comprising a DNA sequence encoding the antigen of the β -coronavirus according to any one of claims 1-7.
- 30 17. A β -coronavirus RNA vaccine, comprising a recombinant vector comprising an mRNA sequence encoding the antigen of the β -coronavirus according to claims 1-7.
- 35 18. A β -coronavirus viral vector vaccine comprising a recombinant viral vector comprising a nucleotide sequence encoding the antigen of the β -coronavirus according to any one of claims 1-7, alternatively, the viral vector is one or more selected from a group consisting of an adenovirus vector, a poxvirus vector, an influenza virus vector, and an adeno-associated virus vector.
- 40
- 45
- 50
- 55

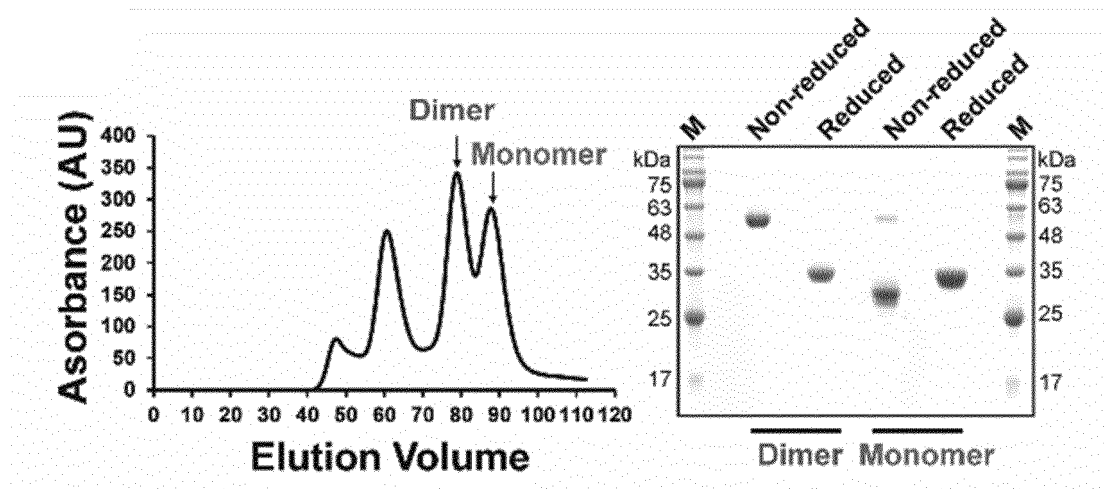


FIG. 1

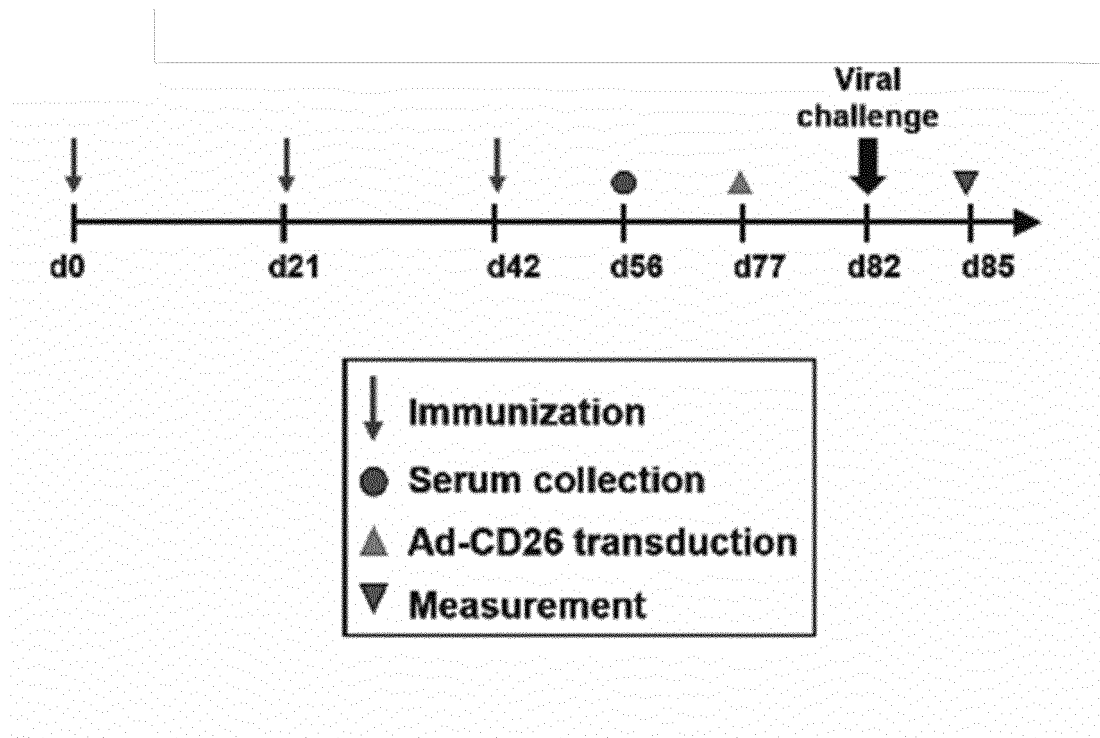


FIG. 2

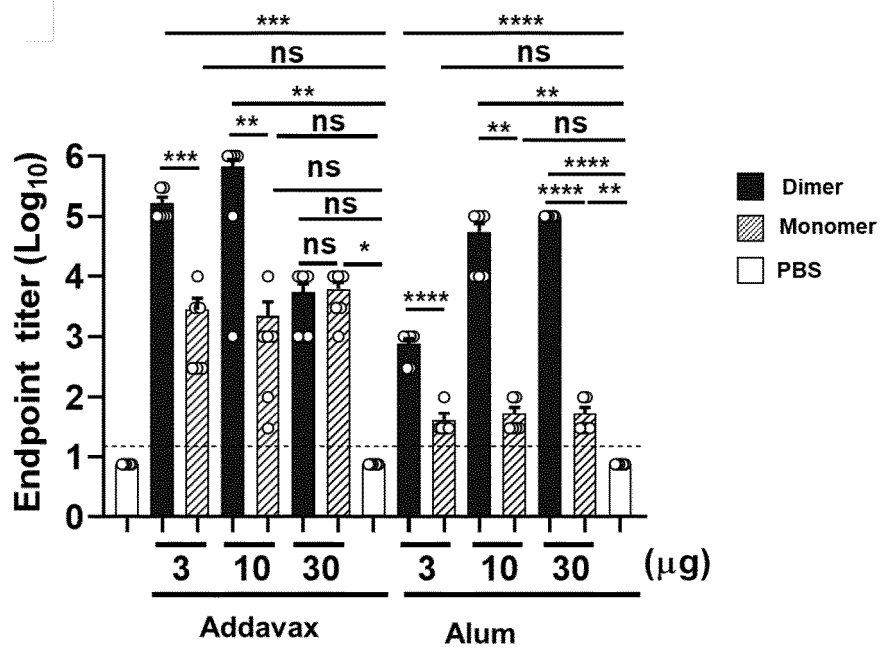


FIG. 3

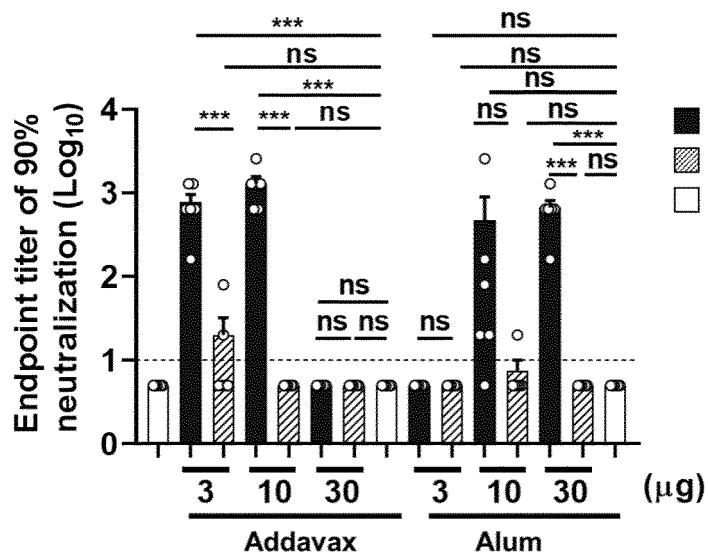


FIG. 4

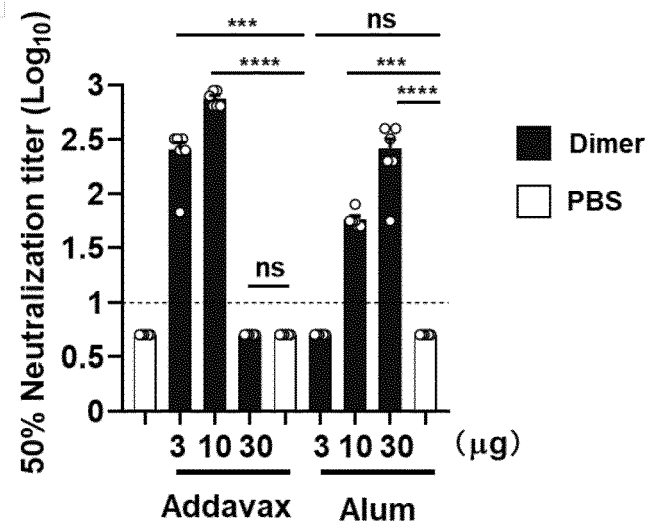


FIG. 5

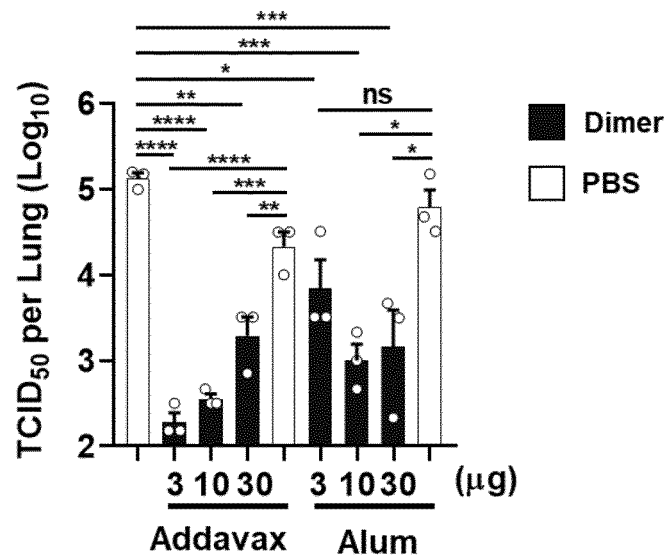


FIG. 6

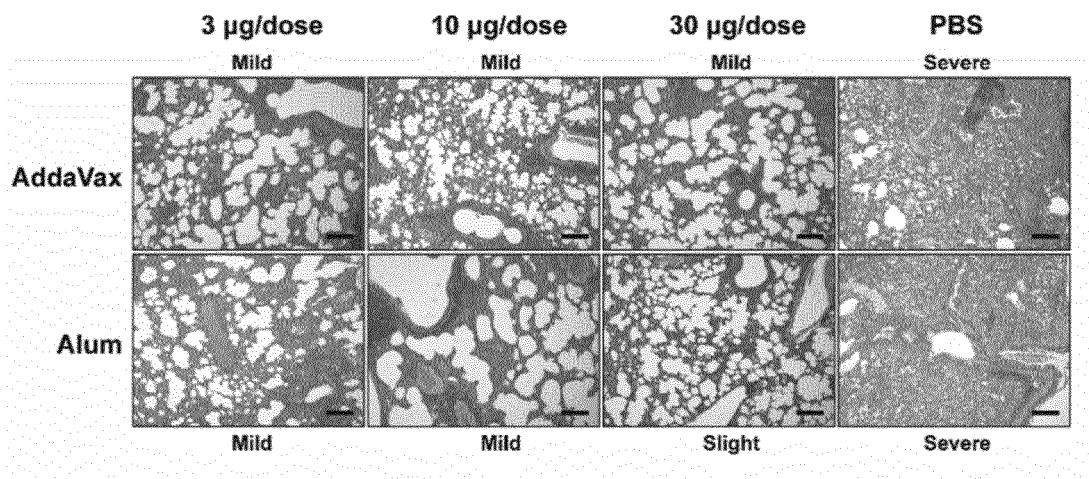


FIG. 7

MERS-CoV-RBD dimer (E367-Y606)

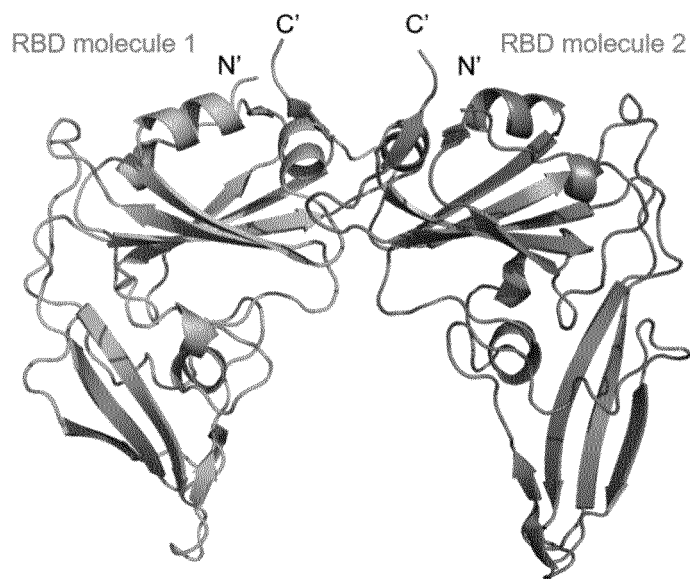


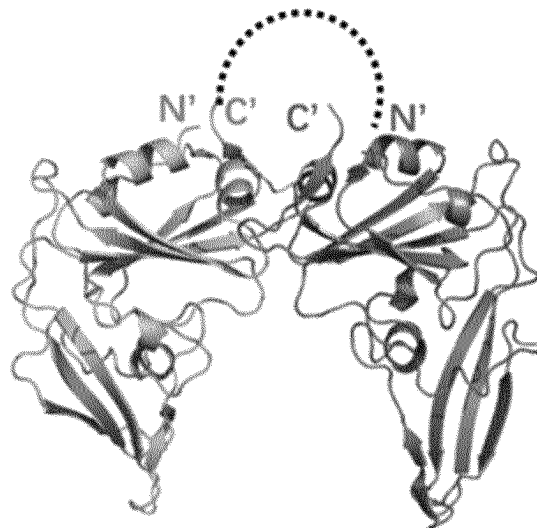
FIG. 8



FIG. 9A



FIG. 9B



C6	V381-L588	-(GGG) ₅	V381-L588
C7	V381-L588	-(GGG) ₄	V381-L588
C8	V381-L588	-(GGG) ₃	V381-L588
C9	V381-L588	GGG	V381-L588
C10	V381-L588		V381-L588

FIG. 9C

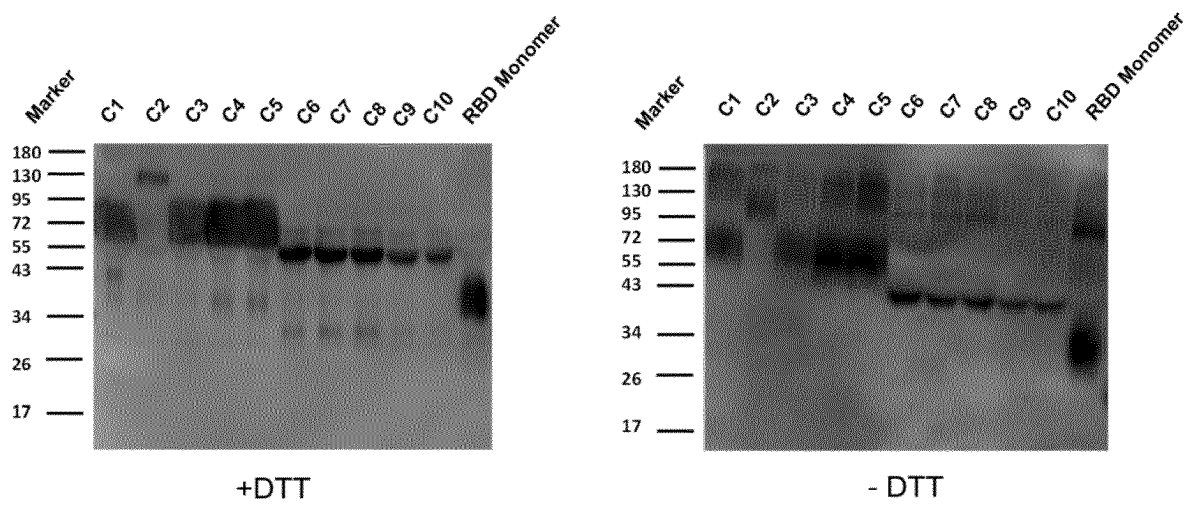


FIG. 10

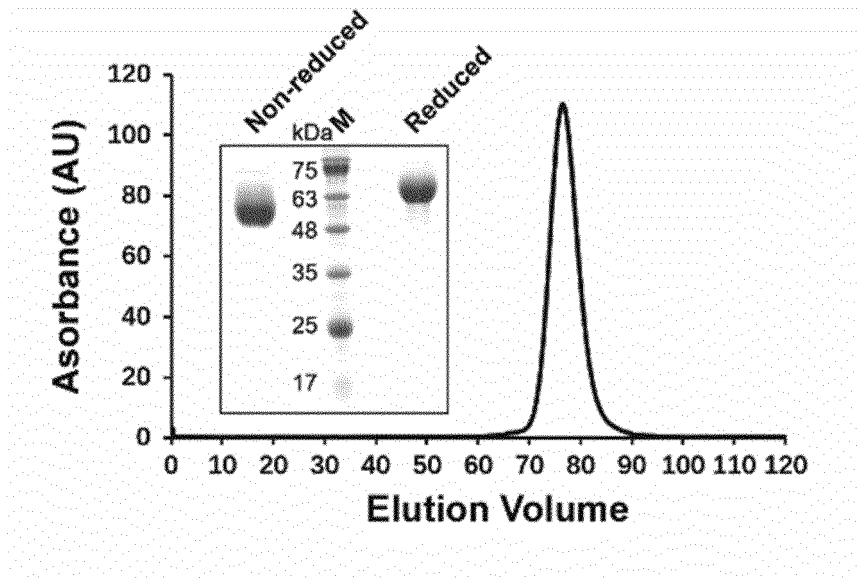


FIG. 11

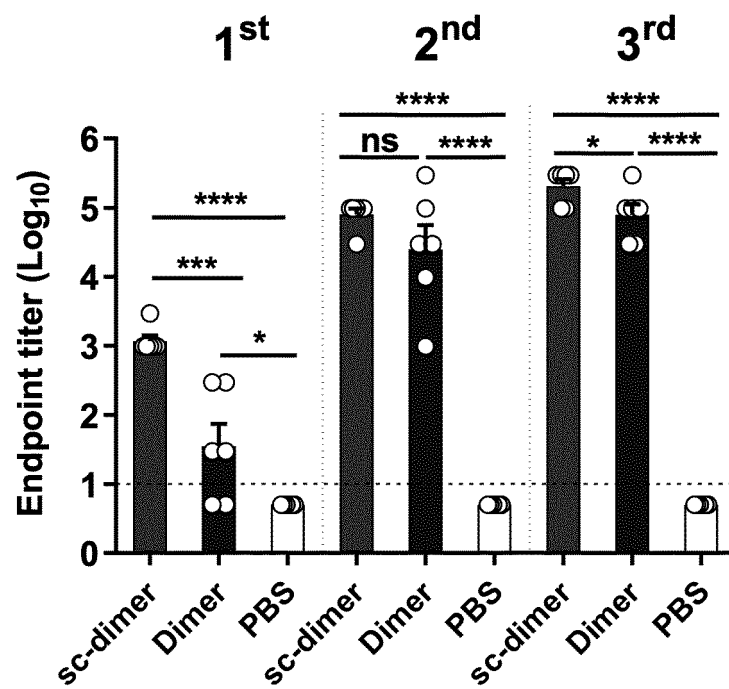


FIG. 12

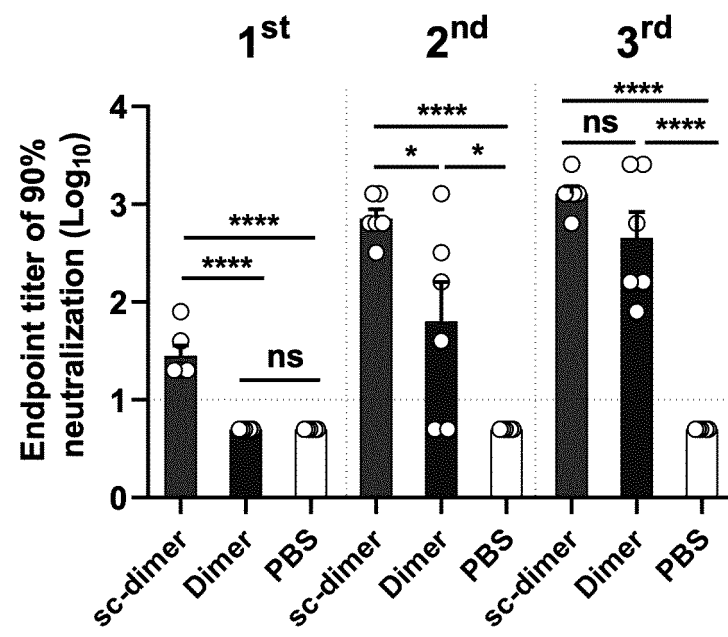
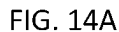


FIG. 13



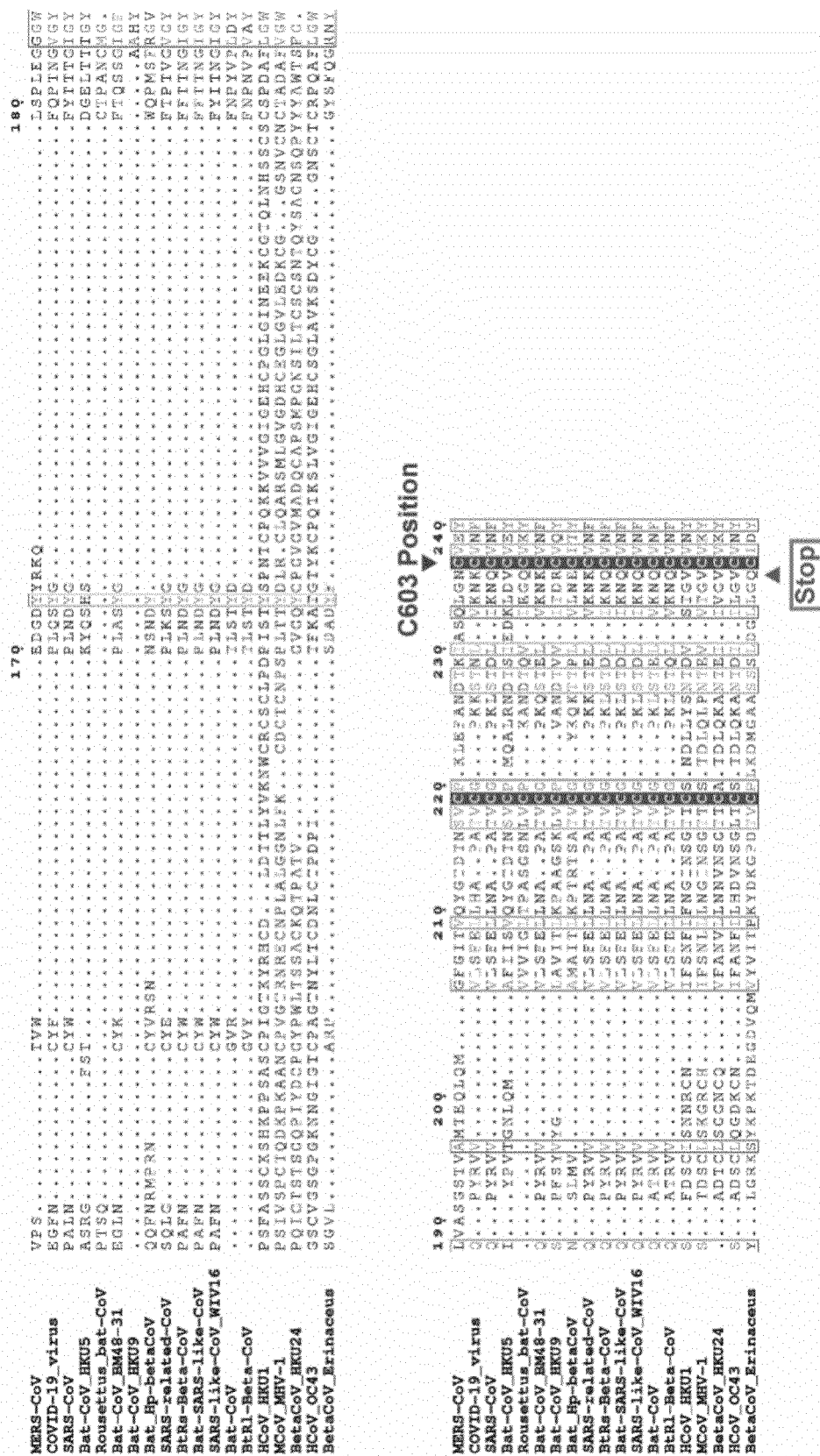


FIG. 14B

SARS-CoV-RBD dimer / 2019-nCoV-RBD dimer (Model)

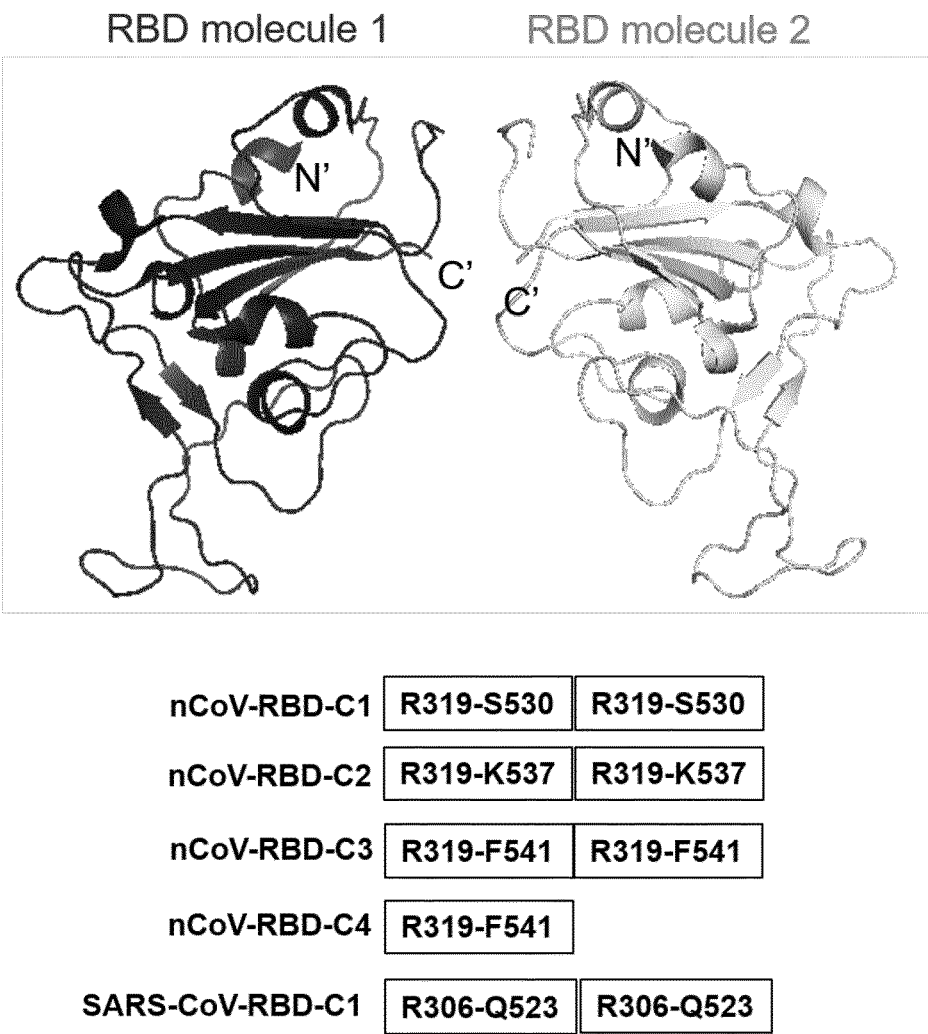


FIG. 15

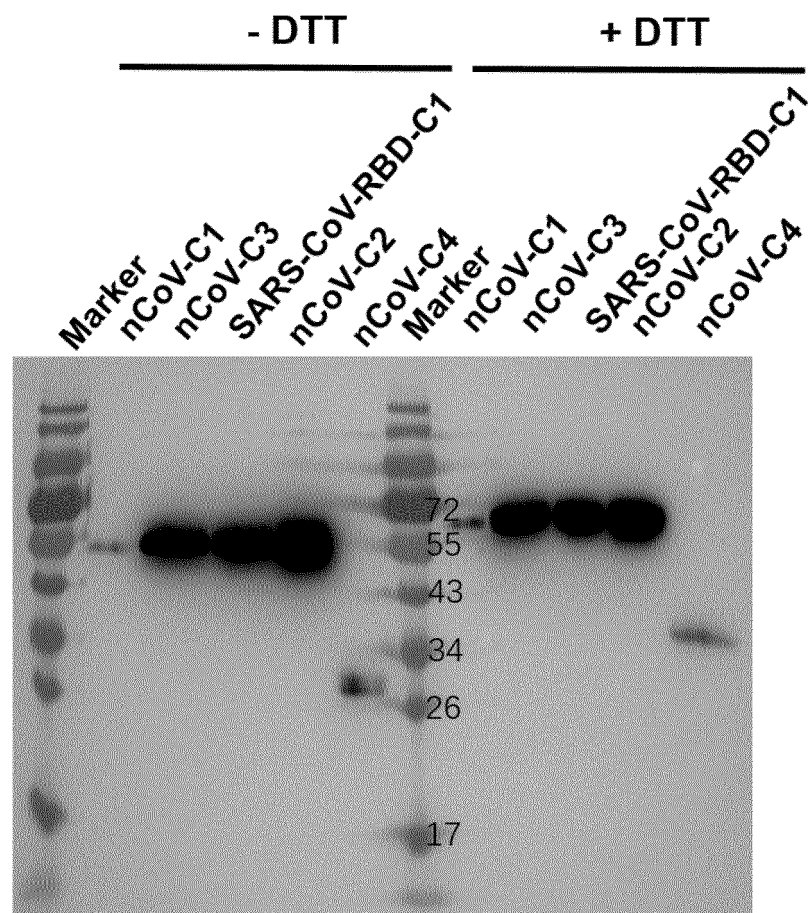


FIG. 16

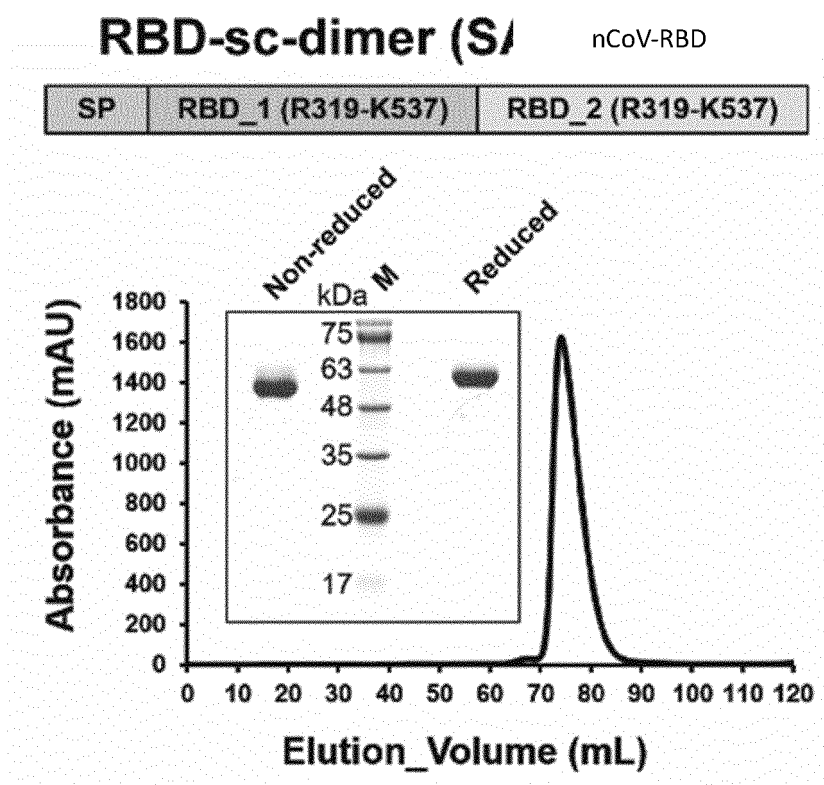


FIG. 17

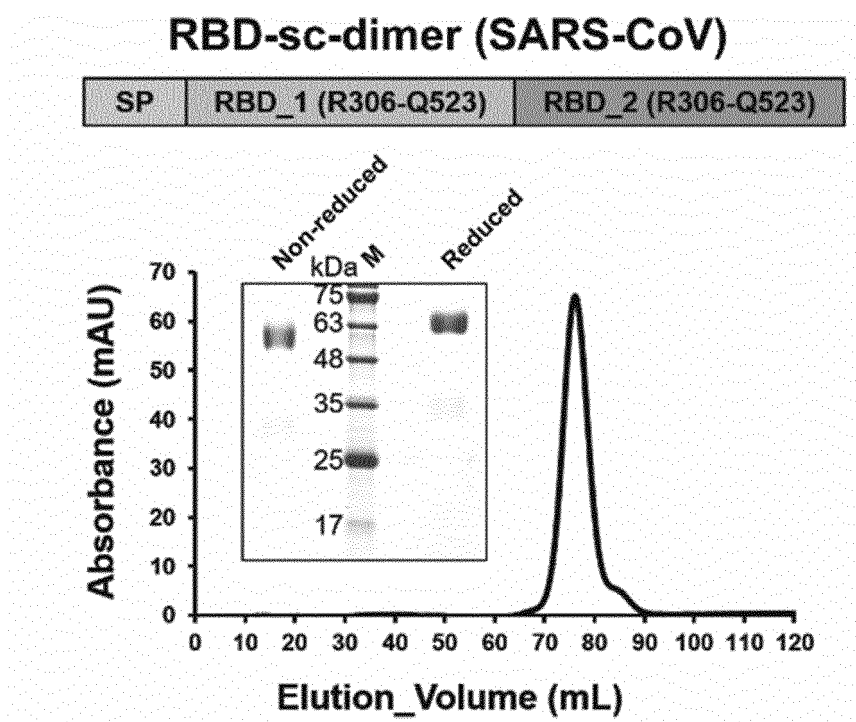


FIG. 18

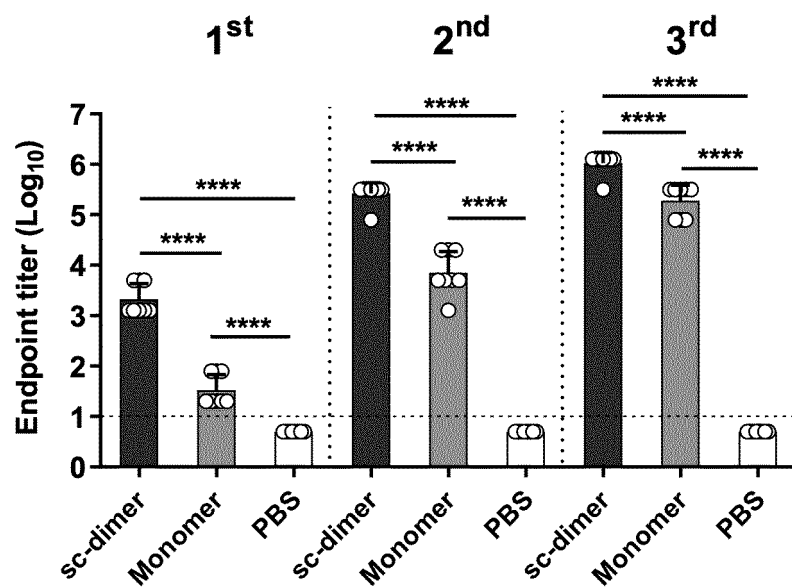


FIG. 19

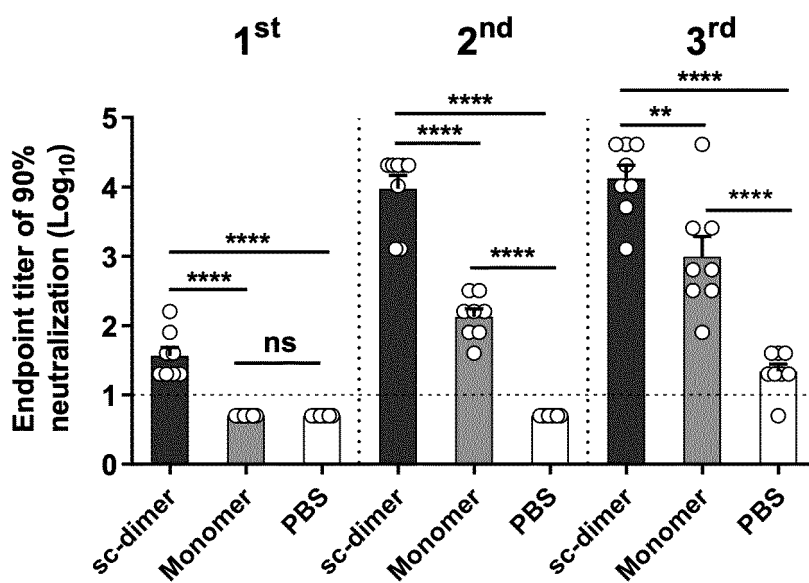


FIG. 20

**Live SARS-CoV-2
neutralization
(2nd immunization)**

Group	Serum ID	NT ₅₀
sc-dimer	1	4096
	2	1024
	3	2048
	4	>4096
	5	>4096
	6	512
	7	4096
	8	4096
Monomer	9	128
	10	256
	11	<16
	12	<16
	13	<16
	14	<16
	15	<16
	16	<16
PBS	17	<16
	18	<16
	19	<16
	20	<16
	21	<16
	22	<16
	23	<16
	24	<16

FIG. 21

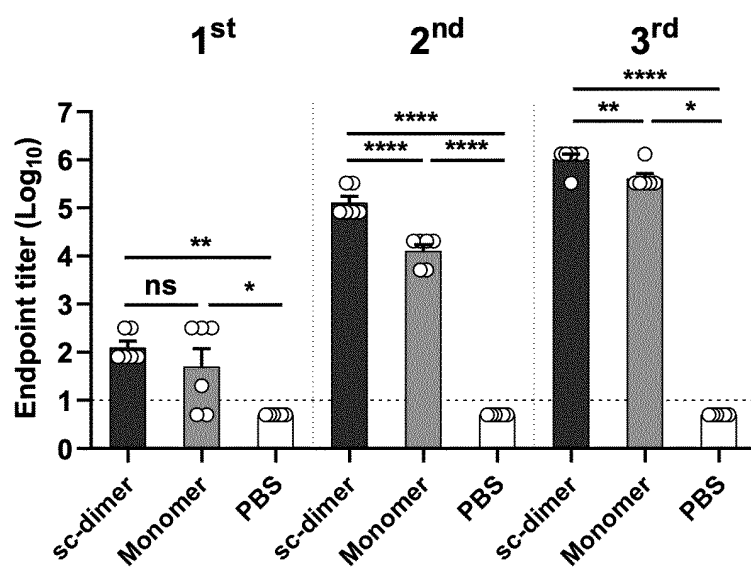


FIG. 22

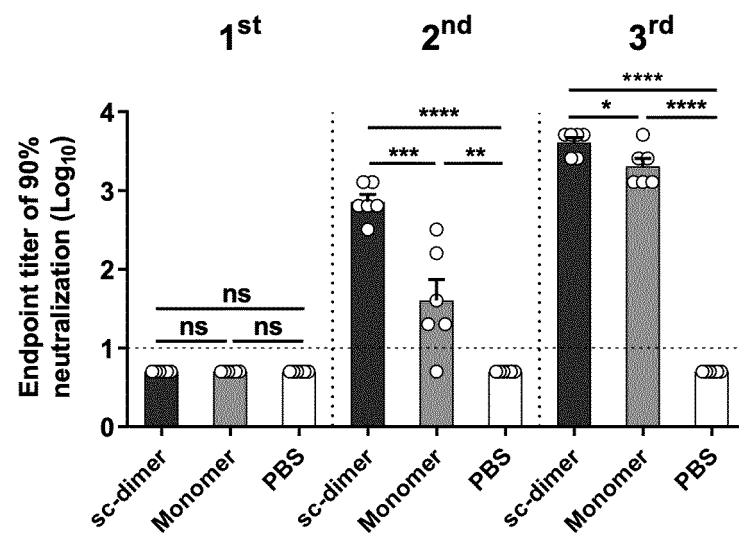


FIG. 23

INTERNATIONAL SEARCH REPORT

International application No.

PCT/CN2020/097775

A. CLASSIFICATION OF SUBJECT MATTER

C07K 19/00(2006.01)i; C12N 15/09(2006.01)i; A61K 39/215(2006.01)i; A61P 31/14(2006.01)i

According to International Patent Classification (IPC) or to both national classification and IPC

B. FIELDS SEARCHED

Minimum documentation searched (classification system followed by classification symbols)

C07K; C12N; A61K; A61P

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched

Electronic data base consulted during the international search (name of data base and, where practicable, search terms used)

CNABS, DWPI, SIPOABS, CNKI, NCBI, ISI Web of Science, GenBank, 中国专利生物序列检索系统: 冠状病毒, 突变, 蛋白, 受体, 结合, 域, 串联, 融合, 重复, 抗原, 疫苗, coronavirus, spike, S, protein, receptor, binding, domain, RBD, tandem, fusion, repeat, antigen, vaccine, 本申请的SEQ ID NOs: 1-8

C. DOCUMENTS CONSIDERED TO BE RELEVANT

Category*	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
Y	Xiaojie Zhu, et al. "Receptor-binding domain as a target for developing SARS vaccines" <i>Journal of Thoracic Disease</i> , Vol. 5, No. Suppl2, 31 August 2013 (2013-08-31), abstract	1-18
Y	Yu-Na Lee, et al. "Cross Protection against Influenza A Virus by Yeast-Expressed Heterologous Tandem Repeat M2 Extracellular Proteins" <i>PLOS ONE</i> , Vol. 10, No. 9, 14 September 2015 (2015-09-14), abstract	1-18
X	CN 107033250 A (DAIRY CATTLE RESEARCH CENTER OF SHANDONG ACADEMY OF AGRICULTURAL SCIENCES; POULTRY INSTITUTE, SHANDONG ACADEMY OF AGRICULTURAL SCIENCE; SHANDONG NORMAL UNIVERSITY) 11 August 2017 (2017-08-11) entire document, in particular claims and abstract	1-3, 5, 8-18

☐ Further documents are listed in the continuation of Box C.
 ☒ See patent family annex.

* Special categories of cited documents:	"T" later document published after the international filing date or priority date and not in conflict with the application but cited to understand the principle or theory underlying the invention
"A" document defining the general state of the art which is not considered to be of particular relevance	"X" document of particular relevance; the claimed invention cannot be considered novel or cannot be considered to involve an inventive step when the document is taken alone
"E" earlier application or patent but published on or after the international filing date	"Y" document of particular relevance; the claimed invention cannot be considered to involve an inventive step when the document is combined with one or more other such documents, such combination being obvious to a person skilled in the art
"L" document which may throw doubts on priority claim(s) or which is cited to establish the publication date of another citation or other special reason (as specified)	"&" document member of the same patent family
"O" document referring to an oral disclosure, use, exhibition or other means	
"P" document published prior to the international filing date but later than the priority date claimed	

Date of the actual completion of the international search

22 July 2020

Date of mailing of the international search report

12 November 2020

Name and mailing address of the ISA/CN

China National Intellectual Property Administration (ISA/
CN)
No. 6, Xitucheng Road, Jimenqiao, Haidian District, Beijing
100088
China

Authorized officer

Facsimile No. (86-10)62019451

Telephone No.

INTERNATIONAL SEARCH REPORT

International application No.

PCT/CN2020/097775

Box No. 1 Nucleotide and/or amino acid sequence(s) (Continuation of item 1.c of the first sheet)

1. With regard to any nucleotide and/or amino acid sequence disclosed in the international application, the international search was carried out on the basis of a sequence listing:
- ☒ forming part of the international application as filed:
 - ☒ in the form of an Annex C/ST.25 text file.
 - ☐ on paper or in the form of an image file.
 - ☐ furnished together with the international application under PCT Rule 13ter.1(a) for the purposes of international search only in the form of an Annex C/ST.25 text file.
 - ☐ furnished subsequent to the international filing date for the purposes of international search only:
 - ☐ in the form of an Annex C/ST.25 text file (Rule 13ter.1(a)).
 - ☐ on paper or in the form of an image file (Rule 13ter.1(b) and Administrative Instructions, Section 713).
2. ☐ In addition, in the case that more than one version or copy of a sequence listing has been filed or furnished, the required statements that the information in the subsequent or additional copies is identical to that forming part of the application as filed or does not go beyond the application as filed, as appropriate, were furnished.
3. Additional comments:

INTERNATIONAL SEARCH REPORT
Information on patent family members

International application No.

PCT/CN2020/097775

Patent document cited in search report	Publication date (day/month/year)	Patent family member(s)	Publication date (day/month/year)
CN 107033250 A	11 August 2017	CN 107033250 B	21 January 2020

Form PCT/ISA/210 (patent family annex) (January 2015)

REFERENCES CITED IN THE DESCRIPTION

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Patent documents cited in the description

- CN 202010085038 [0001]

Non-patent literature cited in the description

- **YAO Y ; BAO L ; DENG W et al.** An animal model of MERS produced by infection of rhesus macaques with MERS coronavirus. *J Infect Dis.*, 2014, vol. 209 (2), 236-242 [0053]
- **CHI H et al.** DNA vaccine encoding Middle East responsive syndrome coronavirus S1 protein induces protective immune responses in mice [J. *Vaccine*, 2017, vol. 35 (16), 2069-2075 [0054]
- **LU J ; DU PLESSIS L ; LIU Z et al.** Genomic Epidemiology of SARS-CoV-2 in Guangdong Province, China. *Cell.*, 2020, vol. 181 (5), 997-1003 [0082]